

## Supplementary Figure Legends

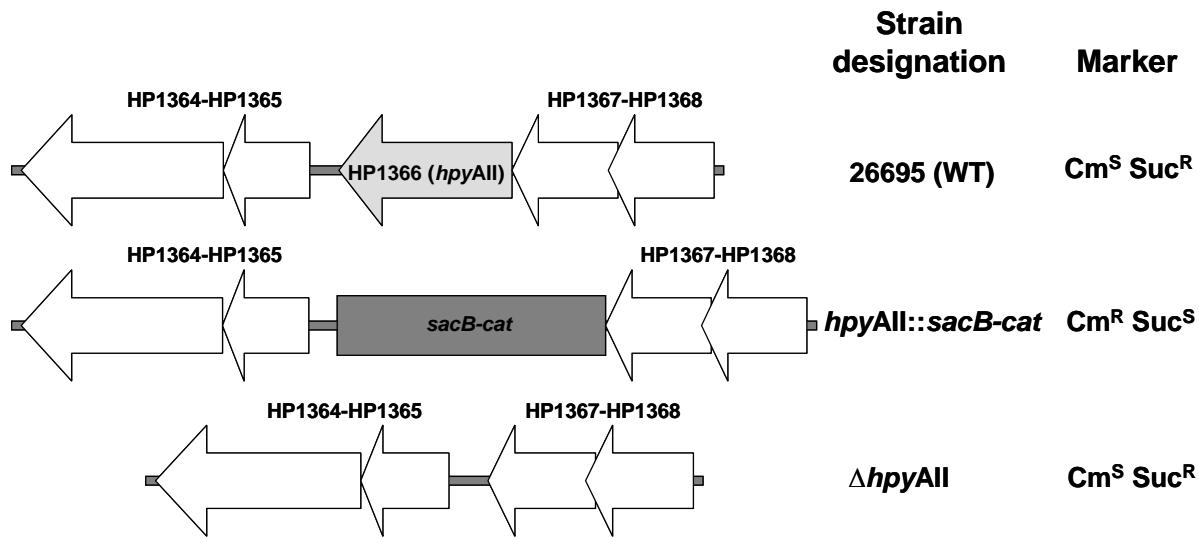
### **Supplementary Figure S1. *sacB*-mediated counter-selection used to construct *H. pylori* type II**

restriction endonuclease-deficient mutants. HP1366 (*hpyAII*), the target restriction endonuclease gene of the wild type 26695 strain, was first replaced by a *sacB-cat* cassette to generate a chloramphenicol-resistant (Cm<sup>R</sup>)/sucrose-sensitive (Suc<sup>S</sup>) mutant strain by transformation and following selection on Cm-containing media. The *sacB-cat* cassette then was deleted from the locus by transformation with a plasmid carrying a 2.1Kb PCR-product with HP1365 and HP1367 sequences and no intervening ORF, and following counter-selection with sucrose-containing media, resulting  $\Delta hpyAII$  (KO-1).

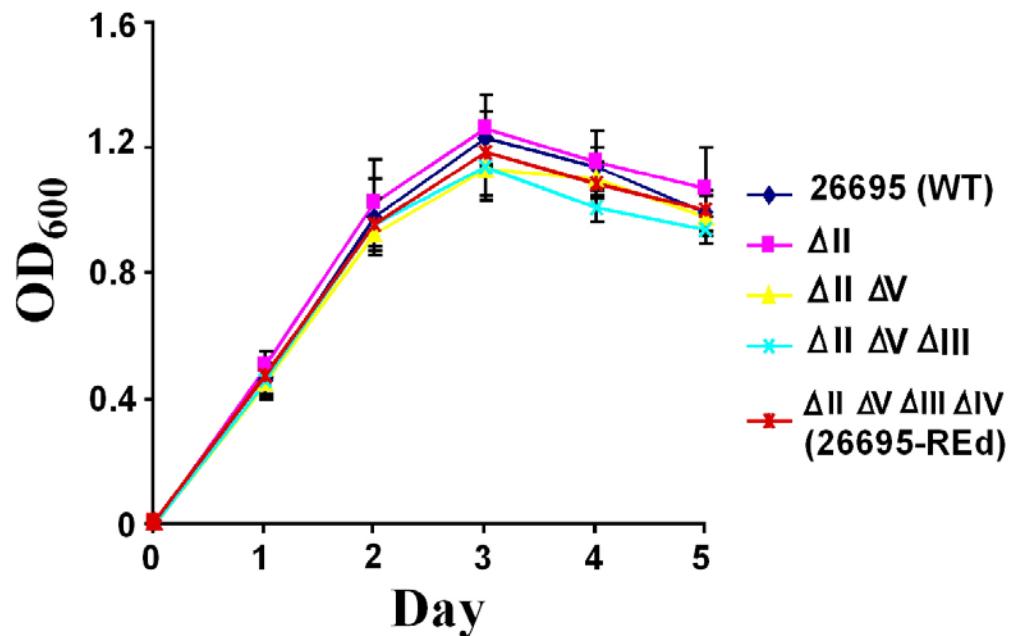
10    **Supplementary Figure S2. Growth of *H. pylori* wild type and the type II restriction endonuclease-deficient mutants.** The wild type strain 26695 and the series of the type II restriction endonuclease-deficient mutants [KO-1, KO-2, KO-3, and KO-4 (26695-REd)] exhibit similar growth during a 5-day incubation in Brucella broth (with 10% NBCS) medium. The experiments were performed independently in triplicate.

15    **Supplementary Figure S3. Survival of *H. pylori* wild type and the type II restriction endonuclease-deficient mutants after UV exposure.** *H. pylori* cells on TSA plates were subject to a range of UV exposures and surviving proportions were determined. The series of the type II restriction endonuclease-deficient mutants [KO-1, KO-2, KO-3, and KO-4 (26695-REd)] exhibit similar survival to the wild type strain 26695. The experiments were performed independently in triplicate.

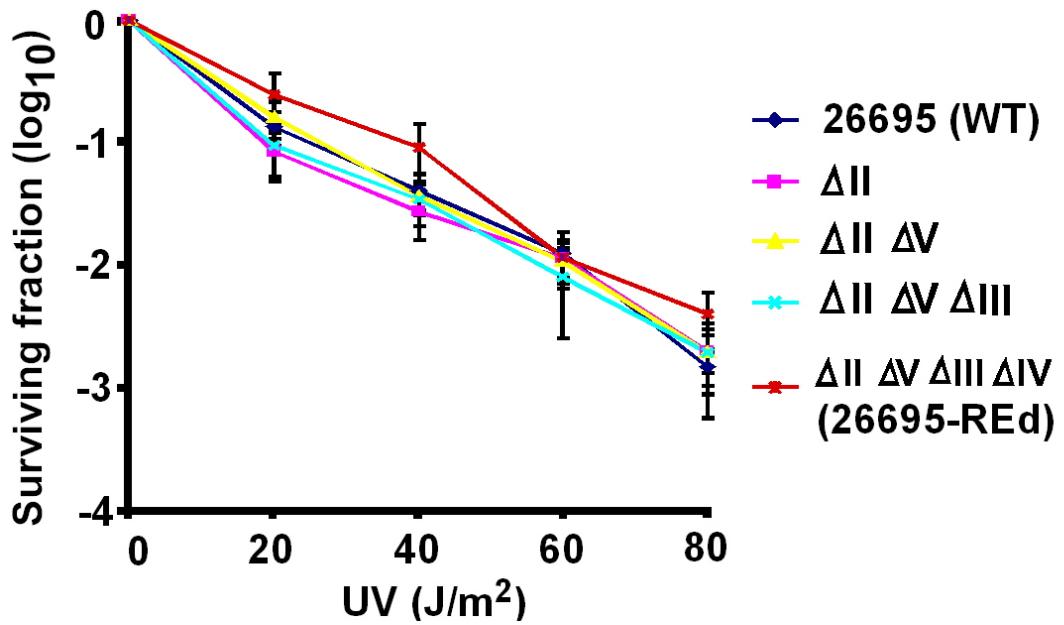
20    **Supplementary Figure S4. Spontaneous mutation frequency of *H. pylori* mutants by evaluating the frequency of an *rpoB* point mutation which confers resistance to rifampin.** The series of the type II restriction endonuclease-deficient mutants [KO-1, KO-2, KO-3, and KO-4 (26695-REd)] exhibit Rif<sup>R</sup> spontaneous mutation rates similar to the wild type strain 26695. The experiments were performed independently in triplicate.



**Supplementary Figure S1.**

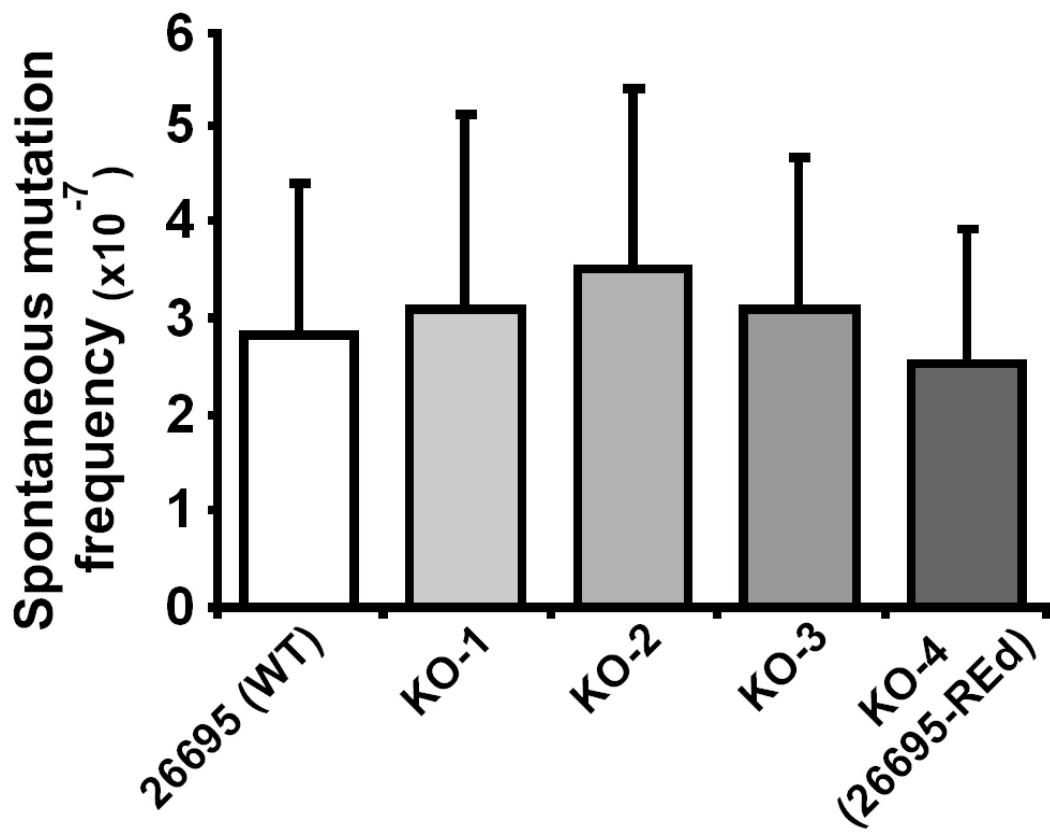


Supplementary Figure S2.



Supplementary Figure S3.

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Supplementary Figure S4.

**Supplementary Table S1: Plasmids and bacterial strains used in this study.**

Plasmid or strain	Relevant characteristics*	Source or reference
pGEM-T easy	Vector to construct gene mutations, Ap <sup>R</sup>	Promega
p801R	pGEM-T easy, 801 bp <i>H. pylori rpsL</i> fragment with A128G point mutation, Ap <sup>R</sup>	This work
pCTB8:Km	pGEM-T easy, <i>H. pylori vacA</i> <sub>60190</sub> :: <i>aphA</i> , Ap <sup>R</sup> , Km <sup>R</sup>	(11)
pAD1-Cat	pGEM-T easy, <i>H. pylori ureA</i> <sub>60190</sub> :: <i>cat</i> , Ap <sup>R</sup> , Cm <sup>R</sup>	(27)
pHel3	Shuttle plasmid, Km <sup>R</sup>	(20)
pKSF	Contains a <i>kan-sacB</i> cassette, Km <sup>R</sup>	(10)
pXZ016	pGEM-T easy, <i>hpyAV</i> upstream flanking region directly fused with <i>hpyAV</i> downstream flanking region, Ap <sup>R</sup>	This work
pXZ017	pGEM-T easy, <i>hpyAII</i> upstream flanking region directly fused with <i>hpyAII</i> downstream flanking region, Ap <sup>R</sup>	This work
pXZ032	pXZ016, <i>sacB-cat</i> cassette inserted between <i>hpyAV</i> upstream and downstream flanking region to construct $\Delta$ <i>hpyAV</i> :: <i>sacB-cat</i> mutation, Ap <sup>R</sup> , Cm <sup>R</sup> , Suc <sup>S</sup>	This work
pXZ033	pXZ017, <i>sacB-cat</i> cassette inserted between <i>hpyAII</i> upstream and downstream flanking region to construct $\Delta$ <i>hpyAII</i> :: <i>sacB-cat</i> mutation, Ap <sup>R</sup> , Cm <sup>R</sup> , Suc <sup>S</sup>	This work
pXZ144	pGEM-T easy, <i>hpyAIII</i> upstream flanking region directly fused with <i>hpyAIII</i> downstream flanking region, Ap <sup>R</sup>	This work
pXZ145	pGEM-T easy, <i>hpyAIV</i> upstream flanking region directly fused with <i>hpyAIV</i> downstream flanking region, Ap <sup>R</sup>	This work
pXZ146	pXZ144, <i>sacB-cat</i> cassette inserted between <i>hpyAIII</i> upstream and downstream flanking region to construct $\Delta$ <i>hpyAIII</i> :: <i>sacB-cat</i> mutation, Ap <sup>R</sup> , Cm <sup>R</sup> , Suc <sup>S</sup>	This work
pXZ147	pXZ145, <i>sacB-cat</i> cassette inserted between <i>hpyAIV</i> upstream and downstream flanking region to construct $\Delta$ <i>hpyAIV</i> :: <i>sacB-cat</i> mutation, Ap <sup>R</sup> , Cm <sup>R</sup> , Suc <sup>S</sup>	This work
<i>E. coli</i>		
XL1-blue	Host for cloning plasmid	Stratagene
<i>H. pylori</i>		
26695	UK-origin isolate (genomic sequence determined)	(55)
J99	USA-origin isolate (genomic sequence determined)	(1)
JP26	Japan-origin isolate (genomic sequence determined)	(3)
J166	USA-origin isolate	(61)
60190	American Type Culture Collection 49503, genome DNA sequence is used as template for pCTB8:Km and pAD1-Cat constructions	(11)
HPXZ285	KO-1, 26695 $\Delta$ <i>hpyAII</i>	This work
HPXZ347	26695 <i>vacA</i> :: <i>aphA</i> , 26695 transformed with pCTB8:Km, Km <sup>R</sup>	This work
HPXZ379	26695 <i>ureA</i> :: <i>cat</i> , 26695 transformed with pAD1-Cat, Cm <sup>R</sup>	This work
HPXZ490	KO-2, 26695 $\Delta$ <i>hpyAII</i> $\Delta$ <i>hpyAV</i>	This work
HPXZ560	KO-3, 26695 $\Delta$ <i>hpyAII</i> $\Delta$ <i>hpyAV</i> $\Delta$ <i>hpyAIII</i>	This work
HPXZ566	KO-4, 26695 $\Delta$ <i>hpyAII</i> $\Delta$ <i>hpyAV</i> $\Delta$ <i>hpyAIII</i> $\Delta$ <i>hpyAIV</i> (26695-REd)	This work
HPXZ621	KO-4 <i>ureA</i> :: <i>cat</i> , HPXZ566 transformed with pAD1-Cat, Cm <sup>R</sup>	This work
HPXZ623	JP26 <i>vacA</i> :: <i>aphA</i> , JP26 transformed with pCTB8:Km, Km <sup>R</sup>	This work
HPXZ624	JP26 <i>ureA</i> :: <i>cat</i> , JP26 transformed with pAD1-Cat, Cm <sup>R</sup>	This work
HPXZ626	J99 <i>vacA</i> :: <i>aphA</i> , J99 transformed with pCTB8:Km, Km <sup>R</sup>	This work
HPXZ627	J99 <i>ureA</i> :: <i>cat</i> , J99 transformed with pAD1-Cat, Cm <sup>R</sup>	This work
HPXZ629	J166 <i>vacA</i> :: <i>aphA</i> , J166 transformed with pCTB8:Km, Km <sup>R</sup>	This work
HPXZ630	J166 <i>ureA</i> :: <i>cat</i> , J166 transformed with pAD1-Cat, Cm <sup>R</sup>	This work
HPXZ641	KO-4 <i>ureA</i> :: <i>cat</i> <i>rpsL</i> (Str <sup>R</sup> ), HPXZ621 transformed with p801R, Cm <sup>R</sup> , Str <sup>R</sup>	This work
HPXZ648	JP26 <i>vacA</i> :: <i>aphA</i> <i>rpsL</i> (Str <sup>R</sup> ), HPXZ623 transformed with p801R, Km <sup>R</sup> , Str <sup>R</sup>	This work
HPXZ649	J99 <i>vacA</i> :: <i>aphA</i> <i>rpsL</i> (Str <sup>R</sup> ), HPXZ626 transformed with p801R, Km <sup>R</sup> , Str <sup>R</sup>	This work
HPXZ650	J166 <i>vacA</i> :: <i>aphA</i> <i>rpsL</i> (Str <sup>R</sup> ), HPXZ629 transformed with p801R, Km <sup>R</sup> , Str <sup>R</sup>	This work

HPXZ660	HPXZ626 strain with a spontaneous Rif <sup>R</sup> mutation, Km <sup>R</sup> , Rif <sup>R</sup>	This work
HPXZ696	26695 <i>vacA::aphA rpsL(Str<sup>R</sup>)</i> , HPXZ347 transformed with p801R, Km <sup>R</sup> , Str <sup>R</sup>	This work
HPXZ698	26695 <i>ureA::cat rpsL(Str<sup>R</sup>)</i> , HPXZ379 transformed with p801R, Cm <sup>R</sup> , Str <sup>R</sup>	This work

\*Ap<sup>R</sup>, Cm<sup>R</sup>, Km<sup>R</sup>, Str<sup>R</sup>, Rif<sup>R</sup>, and Suc<sup>S</sup> represent ampicillin-resistant, chloramphenicol-resistant, kanamycin-resistant, streptomycin-resistant, rifampin-resistant, and sucrose-sensitive, respectively.

**Supplementary Table S2. Oligonucleotides used in this study.**

Oligonucleotide	Primer sequence (5'→3')*
SC-F-XbaI	GCT <u>CTAGA</u> TATAAGCCCATTTCATGCTCC
SC-R-XbaI	GCT <u>CTAGA</u> CTCGAGGC <u>GTGAT</u> ATAGATTGAAAAGTG
S-R-PstI	CCCAAA <u>ACTGC</u> <u>CAGGT</u> TAGCCATTGCCTGC
C-R-PstI	CCC <u>CTGCAG</u> CACTACTCTGACAGAGAGTATA
VacA-F	<u>TGAAAGCGAAAAACAAG</u>
VacA-R	AAGAGAA <u>AGCTTTAACCC</u> TCC
Ure-F	<u>TCAAGTCCAGTCGTGGCCAC</u>
Ure-R	<u>GTGTCTGCTTGCCTATCAA</u>
IIL-F-SacII	TCCC <u>CGCGCG</u> CTCAATAGGTAA <u>TA</u> ACTCTC
IIL-R-SpeI	GGACTAG <u>TTGATAAAATAAAGCGGT</u> GTCTT
IIR-F-SpeI	GGACTAG <u>GTCAAAT</u> ATCCTTTTATT <u>CGC</u>
IIR-R-PstI	AA <u>ACTGCAGATGCCG</u> CTGAATTAGCAAGG
VL-F-SacII	TCCC <u>CGCGCG</u> GTGAA <u>AGGCGATAAAGA</u>
VL-R-SpeI	GG <u>ACTAGTTAAGTTTATTGAA</u> ACTGGCTAT
VR-F-SpeI	GG <u>ACTAGTGCTCTTCATAAGCTACTC</u> TT
VR-R-PstI	AA <u>ACTGCAGGAA</u> ATAGCGAAGTTATTGCCA
IIIL-F-SacII	TCCC <u>CGCGCT</u> CTCATAGAATGATTCCCCATTCC
IIIL-R-SpeI	GG <u>ACTAGTTTGCTCCG</u> CTTAATGTTTTCTTATT
IIIR-F-SpeI	GG <u>ACTAGTATGGTAATCGCG</u> ATTCTAATGAAAT
IIIR-R-PstI	AA <u>ACTGCAGCCTCTCAAAGATTAGCCG</u> CAATG
IVL-F-SacII	TCCC <u>CGCGG</u> ACAGAGGGGAGTTAATGATGTCTC
IVL-R-SpeI	GG <u>ACTAGTAGGCTCTAAAGTAAGCCC</u> ATTCT
IVR-F-SpeI	GG <u>ACTAGTGCCATTAAAGAGTC</u> CTTTGGCAGAT
IVR-R-PstI	AA <u>ACTGCAGGAAGTACCTCAATAACGACAAACA</u>
cataup	TGGATGAATTACAAGACT
catdown	TCAATCTTGTGAATTGC
ureAB-R-1	CGTGGTGGATTATGTGATTATCATTATGG
ureAB-R-2	CTAGAAATCCGCCATTGATCCGTTAAGCGGC
ureAB-R-3	CATTGAAATTACAGAGTTAAGGATCGTGC
ureAB-R-4	CATCATTGACAGCAACGGCTCACGCACGG
ureAB-R-5	CAAATCCGCATAACGGCAATACGCCTTAAAC
ureAB-R-6	TTAGTAATGGTCTTATTCAA <u>ACTGG</u> CTTG
ureAB-R-7	GTGCGGGTGGTGT <u>TTGG</u> TTCTAAATTA
ureAB-L-1	GCCAGGCTCAAACCTTACCGCTGTCCCCT
ureAB-L-2	GATAGTAGTCGCATTAGGCCATCAGCAGG
ureAB-L-3	GTTATGCC <u>TTTCTTCAAGCGGG</u> CC
ureAB-L-4	CAACATATAACAATACA <u>AGTC</u> CTAGCATTGC
ureAB-L-5	CCTCGTTCAAACCATTCCA <u>ATCCACATG</u>
ureAB-L-6	CCGTGTAAAGGA <u>ACTGG</u> CTAGAGAGATTGG
ureAB-L-7	GCCGTGTGCGGACAGCCTCTGTTCTACGC

\* Restriction sites underlined; *Xba*I (TCTAGA), *Sac*II (CCGCGG), *Spe*I (ACTAGT), *Pst*I (CTGCAG).

**Supplementary Table S3. Susceptibility of *H. pylori* wild type and mutant strains to levofloxacin<sup>a</sup>.**

Strain	Genotype	MIC Levofloxacin ( $\mu\text{g/mL}$ )
26695	Wild type	0.074 $\pm$ 0.017
HPXZ285	KO-1 ( $\Delta hpyAII$ )	0.079 $\pm$ 0.017
HPXZ490	KO-2 ( $\Delta hpyAII \Delta hpyAV$ )	0.072 $\pm$ 0.015
HPXZ560	KO-3 ( $\Delta hpyAII \Delta hpyAV \Delta hpyAIII$ )	0.072 $\pm$ 0.015
HPXZ566 <sup>b</sup>	KO-4 ( $\Delta hpyAII \Delta hpyAV \Delta hpyAIII \Delta hpyAIV$ )	0.079 $\pm$ 0.017

5           <sup>a</sup>Each experiment was repeated  $\geq 3$  times.

<sup>b</sup>Also named as 26695-REd.