

SUPPLEMENTAL FIGURE LEGENDS

Fig. S1. A (inset box), The Complex I isolated from bovine heart contains up to 45 different subunits as analyzed by SDS-PAGE using 4-12% Bis-tris gel (MWM denotes molecular weight marker). **B,** Out of these 45 subunits, there are 14 highly conserved core subunits in the complex I from different species. There are 7 core subunits in the hydrophobic subcomplex. Based on the known protein crystal structures of the core subunits from *T. thermophilus* (hydrophilic domain, PDB ID: 3I9V) and *E. coli* (hydrophobic subcomplex, PDB ID: 3RKO), we have produced a model scheme showing the locations of 51 kDa and 75 kDa subunits in the core functional Complex I. 51 kDa subunit (in light cyan color) and 75 kDa subunit (in light green color) reside at the top of the core hydrophilic domain, which contains FMN cofactor (magenta sticks model) and iron-sulfur clusters (orange-yellow spheres model), and hosts the initial step of respiratory electron transfer activities.

Fig. S2. Amino acid sequence of the precursor of the complex I-75 kDa subunit. The regions labeled with bold represent the amino acid residues identified with LC/MS/MS under the non-reduced conditions in the absence of β -ME. The cysteinyl residues involved in S-glutathionylation are highlighted with gray and they are C₂₂₆, C₃₆₇, and C₇₂₇. The cysteinyl residues involved in disulfide linkage are C₅₅₄, and C₅₆₄. The region labeled with a dotted underline is the N-terminal extension (aa 1-23), which acts as an import sequence and does not exist in the mature protein.

Fig. S3. MS/MS spectra of the protonated molecular ions of S-glutathionylated peptides (a) ³⁶¹VDADTLC³⁶⁷TEEVPTAGAGTDLR₃₈₂, and **(b)** ⁷¹³AVTEGAHAVEEPSIC⁷²⁷.

Fig. S4. Effect of complex I-mediated O₂^{•-} generation on the ratio of GSH/GSSG (in A) and the enzymatic activity of complex I (in B) under the conditions of enzyme turnover in the presence of GSH. The reaction mixture is as described in the legend of the Figure 1. The concentrations of GSH/GSSG and the enzymatic activity of complex I were measured after 1 h incubation at 37° C.

Fig. S5. Amino acid sequence of the precursor of the complex I-75 kDa subunit. The regions labeled with bold represent the amino acid residues identified with LC/MS/MS under the reduced conditions in the presence of β -ME. The cysteinyl residues involved in protein radical formation are underlined and they are C₂₂₆, C₅₅₄, and C₇₂₇. The region labeled with a dotted underline is the N-terminal extension (aa 1-23), which acts as an import sequence and does not exist in the mature protein.

Fig. S6. MS/MS spectrum of the triply protonated molecular ion of DMPO-binding peptide ⁵⁴⁴MLFLLGADGGC⁵⁵⁴ITR₅₅₇.

Fig. S7. MS/MS spectra of the doubly protonated molecular ions of S-sulfonated peptides. a, ⁷¹³AVTEGAHAVEEPSIC(SO₃)⁷²⁷. **b,** ⁵⁴⁴M(ox)LFLLGADGGC⁵⁵⁴(SO₃)ITR₅₅₇.

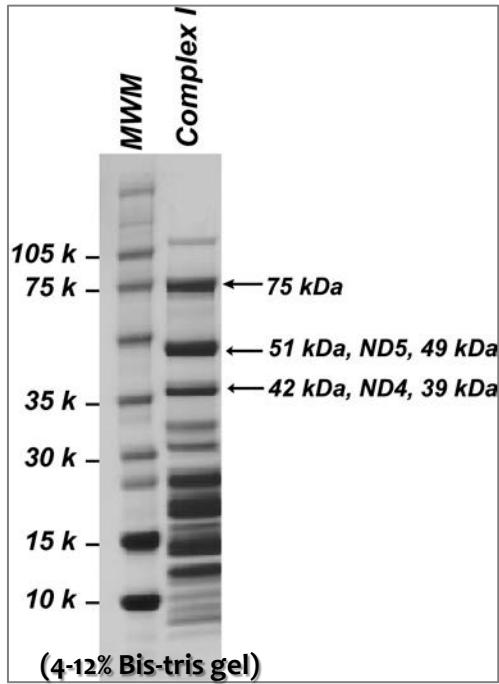
Fig. S8. The homology model of 75 kDa (pale green) subunit of Complex I. FMN (magenta sticks) and iron-sulfur clusters (orange-yellow spheres) are also shown for orientation. There are total 17 highly conserved cysteines in mature mammal 75 kDa subunits. According to our multiple sequence alignment results (data not shown), 11 of the 17 cysteines are identical to the Nqo3 subunit of *T. thermophilus*, and directly involved in the coordinates or ligands of the iron-sulfur clusters, and they are highlighted with magenta sticks. The other 6 cysteines (C₅₃, C₃₆₇, C₅₅₄, C₅₆₄, C₇₁₀ and C₇₂₇) are labeled and highlighted in blue-yellow spheres. Except C₅₃, all other five active cysteines are surface exposed according to the homology model.

Fig. S9. The effect of different dosages of menadione (0-100 μ M) on the HL-1 cellular S-glutathionylation (in A) and protein radical adduct of DMPO (in B) measured by immunomicroscopic fluorescence using anti-GSH or anti-DMPO monoclonal antibody.

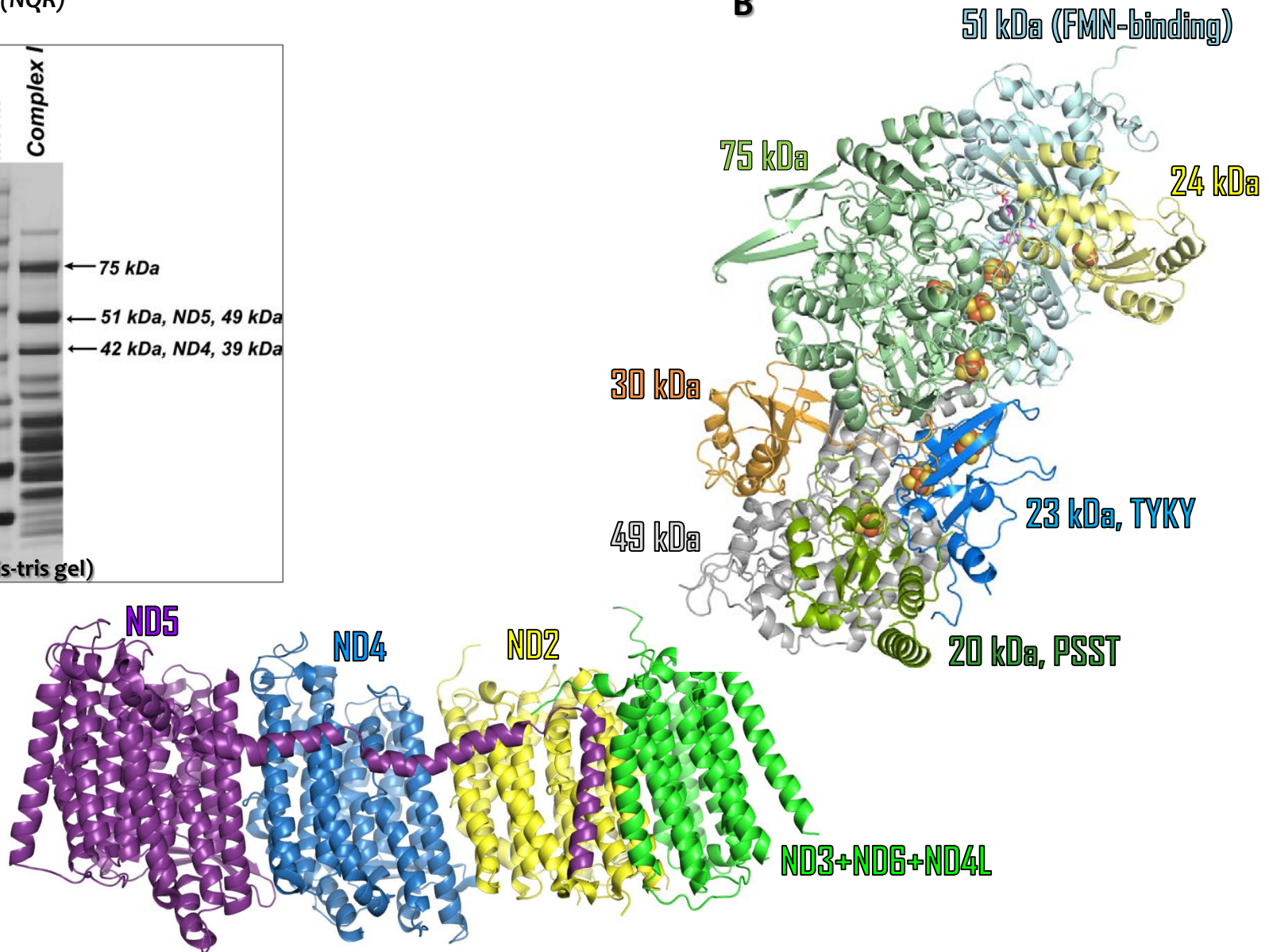
Fig. S10. Amino acid sequence coverage of the precursor of the 75 kDa subunit from the alkylated control complex I as analyzed by nano-LC/MS/MS. The regions labeled with *italic bold* and represent the amino acid

residues identified with LC/MS/MS under the non-reduced conditions in the absence of β -ME. The cysteinyl residues involved in disulfide bond are detected between *C*₅₅₄, and *C*₅₆₄. *Iodoacetamide alkylation was detected at all cysteine residues except C₆₄ and C₇₁₀ which were not observed by MS/MS.* The region labeled with a dotted underline is the N-terminal extension (aa 1-23), which acts as an import sequence and does not exist in the mature protein.

A Complex I (NQR)



B

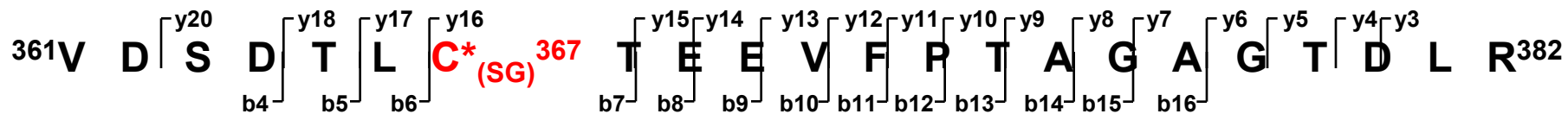


Supplemental Fig. S2

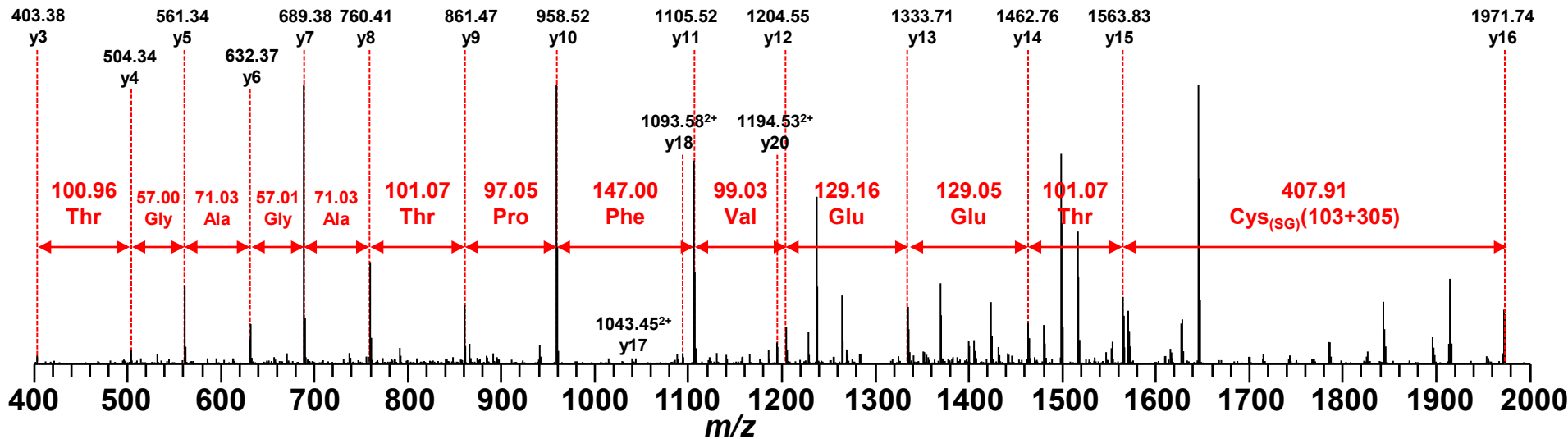
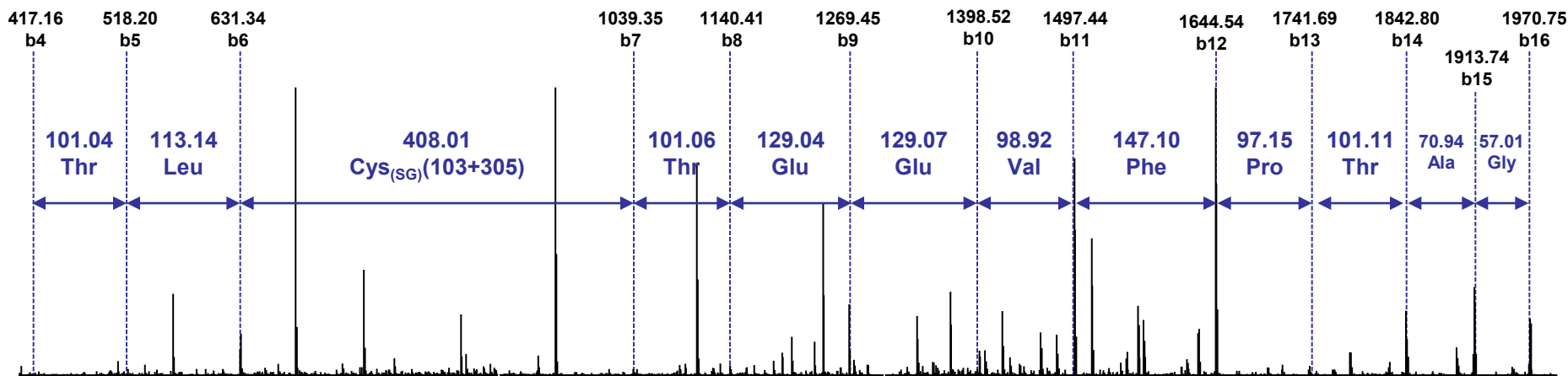
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51 QACEKVGMOI PRFCYHERLS VAGNCRMCLV EIEKAPKVVA ACAMPVMKGW
101 NILTNSEKTK KAREGVMEFL LANHPLDCPI CDQGGCEDLQ DQSMFSGSDR
151 SRFLEGKRAV EDKNIGPLVK TIMTRCIQCT RCIRFASEIA GVDDLGGTTGR
201 GNDMQVGTYYI EKMFMSSELSG NIIDI^CCPVGA LTSKPYAFTA RPWETRKTES
251 IDVMDAVGSN IVVSTRTGEV MRILPRMHED INEEWISDKT RFAYDGLKRQ
301 RLTEPMVRNE KGLLTHTTWE DALSRVAGML QSFQGNVAA IAGGLVDAEA
351 LIALKDLLNR VSDTL^CCTEE VFPTAGAGTD LRSNYLLNTT IAGVEEADV
401 LLVGTNPRFE APLFNARIRK SWLHNDLKVA LIGSPVDLTY RYDHLGDSPK
451 ILQDIASGSH PFSQVLQEAK KPMVILGSSA LQRNDGAAIL AAVSNIAQKI
501 RTSSGVTGDW KVMNILHRIA SQVAALDLGY KPGVEAIQKN PPKMLFLLGA
551 DGGCITRQDL PKDCFIVYQG HGGDVGAPIA DVILPGAAYT EKSATYVNTE
601 GRAQQTKVAV TPPGLAREDW KIIRALSEIA GMTLPYDTLD QVRNRLEEVS
651 PNLVRYDDVE GANYFQQASE LSKLVNQQLL ADPLVPPQLT IKDFYMTDSI
701 SRASQTMKAC VKAVTEGAHA VEEPSIC

Sequence coverage=511/704=72.58%

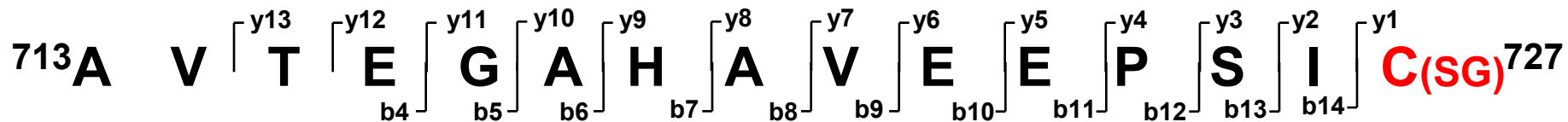
Supplemental Fig. S3a



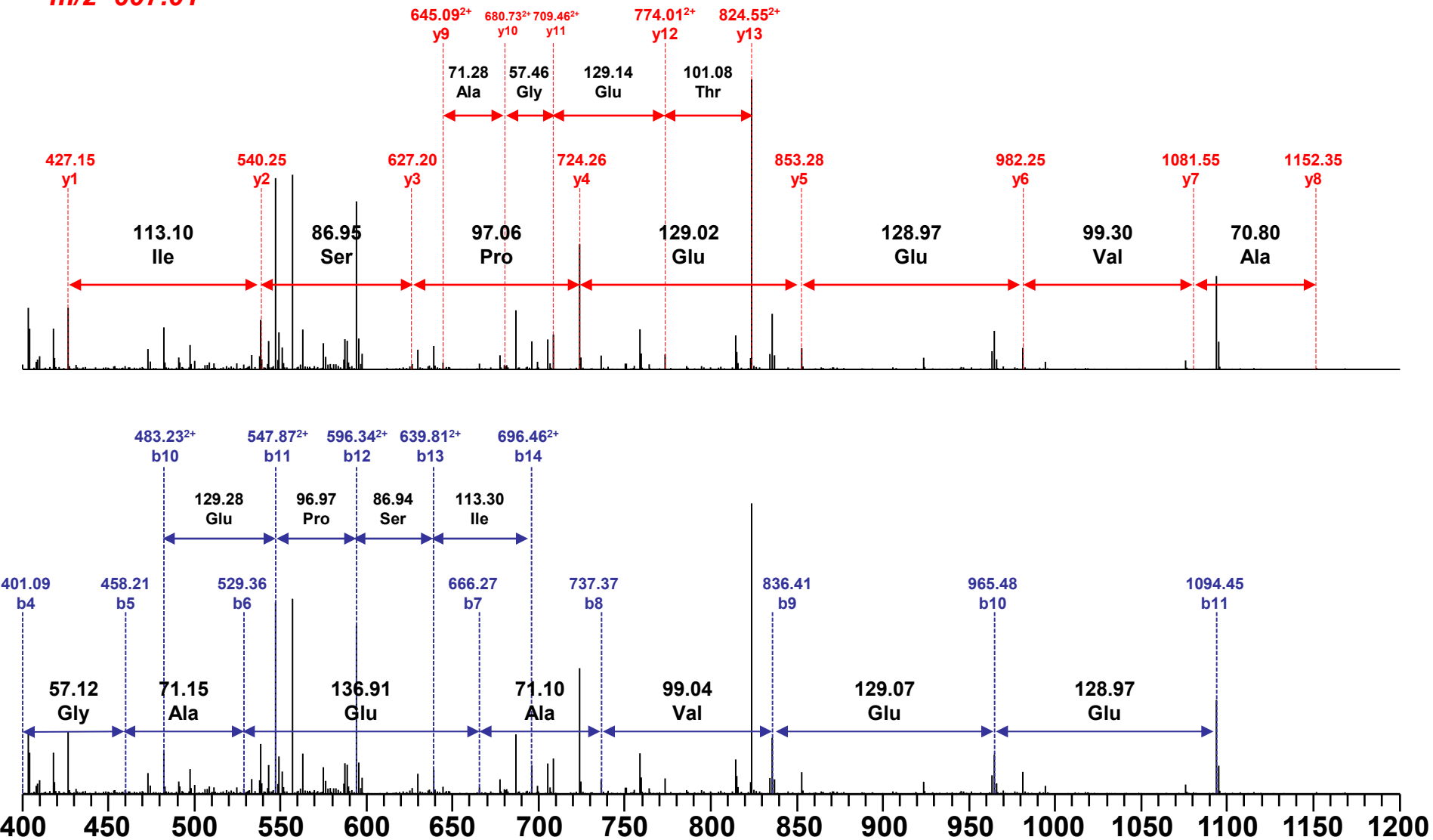
m/z=1301.85²⁺



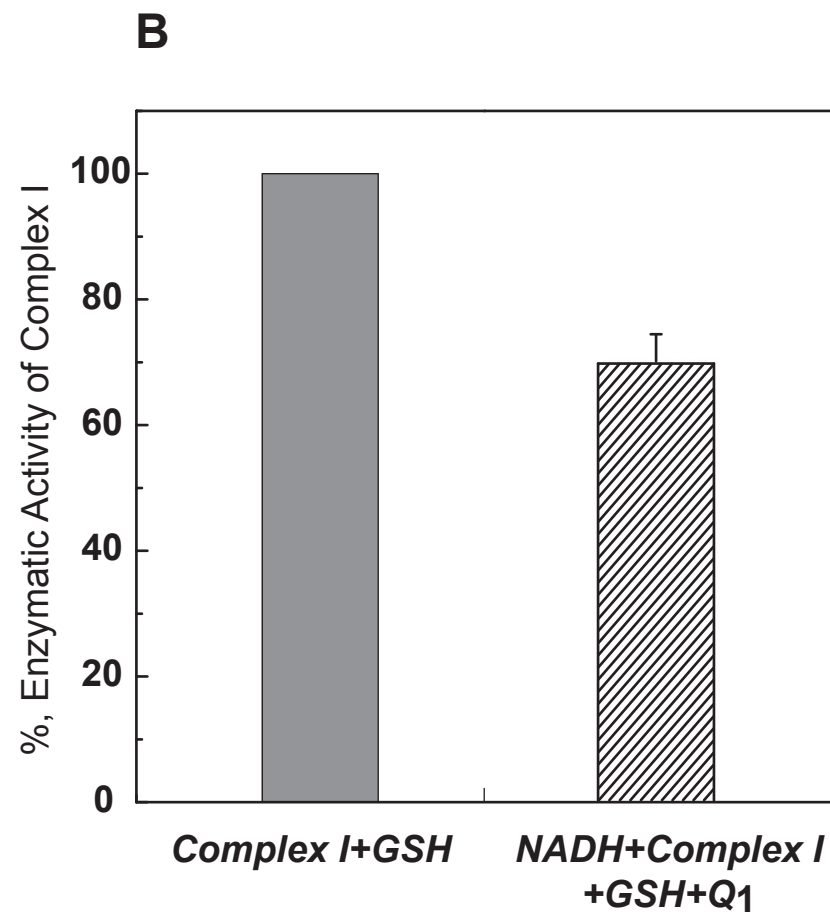
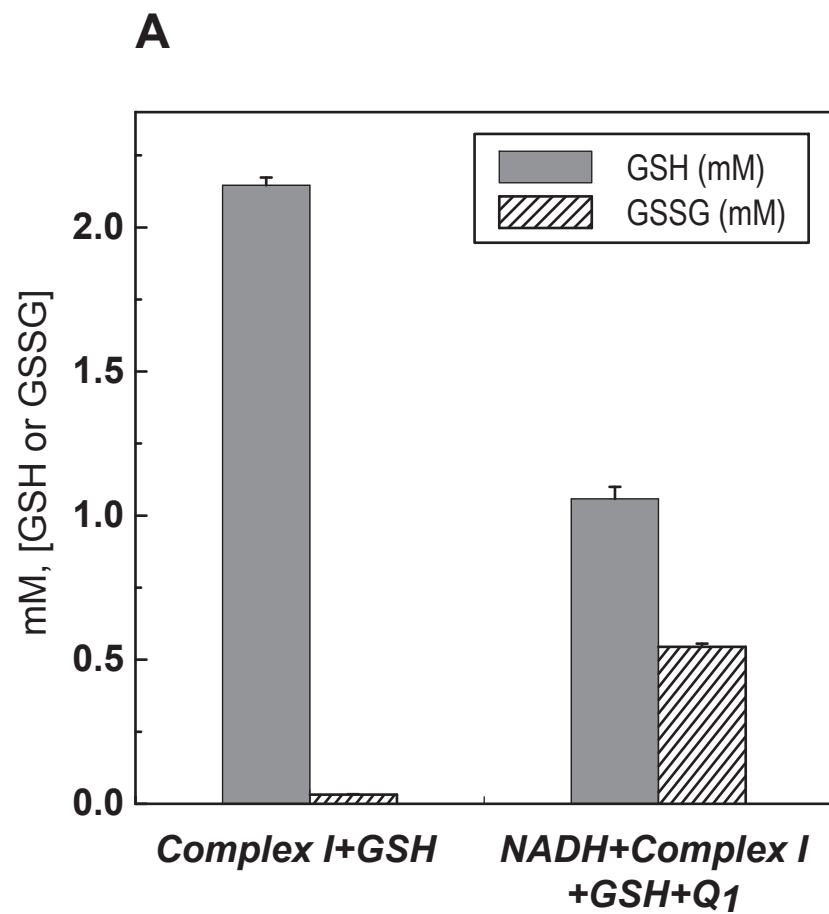
Supplemental Fig. S3b



$m/z=607.01^{3+}$



Supplemental Fig. S4

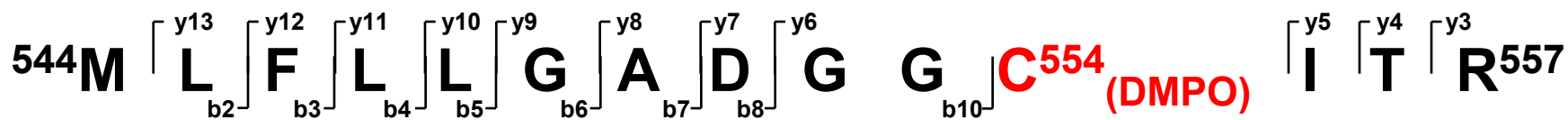


Supplemental Fig. S5

1 MLRIPVRKAL VGLSKSSKGC VRTTATAASN LIEVFVDGQS VMVEPGTTVL
51 QACEKVGMI PRFCYHERLS VAGNCRMCLV EIEKAPKVVA ACAMPVMKGW
101 NILTNSEKTK KAREGVMEFL LANHPLDCPI CDQGGECDLQ DQSMFSGSDR
151 SRFLEGKRAV EDKNIGPLVK TIMTRCIQCT RCIRFASEIA GVDDLGGTGR
201 GNDMQVGTYI EKMFMSSELSG NIIDICPVGA LTSKPYAFTA RPWETRKTES
251 IDVMDAVGSN IVVSTRTGEV MRILPRMHED INEEWISDKT RFAYDGLKRQ
301 RLTEPMVRNE KGLLTHTTWE DALSRVAGML QSFQGNVAA IAGGLVDAEA
351 LIALKDLLNR VSDTLCTEE VFPTAGAGTD LRSNYLLNTT IAGVEEADV
401 LLVGTNPRFE APLENARIRK SWLHNDLKVA LIGSPVDLTY RYDHLGDSPK
451 ILQDIASGSH PFSQVLQEAK KPMVILGSSA LQRNDGAAIL AAVSNIAQKI
501 RTSSGVTGDW KVMNILHRIA SQVAALDLGY KPGVEAIQKN PPKMLFLLGA
551 DGGCITRQDL PKDCFIVYQG HHGDVGAPIA DVILPGAAYT EKSATYVNTE
601 GRAQQTKVAV TPPGLAREDW KIIRALSEIA GMTLPYDTLD QVRNRLEEVS
651 PNLVRYDDVE GANYFQQASE LSKLVNQQLL ADPLVPPQLT IKDFYMTDSI
701 SRASQTMKC VKAVTEGAHA VEEPSIC

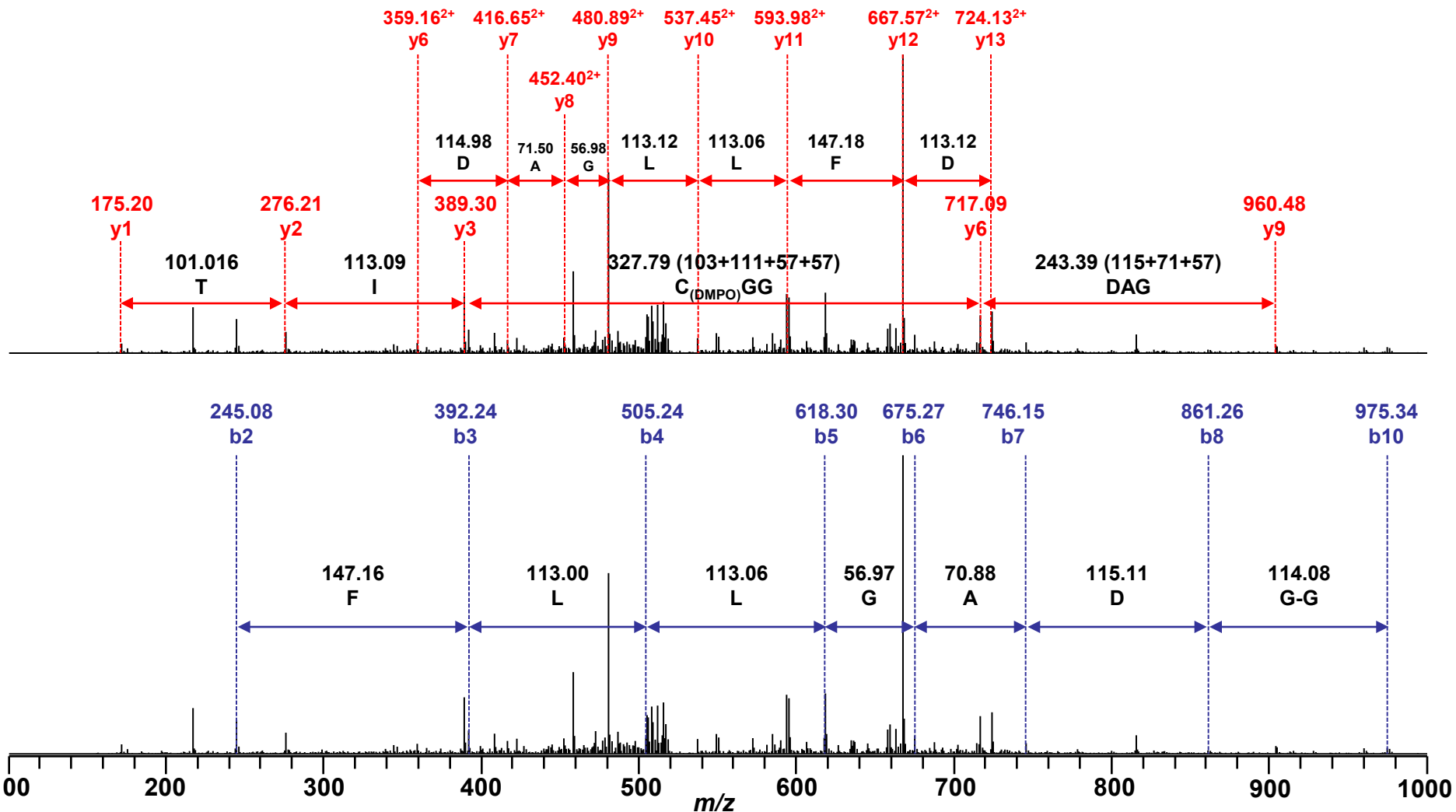
Sequence coverage=679/704 =96.44%

Supplemental Fig. S6

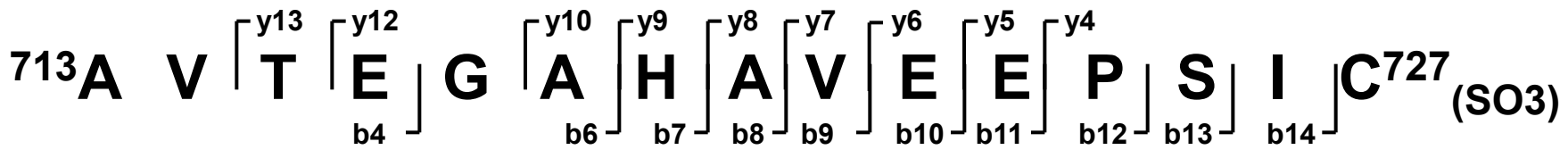


$m/z_{(theoretical)} = 526.6108^{3+}$

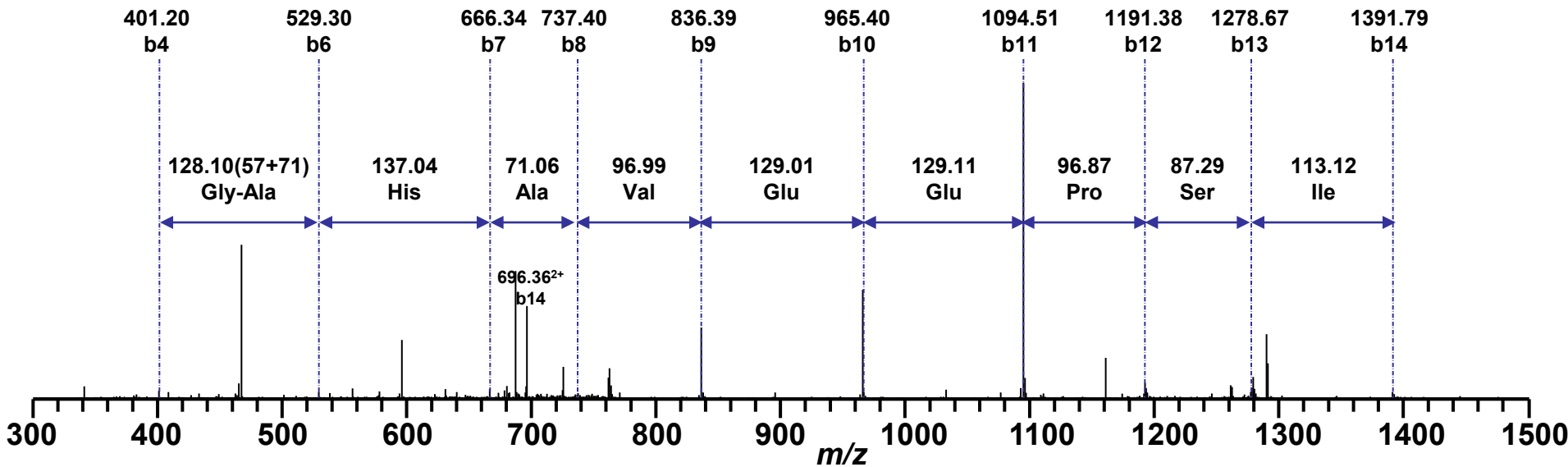
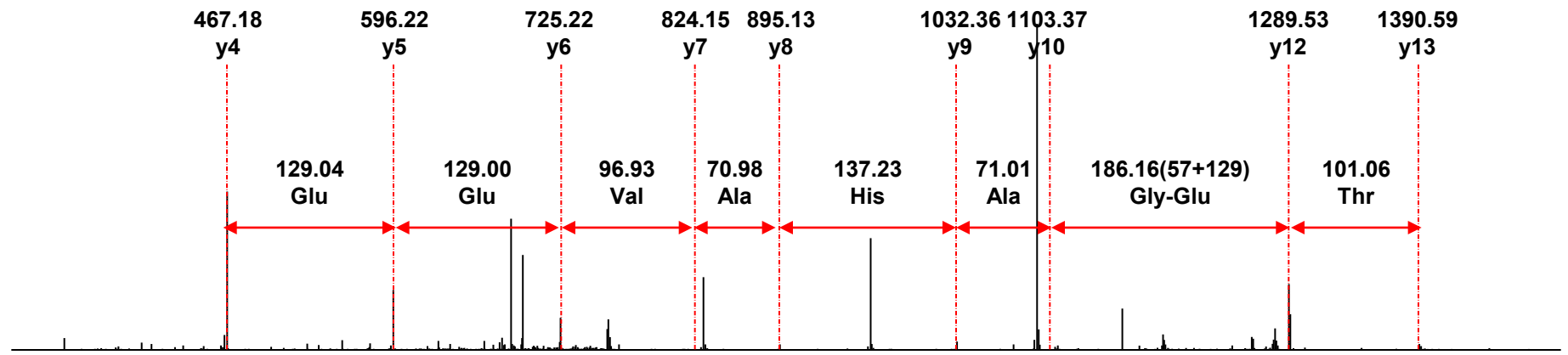
$m/z_{(measured)} = 527.03^{3+}$



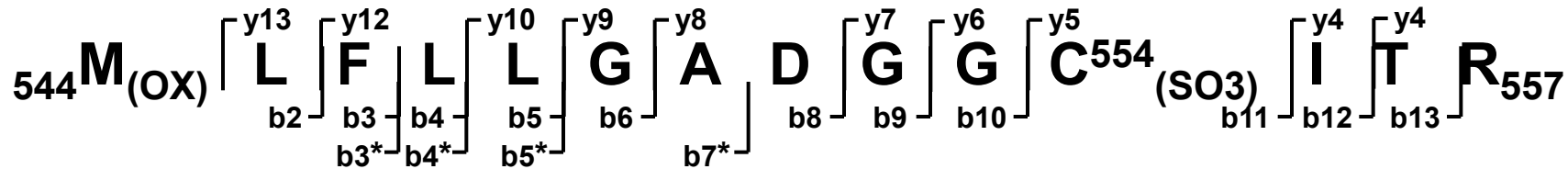
Supplemental Fig. S7a



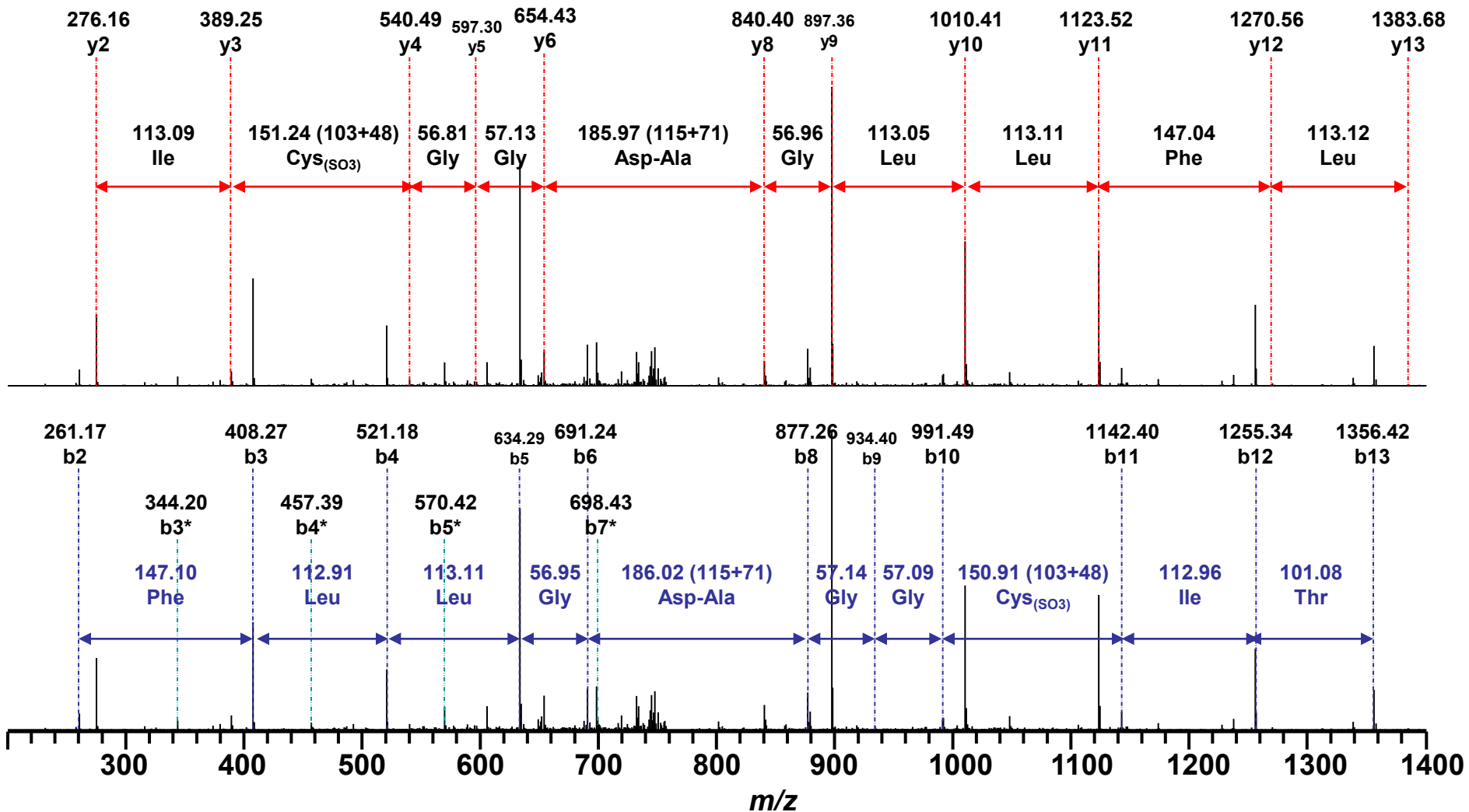
m/z=766.18²⁺



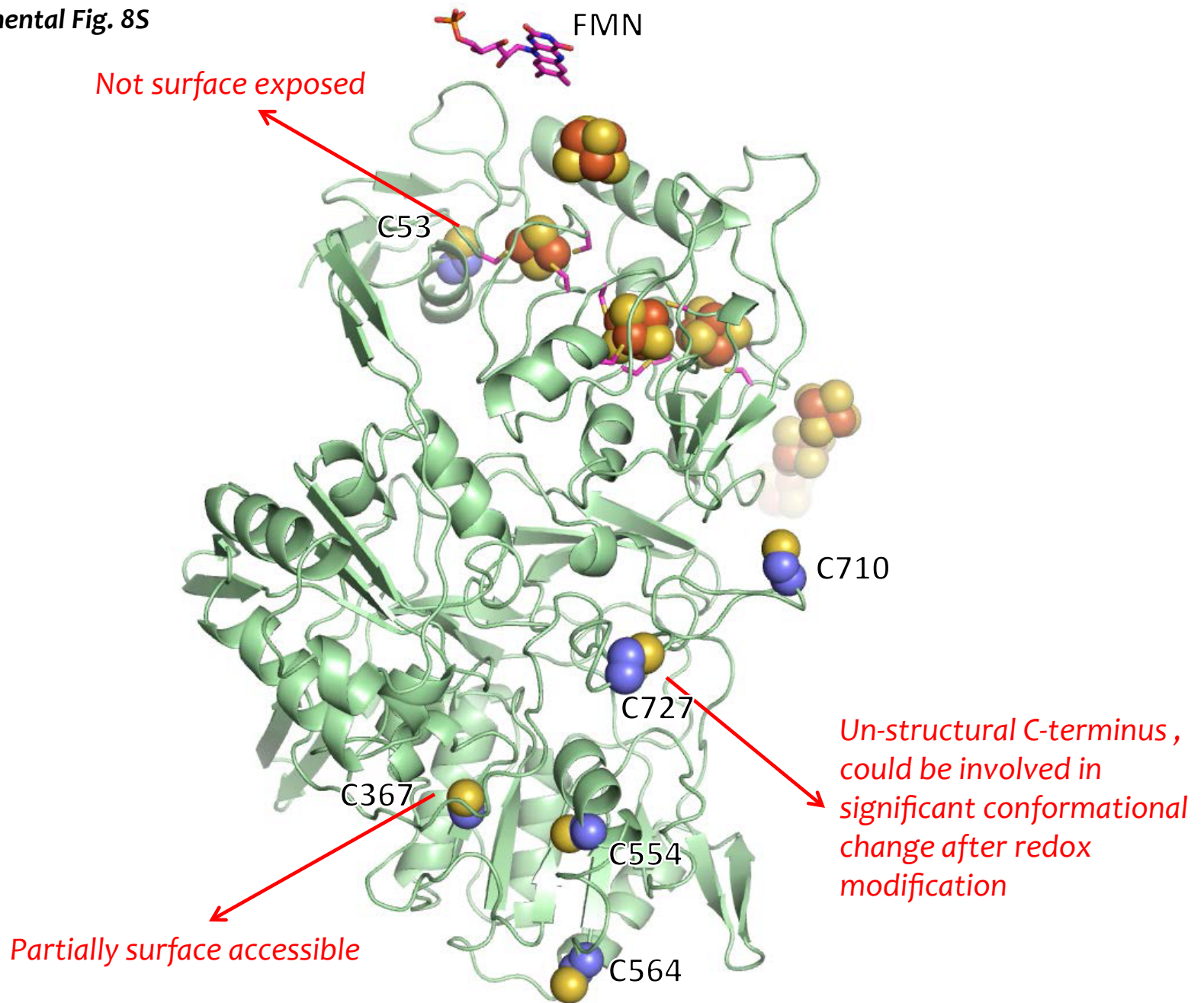
Supplemental Fig. S7b



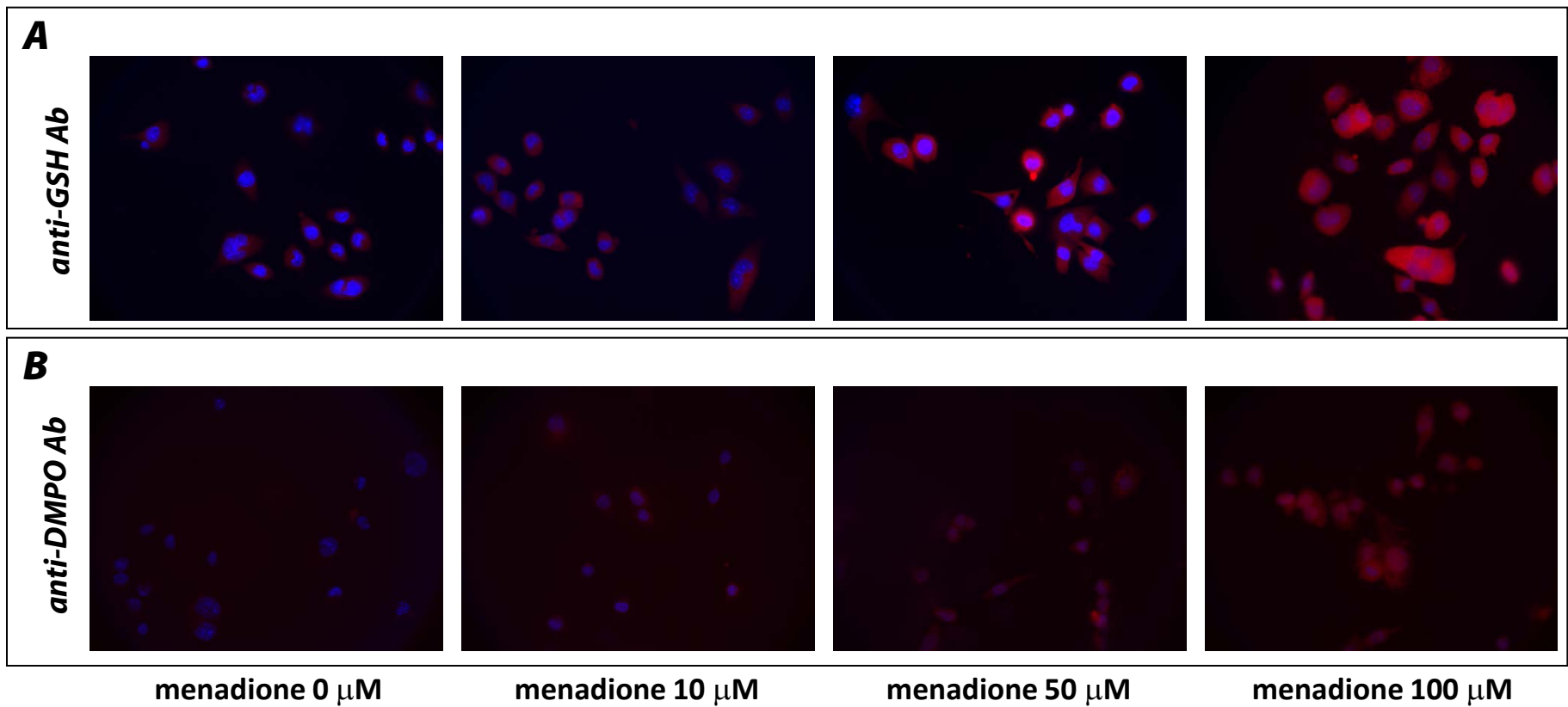
$m/z=780.83^{2+}$
 $bn^*=bn-\text{CH}_4\text{SO}$



Supplemental Fig. 8S



Supplemental Fig. S9



Red: Alexa Fluor 594 -conjugated goat anti-mouse IgG
Blue: DAPI staining of nuclear

Supplementary Fig. S10

1 MLRIPVRKAL VGLSKSSKGC VRTTATAASN LIEVFVDGQS VMVEPGTTVL
51 QACEKVGMQI PRFCYHERLS VAGNCRMCLV EIEKAPKVVA ACAMPVMKGW
101 NILTNSEKTK KAREGVMEFL LANHPLDCPI CDQGGECDLQ DQSMMFGSDR
151 SRFLEGKRAV EDKNIGPLVK TIMTRCIQCT RCIRFASEIA GVDDLGTTR
201 GNDMQVGTYI EKMFMSLSG NIIDICPVGA LTSKPYAFTA RPWETRKTES
251 IDVMDAVGSN IVVSTRTGEV MRILPRMHED INEEWISDKT RFAYDGLKRQ
301 RLTEPMVRNE KGLLTHTTWE DALSRVAGML QSFQGNVAA IAGGLVDAEA
351 LIALKDLLNR VSDTLCTEE VFPTAGAGTD LRSNYLLNTT IAGVEEADV
401 LLVGTNPRFE APLFNARIRK SWLHNDLKVA LIGSPVDLTY RYDHLGDSPK
451 ILQDIASGSH PFSQVLQEAQ KPMVILGSSA LQRNDGAAIL AAVSNIAQKI
501 RTSSGVTGDW KVMNILHRIA SQVAALDLGY KPGVEAIQKN PPKMLFLLGA
551 DGGCITRQDL PKDCFIVYQG HHGDVGAPIA DVILPGAAYT EKSATYVNTE
601 GRAQTKVAV TPPGLAREDW KIIRALSEIA GMTLPYDTLD QVRNRLEEVS
651 PNLVRYDDVE GANYFQQASE LSKLVNQQLL ADPLVPPQLT IKDFYMTDSI
701 SRASQTMACK VKAVTEGAHA VEEPSIC

Sequence coverage = 634/704=90.05%

Supplementary Table

A. Detailed MS/MS map of ${}_{544}\text{M}_{(\text{OX})}\text{LFLLGADGGC}_{(\text{SO}_2)}{}^{554}\text{ITR}_{557}$

Δm between y_n and y_{n-1}	Fragment Ion	Measured m/z	Sequence	Measured m/z	Fragment Ion	Δm between b_n and b_{n-1}
			Met _(OX)			
			Leu	261.02	b₂	
146.76	y₁₂	1254.59	Phe	408.15	b₃	147.13
113.36	y₁₁	1107.83	Leu	521.28	b₄	113.13
113.14	y₁₀	994.47	Leu	634.27	b₅	112.99
56.81	y₉	881.33	Gly	691.39	b₆	57.12
186.2 (115+71)	y₈	824.52	Ala			
			Asp	877.41	b₈	186.02 (71+115)
56.88	y₆	638.32	Gly	934.33	b₉	56.92
57.17	y₅	581.44	Gly	991.42	b₁₀	57.09
135 (103+32)	y₄	524.27	Cys_(SO₂)	1126.41	b₁₁	134.99 (103+32)
113.19	y₃	389.27	Ile	1239.46	b₁₂	113.05
	y₂	276.08	Thr	1340.80	b₁₃	
			Arg			

B. Detailed MS/MS map of ${}^{713}\text{AVTEGAHAVEEPSIC}_{(\text{SO}_2)}{}^{727}$

Δm between y_n and y_{n-1}	Fragment Ion	Measured m/z	Sequence	Measured m/z	Fragment Ion	Δm between b_n and b_{n-1}
			Ala			
			Val			
99.82	y₁₃	687.50 ⁺²	Thr	272.26	b₃	
128.98	y₁₂	637.59 ⁺²	Glu	401.22	b₄	128.96
57.78	y₁₁	573.13 ⁺²	Gly	458.61	b₅	57.39
71.11	y₁₀	1087.48	Ala	529.29	b₆	70.68
137.08	y₉	1016.37	His	666.41	b₇	137.12
70.75	y₈	879.29	Ala			
99.23	y₇	808.54	Val	836.44	b₉	170.03 (71+99)
129.07	y₆	709.31	Glu	965.40	b₁₀	128.96
129.01	y₅	580.24	Glu	1094.48	b₁₁	129.08
	y₄	451.23	Pro	1191.53	b₁₂	97.05
			Ser	1278.58	b₁₃	113.28
			Ile	696.43 ⁺²	b₁₄	
			Cys_(SO₂)			