Edgerton-Morgan and Oakley, http://www.jcb.org/cgi/content/full/jcb.201203115/DC1

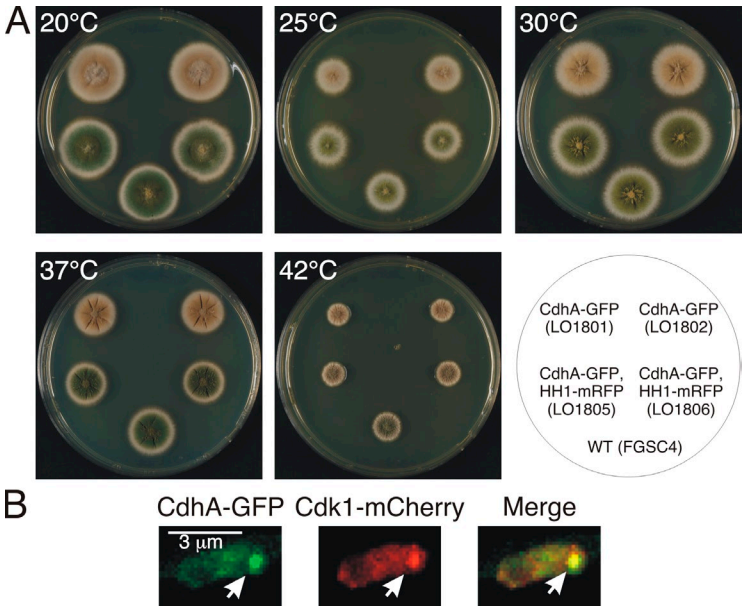

Figure S1. **CdhA-GFP fusions support normal growth.** (A) Strains expressing CdhA-GFP and a control WT strain were inoculated onto complete media and incubated at the temperatures shown. The times of incubation were different at the different temperatures to partially compensate for different growth rates at the different temperatures. Strains expressing CdhA-GFP grew as well as the WT strain at all temperatures. (B) Projections of z-series stacks of CdhA-GFP and Cdk1-mCherry at 25°C in an *mipA*$^{+}$ strain. CdhA and Cdk1 colocalize at the SPB. Cdk1 has previously been shown to be concentrated at the SPB (arrows), and CdhA is concentrated at the same spot.

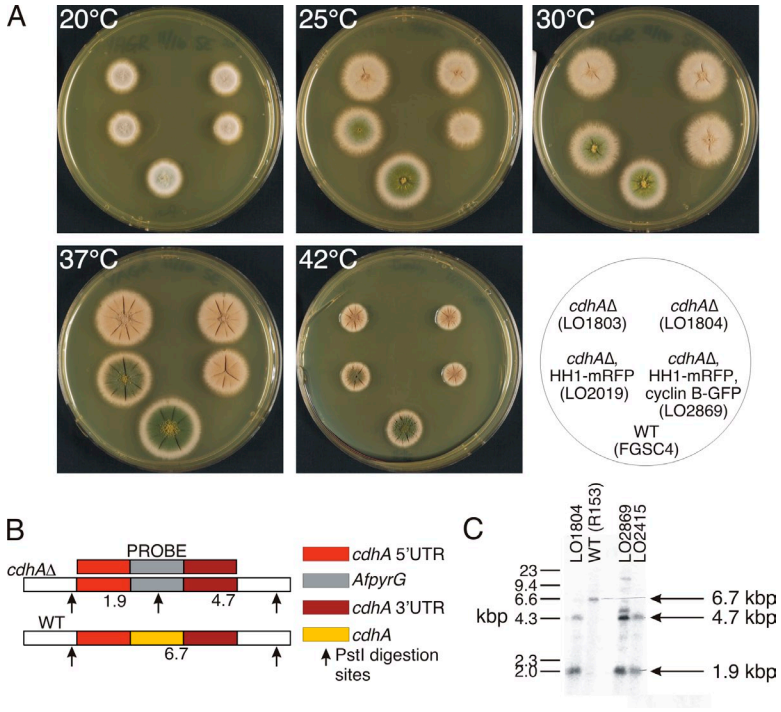

Figure S2. ***cdhA* deletion strains are viable.** (A) Strains containing *cdhA*Δ as well as a WT control strain were inoculated onto complete media and incubated at the temperatures shown. The times of incubation were different at the different temperatures to partially compensate for different growth rates at the different temperatures. *cdhA*Δ strains are slightly temperature sensitive at 42°C. (B and C) Southern hybridizations confirm that *cdhA* is not essential. (B) The *cdhA::AfpyrG* fusion PCR product was radioactively labeled and used as a probe. Arrows show the predicted restriction sites for PstI. UTR, untranslated region. (C) The probe was hybridized to DNA prepared from a putative *cdhA*Δ transformant (LO1804) as well as two other strains that are progeny of crosses predicted to carry *cdhA*Δ (LO2869 and LO2415). All these strains showed bands of hybridization of the sizes predicted for *cdhA*Δ (1.9 and 4.7 kbp vs. the WT band of 6.7 kbp). This verifies that *cdhA* is not essential, and *cdhA*Δ stains are viable. The extra bands in the LO2869 lane are a result of the probe hybridizing to CB-GFP-*AfpyrG* (5.3 and 12.6 kbp), which this strain also carries. Radioactively labeled bacteriophage λ-DNA digested with HindIII was not used in this gel; however, a picture of the gel was taken before the addition of probe so that the sizes could be determined. These are marked (in kilobase pairs) to the left of the gel. R153 was used as a WT control in this experiment.

Table S1. ***A. nidulans* strains used in this study**

Strain no.	Genotype
FGSC4	Glasgow WT
LO1438	*pyrG89; nimE-GFP-AfpyrG; hhoA-mRFP-AfpyrG; pyroA4; nkuA::argB; riboB2; yA2*
LO1439	*pyrG89; nimE-GFP-AfpyrG; hhoA-mRFP-AfpyrG; pyroA4; nkuA::argB; mipA*-D159
LO1501	*pyrG89; pabaA1; pyroA4; nkuA::argB; riboB2; fwA1*
LO1516	*pyrG89; pyroA4; nkuA::argB; riboB2; hhoA-mRFP-AfriboB*
LO1801/1802	*pyrG89; cdhA-GFP-AfpyrG; pabaA1; pyroA4; nkuA::argB; riboB2; fwA1*
LO1803/1804	*pyrG89; cdhA::AfpyrG; pabaA1; pyroA4; nkuA::argB; riboB2; fwA1*
LO1805/1806	*pyrG89; cdhA-GFP-AfpyrG; pyroA4; nkuA::argB; riboB2?; hhoA-mRFP-AfriboB*
LO2019	*pyrG89; cdhA::AfpyrG; pyroA4; nkuA::argB; riboB2; hhoA-mRFP-AfriboB*
LO2415	*pyrG89; cdhA::AfpyrG; pyroA4; nkuA::argB; riboB2*
LO2869	*pyrG89; nimE-GFP-AfpyrG; cdhA::AfpyrG; pabaA1; pyroA4; nkuA::argB; riboB2; hhoA-mRFP-AfriboB; fwA1*
LO3317	*pyrG89; nimE-GFP-AfpyrG; pabaA1; pyroA4; riboB2; hhoA-mRFP-AfriboB*
LO5442-5444	*pyrG89; nimE-GFP-AfpyrG; cdhA::AfpyrG; pabaA1; pyroA4?; nkuAΔ?; riboB2?; hhoA-mRFP-AfriboB; mipA*-D159*; fwA1*
LO6168	*pyrG89; cdhA-GFP-AfpyrG; pabaA1; pyroA4; nkuAΔ?; nimX-mCherry-AfriboB; fwA1*
LO6256	*pyrG89; cdhA-GFP-AfpyrG; nup49-mCherry-AfpyrG; pabaA1; pyroA4?; nirA14?; argB2?; riboB2?; nimX-mCherry-AfriboB; mipA*-D159*; chaA1?; fwA1*
R153	*pyroA4; wA3*

Question marks indicate alleles that were present in one of the parents of a cross but have not been tested in the strains shown.