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Table S1. Bacterial strains and plasmids used in this study

Strains or plasmids	Description	Source
<i>Xylella fastidiosa</i> subsp. <i>fastidiosa</i>		
Temecula1	Wild type, Pierce's disease strain	(Guilhabert <i>et al.</i> , 2001)
TAM22	Temecula1, NS1::Cm ^r	(Matsumoto <i>et al.</i> , 2009)
TAM103	Temecula1, <i>xatA3</i> ::Cm ^r	This study
TAM103/pAM61	TAM103 with pAM61- Cm ^r , Gm ^r	This study
<i>Escherichia coli</i>		
BL21(DE3)	F ⁻ <i>dcm</i> <i>ompT</i> <i>hsdS_B(r_B⁻ m_B⁻) gal</i> (λ DE3)	Novagen
EAM1	DH5 α derivative; Sp ^r St ^r <i>attP_{HK022}::(P_{LlacO-1}-PD1607)</i>	(Matsumoto and Igo, 2010)
DH5 α	F ⁻ <i>endA1</i> <i>recA1</i> <i>gyrA96 thi-1 hsdr17(r_K⁻ m_K⁺) relA1 supE44</i> Δ (<i>lacZYA-argF</i>) <i>U169</i> Φ 80 <i>lacZ</i> Δ <i>M15 deor phoA</i> λ ⁻	Lab collection
TOP10	F ⁻ <i>mcrA</i> Δ (<i>mrr-hsdrMS-mcrBC</i>) Φ 80 <i>lacZ</i> Δ <i>M15</i> Δ <i>lacX74 recA1 araD139</i> Δ (<i>ara-leu</i>) <i>7697 galU galK rpsL</i> (St ^r) <i>endA1 nupG</i>	Invitrogen
UT5600	F- <i>ara-14 leuB6 azi-6 lacY1 leu-6</i> <i>proC14 tsx-67 entA403 trpE38 rfbD1</i> <i>rpsL109 xyl-5 mtl-1 thi1</i> Δ (<i>ompT-fepC266</i>) Δ <i>ompP</i>	(Elish <i>et al.</i> , 1988; Kaufmann <i>et al.</i> , 1994)
Plasmids		
pAM34	<i>xatA</i> ⁺ (2.65 kb) in pCR-Blunt II-TOPO, Km ^r	This study
pAM53	<i>xatA</i> passenger domain in pCR-Blunt II-TOPO, Km ^r	This study
pAM54	<i>xatA</i> from pAM53 in pET-29b, Km ^r	This study
pAM61	<i>xatA</i> ⁺ (3.5 kb) in pBBR1MCS-5, Gm ^r	This study
pAM109	<i>xatA3</i> ::Cm ^r in pCR-Blunt II-TOPO; Cm ^r , Km ^r	This study
pBBR1MCS-5	Broad-host range cloning vector, Gm ^r	(Kovach <i>et al.</i> , 1995)
pCR-Blunt II-TOPO	Blunt PCR cloning vector, Km ^r	Invitrogen
pET-29b(+)	Overexpression vector, Km ^r	Novagen
pLBT0528	<i>xatA</i> (3.5 kb) in pCR-Blunt II-TOPO	This study
pRL1342	RSF1010 replicon; Cm ^r , Em ^r	(Wolk <i>et al.</i> , 2007)

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4 **Table S2: Primers**

Primer name	Primer sequence (5'→3')	Restriction Sites	Source or reference
For generating of <i>xatA</i> mutant and complementation			
Cm-f	GG <u>ACTAGT</u> CCAATCAGCGACACTGAATA <u>CGG</u>	SpeI	(Matsumoto <i>et al.</i> , 2009)
Cm-r	GG <u>ACTAGT</u> CCTCACTTATT <u>CAGGC</u> GTAGCAC	SpeI	(Matsumoto <i>et al.</i> , 2009)
PD0528F-Spe	CAA <u>ACTAGT</u> AACAGAACATGACGTGACGC	SpeI	This study
PD0528R-Spe	CAG <u>ACTAGT</u> AACAGCGCTATTGGGCGATT	SpeI	This study
PD0528_fwd	CGCCAACGTTTATT <u>CGAATTGCC</u>		This study
PD0528_rev	GAGTAGTT <u>GCAGCTTAGACTTCATCC</u>		This study
For generating the XatA passenger domain for antibody production			
PD0528-29	CAT <u>ATGAA</u> CTGCCATCC <u>ATCGGAG</u>	NdeI	This study
PD0528-470	CT <u>CGAGCGCGT</u> CTAT <u>CTGACAGTGTG</u> TG	XhoI	This study

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8 **Figure S1.**9 **Phylogenetic tree of XatA homologs.**

10 Sequences exhibiting homology to the XatA passenger domain (amino acids 30-426) with E-
11 values above 2.00E-05 were aligned with MUSCLE (<http://www.biomedcentral.com/1471-2105/5/113>) and visually screened in Jalview to remove redundancies and fragmentary
12 sequences. The resulting pool of sequences was used to generate 1000 maximum likelihood
13 bootstrapped phylogenetic trees using RAxML with the JTT substitution model
14 (<http://bioinformatics.oxfordjournals.org/content/22/21/2688.long>) and visualized using FigTree.
15 Numbers by nodes are bootstrap values. The *X. fastidiosa* Temecula1 homologs are in gray. The
16 followings proteins were used for the analysis: XatA orthologs [Xf Temecula-XatA
17 (NP_778752.1), Xf M23-XatA (YP_001829270.1), Xf M12-XatA (YP_001775231.1), Xf
18 Dixon-XatA (ZP_00651219.1), Xf Ann1-XatA (ZP_00682978.1), Xf CVC-XF1265/XF1264
19 (composite protein generated from DNA sequence by removing a single nucleotide at position
20 1,220,709, which results in a protein of 807 amino acids.)]; XatB orthologs [Xf Temecula1-XatB
21 (NP_779577.1), Xf Ann1-XatB (ZP_00680532), Xf GB514-XatB (ADN62223.1), Xf CVC-
22 XatB (NP_299628.2), Xf Dixon-XatB (ZP_00652420.1)]; XatC orthologs [Xf Temecula1-
23 XatC (NP_779013.2), Xf M23-XatC (YP_001829549.1), Xf GB514-XatC (ADN63803.1), Xf
24 Ann1- XatC (ZP_00680178.1), Xf CVC-XatC (NP_299303.2), Xf Dixon-XatC
25 (ZP_00652050.1), Xf CVC-XF2069 (NP_299348.1)]; Other bacteria [NEIELOOT_00780
26 (*Neisseria elongata* subsp. *glycolytica*; ZP_06733955.1), NEIFL0001_2154 (*Neisseria*
27 *flavescens* SK114; ZP_04757153.1), PROSTU_00583 (*Providencia stuartii* ATCC 25827;
28 ZP_02958823.1), PFL_2727 (*Pseudomonas fluorescens* Pf-5; YP_259834.1), Rmar_2382

30 (*Rhodothermus marinus* DSM 4252; YP_003291648.1), AGROH133_04441 (*Agrobacterium* sp.
31 H13-3; YP_004278026), CPn0796 (*Chlamydia pneumoniae* CWL029; NP_224991.1)].

