

Supporting Information

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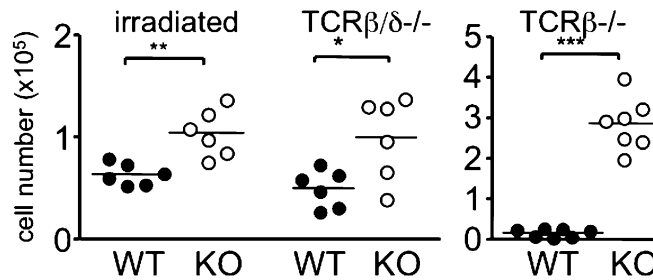


Fig. 51. Naive CD8 T cells derived from MHC II^{-/-} mice expand greater than WT naive CD8 T cells. A total of 1×10^6 naive Thy1.1 WT and MHC II^{-/-} CD8 T cells were transferred into indicated lymphopenic recipients. Recipients were killed at 7 d posttransfer. Donor CD8 T-cell numbers in the lymph nodes (LNs) were enumerated by flow analysis. Each symbol represents an individually tested mouse.

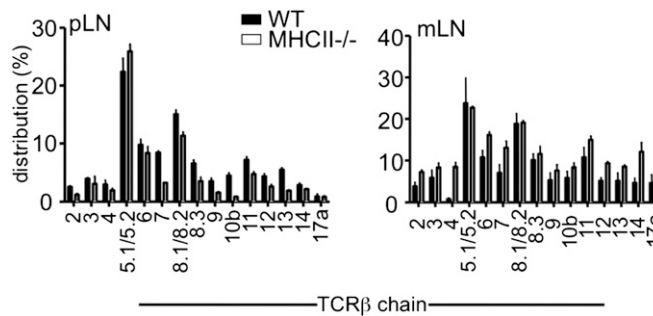


Fig. 52. TCR-V β expression of WT and MHC II^{-/-} naive CD8 T cells after transfer into lymphopenic recipients. A total of 1×10^6 naive Thy1.1 WT and MHC II^{-/-} CD8 T cells were transferred into TCR β ^{-/-} recipients. TCR distribution of the Thy1.1⁺ donor CD8 T cells in the peripheral LNs and mesenteric LNs were examined at 7 d after transfer. The results shown are representative of three individually tested recipients.

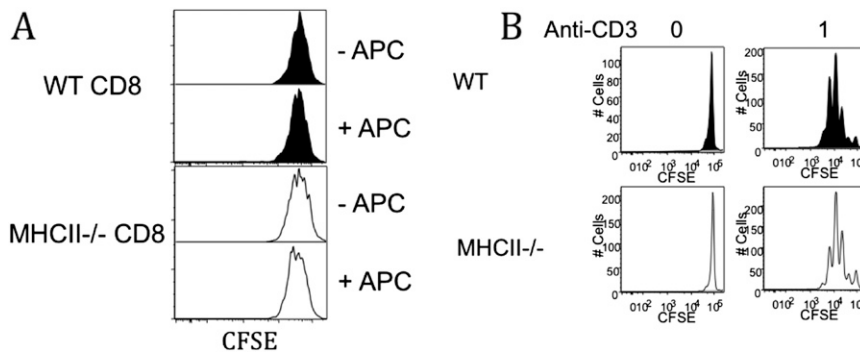


Fig. 53. In vitro proliferation of WT and MHC II^{-/-} CD8 T cells (A) WT and MHC II^{-/-} CD8 T cells transferred into TCR β ^{-/-} recipients were FACS-sorted from the recipients at 7 d posttransfer, CFSE-labeled, and cultured with MHC II-expressing splenocytes for 3 d. (B) FACS-sorted naive WT and MHC II^{-/-} CD8 T cells were stimulated with immobilized anti-CD3 and anti-CD28 (0.5 μ g/mL) mAbs for 3 d. CFSE profiles were subsequently analyzed by FACS.

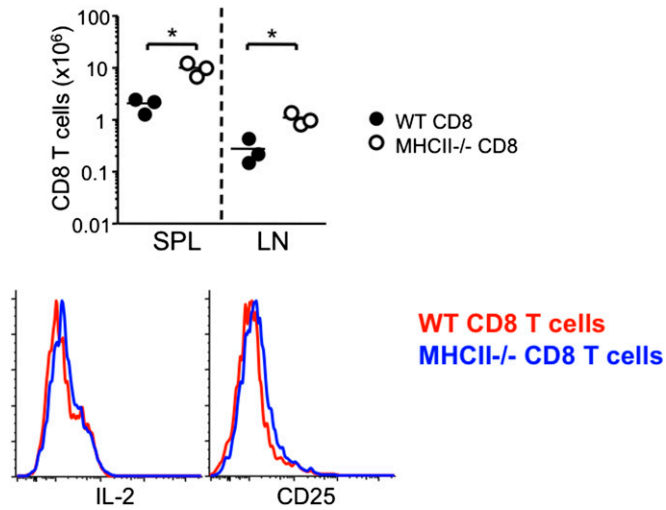


Fig. 54. T-cell responses at 14 d posttransfer. WT and MHC II^{-/-} CD8 T cells transferred into TCR- $\beta^{-/-}$ recipients were analyzed at 14 d posttransfer. Total cellularity, surface CD25 expression, and intracellular IL-2 expression were examined.

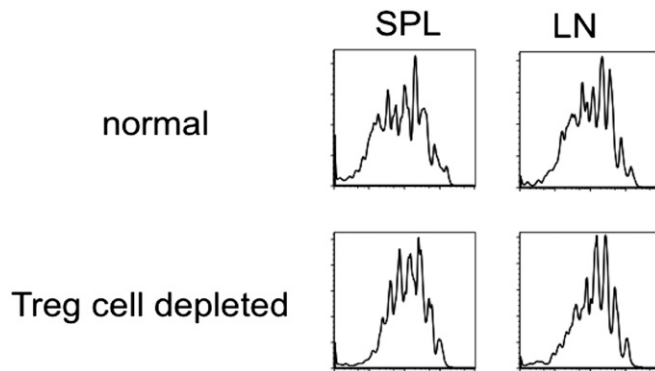


Fig. 55. Regulatory T cells have little role in CD8 T-cell hyperresponses. WT mice were treated with anti-CD25 Abs (250 μ g per mouse) every 3 d to deplete CD25⁺ regulatory T cells for 3 wk. Naïve CD8 T cells were then sorted from the Ab-treated mice and transferred into Rag^{-/-} recipients. CFSE profiles of the donor cells in the spleen and LN were examined at 7 d after transfer. The results shown are representative of three individually tested recipients.

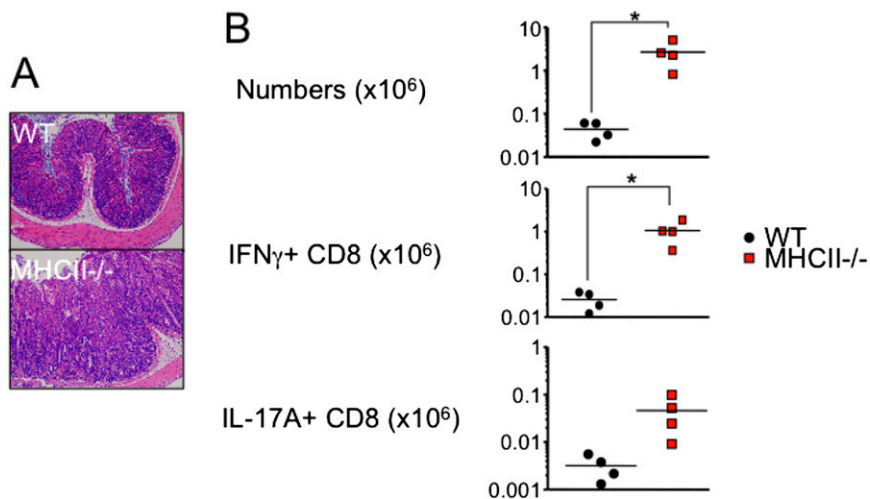


Fig. 56. Severe inflammation in the colon after MHC II^{-/-} CD8 T-cell transfer. (A) A total of 1×10^6 naïve Thy1.1 WT and MHC II^{-/-} CD8 T cells were transferred into TCR- $\beta^{-/-}$ recipients. Shown is H&E staining of the colon tissues at 8 wk after transfer. (Original magnification, 10 \times). (B) mLN cells were isolated at 5 wk posttransfer, and cytokine expression was analyzed by intracellular cytokine staining. Total IFN- γ - and IL-17A-producing Thy1.1 CD8 T cells were enumerated. Each symbol represents an individually tested mouse. * $P < 0.05$.

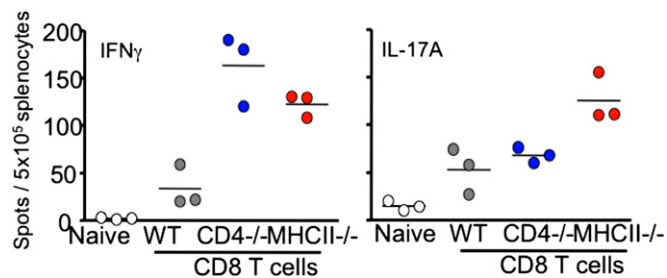


Fig. 57. Cytokine expression of hapten-sensitized CD8 T cells. WT, CD4^{-/-}, and MHC II^{-/-} mice were sensitized with dinitrofluorobenzene as described in *Materials and Methods*. Draining LN CD8 T cells were isolated on day 5. Then 1×10^7 cells were transferred into naive WT mice, and the recipients were subsequently challenged on each side of both ears with dinitrofluorobenzene. After 16 h, splenocytes were stimulated with hapten-pulsed syngenic T-cell-depleted splenocytes for enzyme-linked immunosorbent spot analysis. Each symbol represents an individually tested mouse.

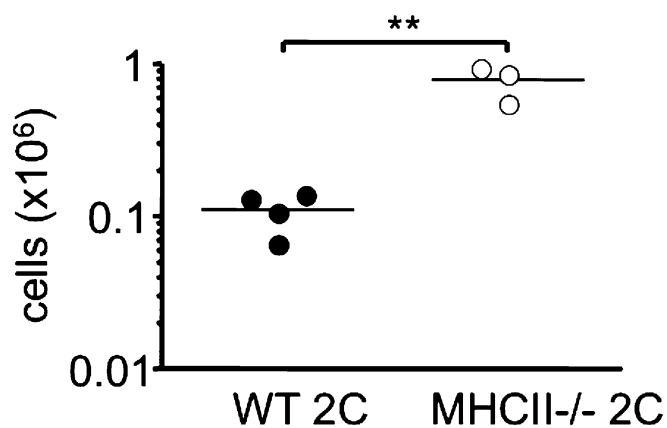


Fig. 58. Proliferative responses of MHC II^{-/-} 2C TCR Tg cells. WT of MHC II^{-/-} 2C TCR Tg naive CD8 T cells were FACS-sorted and transferred into Rag^{-/-} recipients. Donor T-cell recovery was examined at 7 d posttransfer. Data shown represent individually tested recipients (LNs).

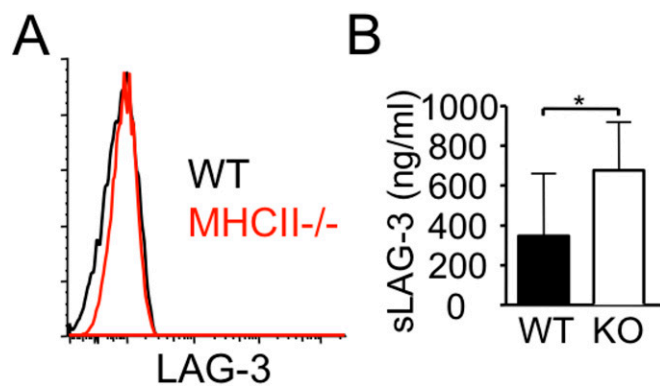


Fig. 59. LAG-3 expression in MHC II^{-/-} CD8 T cells. (A) Naive WT and MHC II^{-/-} CD8 T cells were stained for surface expression of LAG-3. (B) Soluble LAG-3 was examined at 7 d after transfer into Rag^{-/-} recipients by ELISA. * $P < 0.05$.