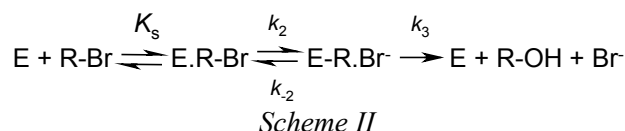


APPENDIX

A single mutation in a tunnel to the active site changes the mechanism and kinetics of product release in haloalkane dehalogenase LinB.

**Lada Biedermannová, Zbyněk Prokop, Artur Gora, Eva Chovancová, Mihály Kovács
Jiří Damborský and Rebecca C. Wade**

Multiple turnover analysis. Simple binding kinetics cannot be applied for the second product of 1,2-dibromoethane conversion, since 2-bromoethanol is a substrate and undergoes reaction with both LinB WT and LinB L177W. Multiple turnover analysis was therefore performed for the mixing of 2-bromoethanol with LinB WT and LinB L177W using the stopped-flow fluorescence technique (Figure 7). In this experiment, the observed quench of the fluorescence signal is not associated with the formation of an enzyme-substrate complex (E.R-Br) during the binding step, but it is connected with the second step, nucleophilic attack on the substrate molecule, followed by formation of the alkyl-enzyme intermediate (E-R) and cleavage of the bromide ion (Br⁻), which induces the fluorescence quench by interaction with the halide-stabilizing tryptophan (Scheme II). The transient kinetic data show that only the steady-state fluorescence is observed upon mixing 2-bromoethanol with LinB WT (Figure 7A), indicating that all the steps involved in the formation of the enzyme complex with 2-bromoethanol and the subsequent formation of the alkyl-enzyme intermediate are fast processes, occurring within the dead time of the instrument (0.5-5 ms).



In LinB L177W, the relaxation of the fluorescence signal was significantly slower, showing an exponential kinetic phase upon mixing with 2-bromoethanol (Figure 7B). The observed rate constants for each concentration (k_{obs}) were obtained by fitting the fluorescence traces to single exponentials. The solution for k_{obs} in terms of rate constants (Equation 8) was derived from Scheme II.

$$k_{\text{obs}} = \frac{[\text{S}] \cdot k_2}{K_s + [\text{S}]} + k_{-2} + k_3$$

Equation 8

The dependence of k_{obs} on 2-bromoethanol concentration showed saturation, thus $k_2 = 40 \pm 10 \text{ s}^{-1}$ and $K_s = 230 \pm 110 \text{ mM}$ could be determined separately (Figure 7C). The formation of the Michaelis complex appears to be fast process described by a rapid equilibrium with dissociation constant K_s . The dependence of the fluorescence quench amplitudes on 2-bromoethanol concentration indicates that the L177W mutation significantly changes the mechanism of the 2-bromoethanol reaction kinetics (Figure 7D). While a part of sigmoid dependence was observed for LinB WT, indicating cooperative behaviour, a simple hyperbolic dependence indicating classical Michaelis-Menten kinetics with an equilibrium constant $K_{\text{eq}} = 26 \pm 3 \text{ mM}$ was observed for LinB L177W.

The multiple turnover data indicate that both the binding and the release of 2-bromoethanol are fast

one-step processes reaching rapid equilibrium and that they cannot be rate determining for the conversion of 1,2-dibromoethane by either LinB WT or LinB L177W. The L177W mutation changed the reaction mechanism and the rate of the second kinetic step during the 2-bromoethanol conversion, but did not affect the kinetics of 2-bromoethanol release after the initial 1,2-dibromoethane dehalogenation.

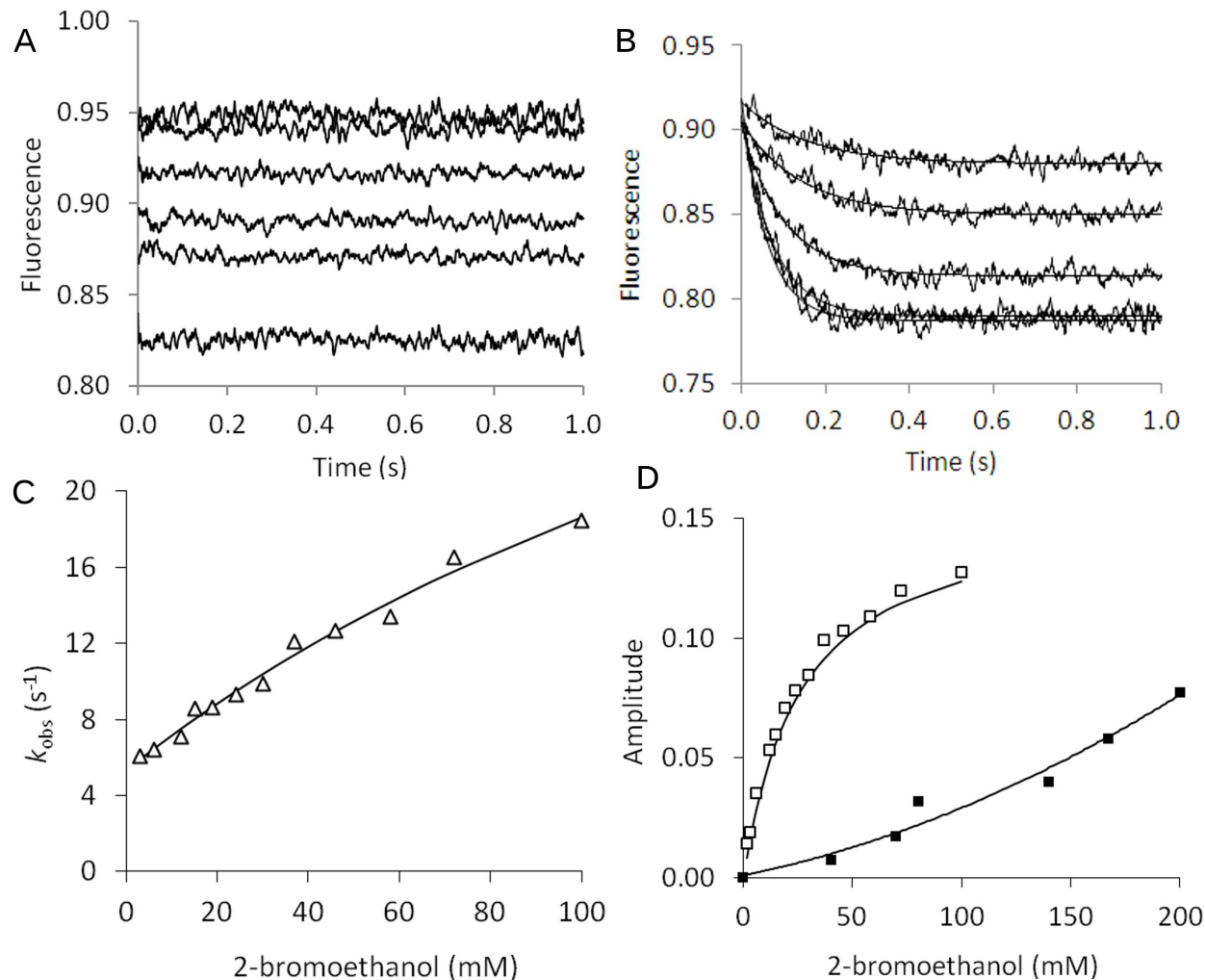


FIGURE 7. Stopped-flow analysis of the mixing of 2-bromoethanol with LinB WT and LinB L177W. Fluorescence traces obtained upon mixing 15 μM LinB WT with 2-bromoethanol to a final concentration of 0 - 200 mM (A). Fluorescence traces obtained upon mixing 30 μM LinB L177W with 2-bromoethanol to a final concentration of 0 - 100 mM (B), solid lines represent the best fit to the data by using a single exponential equation. The dependence of the observed rate constants on the concentration of 2-bromoethanol (C), the solid line represents the best fits obtained by using Equation 8. The dependence of the fluorescence quench amplitude on the concentration of 2-bromoethanol (D) in reaction with LinB WT (black squares) and LinB L177W (open squares). Solid lines represent the best fit by using: (i) the hyperbolic equation $A = A_{lim} \cdot [S] / K_{eq} + [S]$ for the LinB L177W data, in which A is the amplitude of fluorescence quench, A_{lim} is the limiting (maximum) amplitude, K_{eq} is the equilibrium constant and S is the concentration of 2-bromoethanol, and (ii) the Hill equation $A = A_{lim} \cdot [S]^n / K_{eq}^n + [S]^n$ for LinB WT in which n is the Hill coefficient. A unique solution for the individual parameters of the Hill equation cannot be obtained.