

Table S3. Primers used in this study.

	Primer name	Primer sequence (5' → 3') <sup>c-f</sup>	Type <sup>g-i</sup>	Restriction site(s)	Reference
<b>A. Primers used to generate chromosomal deletion alleles</b>	EF1094 sew-f <sup>a1</sup>	CTGTCCCAGGCTcTCATGCTTTATTTTTAAGGAGGAAGCA/ TAGTATGACGAAAAGGCTAAACATACTAAAAAAAAGAG	DS-f		(1)
	EF1094 sew-r <sup>a1</sup>	CTCTTTTTTTAGTATGTTTAGCCTTTTCGTCATACTA/ TGCTTCCTCCTTAAAAATAAAGCATGA <sub>g</sub> AGCCTGGGACAG	US-r		(1)
	EF1094e-f3 <sup>a2</sup>	<u>gctctaga</u> ATAACGTGACTTCTGGAGAGTATGc <sub>i</sub> TATGC	US-f	<b>XbaI</b>	(1)
	EF1094e-r3	<u>gctctaga</u> CGCGGAACAAAGGAGCTAAAAACTTAATAGAATA	DS-r	<b>XbaI</b>	(1)
	HNV001	tccccgggtctagaACGTTGTTGAAATCCTTGATG	US-f	<b>XbaI, SmaI</b>	This study
	HVN002	TGCGTTTTTCATTGATTACACACT/ATCATTCTCATCTGTTATCATTGAG	US-r		This study
	HVN003	CTCAAATGATAACAGATGAGAATGAT/AGTGTGTAATCAATGAAAAACGCA	DS-f		This study
	HVN004	gcggtctgacgcgtTACCATCCCAATGATTAGCAGGCC	DS-r	<b>SalI</b>	This study
	HVN005	CCAAATGACTACCATCCCAAT/ATCATTCTCATCTGTTATCATTGAGCGG	US-r		This study
	HVN006	CCGCTCAAATGATAACAGATGAGAATGAT/ATTGGGATGGTAGTCATTG	DS-f		This study
	HVN007	CGGTGATGCTAGAAATTAAGCATT/ATCATTCTCATCTGTTATCATTGAG	US-r		This study
	HVN008	CTCAAATGATAACAGATGAGAATGAT/AATGCTTAATTTCTAGCATCACCG	DS-f		This study
	HVN009	tccccgggtctagaAATGTTCAAACAACATTAGTGG	US-f	<b>XbaI, SmaI</b>	This study
	HVN010	CCAAATGACTACCATCCCAAT/CATTGATTACACACTCCCTTCTGG	US-r		This study
	HVN011	CCAGAAGGGAGTGTGTAATCAATG/ATTGGGATGGTAGTCATTG	DS-f		This study
	HVN012	gcggtctgacgcgtAATAAGTAGCAAGACTGCGCCAC	DS-r	<b>SalI</b>	This study
	HVN013	CGGTGATGCTAGAAATTAAGCATT/CATTGATTACACACTCCCTTCTGG	US-r		This study
	HVN014	CCAGAAGGGAGTGTGTAATCAATG/AATGCTTAATTTCTAGCATCACCG	DS-f		This study
	HVN015 <sup>a3</sup>	tccccgggtctagaAAAGCTAGcAACCTAGTAGTG	US-f	<b>XbaI, SmaI</b>	This study
	HVN016	CTTCGGTGTGCTAGAAATTAAGCATT/AATTGCTTCATTTCTCTCT	US-r		This study
HVN017	AGAGAGAAAATGAAGCAATT/AAATGCTTAATTTCTAGCATCACCGAAG	DS-f		This study	
HVN018	gcggtctgacgcgtTCGATTGATTGGCCCTTTGTCCG	DS-r	<b>SalI</b>	This study	
HVN078	CTCTTTTTTTAGTATGTTTAGCC/ATCATTCTCATCTGTTATCATTGAG	US-r		This study	
HVN079	CTCAAATGATAACAGATGAGAATGAT/GGCTAAACATACTAAAAAAAAGAG	DS-f		This study	
<b>B. Primers used to generate chromosomal MIDAS motif mutant allele</b>	HVN226	<u>gctctaga</u> CAGAGCAGCCATTACAACGA	US-f	<b>XbaI</b>	This study
	HVN227	ccgctcgagCCTGAACAGGACCCACACTT	DS-R	<b>XhoI</b>	This study
	HVN228 <sup>b</sup>	GGATTTGGTCTTAGTCGTT(g)GcCTGGgCCGGAgcTATGAATGAAAACAATCGGATT	DS-f		This study
	HVN229 <sup>b</sup>	AATCCGATTGTTTTTCATTCATAg <sub>c</sub> TCCGGcCCAGg <sub>c</sub> (c)AACGACTAAGACCAAATCC	US-r		This study
	HVN241	GGATTTGGTCTTAGTCGTTGcCTGGgCCG	SDM (sense)		This study
	HVN242	CGGcCCAGgCAACGACTAAGACCAAATCC	SDM (antisense)		This study

	Primer name	Primer sequence (5' → 3') <sup>c-f</sup>	Type <sup>g-i</sup>	Restriction site(s)	Reference
<b>C. Primers used to generate <i>E. faecalis</i> expression constructs</b>	HVN080	<i>gcgggatcc</i> ATAACAGTAGAGGATTCTGCTAAA	f	<i>Bam</i> HI	This study
	HVN145	<i>ccgccgctcgag</i> ATAAACTCATGGAGCAATTGG	f; US-f	<i>Xho</i> I	This study
	HVN147	<i>tccccccggg</i> TCATTTCTCTCTCTCTTTTGG	r	<i>Xma</i> I	This study
	HVN150	<i>tccccccggg</i> TAAAGCATTTTCTTTTCTACG	r	<i>Xma</i> I	This study
	HVN184	<i>tccccccggg</i> CTACTTTGGTTTTCTGGTCGTC	r	<i>Xma</i> I	This study
	HVN201	<i>aaaAGGCCT</i> GCTAATCATTGGGATGGTAGTC	f	<i>Stu</i> I	This study
	HVN230	<i>agcttCCTGCAGG</i> TGCTTTTACTTCAGTTAATGTATAGCG	r	<i>Sbf</i> I	This study
<b>D. Primers used to generate <i>E. coli</i> expression constructs</b>	EF1093A Fw	<i>gcgggatcc</i> GAAGAAAATGGGGAGAGCGC	f	<i>Bam</i> HI	(2)
	EF1093A Rev	<i>gcgggagctt</i> aGGTACCTTTGTGTTTGTGG	r	<i>Sac</i> I	(2)
	HVN107	<i>ccgcatgc</i> GCATCGCAAGCAAGCGTTCAAG	f	<i>Sph</i> I	This study
	HVN108	<i>ccctgcag</i> ttaAATAGAACGTTCTTCATTTG	r	<i>Pst</i> I	This study
	HVN114	<i>gcgggatcc</i> ATGAATGGTCGGACAACGTTTCAGCC	f	<i>Bam</i> HI	This study
	HVN117	<i>ccgtcgact</i> taCAAGCGTCCTATGCCACCAGTTTCAGG	r	<i>Sal</i> I	This study
<b>E. Primers used in pGCP123 construction</b>	GP133	<i>gctgcatgctc</i> gatttttataaaacgtctc		<i>Sph</i> I	This study
	GP134	<i>gctagatctt</i> ccgtcgatactatgttatacg		<i>Bgl</i> II	This study
	GP135	<i>gctagatctg</i> tataaacgacggccag		<i>Bgl</i> II	This study
	GP136	<i>gctgcatgcc</i> caggaacagctatgac		<i>Sph</i> I	This study
<b>F. Primers used in pGCP213 construction</b>	oJoy80	<i>gatatgattac</i> gccaaagcttggtagcagctcggatcc	5'-P	<i>Cla</i> I	This study
	oJoy81	<i>gatggatagct</i> gttccctgtgtgaaattgtatccg	5'-P	<i>Cla</i> I	This study
	GP240	<i>ctgcagatate</i> catcacacaggccttggcggccgctcgatgc	SDM (sense)	<i>Stu</i> I	This study
	GP241	<i>gcatgctcgagc</i> ggccccaaggcctgtgtgatggatatctgcag	SDM (antisense)	<i>Stu</i> I	This study
	GP224	<i>gtacgcgt</i> cccgtagaaaagatcaagg		<i>Mlu</i> I	This study
	GP225	<i>gctagatctg</i> gcaagtgtagcgtcac		<i>Bgl</i> II	This study
	GP226	<i>gtacgcgtt</i> ccctcccttaacttacttataaat		<i>Mlu</i> I	This study
	GP227	<i>gctagatctt</i> tattaatcgcaacatcaaacc		<i>Bgl</i> II	This study

<sup>a1-a3</sup> Primer was designed to anneal to *E. faecalis* V583 sequence, resulting in the following DNA mutations in OG1RF strains: 1) C to T mutation in the *ebpC-srtC* intergenic region in SrtC<sup>-</sup>, 2) silent mutation in A386 of EbpC in SrtC<sup>-</sup>, and 3) silent mutation in S155 and a M159V transmutation in EbpB in EbpC<sup>-</sup>.

<sup>b</sup> An aberrantly added base in HVN228/HVN229 is in parentheses and was removed using SDM with primers HVN241/HVN242 to yield pSJH-509.

<sup>c</sup> Uppercase sequence anneals to the OG1RF chromosome.

<sup>d</sup> A forward slash is inserted in primers where chromosomal sequence was deleted.

<sup>e</sup> XbaI, SalI, AatII, SbfI, BglII, and PstI restriction sites are underlined and bolded; SmaI, XhoI, XmaI, BamHI, and SphI restriction sites are shown in bolded, italics type.

<sup>f</sup> MIDAS motif sequence is underlined but not bolded.

<sup>g</sup> Primers are categorized as forward (f) or reverse (r) (or sense or antisense for SDM) relative to the direction of *ebp* transcription.

<sup>h</sup> US (upstream) or DS (downstream) refers to the relevant splice segment for SOE-PCR primers.

<sup>i</sup> 5'-P denotes 5' phosphorylation of the primer.

## REFERENCES

1. **Kline KA, et al.** 2009. *J. Bacteriol.* **191**:3237-3247.
2. **Sillanpää J, Xu Y, Nallapareddy SR, Murray BE, Höök M.** 2004. *Microbiology.* **150**:2069-2078.