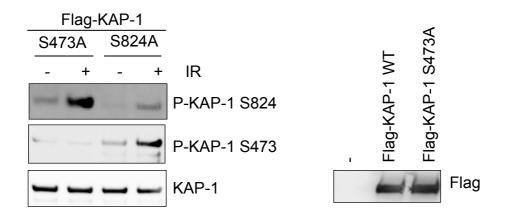


Sequence		Expectation	Site	No.
	Ions	Value		Occasions
	Score			Identified §
Ac-AASAAAASAAASAASG pS PGPGEGSAGGEKR/S	101	4.2×10^{-8}	S19	16
${\tt STAPSAAASASASAAASSPAGGGAEALELLEHCGVCR}$	56	6.8×10^{-3}	S50	17
K/LLA S LVKR/L	58	2.4×10^{-4}	S258	2
K/QG pS GSSQPMEVQEGYGFGSGDDPYSSAEPHVSGVKR/S	73	6.5×10^{-5}	S437	4
K/QGSGS pS QPMEVQEGYGFGSGDDPYSSAEPHVSGVKR/S	70	1.3×10^{-4}	S440	4
R/pSGEGEVSGLMR/K	68	3.8×10^{-5}	S473	>25
R/V pS LERLDLDLTADSQPPVFK/V	52	2.8×10^{-3}	S489	8
R/LAp(SPS)GSTSSGLEVVAPEGTSAPGGGPGTLDDSATICR/V	57	2.4×10^{-3}	S594	17
			or	
			S596	5
R/LQEKL p\$ PPYSSPQEFAQDVGR/M	79	6.0×10^{-6}	S752	17
K/LSPPYp(SS)PQEFAQDVGR/M	68	8.6×10^{-5}	S757	14
			or	
			S756	8
K/FSAVLVEPPPMSLPGAGL p(SS)QELSGGPGDGP/-	41	7.1×10^{-2}	S824	10
			or	
			S823	4

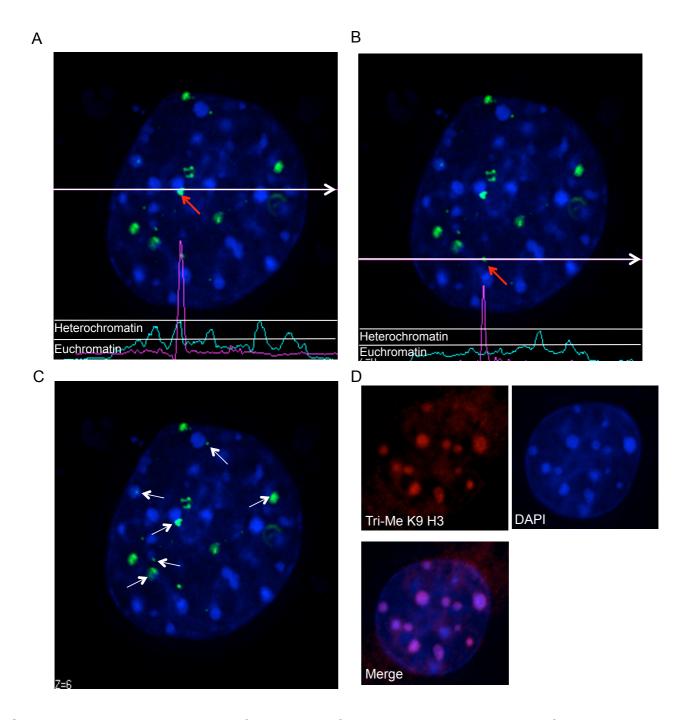
1100

1300

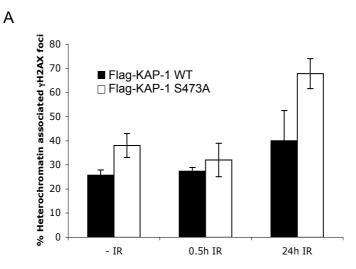
Supplementary Figure 1. A. 293T cells were transfected with Flag alone or Flag-KAP-1 WT and harvested one hour after treatment with IR (10Gy). Flag-KAP-1 was immunoprecipitated, resolved on an SDS PAGE gel and stained with Colloidal Coomassie G-250. The band corresponding to Flag-KAP-1 was excised and used for mass spectrometric phospho-peptide and phospho-residue mapping. B. The doubly charged precursor peptide at m/z 722.82 which represents KAP-1₄₇₁₋₄₈₃ (doubly charged 1444.67 peptide) was selected for fragmentation in a nanoESI-MSMS analysis. The fragmented ions matched the sequence of the KAP-1 $_{471-483}$ peptide where serine-473 is phosphorylated, the phospho-serine was detected between ions b_2 and b_3 and also redundantly between ions y_{10} and y_{11} -98. C. Detected KAP-1 phosphopeptides and phosphosites. § The number of times the phosphosite has been reported according to www.phosphosite.org.



Supplementary Figure 2. A. HeLa cells were transfected with the indicated constructs and mock-treated or treated with 6 Gy IR. Cell extracts were taken 1 h post-IR and immunoblotting performed with the indicated antibodies. B. HeLa cells were treated as in A. (in the absence of IR) and immunoblotted with Flag antibodies.



Supplementary Figure 3. Identifying γ H2AX foci associated with regions of heterochromatin. A-C. NIH3T3 cells were transfected with Flag-KAP-1 S473A. After 24 hours cells were treated with 2 Gy IR and harvested 24 hours post-IR. Cells were stained with γ H2AX (green) and DADI (blue). High resolution images were catpured using a Delta Vision PDV microscope, images were deconvolved using softWoRx Suite software. Signal intensity chromatographs are shown at the base of A. and B. Pink lines indicate γ H2AX signal and light blue lines indicate DAPI signal. A. Shows the chromatograph of a γ H2AX foci (indicated by a red arrow) associated with heterochromatin. B. Shows the chromatograph of a γ H2AX foci (indicated by a red arrow) not associated with heterochromatin. C. Shows an image with the heterochromatin-associated γ H2AX foci indicated by white arrows. D. NIH3T3 cells were fixed and stained with trimethylated K9 histone H3 and DAPI.

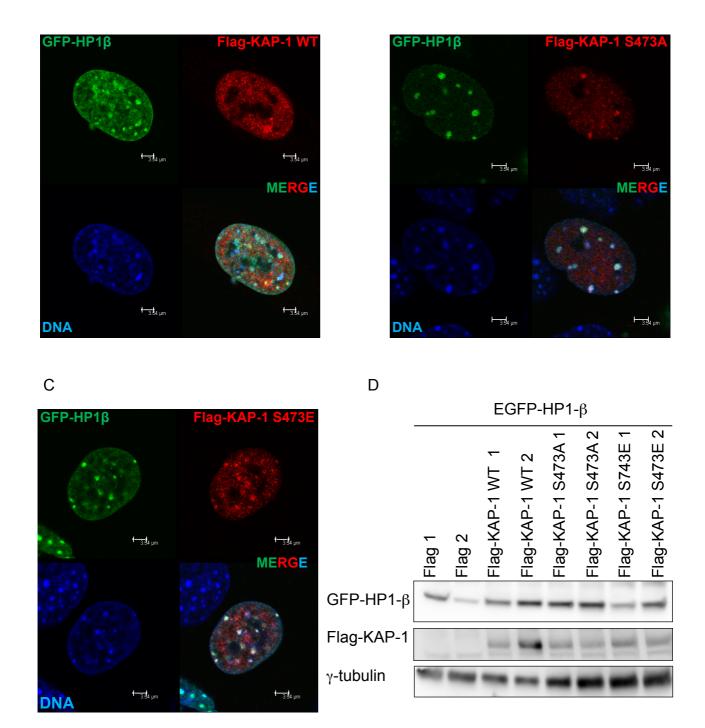


В

	KAP-1 WT - IR	KAP-1 WT 0.5hIR	KAP-1 WT 24h IR	KAP-1 S473A - IR	KAP-1 S473A 0.5h IR	KAP-1 S473A 24h IR
Total γH2Ax foci/cell	0.38	29	1.82	0.82	29	4.5
γH2Ax foci/cell associated with HC	0.10	7.83	0.69	0.31	9.67	3.03
% HC associated γ H2Ax foci	25.67	27.33	40	38	32	67.8

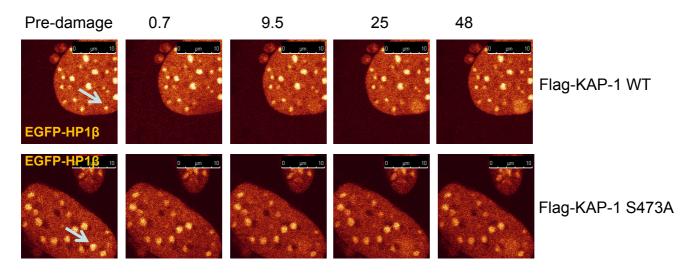
Supplementary Figure 4. > 67% of unrepaired γ H2AX foci are associated with heterochromatin in cells expressing S473A. Growth-arrested confluent NIH3T3 cells were transfected with Flag-KAP-1 WT or S473A. After 24 hours cells were exposed to 2 Gy IR and harvested at either 0.5 or 24 hour/s post-IR. Cells were immunostained for γ H2AX and DAPI and were analysed as described in Supplementary Figure 3. Briefly, captured images were examined and the total number of γ H2AX foci and γ H2AX foci associated with heterochromatin were counted. From this the percentage of heterochromatin associated γ H2AX foci was calculated. A. Graphical representation of the percentage of heterochromatin associated γ H2AX foci. B. A table showing the data values collected.

A B



Supplementary Figure 5. Stable expression of EGFP-HP1- β and Flag-KAP-1 or, S473A in MEF cells. Immunofluorescence images of fixed MEFs stably expressing EGFP-HP1 β and FLAG-KAP1 WT (A), S473A (B) or S473E (C). KAP-1 is visualised using anti-FLAG staining and DNA using DAPI. D. Expression of EGFP-HP1- β , Flag-KAP-1 WT, S473A and S473E in MEFs as shown by immunoblotting with the indicated antibodies.

Time (s) after damage

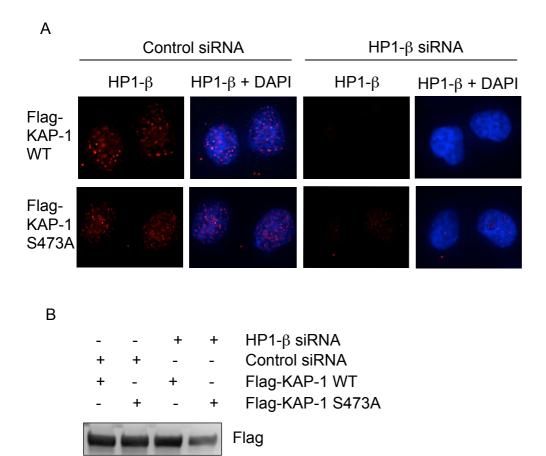


В

Results of FRAP one phase association curve fits

		Undamaged Damaged		95 % confidence intervals	
Flag-KAP-1 WT	Half time of recovery	5.5	4.5	5.1 to 6.1	4.3 to 4.8
	Plateau of recovery	68.6	80.5	67.7 to 69.5	79.9 to 81.1
Flag-KAP-1 S473A Half time of recovery		5.06	4.4	4.7 to 5.5	4.2 to 4.7
	Plateau of recovery	74.6	79.4	73.8 to 75.5	78.8 to 80.0

Supplementary Figure 6. EGFP-HP1- β fluorescence recovery. A. Time-lapse imaging of EGFP-HP1 β following 405nm laser activated DNA damage in living cells transiently expressing Flag-KAP1 WT or S473A. B. Results of the fits to the EGFP-HP1 β FRAP recovery curves shown in Figure 5.



Hp1-β

Non-specific

Supplementary Figure 7. A, B. HeLa cells were transfected with control or HP1- β siRNA 24 h before transfection with Flag-KAP-1 WT or S473A. Cells were fixed or extracted 48 h post-siRNA transfection and immunofluorescence (A) or immunoblotting (B) was carried out with the indicated antibodies.