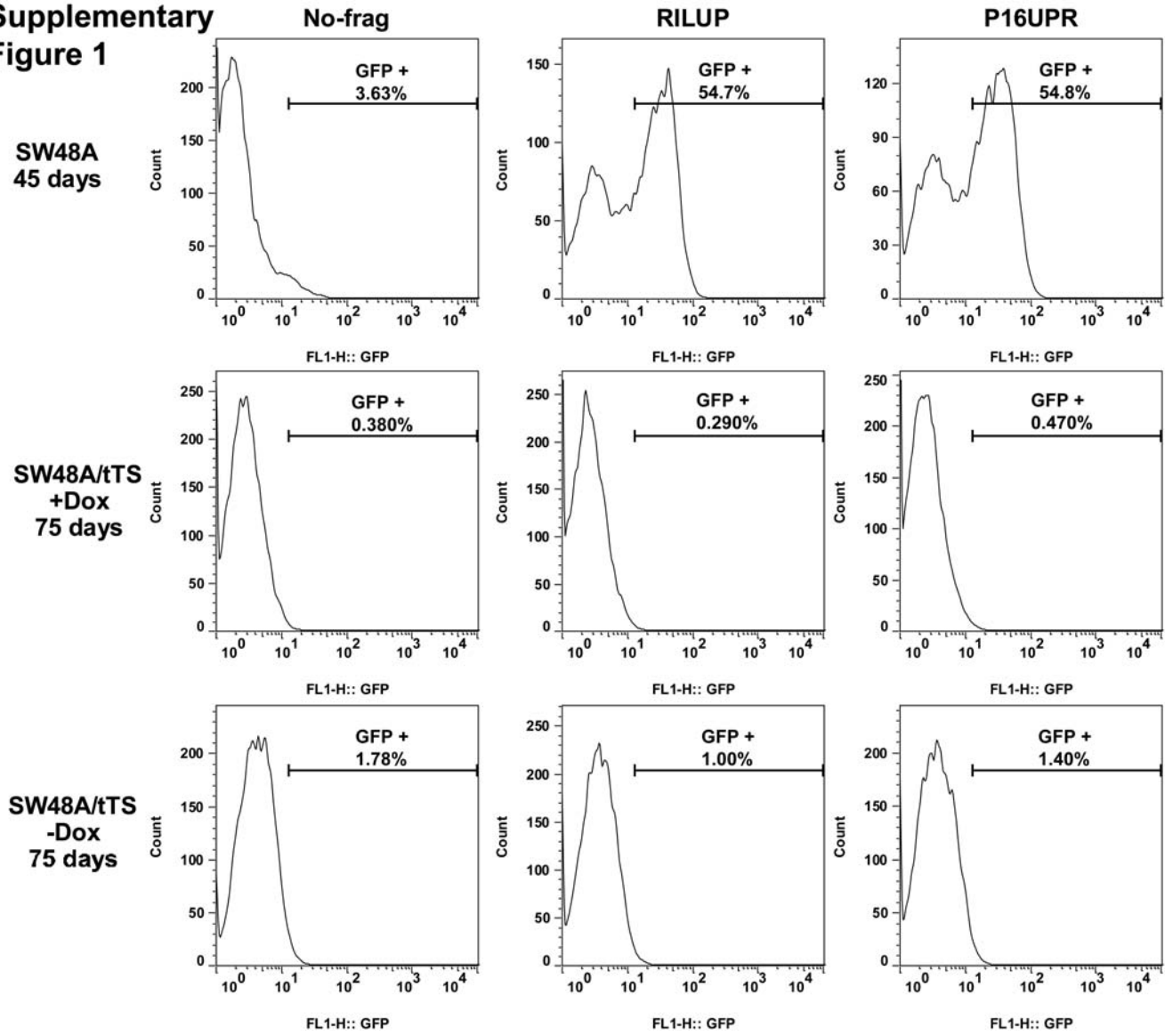
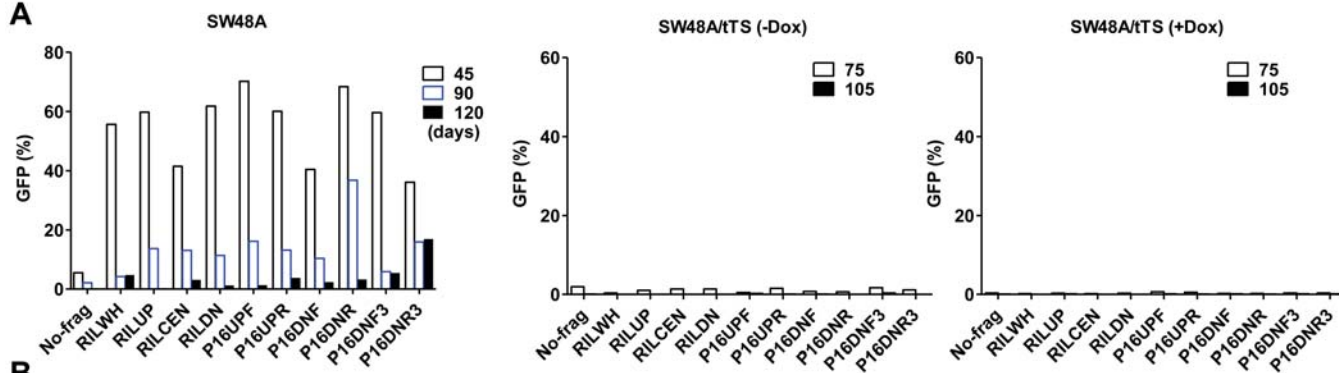


**Supplementary  
Figure 1**

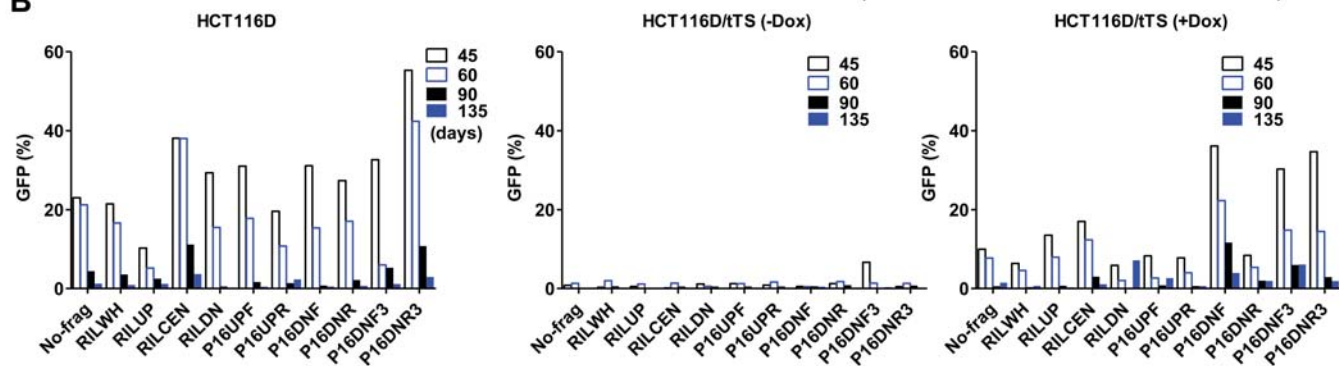


## Supplementary Figure 2

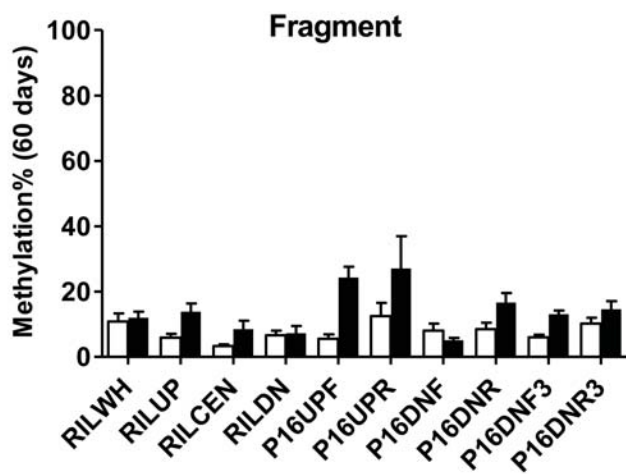
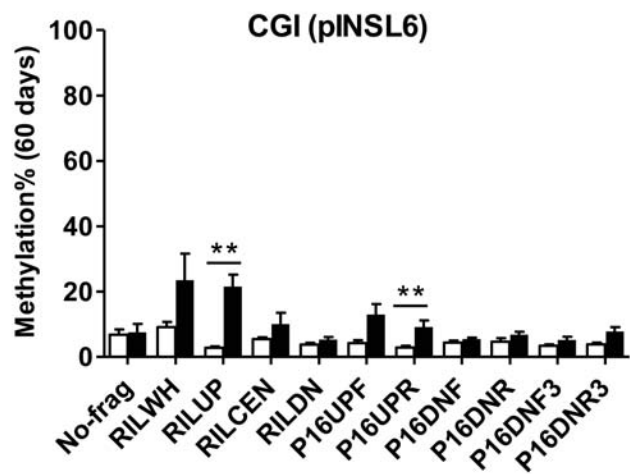
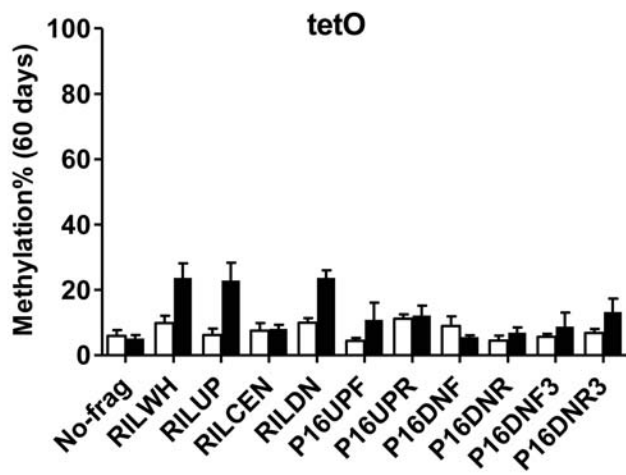
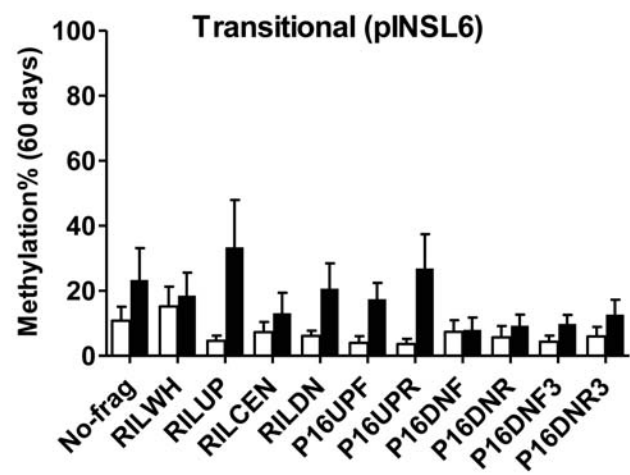
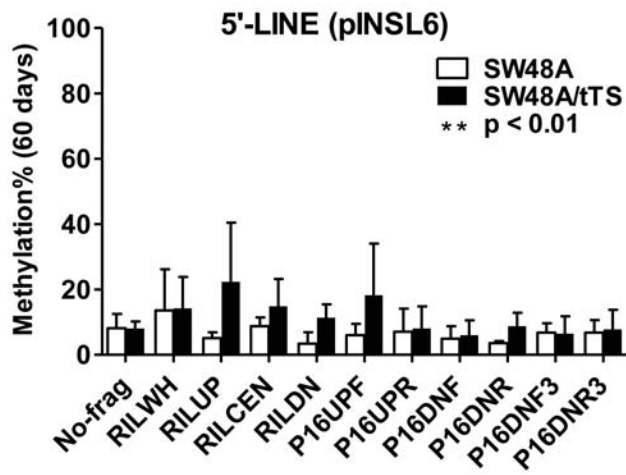
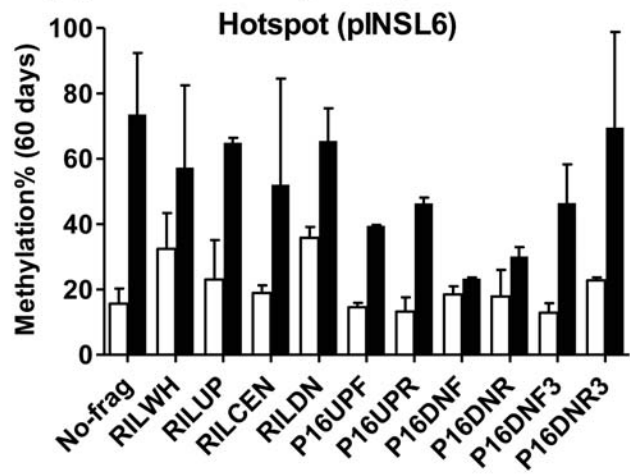
**A**



**B**



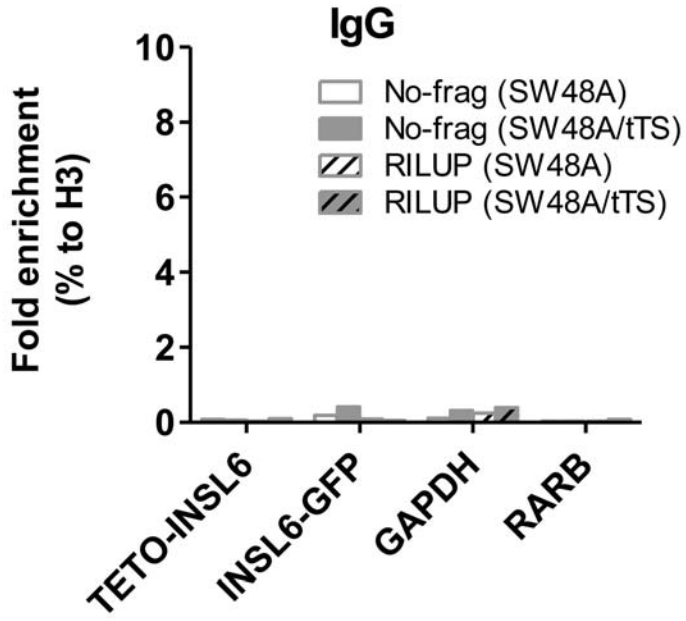
# Supplementary Figure 3



# Supplementary Figure 4

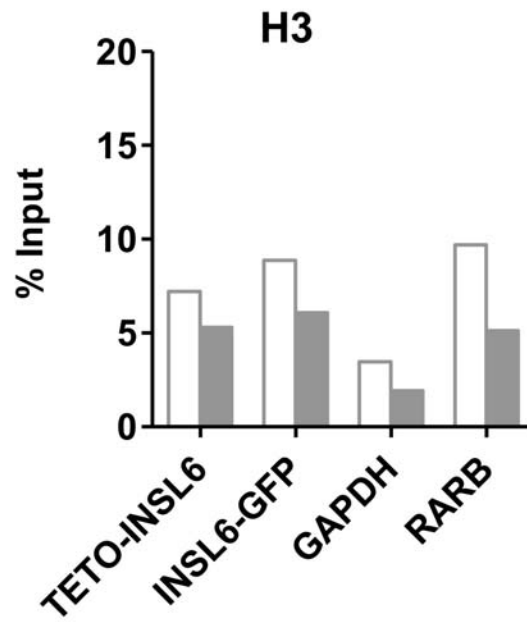
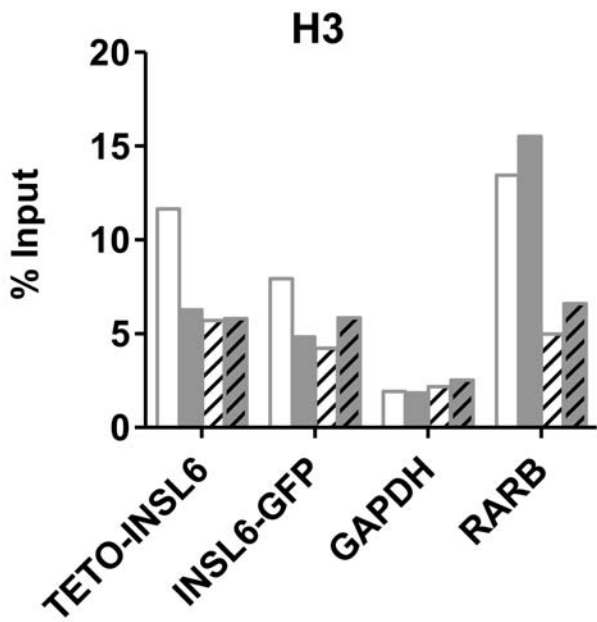
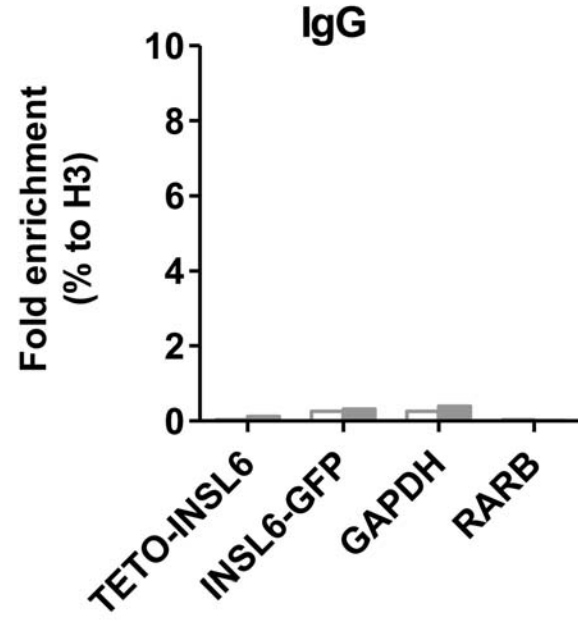
**A**

**75 days**



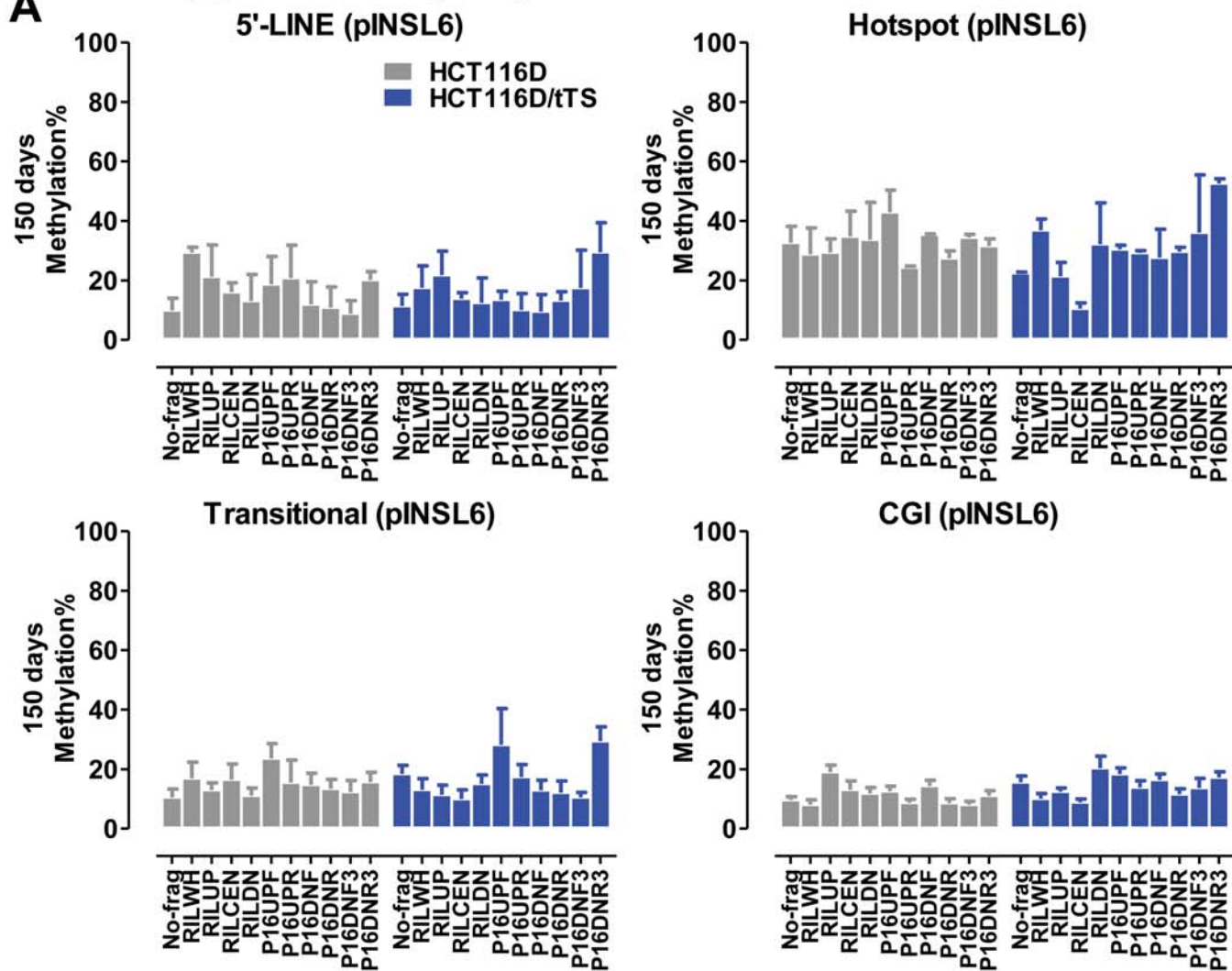
**B**

**180 days**

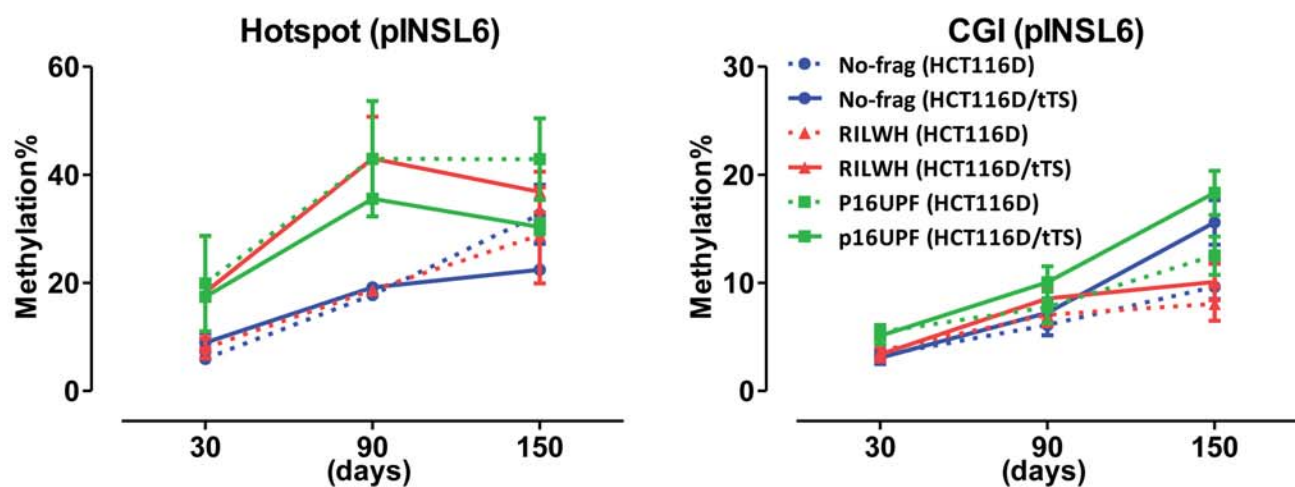


# Supplementary Figure 5

**A**



**B**



**Supplementary figure 1.** Flow cytometry graphs to exemplify transgene expression in Flp-in or Flp-in/tTS cells at the earliest available time points (45 days for SW48A and 75 days for SW48A/tTS). Three transgenes (No-frag, RILUP and P16UPR) are shown here, and please refer to Supplementary Fig. 2 for more transgenes and other time points.

**Supplementary figure 2.** The function of tTS in accelerating GFP silencing. GFP expression was measured by flow cytometry. Stable single clones were sampled at several time points from 45 days to 135 days after transfection. For Flp-in/tTS host cells, clones were cultured in media with and without doxycycline (Dox) respectively. (A) SW48A and SW48A/tTS as host cells; (B) HCT116D and HCT116D/tTS as host cells. GFP expression in long-term cultured tTS-containing clones presented partial inducibility in response to doxycycline, and maintaining clones under doxycycline did not prevent GFP from silencing in the long run.

**Supplementary figure 3.** Comparing the methylation profiles of pINSL6 in SW48A/tTS and SW48A cells (60 days). \*\*, significantly different methylation of each transgene between host cells ( $p < 0.01$ ).

**Supplementary figure 4.** ChIP-qPCR to analyze the H3 occupancy and IgG affinity in pINSL6. All the values of fold enrichment were normalized to H3 and H3 occupancy was calculated as the percentage of input. GAPDH and RARB as the control regions. TETO-INSL6 and INSL6-GFP are targets designed at the 5'-end and 3'-end of pINSL6 in order

to distinguish it from the endogenous one. (A) Two transgenes (No-frag and RILUP) examined in SW48A and SW48A/tTS at 75 days. (B) The transgene (No-frag) examined in SW48A and SW48A/tTS at 180 days.

**Supplementary figure 5.** Methylation profiles of pINSL6 of transgenes in HCT116D and HCT116D/tTS. Pyrosequencing was used to detect regional methylation patterns as described before. (A) Methylation in HCT116D/tTS and HCT116D, sampled at 150 days. Error bar, SEM. (B) Gradual accumulation of transgene methylation. Three time points (30, 90 and 150 days) are shown for three transgenes (No-frag, RILWH and P16UPF).

**Supplementary table 1.** PCR primers and sequencing primers used for bisulfite-sequencing and pyrosequencing.

Name	Step	Forward	Reverse	Sequencing	Target Sequence	Ta (°C)	Size (bp)
BSP	1°	GGTAYGATGGAAAGTAAG AATAG	TCCAAC <del>T</del> CRACCAAAATAA ACAC	NA	NA	50-58(53)	642
INSGFP- P1	1°	GGTAYGATGGAAAGTAAG AATAG	TCCAAC <del>T</del> CRACCAAAATAA ACAC			53	642
	2°	GTAAGATGGAAAGTAAGA ATAGA	U- AAAAACCCCCTATCATCTC T	TAAGTTTATAAATGGAA ATA AGGTTTTTGT <del>TTTT</del> TATGA AG	GGTYGATTAATATYGTG AAA YGGGGGTTTTAGGAAAG AGGTTATAYGTTTTAT	50-61(58)	128
INSGFP- P2	1°	GGTAYGATGGAAAGTAAG AATAG	TCCAAC <del>T</del> CRACCAAAATAA ACAC			53	642
	2°	*U- TAGGAGTAGATAGGGAGT AGG	CTCCACCAAAATAAACAC	AACCCCAA <del>A</del> CTAATCCT	CCRCRTTATACAATAAC	50-61(58)	200
INSGFP- P3	1°	GGTAYGATGGAAAGTAAG AATAG	TCCAAC <del>T</del> CRACCAAAATAA ACAC			53	642
	2°	U- TGAGGGAGAAGAAGTTTT ATTAA	CCCCCTAATTTAATTCATT T	TCCCTCCACCTAAAATA C AATAAAAAATTCCTCAC CA	RTTTCTCR <del>TTCT</del> TATTTA AACRTAAAAACA	54	174
INSGFP- P4	1°	GGTAYGATGGAAAGTAAG AATAG	TCCAAC <del>T</del> CRACCAAAATAA ACAC			53	642
	2°	U- TTTTGGTTTTTATTTTAGG AGTTAG	CCCCTTACTACCATAATA AC	CCAACCCTACTCCCTAT C	TACRCCTACRCCAAACC RAACRACRCRCAC	50-62(58)	329
tetOINS- P1	1°	TTYGGGATTTGGTATTTAG TTG	CCGGAAACTCATAACCTA AACTC			50-58(55)	707
	2°	AGGGATTTGGTATTTAGTT GGTAATT	U- ACCCATACCTCCCTATAAAA CCAATAA	TGGTAATTA <del>A</del> AATATGTG GTG	AAAYGTTTAAGTGTGTA TATGGTTATGTGYGTTTA GTTA	50-62(58)	129
tetOINS- P2	1°	TTYGGGATTTGGTATTTAG TTG	CCGGAAACTCATAACCTA AACTC			50-58(55)	707
	2°	TTGAGGTATTTGGGATGG AAATT	U- CAAATCCTCTAAAAATCCC TATCACA	GGATGGAAATTTGGTTT T	TTYGGTTAGATAA	50-62(58)	99
tetOINS- P3	1°	TTYGGGATTTGGTATTTAG TTG	CCGGAAACTCATAACCTA AACTC			50-58(55)	707
	2°	GATAATTTGTTTAGTTAG GA	U- AAACTCATAACCTAAACTC	TTTATTATTGATAGGGA GTAAAT	TYGATATAYGTTTTTTAT TA	50-58(58)	390
RILWH-P1	1°	ATGAAGAATTTGTTTAGGG TTAGG	U- AAACATTCAAACCTTCTCT TACTACTTA	AGATGTAAGGGTTAGAT ATA	YGYGTTGATATTGATTAT TG	50-58.9	198
RILWH-P2	1°	GTTTAGTGTGGTGGAATTT TGTAG	AACRCRAACCCAAACCC AAC			58-62(58)	487
	2°	GGTGGTTTGAGAGGAGTT TGTA	U- TATCCACACCCACCCTTTA	TTTTTATAAGGTAGGA TGG	GTYGTAGGTGTGTTAGT YGGGTTTAT	50-62(60)	401



			AAATC					
RILUP-P2	1°	TAGTGTGGTGGAAATTTGT AGA	CACTATACTAAAACTACC CCAA				52-62(55)	394
	2°	AGAAGGTTTGAATGTTTTG GGGAAGTA	U- TACCCCAACTCAAATACCT CCTCATAA	TTTTTTATAAGGTAGGA TGG	YGYGTTGATATTGATTAT TG		50-62(58)	286
RILCEN-P	1°	GTTTAGTGTGGTGGAAATT TGTA	ACCCTCCCCTTCRAAAAA CTCT				50-62(55)	319
	2°	AGGTGTAGATAGTTGGGT TTGGG	U- CACACCCCACTCAACTC TC	AAGGTTAGAGTAGGATT TAG	GTYGTYGGGGTYGTTYG AAYGYGGGGATT		50-62(55)	113
RILDN-P1	1°	ATGAAGAATTTGTTTAGGG TTAGG	U- ATCCACAATAACTCCCCCA ATA	AGATGTAAGGGTTAGAT ATA	YGYGTTGATATTGATTAT TG		58	333
RILDN-P2	1°	TAGTGTGGTGGAAATTTGT AGA	CRAAAACCCCAAACCTCC CTAAA				58	421
	2°	U- TTTGGGGGAGTTATTGTG GAT	CAAAACCCCAAACCTCCC TAAAT	CCCCTAACCCATCTC	CRCRAATCCTTCCCRAA TCCAC		60	196
P16UPF-P1	1°	ATGAAGAATTTGTTTAGGG TTAGG	U- TAAACACCCTCCACTAATC A	AGATGTAAGGGTTAGAT ATA	YGYGTTGATATTGATTAT TG		58	96
P16UPF-P2	1°	TGAAGAATTTGTTTAGGGT TAGG	ATCTCTCCACCACCCTCC				50	548
	2°	TTTTAGGTTGGAGTGTAAAT G	U- CTCTATAATCCCAACATTC T	GTAGTTGGGATTATAGG TAT	GYGTTATTAAGTTTYGTT AATTTTG		50-55	243
P16UPF-P3	1°	TGAAGAATTTGTTTAGGGT TAGG	ATCTCTCCACCACCCTCC				50	548
	2°	YGTTATGTTGGTTAGGTTT	U- ATCTCTCCACCACCCTCC A	TTAGAATGTTGGGATTA TAG	AYGTGAGTTATYGTATTY GGATTTT		52-58	207
P16UPR-P1	1°	ATGAAGAATTTGTTTAGGG TTAGG	U- ATCCCTTCCCCCTTTATA ATTAC	AGATGTAAGGGTTAGAT ATA	YGYGTTGATATTGATTAT TG		58	166
P16UPR-P2	1°	TGAAGAATTTGTTTAGGGT TAGG	CACACRTACRCCACCATA ACCAA				50	501
	2°	GGGGTTGTTGTGAGTTTA AATGAT	U- TACCCACCATAACCAACT AATT	AATTTTAGTATTTTGGG AAG	TYGAGGYGGGTAGATTA TTTGAGG		50-55	306
P16DNF-P1	1°	ATGAAGAATTTGTTTAGGG TTAGG	U- ACCACAAATTTCCACTAAT AATCC	AGATGTAAGGGTTAGAT ATA	YGYGTTGATATTGATTAT TG		58	114
P16DNF-P2	1°	TGAAGAATTTGTTTAGGGT TAGG	CTAAAACATAAACTCCAAC CAC				58	601
	2°	GGAGGGATTATTAGTGGA AATTTG	U- TTCTTTCCACAAAATCTCA CTCTA	AGGTTGAGGTAGGAGA AT	YGTTTGAATTYGGGAGG TTG		60	370
P16DNR-	1°	ATGAAGAATTTGTTTAGGG	U-	AGATGTAAGGGTTAGAT	YGYGTTGATATTGATTAT		58	466

P1		TTAGG	CCCTTATAATCCCTTCACT TTAAA	ATA	TG		
P16DNR- P2	1°	AATTTYGGTTYGAGTTTAG GGTAT	TCAAACRATTCTCCTACCT CAACC			58	335
	2°	GTGTTGGATATTAGGAGG GATTAT	U- CCACCCCCCACTACTTTTT ATAT	TGATAGAAATTATTTAG AAG	YGGTYGGGYGYGGTGT TTAYGTTTTGTAA	58-62	241
EGFP-P	1°	AAGGAAGAAGGTAATTATA AGA	U- CCAACCTTATACCCCAAAAT	GGAAGAAGGTAATTATA AGA	TTYGYGTGAGGTGAAG TTYGAGGGYGATAT	50-52	124
HYG-P	1°	AGGGTGTTAAGTTGTAAG ATT	U- TCCCCCATATAAAATCAC	AATTGTTAGTTGTTTTGT AG	TYGGTYGYGGAGGTTAT GGATGYGATYTTGYGG TYGATTTT	50-52	174
siteD-P1	1°	GTAAGAGTAGAAATAGGG GAAGA	U- CATCTCCCCTTAACCTCACA A	AATTAGTTTTTTTTTTAT GA	YGGAGTAAGGATGGAGA ATAAGGATATYGGTTTA	50-58	277
siteD-P2	1°	AGAGATGTAGTAGGTAGT TGGATATA	U- AATTAACCAACTACTTAC ATATTC	TTTTATAGTATAAGTTTA TAGGG	TAYGAAGAATGAATTTTG AGAATGTTTGG	50-58	75
siteA-P1	1°	AGTGATTTGGGGATAAG A	U- TCTTTCACCACTTAATTA TC	TTATGATAGATATTTGG TAA	AYGATGGAAATGT	50-52	78
siteA-P2	1°	AGTTTAGGTTTTTGAATG TAG	U- TTATTTACTCACCTTCTTTA CAT	TGTATTTAGGTTAATTTA TG	TTGTATTTGAATTYGAYG TTAA	50-52	243
Zeo-P1	1°	AAGTAGTGATTGGGAAAA T	U- CATTCCCCATTCAAATA	ATTGGGAAAATTTTGG	YGTTATTTAATTTAATYG TT	50	138
Zeo-P2	1°	GGTTGATTAATTTTTTTAT TTATGT	U- AAACTTCCAATAATAACT TCTC	TTTTTTTTATTTATGTAG AG	GTYGAGGTGTTTYGGT TT	50-52	120

\*U-: Biotin-labeled universal primer (GGGACACCGCTGATCGTTTA).

**Supplementary table 2.** Realtime-PCR primers and probes used for ChIP analysis.

Name	Forward	Reverse	Probe	Amplicon(bp)
SiteA	TGCTGTATTTGAATCCGACGTT	GCCTTGGATGGAAGAACAAATC	ATCTGAAAGGAATCC CT	69
SiteD	AACTTCACAAACATATACTCTGCT GAT	GACGGAGCAAGGATGGAGAA	CTAGACCGGTGTCCT T	67
TETO-INSL	GGCGTGTACGGTCATACTTAAGG	GGTGAGATAATTTTGCTCAGTT AGGA	TCTCTCGAGCCCCTT C	85
INSL-GFP	GCCGACCGCCATTGC	ACTGCAGAATTCGAAGCTTACT TAGA	TCACAGGAGATCTGC	79
ACTB	TCCCCTCCTTTTGCGAAAA	CGGCCAACGCCAAAAC	AGCGAGATTGAGGAA GA	78
GAPDH	CTGCTCCTCCTGTTTCGACAGT	TCACCTGGCGACGCAA	AGCCGCATCTTC	52
RARB	TTGGAAGGAGAACTTGGGATCTT	AGGCTTGCTCGGCCAATC	CTGGGAACCCCC	85

**Supplementary table 3.** Fragments subcloned into transgenes.

Fragments	Distance to TSS (Gene)	Subcloned length (bp)	CG sites	Orientation
RILWH	-713 to +664 (RIL)	1421	96	Forward
RILUP	-713 to -378 (RIL)	336	11	Forward
RILCEN	-406 to +237 (RIL)	643	66	Forward
RILDN	+214 to +664 (RIL)	451	20	Forward
P16UPF	-1752 to -804 (P16)	980	33	Forward
P16UPR	-1752 to -804 (P16)	980	33	Reverse
P16DNF	+768 to +1248 (P16)	511	18	Forward
P16DNR	+768 to +1248 (P16)	511	18	Reverse
P16DNF3	(+768 to +1248)x3 copies (P16)	1485	18x3	Forward
P16DNR3	(+768 to +1248)x3 copies (P16)	1485	18x3	Reverse