

Figure S1. Enhanced stress tolerance in *nrt1.5* mutants is nitrate-dependent. Seedlings were germinated on plates with 5mM (NH₄)₂succinate, then were transferred to the control plates (left), plates supplemented with 50 mM NaCl (middle), or plates with 50 μ M CdCl₂ (right). Plants grew vertically for an additional 6 days before imaging (A) and determination of root elongation rate (B) as described in Methods. Values are mean \pm SD, n=8.



Figure S2. *nrt1.8* is sensitive to drought and salt stresses and neutralizes phenotypes observed in *nrt1.5* mutants. (A) Drought tolerance assay were performed as described in Methods. C×W represents hybrids from Col-0 and Ws crosses, *dm1* represents *nrt1.5-3/nrt1.8-1* and *dm2* represents *nrt1.5-4/nrt1.8-1*. (B) 6-day-old seedlings were transferred to $\frac{1}{4} \times$ basal medium supplemented with 25 mM NO₃⁻(left) or 25 mM NO₃⁻ plus 50 mM NaCl (right), and grown for another 6 days. (C) 5-day-old seedlings were transferred to $\frac{1}{2} \times$ MS or $\frac{1}{2} \times$ MS supplemented with 125 mM NaCl, pictures were taken after 7 days. (D) and (E) Root elongation after transfer to treatments in (B) and (C). Values are mean \pm SD, n=8. ** *P* < 0.01.



Figure S3. Increased *NRT1.8* expression in *nrt1.5* mutants. Plants exposed to 200 mM NaCl for 6h or 10% PEG for 12h or under control condition were subjected to root and shoot sampling. Quantitative RT-PCR was performed to determine *NRT1.8* expression in *nrt1.5* mutants (A) or *NRT1.5* expression in *nrt1.8-2* mutant (B). Data were normalized to *Actin2*, values are mean \pm SD, n =3. Asterisks indicate significant difference between wild type and the mutant lines at *P*< 0.01(**) by *t*-tests.



Figure S4. Chloride allocation in *nrt1.5* mutants. Plants were grown hydroponically for 4 weeks and then exposed to 50 mM NaCl or 20 μ M CdCl₂ for 3 days. Chloride concentration in both shoots and roots were determined by Ion Chromatography (IC). Values are mean \pm SD, n = 8. Asterisks indicate significant difference between wild type and mutant lines at *P*< 0.01(**) by *t*- tests.



Figure S5. Nitrate-dependent growth of *nrt1.5* mutants. (A) 5-day-old seedlings were transferred to $\frac{1}{4} \times basal$ plates with 2.2 mM NO₃⁻(upper left, Ctl), 2.2 mM NO₃⁻ plus 50 mM NaCl (upper middle, Na⁺), 2.2 mM NO₃⁻ plus 50 µM CdCl₂ (upper right, Cd²⁺), 25mM NO₃⁻(lower left, NO₃⁻), 25mM NO₃⁻ plus 50mM NaCl (lower middle, $NO_3^- + Na^+$) or 25mM NO_3^- plus 50 μ M CdCl₂ (lower right, $NO_3^- + Cd^+$). Photographs were taken after an additional growth for 6 days. (B) Quantification of the primary root elongation after transfer to treatments in (A), values are mean \pm SD, n=8. Asterisks indicate significant difference between wild type and mutant lines at P < 0.05 (*) or 0.01 (**) by *t*-tests.