

Supplemental Figure 1. *bex5* Shows Enhanced Sensitivity to BFA.

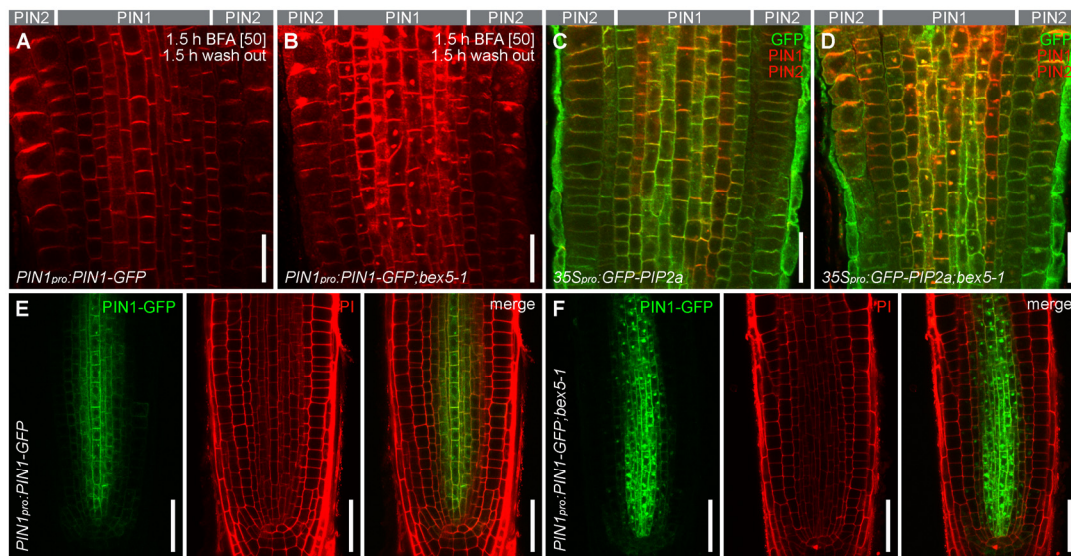
(A) to (G) The *bex5-1* mutation causes protein hyper-accumulation into BFA bodies. PIN1-GFP and PIN2-GFP accumulate in larger BFA bodies in the *bex5-1* mutant treated for 1.5-h with 25 µM BFA (n = 150 BFA bodies from 10 roots; red stars, t-test p-value = 1.33E-26; black stars, t-test p-value = 1.15E-25) (A). Immunolocalization of PIN1-GFP and ARF1 (B) and (C), PM-ATPase and PIN2 (D) and (E) or GFP-PIP2a, PIN1, and PIN2 (F) and (G) in BFA-treated (1.5-h; 25 µM) controls (B), (D) and (F) and *bex5-1* (C), (E) and (G) roots show defective protein accumulation in enlarged BFA bodies in *bex5-1*.

(H) BFA-induced agglomerations have abnormal morphology in the *bex5-1* mutant. They are spread over large areas of the cell (white border), are less compact and the vesicles themselves have an impaired morphology. Seedlings were treated for 1 sh with 50 µM BFA.

(I) and (J) 5-day-old *PIN1_{pro}:PIN1-GFP;bex5-1* seedlings (J) are smaller than *PIN1_{pro}:PIN1-GFP* (I).

(K) Evaluation of root length of 7-day-old *PIN1_{pro}:PIN1-GFP* and *PIN1_{pro}:PIN1-GFP;bex5-1* seedlings grown on 5 µM and 7.5 µM BFA (n = 128).

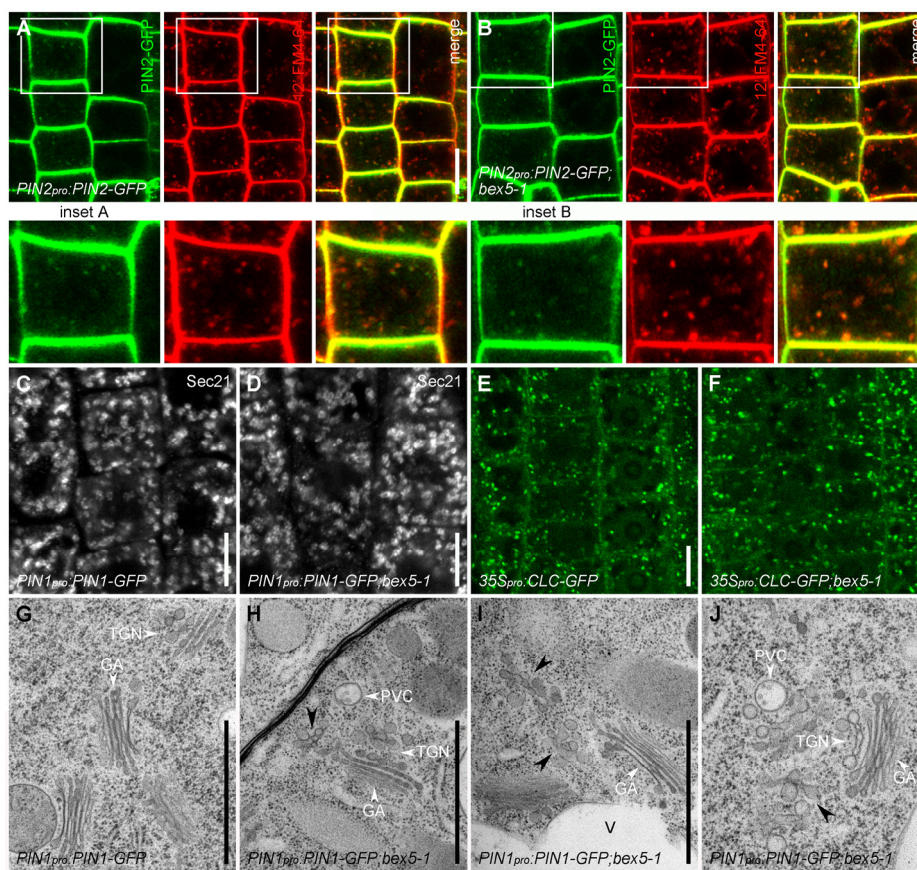
N, Nucleus. Scale bars are 10 µm (B) to (E), 20 µm (F) and (G). Error bars represent SE.



Supplemental Figure 2. *bex5* Is Defective in Exocytosis and Transcytosis of PM Proteins.

(A) to (D) Immunolocalizations show abnormal intracellular accumulation of PIN1 (red color in the vasculature cells) ([A] to [D]), PIN2 (red color in the epidermal and cortex cells) ([A] to [D]) and GFP-PIP2a (green color throughout the root) ([C] and [D]) in the *PIN1_{pro}:PIN1-GFP;bex5-1* (B) and *35S_{pro}:GFP-PIP2a;bex5-1* (D) lines treated for 1.5-h with 50 μ M BFA followed by a 1.5-h wash out in liquid MS medium. In contrast, no or very little accumulation of these proteins could be observed in the *PIN1_{pro}:PIN1-GFP* (A) or *35S_{pro}:GFP-PIP2a* (C) lines.

(E) and (F) Transcytosis of PIN1-GFP is defective in the *bex5-1*. Wild-type seedlings treated for 16-h with 50 μ M BFA show no intracellular accumulation of PIN1-GFP into BFA bodies (E). In contrast, *bex5-1* shows pronounced intracellular PIN1-GFP accumulation (F). Short PI staining (red) shows that long BFA treatment does not interfere with cell viability. Scale bars are 20 μ m (A) to (D) and 40 μ m (E) and (F).



Supplemental Figure 3. *bex5* Shows Abnormal Endosomal Compartments.

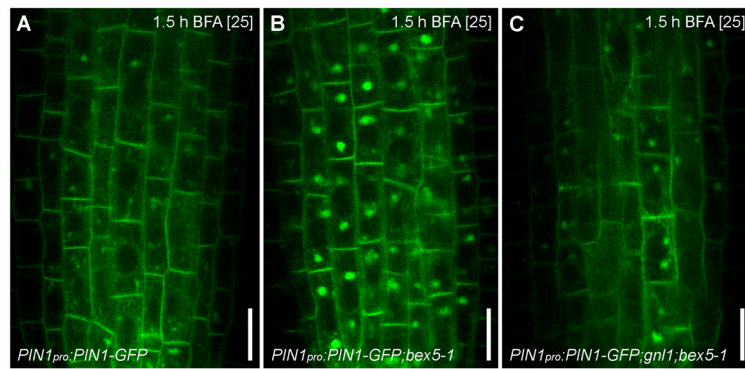
(A) and (B) PIN2-GFP (green) and FM4-64 (red; 2 μ M) label slightly larger endosomal compartments in the *bex5-1* epidermal cells (B) as compared to the control (A). Insets represent enlargements of the boxed regions from the corresponding panels.

(C) and (D) Immunolocalization of Sec21 in *PIN1_{pro}:PIN1-GFP* (C) and *PIN1_{pro}:PIN1-GFP;bex5-1* (D) show that Golgi morphology is not affected in the *bex5-1* mutant.

(E) and (F) Live imaging of CLC-GFP localization in wild-type (E) and *bex5-1* mutant (F).

(G) to (J) *PIN1_{pro}:PIN1-GFP* (G) and *PIN1_{pro}:PIN1-GFP;bex5-1* (H) to (J) cell ultrastructure reveals that *bex5-1* mutant has abnormally fused, shaped and sized vesicles, mainly localized close to one side of the GA-TGN/EE complex. The black arrowheads indicate the abnormal clusters of vesicles in the *bex5-1* mutant.

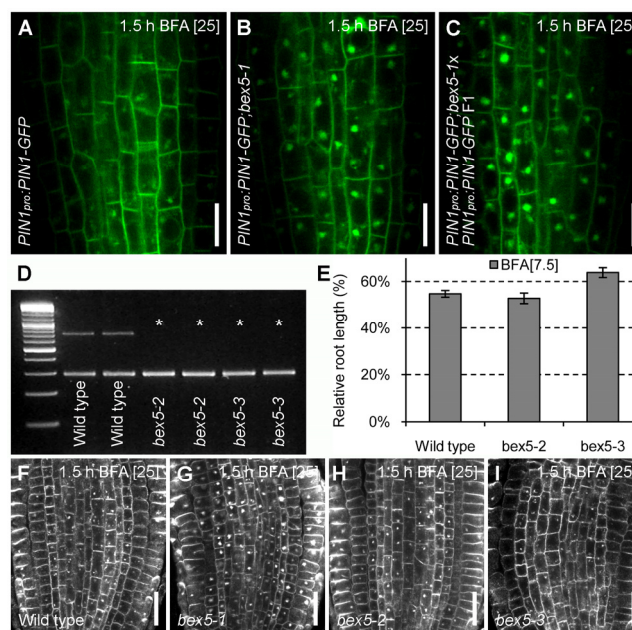
PVC, prevacuolar compartment. Scale bars are 10 μ m (A) to (F) and 1 μ m (G) to (J).



Supplemental Figure 4. *gnl1* Is Epistatic to *bex5*.

(A) to (C) Live imaging of PIN1-GFP in the vasculature of wild-type (A), *bex5-1* (B) and *gnl1*;*bex5-1* (C) roots treated for 1.5-h with 25 μM BFA. Note that *gnl1* mutation reduces the abnormal BFA-induced PIN1-GFP intracellular agglomeration in the *bex5-1* mutant (C).

Scale bars are 10 μm.



Supplemental Figure 5. *bex5* Is a Dominant RabA1b Mutant.

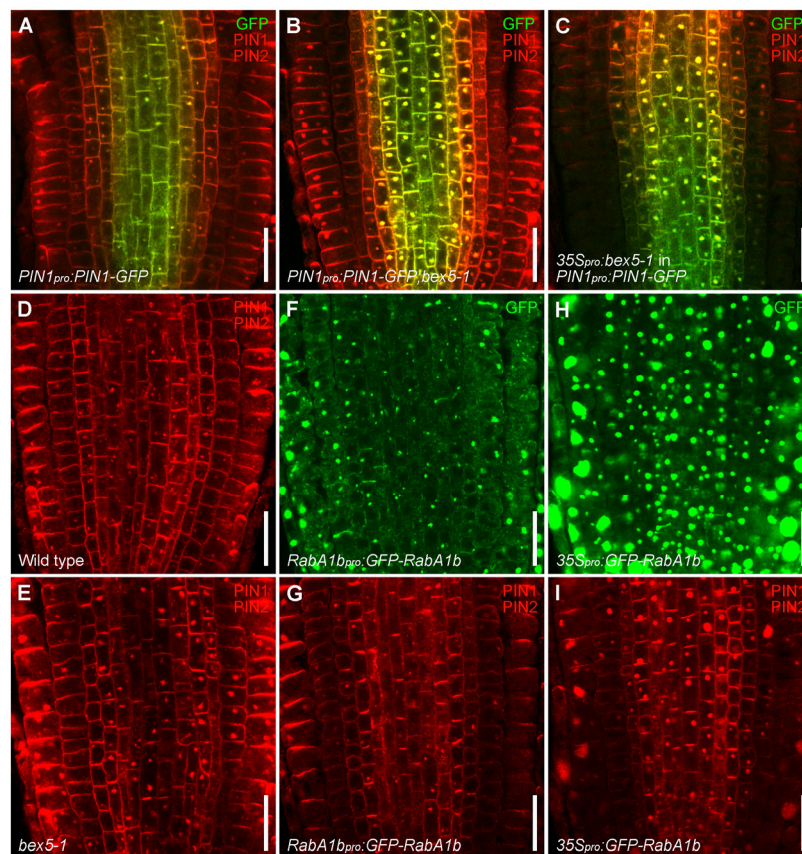
(A) to (C) Live imaging of PIN1-GFP in the vasculature of *PIN1_{pro}::PIN1-GFP;bex5-1* (B) and *PIN1_{pro}::PIN1-GFP;bex5-1* crossed with *PIN1_{pro}::PIN1-GFP* and analyzed in the F1 generation (C) show similar subcellular sensitivity to a 1.5-h treatment with 25 μ M BFA. In contrast, *PIN1_{pro}::PIN1-GFP* (A) shows reduced intracellular PIN1-GFP accumulation into BFA bodies.

(D) *bex5-2* and *bex5-3* mutants are complete knockouts (asterisk), as indicated by RT-PCR. Tubulin was used as control (lower band).

(E) Five-day-old *bex5-2* and *bex5-3* mutants do not show hypersensitivity to the BFA-mediated inhibition of root growth (n = 60).

(F) to (I) Immunolocalization of PIN1 and PIN2 in wild-type (F), *bex5-1* (G), *bex5-2* (H) and *bex5-3* (I) seedlings treated for 1.5-h with 25 μ M BFA. BFA-induced accumulation of PIN1 and PIN2 in *bex5-2* and *bex5-3* is reduced in comparison with their accumulation in *bex5-1*.

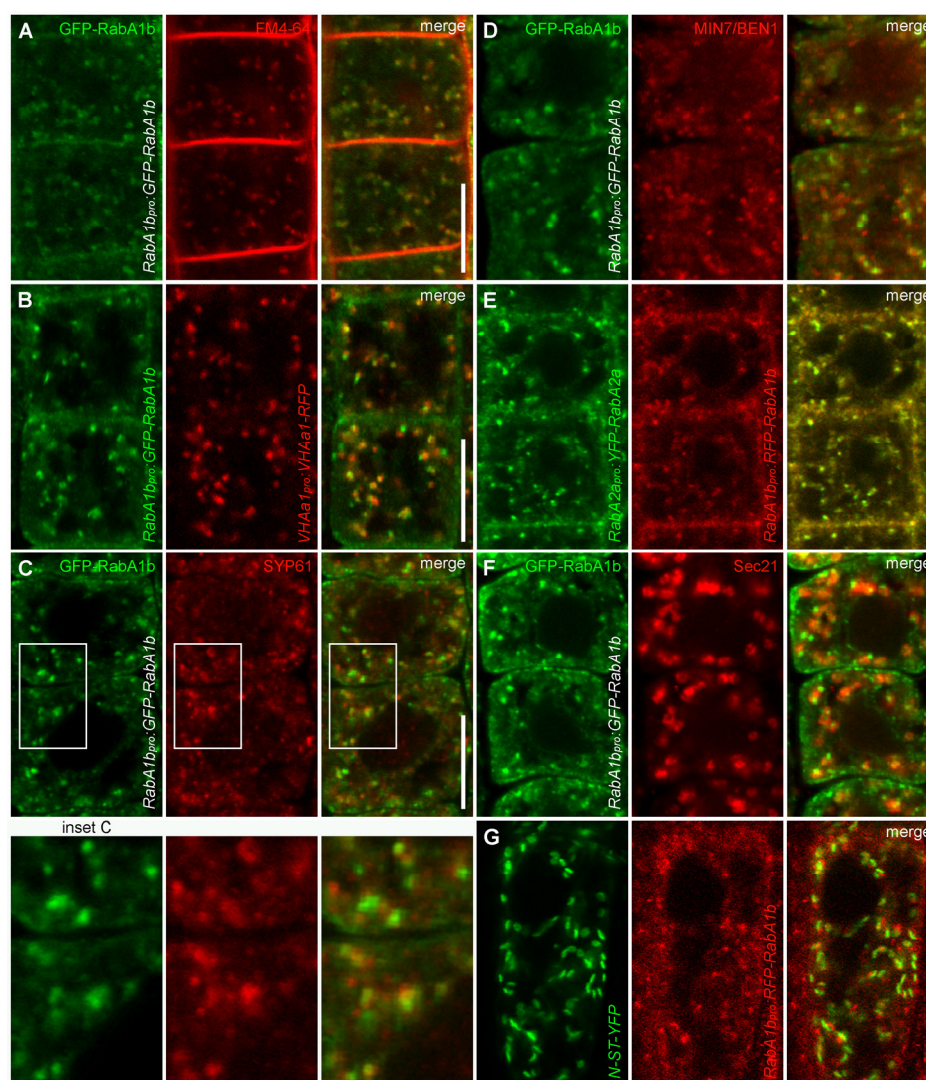
Scale bars are 10 μ m (A) to (C) and 20 μ m (F) to (I). Error bars represent SE.



Supplemental Figure 6. *BEX5* Encodes *RabA1b*.

(A) to (I) Immunolocalization of PIN1 (red color in the vasculature) and PIN2 (red color in the epidermal and cortex cells) in *PIN1_{pro}:PIN1-GFP* (A), *PIN1_{pro}:PIN1-GFP;bex5-1* (B), *PIN1_{pro}:PIN1-GFP;35S_{pro}:bex5-1* (C), wild-type (D), *bex5-1* (E), *RabA1b_{pro}:GFP-RabA1b* WT (G) and *35S_{pro}:GFP-RabA1b* WT (I) seedlings treated for 1.5-h with 25 μ M BFA show that *PIN1_{pro}:PIN1-GFP;35S_{pro}:bex5-1* (C), and *35S_{pro}:GFP-RabA1b* WT (I) display similar BFA-induced abnormal accumulation of PIN1 and PIN2 as *PIN1_{pro}:PIN1-GFP;bex5-1* (B) or *bex5-1* (E), respectively. Immunolocalization of PIN1-GFP (green color in the vasculature) is shown in *PIN1_{pro}:PIN1-GFP* (A), *PIN1_{pro}:PIN1-GFP;bex5-1* (B) and *PIN1_{pro}:PIN1-GFP;35S_{pro}:bex5-1* (C). Immunolocalization of *GFP-RabA1b* WT is shown for *RabA1b_{pro}:GFP-RabA1b* WT (F) and *35S_{pro}:GFP-RabA1b* WT (H).

Scale bars are 20 μ m.



Supplemental Figure 7. BEX5 Localizes at a TGN/EE Compartment.

(A) and (B) Live imaging of *RabA1b_{pro}:GFP-RabA1b* stained with FM4-64 (2 μ M; red) (A) or crossed with *VHAa1_{pro}:VHAa1-RFP* (red) and analyzed in F1 generation (B) shows that GFP-RabA1b (green) partly colocalizes (merge; yellow) with both markers in the root.

(C) and (D) Immunolocalizations show that GFP-RabA1b (green) partly colocalizes (merge; yellow) with SYP61 (red) (C) or MIN7/BEN1 (red) (D). Insets represent magnifications of the boxed regions from the corresponding panels.

(E) YFP-RabA2a (green) shows complete colocalization with RFP-RabA1b (red) in a *RabA2a_{pro}:YFP-RabA2a;RabA1b_{pro}:RFP-RabA1b* F1 generation cross.

(F) and (G) GFP-RabA1b (green) (F) or RFP-RabA1b (red) (G) are distinct of Sec21 (red) (F) and N-ST-YFP (green) (G) although they are often close together. Note that the predominant punctuate shape of RabA1 endosomes is different from that of Sec21 and N-ST endosomes. Immunolocalization (F). Live imaging of *35S_{pro}:N-ST-YFP;RabA1b_{pro}:RFP-RabA1b* F1 cross (G). Scale bars are 10 μ m.