Protonated polynucleotide structures. 16. Thermodynamics of the melting of the acid form of polycytidylic acid

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ABSTRACT

A phase diagram (pH, ionic strength, temperature) for the double helical form of poly(C) is presented. The thermodynamic analysis of these data shows that poly(C) behaves essentially as cytidine, if the electrostatic (ionic strength) contributions and the free energy of double helix formation are considered and taken into account.

INTRODUCTION

Polynucleotide structures which form upon protonation are rather rare. Two such homopolymer structures have been studied extensively: $\texttt{poly}(\mathtt{A}')$. $\texttt{poly}(\mathtt{A}'')$ $\overset{\text{--}}{\text{--}}$ and the semi-protonated $poly(C) \cdot poly(C^+)^{4-10}$. While the first of the complexes shows only one acid transition, evidence has been presented^{4,7} that two protonation processes take place in the case of $poly(C)$. It had been shown⁷ that this second protonation of poly(C) is responsible for the bell-shaped T_m vs. pH profile (Fig. 1). It also had been argued⁷ that above 80° C "the cytosine residues behave essentially as in the monomer state", a statement which had been contested by Wrobel et al.¹⁰

RESULTS

Phase diagram of the acid form of $poly(C)$

All published^{7,11,12} and unpublished T_m and titration data on poly(C) accumulated in this laboratory in the last years have been summarized in the three dimensional phase diagram in Fig. 1. It includes some 200 T_m measurements and 50 pK determinations. The vertex at about 80°C is at pH 4.5 in 0.01 M $[Ma^+]$ and at pH 3.7 in 1.0 M $[Na^+]$. Above these pH values thermal denaturation yields neutral poly(C), below protonation takes place during denaturation and $poly(c^+)$ is formed.

Fig 1: Phase diagram of the acid form of poly(C) as a function of pH, temperature and ionic strength (force ionique). red: isotonic (constant ionic strength), green: isothermic (constant temperature), blue: isoprotonic (constant pH).

Thermodynamics of the thermal dissociation of the acid form of $poly(C)$

The protonation of $poly(C)$ takes place in two steps $4,7$ with the double stranded semi-protonated form as intermediate. The following reactions take place between pH 7 and pH 2.5: On the basic side we have:

$$
2 \text{ poly}(C) + H^{\dagger} \Longleftrightarrow \text{poly}(C) \cdot \text{poly}(C^{\dagger})
$$
 (B1)

$$
K_B = \frac{\left[(c^+ \cdot c)_d \right]}{\left[c_g \right]^2 \left[H^+ \right]}
$$
 (B2)

and on the acid side we have:

$$
poly(C) \cdot poly(C^+) + H^+ \longrightarrow 2 poly(C^+) \tag{A1}
$$

$$
K_{\mathbf{A}} = \frac{\begin{bmatrix} c_{\mathbf{S}}^{+} \end{bmatrix}^{2}}{\begin{bmatrix} c^{+} \cdot c \end{bmatrix} \begin{bmatrix} \mathbf{I}^{+} \end{bmatrix}}
$$
 (A2)

We also assume an intrinsic ionisation without formation of a double stranded structure

$$
poly(C) + H^+ \longrightarrow poly(C^+)
$$
 (C1)

$$
K_C = \frac{\begin{bmatrix} c_s^+ \end{bmatrix}}{\begin{bmatrix} c_s \end{bmatrix} \begin{bmatrix} \overline{a}^+ \end{bmatrix}}
$$
 (c2)

and an intrinsic double strand formation without protonation

$$
2 poly(C) \longrightarrow poly(C).poly(C)
$$
 (D1)

$$
K_{D} = \frac{\begin{bmatrix} (C \cdot C)_{d} \end{bmatrix}}{\begin{bmatrix} \sigma_{g} \end{bmatrix}^{2}}
$$
 (D2)

In the above equations C_{g} , C_{g}^{+} , $(C \cdot C)_{d}$ and $(C^{+} \cdot C)_{d}$ represent the concentrations of neutral and protonated cytidine in single and double stranded conformation,respectively.

For each of the four equilibria one can write the appropriate free energy changes as follows

 ΔF_A = $-RT1nK_A$ = 2.3 RT pK_A = ΔH_A - T ΔS_A etc.. It can easily be seen that

 $K_A \cdot K_B = K_C^2$ and $pK_A + pK_B = 2 pK_C$ (C3) The two steps of the protonation of $poly(C)$ can be described by eq. (Bi) and (Al) or , alternatively by an appropriate combination of eq. (Cl) and (DI). The total apparent free energy change between the single and double stranded form can be written for the acid and basic branch, respectively, as

$$
\Delta \mathbf{F}_{\mathbf{A}}^{\mathsf{T}} = \Delta \mathbf{F}_{\mathbf{A}} + \Delta \mathbf{F}_{\mathbf{e}} = 2.3 \text{R} \mathbf{T} \mathbf{p} \mathbf{K}_{\mathbf{A}}^{\mathsf{T}} = \Delta \mathbf{F}_{\mathbf{C}} + \Delta \mathbf{F}_{\mathbf{D}} + \Delta \mathbf{F}_{\mathbf{e}} \tag{A3}
$$

$$
\Delta \mathbf{F}_{\mathbf{B}}^{\bullet} = \Delta \mathbf{F}_{\mathbf{B}}^{\bullet} + \Delta \mathbf{F}_{\mathbf{e}}^{\bullet} = 2.3 \text{R} \text{P} \text{P} \mathbf{F}_{\mathbf{B}}^{\bullet} = \Delta \mathbf{F}_{\mathbf{C}}^{\bullet} - \Delta \mathbf{F}_{\mathbf{D}}^{\bullet} + \Delta \mathbf{F}_{\mathbf{e}}^{\bullet} \tag{B3}
$$

where pK_A^* and pK_B^* are the apparent (experimental) acid and basic transition pH, ΔF_{a} is the electrostatic contribution to the total free energy change, $\Delta \text{F}_{\text{D}}$ the free energy of dissociation(positive

Figure 2: van t'Hoff plot of the reduced pK_A or pK_B (= pK' - ΔF_{ρ} / 2.3.RT) as a function of the inverse of the absolute temperature. \bullet experimental data for cytidine of Wrobel et al.¹⁰ extrapolated to zero ionic strength.

	cytidine		cytidine-5'-phosphate			$poly(c)^+$		
Δ H [*]	ΔS ^{**}				ref. ΔH^* ΔS^* ref. ΔH^*		$\overrightarrow{\Delta S}^{**}$	eq.
-4.0	5.0	10	-3.64	8.0	10			
-3.75	8.1	15						
-4.83	2.5	14					7.7 36.2	A1
-4.4	5.0	16					$-15.5 - 25.8$	B1
						-3.9	5.2	C ₁
						11.6	31.0	D ₁

Table I: Thermodynamic parameters for the protonation of cytidine and cytidine-5'-phosphate compared with those of $poly(C)$

+ Note that for $poly(C)$ all values except for $C1$ are per mole base pair. in kcal/mole ** in cal/(mole.degree)

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on the acid side, negative on the basic side), $\Delta \mathbf{F}_c$ the intrinsic protonation free energy change. Substraction of ΔF gives

$$
\Delta F_A = 2.3R \text{Tr} K_A - \Delta F_e = 2.3R \text{Tr} K_A = \Delta F_C + \Delta F_D
$$
 (A4)

$$
\Delta F_B = 2.3RTPK_B^1 - \Delta F_e = 2.3RTPK_B = \Delta F_C - \Delta F_D
$$
 (B4)

 ΔF_a can be evaluated from the ionic strength dependence of the experimental pK values according to Kotin¹³

 ΔF_e = RT (c₁ - c₂. log [Na⁺]) (D3) where c_2 and $c_1 (= c_2/2)$ are constants; these are found to be c_i =0.525 and c_2 =1.05, identical to those obtained for the acid form of $poly(A)^3$. A plot of the reduced pK's (=pK'- $\Delta F_{e}/2.3RT$) against 1/T should give a plot which is independent of the ionic strength. The result in Figure 2 shows the correctness of the choice of variables. From this plot can now be determined the enthalpy and entropy values for the four reactions considered (eq. A1,B1,C1 and DI). The results are summarized in Table ^I and compared with the literature results on cytidine and 51CMP. Since one deals with cooperative transitions, the contributions in stacking at the transition point should be small (of the order of scatter in the points in Figure 2). DISCUSSION

The only complete temperature study (between 10°and 80°C) of pK values of cytidine and its 5'-phosphate and their ionic strength dependence has been published by Wrobel et al.¹⁰. The agreement between their data (large dots on the broken line in Fig.2) and our computed values of $pK_{,1}$ (broken line in Fig. 2) is striking. From these results it is quite clear that melting and titration of $poly(C)$.poly (C^+) can be expressed as the sum of three terms: the electrostatic contribution, the dissociation free energy (positive or negative, depending on the branch) and the free energy of protonation of cytidine. In the absence of double strand formation and ionic strength effects (i.e. above the vertex in Figure ¹ at temperatures higher than 80°C) poly(C) behaves like cytidine according to equation $(C1)$

The dissociation enthalpy $\Delta H_{D} = 11.6$ kcal/(mole base pair) compares favorably with the value determined by Pohl17 for poly(dG-C). In both oases three hydrogen bonds per base pair are broken. Similarly, the entropy values are comparable. These values are considerably larger than those obtained for

other polynucleotide complexes^{18-20,23-25} containing only two hydrogen bonds per base pair. In contrast, Podder²¹ determined

Figure 3: Dissociation enthalpy ΔH_D of various polynucleotide complexes as ^a function of hydrogen bonds formed. $O\bullet$: $poly(A)$. $poly(U)$ and $poly(A)$. $poly(U)$. $poly(U)$ (ref. 18,19); $\bullet: \text{ poly}(1) \cdot \text{poly}(C)$ (ref. 20); $\triangledown: \text{ poly}(dG-C) \cdot \text{poly}(dG-C)$ (17); $\nabla: poly(dA-T).poly(dA-T)$ (ref. 23); A: T₂-DNA (ref. 25); $\blacksquare: \text{ poly}(U)$.poly(U) (ref. 24); $\Delta : \text{ poly}(C)$.poly(C⁺) this work; \blacksquare : GMP-gels (ref. 22). closed symbols: calorimetric data; open symbols: T_{m} data.

This compilation is not exhaustive.

for the interaction of GpGpGpC with CpCpCpG a ΔH_{p} = 5.4 kcal/ (mole base pair) without taking into account end effects and nucleation. A value of 7 to 8 kcal $\sqrt{\mod{p}}$ was found by Chantot²² for the gel formation of guanylic acids when high polymerisation was observed. In this case two hydrogen bonds are formed for each base in ^a tetrameric structure which has ^a total of eight hydrogen bonds. In Figure 3 an attempt is made to correlate these observations. It appears that the dissociation enthalpy $\Delta H_{\rm m}$ depends on the number of hydrogen bonds formed in the polynucleotide structure. Extrapolations indicate the low stability of structures with only one hydrogen bond per base pair (which have not-been observed).

The rather high dissociation enthalpy of poly(C).poly(C⁺) suggests that three hydrogen bonds are formed and that the base pairing scheme (with parallel strands and a shared proton forming the third hydrogen bond) generally accepted $4-8$ may be correct. Recent viscosity data⁹ had been interpreted as a bent-back hairpin structure for $poly(C)$.poly (C^+) which, however, could only form two hydrogen bonds (anti-parallel strands). Kinetic experiments and precise x-ray fibre diffraction data will be needed to definitively clcar this point.

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REFERENCES

- 1) Rich, A., Davies, D.R., Crick, F.H.C. and Watson, J.D. (1961) J.Mol. Biol. 3, 71-86
- 2) Holcomb, D.N. and Timascheff, S.N. (1965) Biorolymers 3, 121-133
- 3) Guschlbauer, W. and Vetterl, Vl. (1969) <u>FEBS-Lett.4</u>, 57-60
- 4) Hartmann, K.A. and Rich,A. (1965)J.Amer.Chem.Soc. 87,2033-2038
- 5) Langridge, R. and Rich, A. (1963) Nature 198, 725-728
- 6) Akinrimisi, E.O., Sander, C., and TS^* 0.P. $0.$ P. (1963) Biochemistry 2, 340-345
- 7) Guschlbauer, W. (1967) Proc.Natl.Acad.Sc.U.S. 57, 1441-1448
- 8) Adler, A., Grossmann, L. and Fasman, G.D. Biochemistry 8, 3846-3853
- 9040-9099
9) Chen,G.C. and Yang, J.T. (1973) <u>Biophys. Chem. 1,</u> 62-72
- 10) Wrobel, A., Rabezenko, A. and Shugar, D. (1970) Acta Biochimica Polonica 17, 339-349
- 11) Thiele, D. and Guschlbauer, W. (1969) Biopolymers 8, 361-378
- 12) Sarocchi,M.Th., Courtois,Y. and Guschlbauer, W. Eur. J. <u>Biochem. 14</u>, 411-421
- 13) Kotin,L. (1963) <u>J. Mol. Biol.</u> 7, 309-311
- 14) Christensen, J.J., Rytting, J.H. and Izatt, R.M. (1970) J. Chem. Soc. B, 1643-1646
- 15) Lewin, S. and Humphreys, D.A. (1966) J.Chem. Soc. B, 210-213
- 16) Sukhorukow, B., Poltev. V. and Blumenfeld, L. (1964) Abh. <u>DAWB, Kl. Med.</u> 381
- 17) Pohl, F. (1974) <u>Eur. J. Biochem.</u> 42, 495-504
- 18) Massoulié,J. (1968) <u>Eur. J. Biochem.</u> 3, 428-438
- 19) Neumann, E. and Ackermann, T. (1969) <u>J. Phys. Chem. 73</u>, 2170-2178
- 20) Ross, P.D. and Scruggs, R.L. (1970) J. Mol. Biol. 45,567-570
- 21) Podd**er, S.K.** (1971) <u>Bur. J. Biochem. 22</u>, 467–477
22) Chantot, J.F. (1972) <u>Arch. Biochem. Biophys. 153</u>, 347–356
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- 23) Scheffler, I.E. and Sturtevant, J.M. (1969) <u>J.Mol.Biol. 42</u>, 577-588
- 24) Heinecke, M., Bode, D. and Schernau, U. (1974) Biopolymers 13, 227-235
- 25) Privalov, P.L., Ptitsyn, O.B. and Birshtein,T.M. (1969) Biopolymers $8, 559-571$