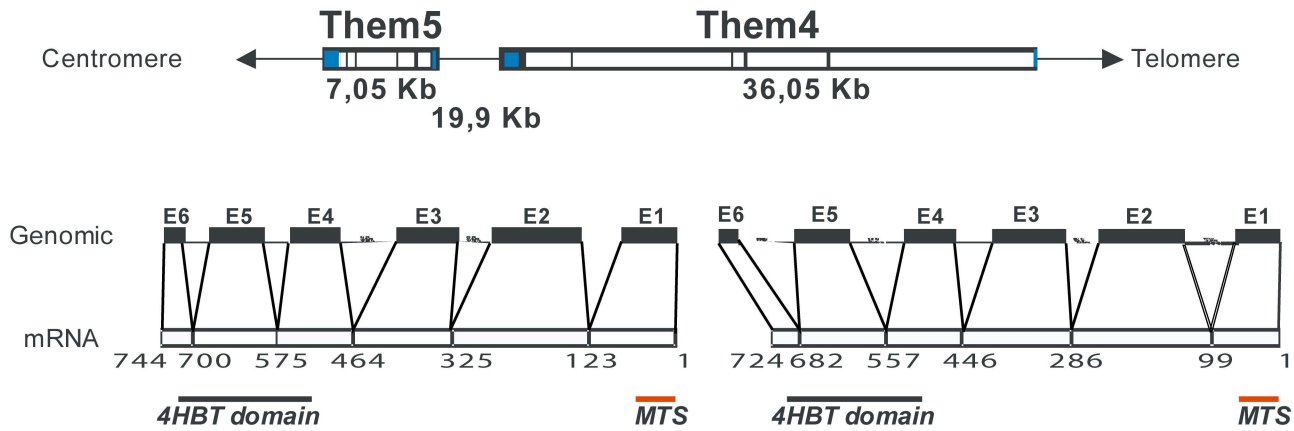


A



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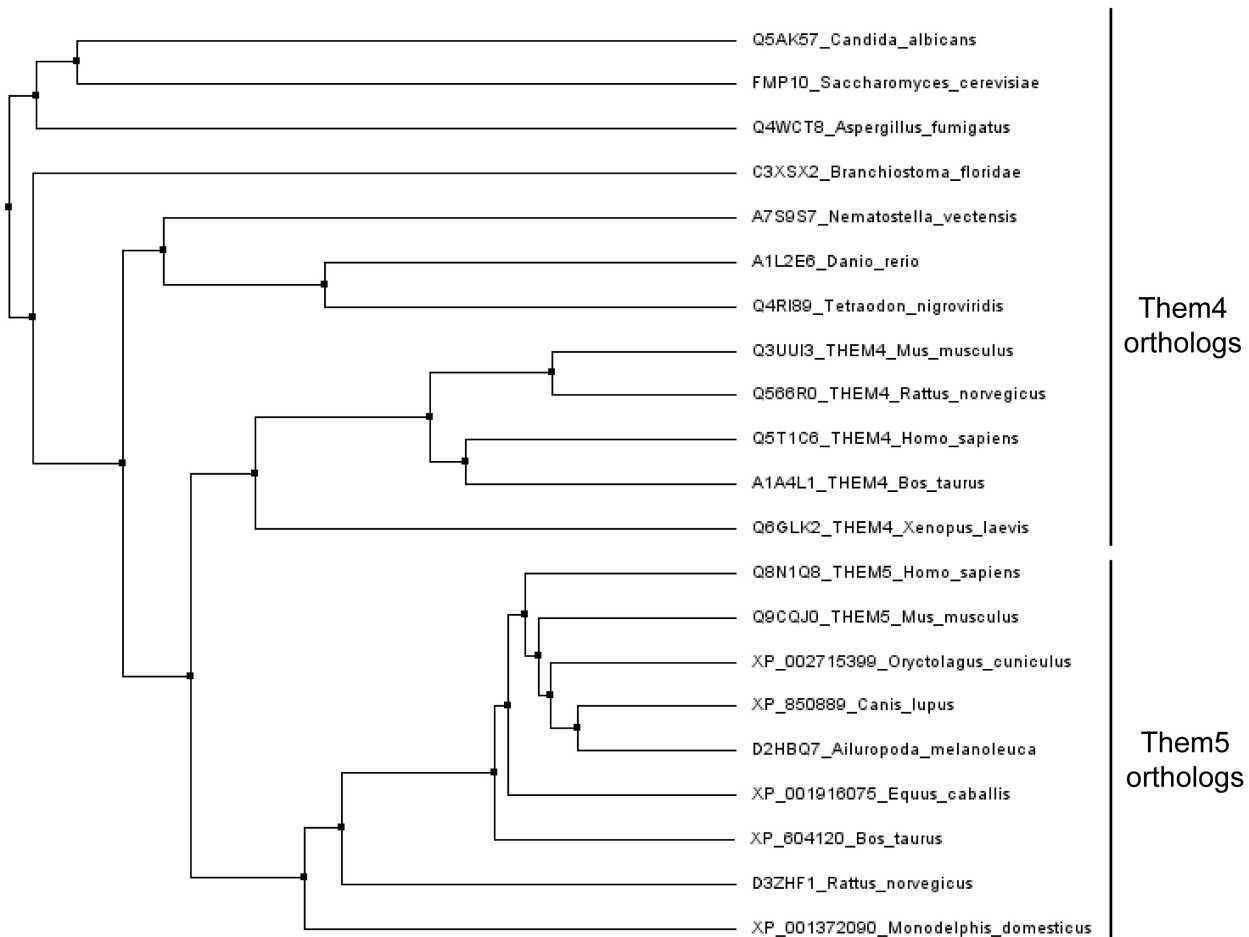


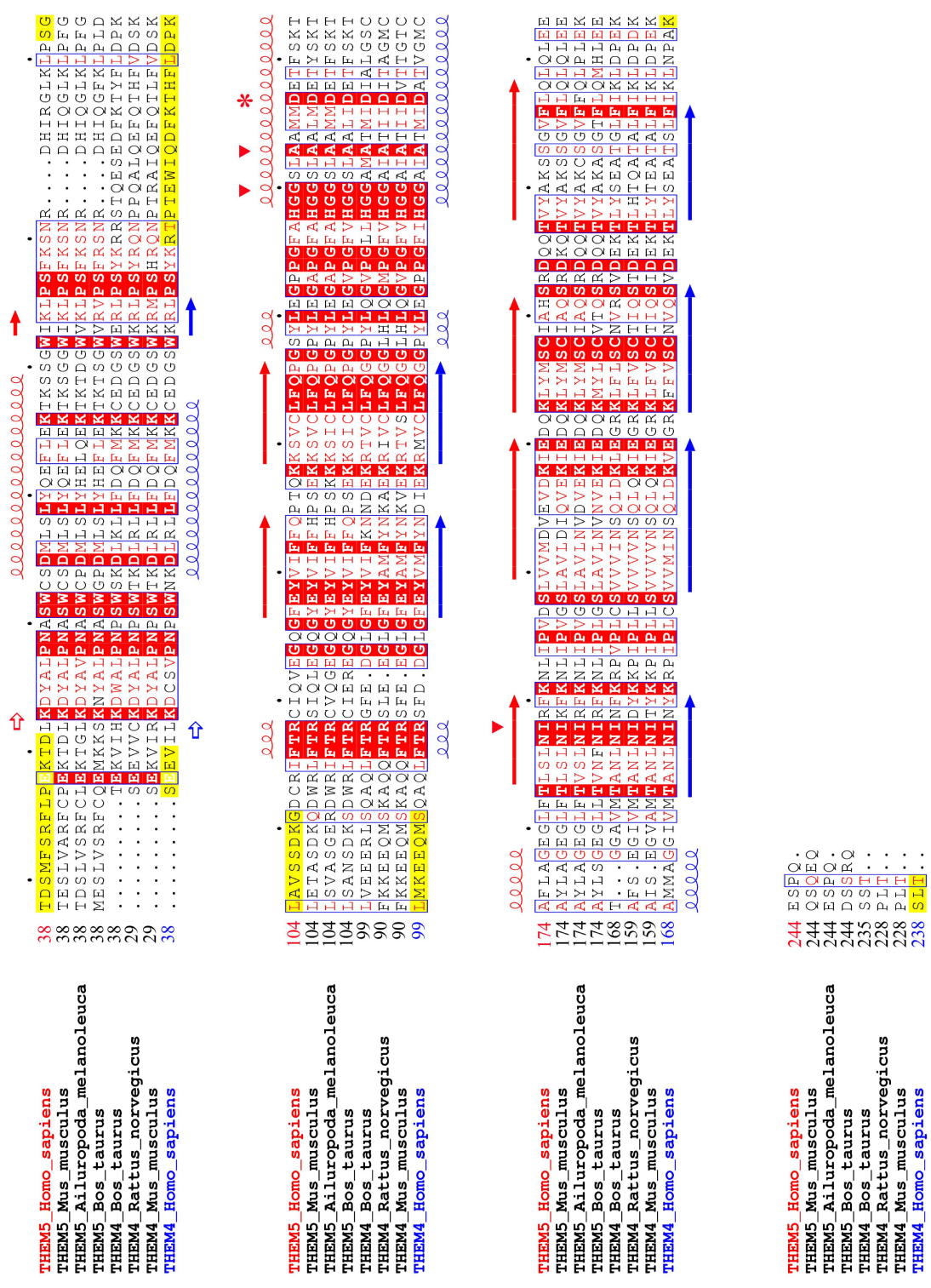
FIG S1. Them5 gene appeared later in evolution but shares similar structure with Them4. (A) Stylized representation of hThem4 and hThem5 genes. Genomic (upper panel) and exon-intron structure and mRNA cartoon representation (lower panel) of hThem4 and hThem5 genes (4HBT - 4-hydroxybenzoyl-CoA thioesterase domain, MTS - mitochondrial targeting sequence). (B) Phylogenetic tree of Them4 and Them5 orthologs in different species. Them4 orthologs are found in lower eukaryotes, such as yeasts and chordata (Them4 orthologs upper group); Them5 orthologs, however, are present only in mammals (lower group).

A

Q6N108 THEM5 Homo_sapiens : MRRCFQVAARLGHRRGLLEAPRILRPLNLPASAFGSSDTSNFRFLPEKTDLKOYALPRAKCSDDLSPVQEFFLEKTK---SSGKIKLPSFKSNR---DHIIRGLKPLSGLAVSSDKGCRIFTRCQIWE : 123
 Q5CQ00 THEM5 Mus_musculus : MLRTSFQVARLVRHKALVRSPPCLLPRVHLASAFGSSDTSILVAFRCEKTDLKOYALPRAKCSDDLSPVQEFFLEKTK---SSGKIKLPSFKSNR---DHIIOGLKLPFLGLETASDKQWRUFRTRIOLE : 123
 Q5CQ03 THEM4 Callithrix_jacchus : MRRKGFQVVARLGHRRGLLGCAPRILPLGNLPASAFGSSDTSVPSRLCEKTDLKOYALPRAKCSDDLSPVQEFFLEKTK---SSGKIKLPSFKSNR---DHIIRGFKLPPGLGVSSDKQWRUFRTRIOLE : 123
 XP 001916075 Equus_caballus : MRRGFVAAARLSHRVLVPGVPHVPLRFLTSASAFGSSDTSILVSRFCVKEKTDLKOYALPRAKCPDMLSPVQEFFLEKTK---ADGKIKLPSFKSNR---DHIIOGLKIPSGFAVSSDKQWRUFRTRIOAE : 123
 XP 002715399 Oryctolagus_cuniculus : MRRGVOAARLGHPCALPGASCPLRDLASAFASDMLISRFLCOKTDLKOYALPRAKCPDMLSPVQEFFLEKTK---SSGKIKLPSFKSNR---DHIIOGLKPLSGLAVTSDKGWRUFRTRICPAE : 123
 XP 8500899 Canis_lupus : MRRGFOAAVRLGQRHAFPGQVPLRNLNLTSAFGSSDTSILVSRFCPEKTDLKOYALPRAKCPDMLDLDVQEFFLEKTK---TDGKIKLPSFKSNR---DHIIOGLKPLSGLAVSSDKSMDWRUFRTRIOWE : 123
 XP 604120 Bos_taurus : MLRKGFGVAGRLSQHRVLPVGPVRLVPLNLTSAFGSSDTSILVSRFLCEKTLGKOYALPRAKCPDMLSPVQEFFLEKTK---TSQWVRYEFSKSNR---DHIIOGLKPLSGLAVSGERQWRUFRTRICVQCE : 123
 XP 001372090 Monodelphis_domestica : MLRSCAARLRTLICALPPVGRRLPQSLRPRPELRSFSEEVILKOCVSNPSSMUKLRLDFDFMKKCE---DGEKIKLPSFKSNR---DHIIOGLKPLDLANSDKSMDWRUFRTRICIERE : 123
 Q571C6 THEM4 Homo_sapiens : QST71C6 THEM4 Homo_sapiens : MLRSCAMRLTLGAT---PARRPGAARLFSSEKVIKROYALPMSYKDLRLDFDFMKKCE---DGSKRLPSFKSNR---DHIIOGLKPLDLKMEEQMSOALPTFSF-DD : 117
 Q56680 THEM4 Rattus_norvegicus : MLRSCAMRLTLGAT---PARRPEATRLEFSEEVKROYALPMSYKDLRLDFDFMKKCE---DGSKRLPSFKSNR---DHIIOGLKPLDLKMEEQMSOALPTFSF-DD : 117
 A1A4L1 THEM4 Bos_taurus : MLRSCTAGLRSIUALRREGAPRLSMDLPAPRLRFLFSKHVHROYALPMSYKDLRLDFDFMKKCE---DGSERLPSFKRSRSTQESDFKTYFLDPLKVEERLSOALPTFSF-FE : 108
 A1A4L1 THEM4 Bos_taurus : A1A4L1 THEM4 Bos_taurus : NYIYVDDEPMDVYMEAEKTSHEIFNOLKREAE---NSMGRWVLUKLKPR---CGDTRLFRGVTGH : 63
 A1A2H6 THEM4 Xenopus_laevis : Q6GLK2 THEM4 Xenopus_laevis : PROOQNLHSLPCSYTAPKFYSOEAPRQVGPVPTLTKSNLLDYNKMEGSK---SGTKRKLPSYNAI---VHVIRG---VPPVNRKRRLFRNLDDQ : 104
 A1A2H6 THEM4 Xenopus_laevis : A1A2H6 THEM4 Xenopus_laevis : MRASLKTNIPSAISLWTFSSQLQFSCPEPSEPNRKPVDHVNACQDTEITVENKTKRPLPSYNRS---IKHATGGIYISKLIOAKARUFRTRVWFEQ : 99
 Q4R189 Tetraodon_nigroviridis : Q4R189 Tetraodon_nigroviridis : MVRPRLFSLPSSSGCPENRHRVYEHNSOCEAEATGGGKQKGPURRIPESYNRS---LKYATGGVYLSKLIOSKARUFRTRISKEA : 77

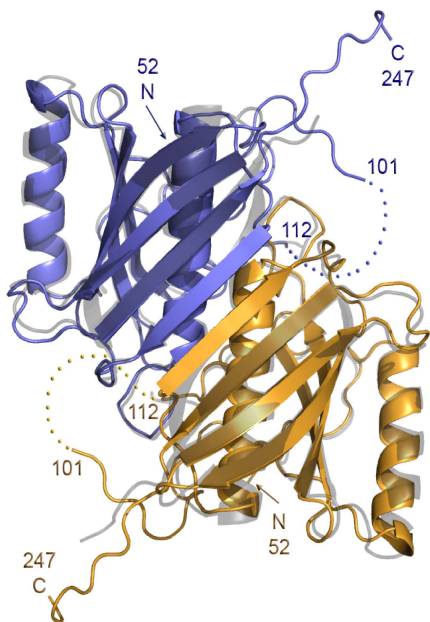
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 Q5CQ00 THEM5 Mus_musculus : GQTEYVIFHPFSEKSVGLCPQPYLCAQFHHGSLALMHDETFYSKTYLAEGGLTLSANRKFMLNLIIVGSLAVLDIQVEIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---EESQEQ- : 248
 Q5CQ03 THEM4 Callithrix_jacchus : GQTEYVIFHPP-RESVGLCPQPYLCAQFHHGSLAMHDETFYSKTYLAEGGLTQSHRFRMLNLIIVGLVMDIIVELIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---EESQEQ- : 247
 XP 001916075 Equus_caballus : GQTEYVIFHPTKRSVGLCPQPYLCSGCHHGSSLAMHDETFYSKTYLAEGGLTLSANRFRKXVQIIVDGLSCLLEIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---EDSSP- : 247
 XP 002715399 Oryctolagus_cuniculus : GQTEYVIFHPSEKSVGLCPQPYLCAQFHHGSLAMHDETFYSKTYLAEGGLTLSHRFRMLNLIIVGSLAVLNHVEIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---EESIQ- : 247
 XP 8500899 Canis_lupus : GQTEYVIFHPSKSVGLCPQPYLCAQFHHGSLAMHDETFYSKTYLAEGGLTSHRFRMLNLIIVGSLAVLNHVEIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---EESIQ- : 247
 XP 604120 Bos_taurus : GQTEYVIFHPSEKSVGLCPQPYLCAQFHHGSLAMHDETFYSKTYLAEGGLTSHRFRMLNLIIVGSLAVLNHVEIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---KEEPQ- : 247
 XP 001372090 Monodelphis_domestica : GQTEYVIFHPSEKSVGLCPQPYLCAQFHHGSLAMHDETFYSKTYLAEGGLTSHRFRMLNLIIVGSLAVLNHVEIDQKLYMESCIAHSNDQOITYAKSSGVLOLOLE---EESRQ- : 248
 Q571C6 THEM4 Homo_sapiens : GLEFYVMEINDIEIRHWGLCPQPYLCPGFHGGALTHDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 240
 Q5CQ03 THEM4 Mus_musculus : GLEFYVMEYKVEIRVYSLQGLHICGVGFVHGGALTIIDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 230
 Q56680 THEM4 Rattus_norvegicus : GLEFYVMEYKVEIRVYSLQGLHICGVGFVHGGALTIIDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 230
 A1A4L1 THEM4 Bos_taurus : GLEFYVMEYKVEIRVYSLQGLHICGVGFVHGGALTIIDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 237
 Q6GLK2 THEM4 Xenopus_laevis : GKNFYVMEYKVEIRVYSLQGLHICGVGFVHGGALTIIDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 188
 A1A2H6 THEM4 Xenopus_laevis : GATFYVMEYKVEIRVYSLQGLHICGVGFVHGGALTIIDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 222
 Q4R189 Tetraodon_nigroviridis : GAAFYVMEYKVEIRVYSLQGLHICGVGFVHGGALTIIDITVITGTCISEG-VAMTANRHTYRPIELCWHVINSQLDVGGRRFFVSNVSDKEXLISEATSLIKLMP---AKSLT- : 214

B

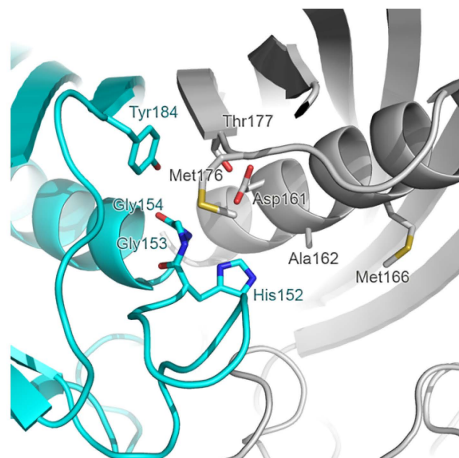


B

C



D



E

Steady-state kinetic constants for hThem5 active site mutants

Enzyme	Substrate	K_m (μM)	k_{cat} (s^{-1})	k_{cat}/K_m ($\text{M}^{-1}\text{s}^{-1}$)
hThem5 T183A	C18:2	0.6	0.2	3.6×10^5
hThem5 G160A	C18:2	7.6	0.3	4.6×10^4
hThem5 G159A/T183A	C18:2	<i>n.d.</i> *	0.1	<i>n.d.</i>

* - not possible to determine K_m

FIG S2. Protein sequence and structural analysis of Them4/5 and their orthologs.

(A) ClustalW multiple sequence alignment of Them4/5 and their orthologs in other species used for ConSurf computational analysis (<http://consurf.tau.ac.il/>). The alignment shows conserved residues (highlighted in dark-grey and grey). Also note the lack of conservation in mitochondrial targeting sequences (N-terminal part of the sequences) between Them4 and Them5 orthologous groups. (B) ClustalW multiple sequence alignment of selected Them5 and Them4 sequences (without predicted MTS). Conserved residues are boxed in red, secondary structure elements in the $\Delta 34$ Them5 and $\Delta 36$ Them4 crystal structures are indicated at the top and bottom of the alignment, respectively. The first residues present in the crystallographic models are highlighted with open arrows. Disordered stretches with no electron density, not included in the models, are shown in yellow shading; mutations used in this study are highlighted with red triangles (dimerization) and a red asterisk (enzymatically inactive). (C) Cartoon representation of superimposed crystal structures of $\Delta 34$ Them5 (blue and orange) and $\Delta 36$ Them4 (gray, transparent). N- and C-terminal residues in the $\Delta 34$ Them5 structure are labeled, and disordered stretches not included in the model are shown as dotted lines. (D) Cartoon representation of the putative $\Delta 36$ Them4 active site. Homodimer subunits are shown in cyan and gray, residues expected to be involved in catalysis and substrate recognition displayed as sticks (atom colors). (E) Steady-state kinetic constants for hThem5 active site mutants in the hydrolysis reaction with linoleyl-CoA, obtained at 37°C, pH 7.5. T183A mutant displays reduced turnover and k_{cat} (due to the interferes with catalysis), G160A mutant has reduced turnover and k_{cat} (due to the sterical interference in the active site), and G159A/T183A mutant has no activity.

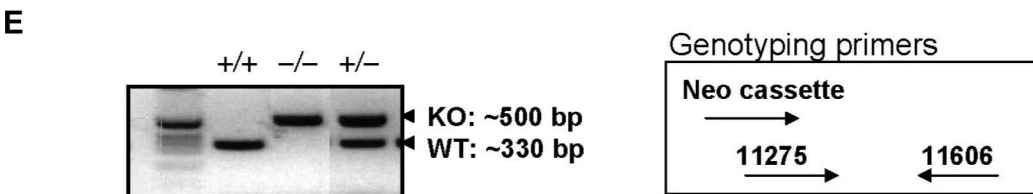
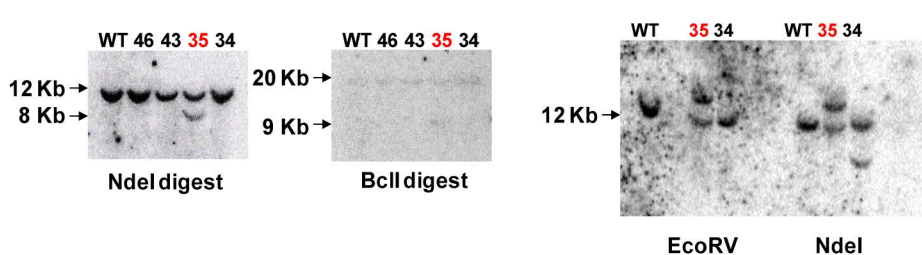
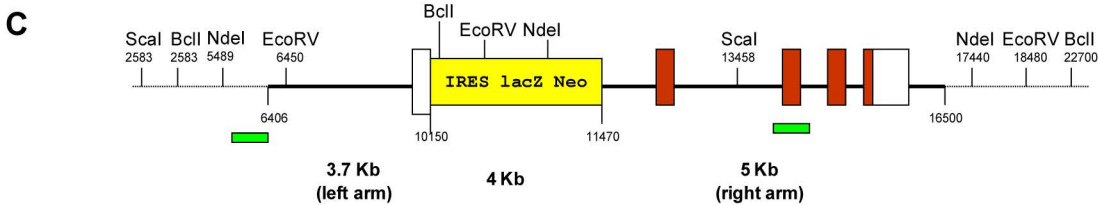
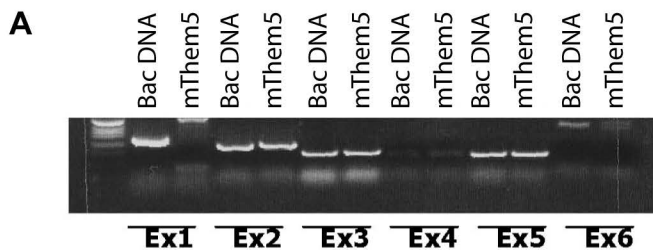
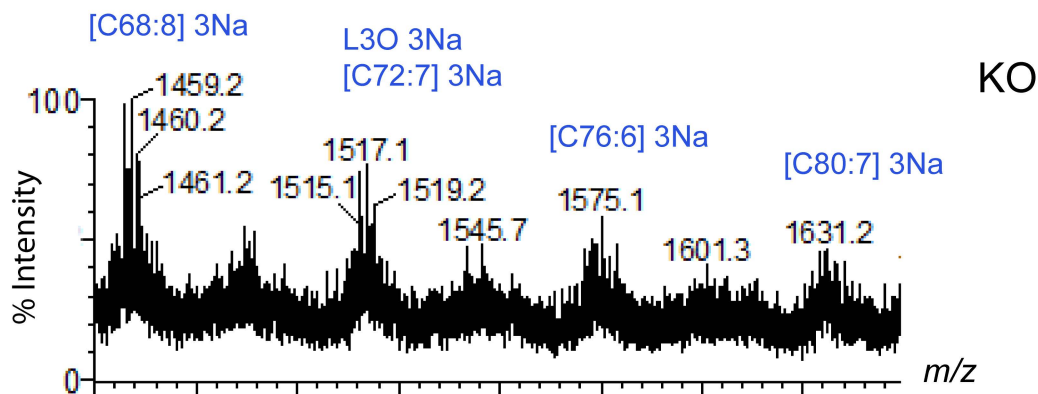
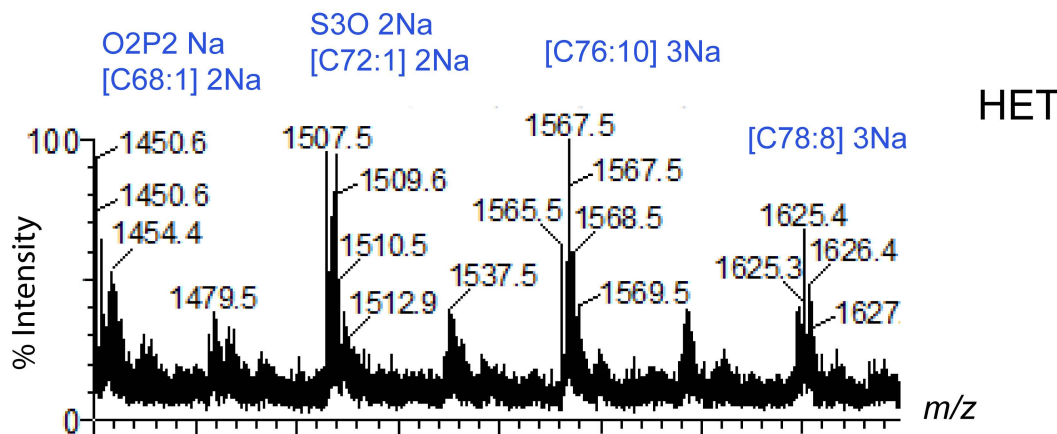
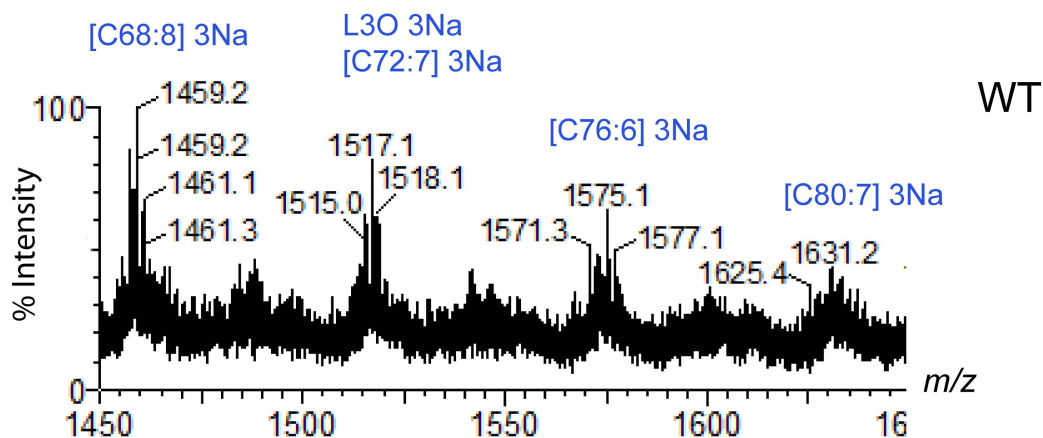
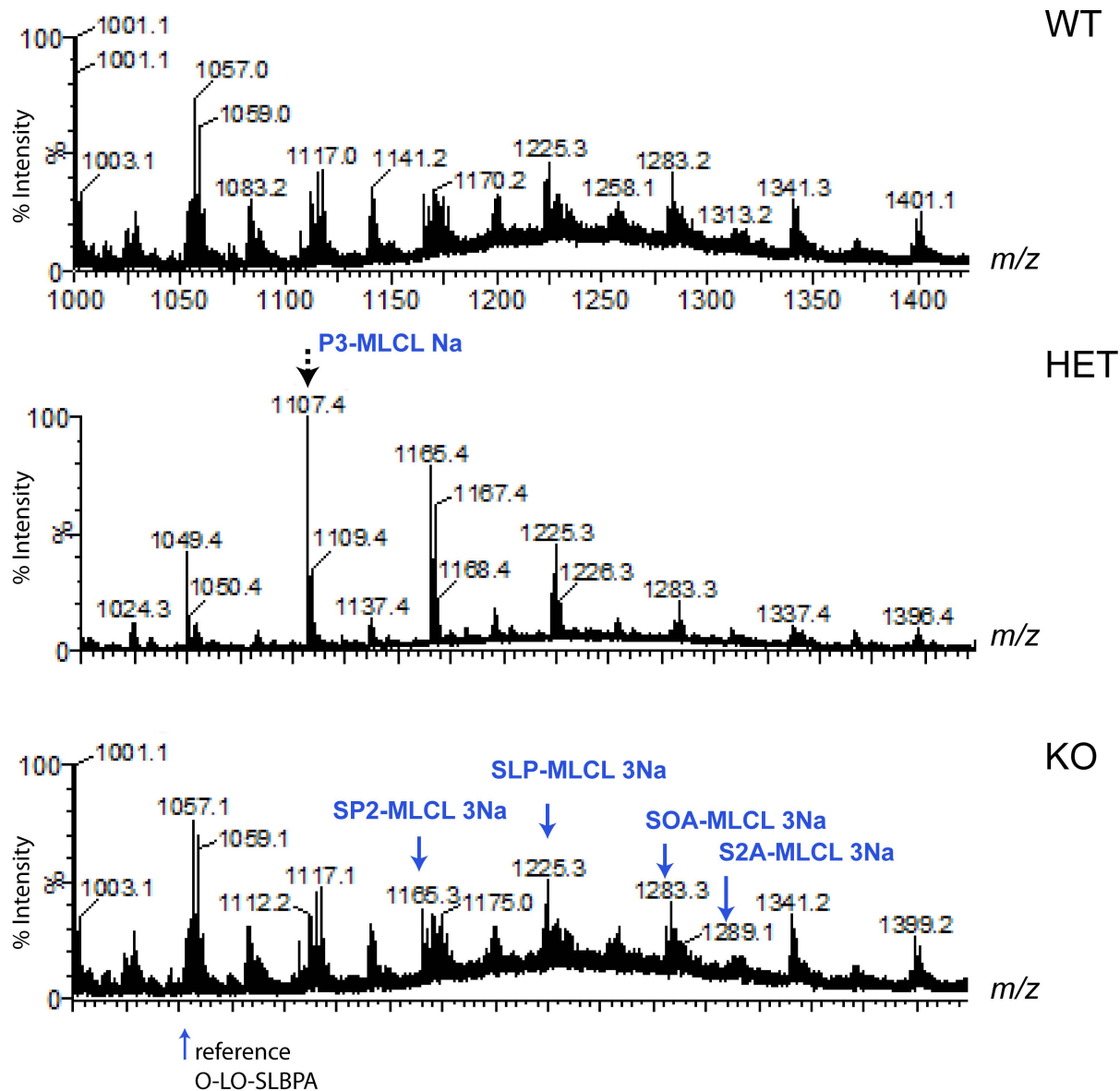


FIG S3. Generation of *Them5* knockout mice. (A) Presence of *mThem5* genomic DNA in BAC clone was verified by PCR of individual exons (cloned *mThem5* cDNA is positive control). BAC clone was used as PCR template for generating left and right homology region of targeting vector. (B) Transcript structure of *mTHEM5* gene in mice. White areas represent untranslated regions (Ensembl database). (C) Targeted *mThem5* allele. A targeting vector was generated that contains a 3.7-kb 5' homology region, an IRES/*lacZ*/neo cassette, and a 5-kb 3' homology region. A genomic DNA fragment of about 1.3 kb, including the ATG start codon in exon 1 and the full sequence of exon2, is deleted in the targeting vector. The targeting vector was linearized with *NotI* and electroporated into 129/Ola ES cells. (D) Screening of ES cell clones was performed by Southern blotting. DNA was digested with *EcoRV* and probed with an external probe (sequence 16980-17663). An internal probe was then used on *NdeI* digested DNA (sequence 9652-10154) for further characterization of ES cell clones positive for homologous recombination. Correctly targeted ES cells (highlighted in red) were used to generate chimeras. Male chimeras were mated with wild-type C57BL/6 females to obtain *Them5*^{+/-} mice, which were intercrossed to produce *Them5* homozygous mutants. (E) Progeny were genotyped for the presence of a targeted allele by multiplex PCR. The following primers were used for genotyping: P1-as 5'-GCA GCA GGC TGA ACT GAC TGA GG-3'; P2/KO-s 5'- GCT GCC TCG TCC TGC AGT TCA TTC-3''; P3/WT-s 5'-CAG GCG GCT GGA TTA AAC TAC C-3'. One reaction amplifies a 500-bp fragment from the targeted allele and the second reaction amplifies a 330-bp fragment from the wild-type allele.

A

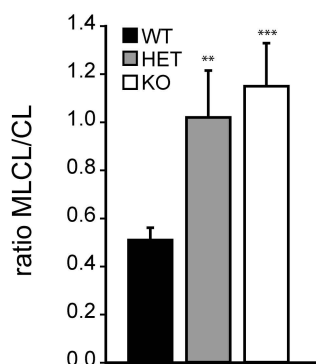


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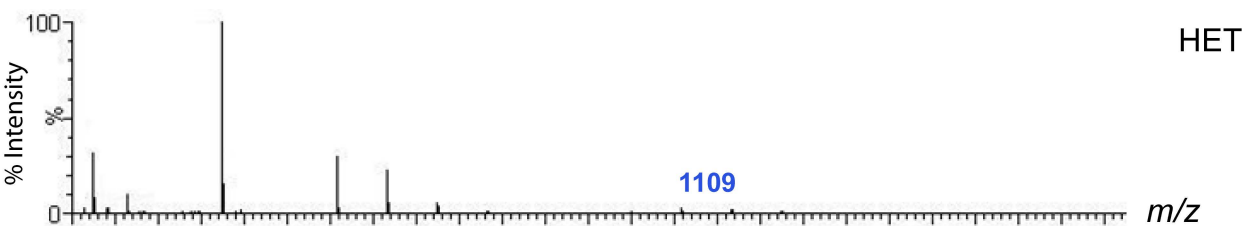
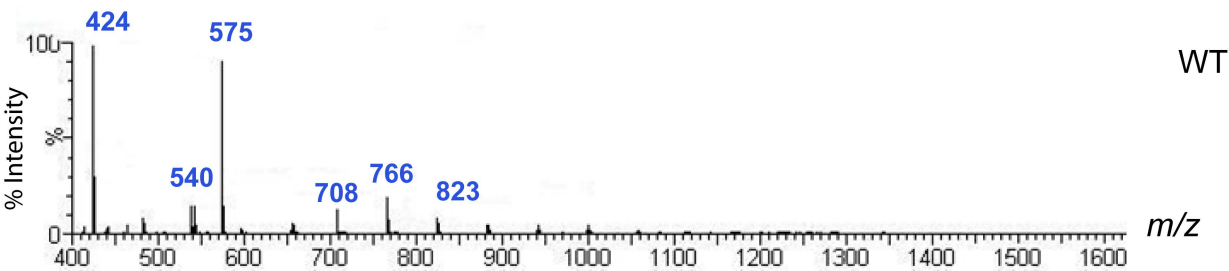


C

Major MLCL vs major CL



D



E

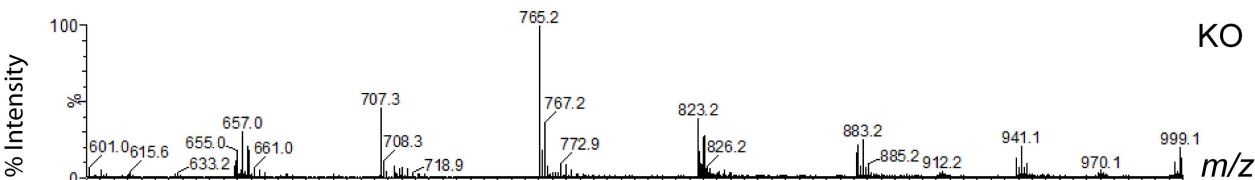
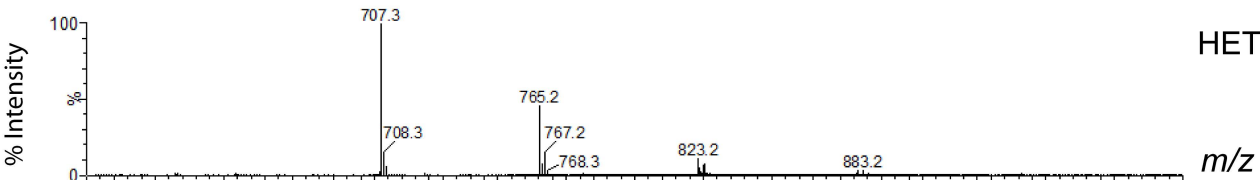
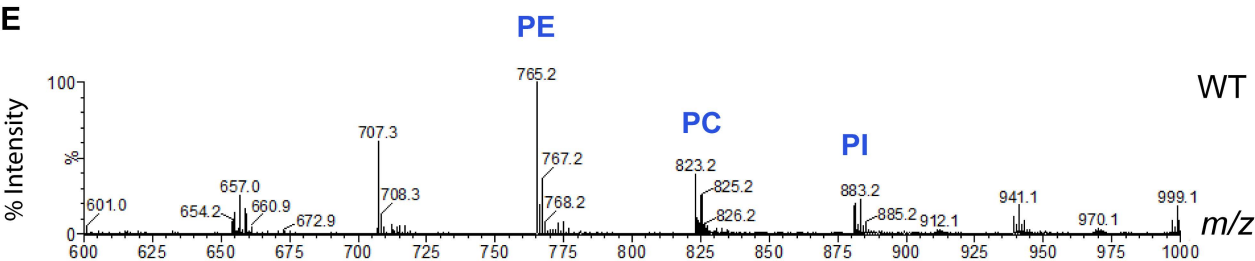


FIG S4. Mass spectrometry analysis of phospholipid composition. Lipid extracts were prepared from Them5 WT (upper panel), HET (middle) and KO (lower) mouse liver mitochondria and analyzed by mass spectrometry in the positive mode [6]. (A) Detailed species composition in the cardiolipin region, analyzed by MS in the positive mode. The lipid profile shows limited, predominantly quantitative differences in the CL profile between Them5 WT (top panel) and KO (bottom panel) liver mitochondria. P – palmitoyl, S – stearoyl, O – oleoyl, L – lineoyl, A – arachidoyl. (B) Species composition in the MLCL/CL region, as analyzed by MS in positive mode. P – palmitoyl, S – stearoyl, O – oleoyl, L – lineoyl, A – arachidoyl. (C) Increase in major monolysocardiolipin (MLCL) over cardiolipin (CL) levels upon Them5 ablation. (D) Total spectra analysis of WT and KO samples without major changes between WT and KO. (E) Species composition in the phospholipid region does not show major differences between WT (top) and KO (bottom) samples. Representative MS profiles are shown. PE – phosphatidylethanolamine, PC – phosphatidylcholine, PI – phosphatidylinositol.

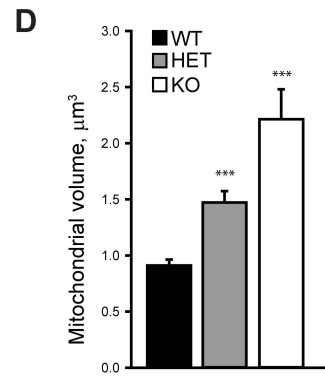
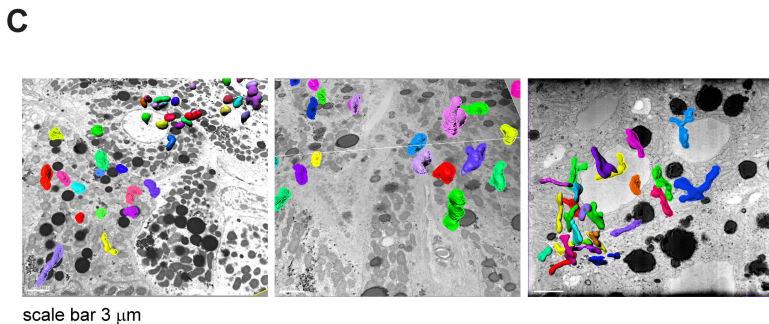
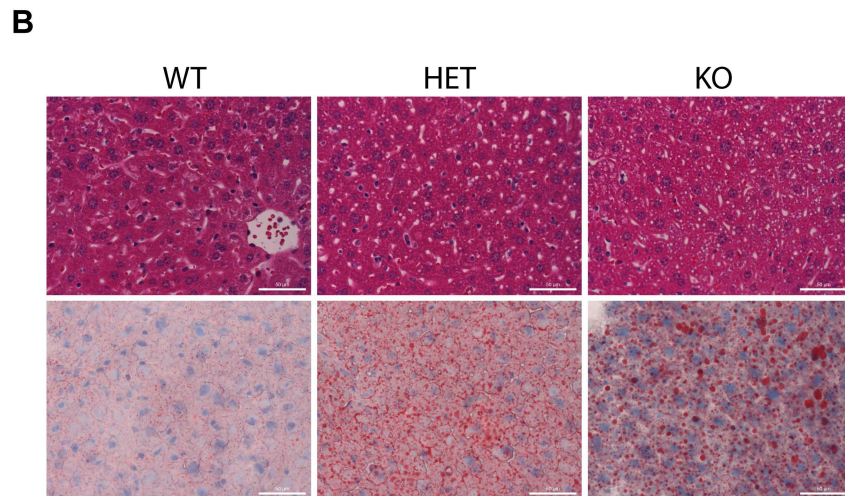
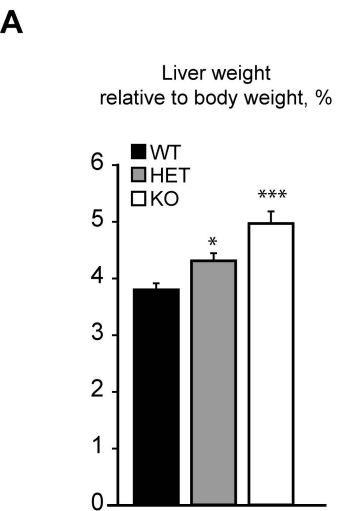


FIG S5. Loss of Them5 leads to fatty liver development and changes in mitochondria morphology. (A-B) Effect of Them5 loss on fatty liver development, reflected in (A) increased liver weights in Them5 HET and KO mice, as compared to WT littermates ($n=8-12$), and (B) progressive development of liver steatosis in Them5 HET and KO mice. Hematoxylin-eosin staining (top) and Oil Red O staining (bottom) of liver sections from 7-month-old mice (right). (C) Representative electron micrographs of liver cells with 3D reconstructed mitochondria (male mice 3-4 months of age, fasted). (D) Increased mitochondria volume in *Them5*^{-/-} and *Them5*^{+/-} hepatocytes of fasted mice, compared to WT controls (min. 15 mitochondria per cell/mouse were reconstructed, 3 cells/per mouse, $n=2$).