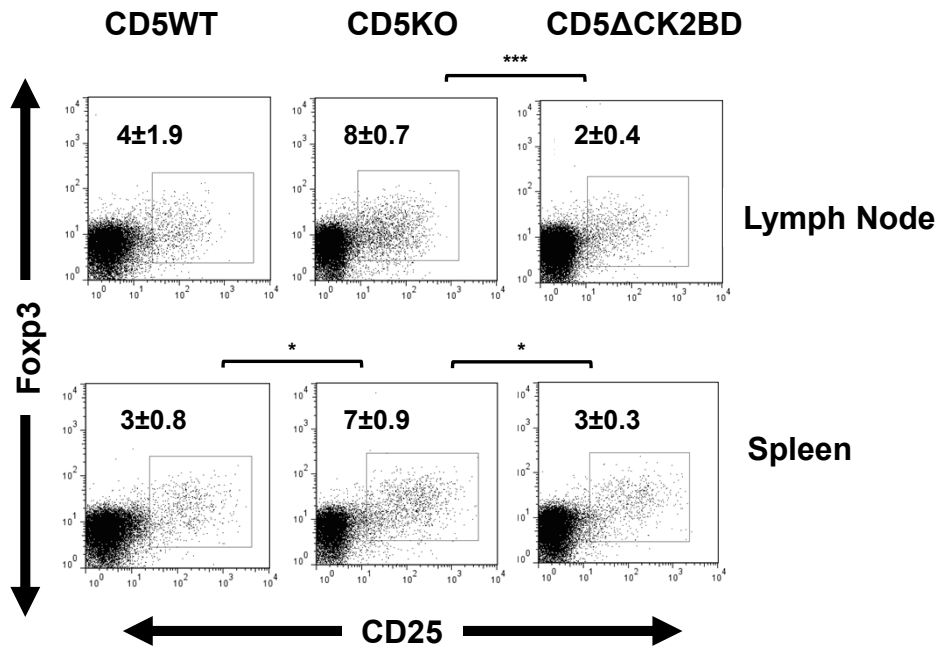
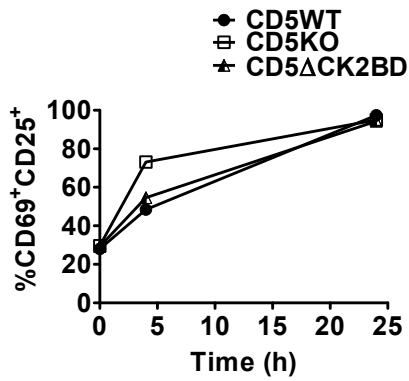
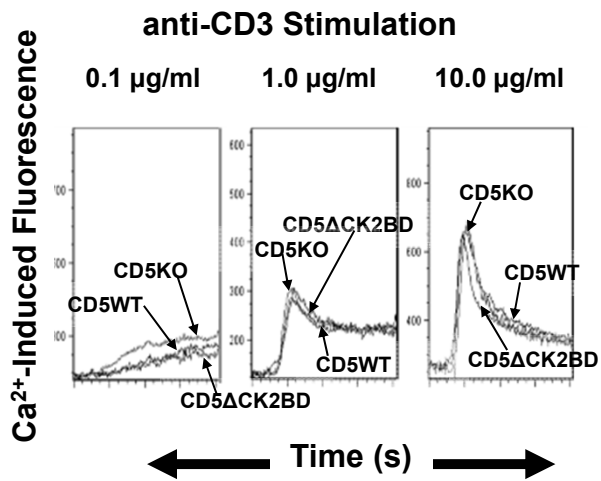


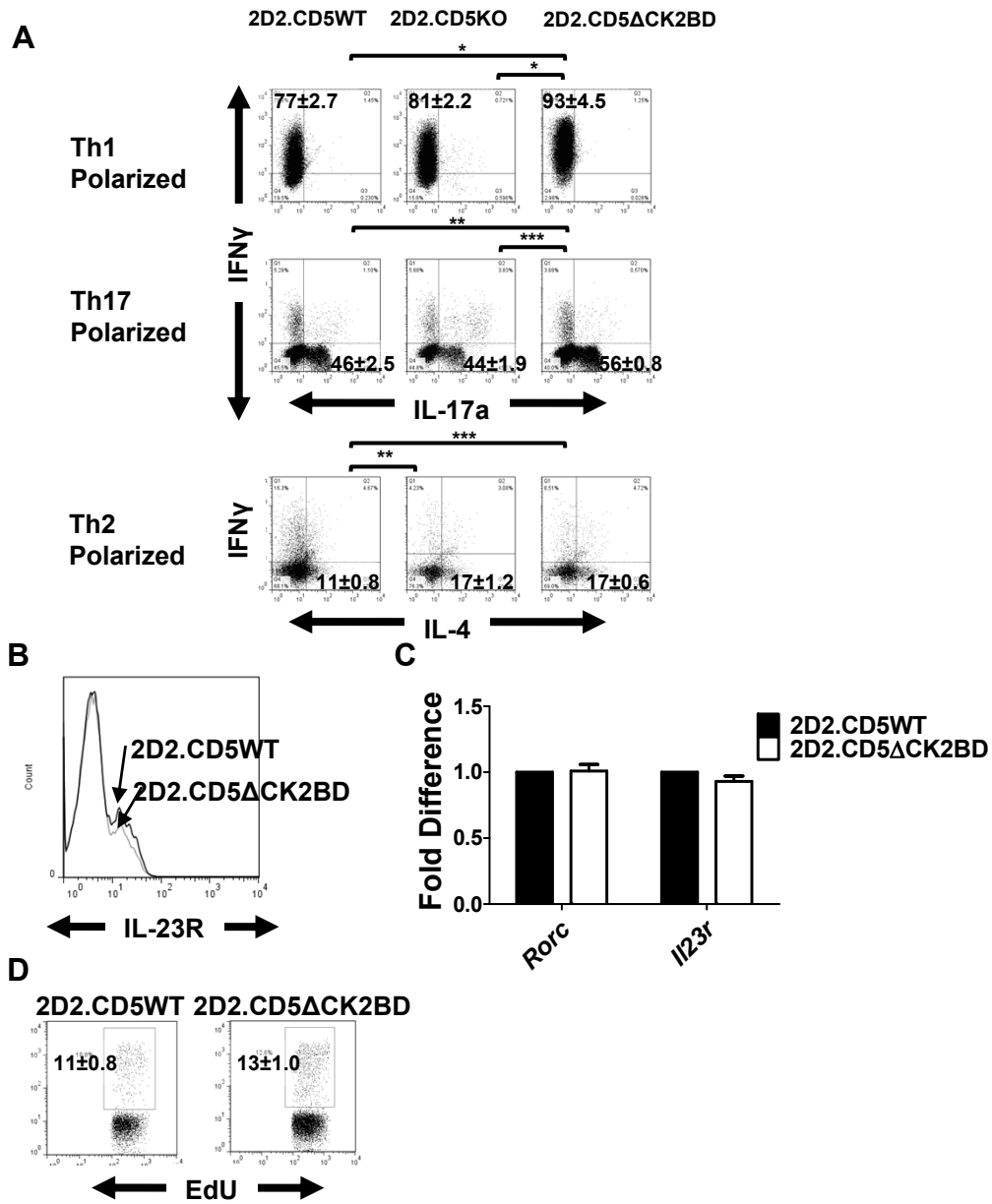
SUPPLEMENTAL FIGURE 1. *A*, Treg populations in lymph nodes and spleens of CD5WT, CD5KO or CD5 Δ CK2BD mice. Data \pm SEM represent the averages of 3-5 mice/group. * p <0.05, *** p <0.001 with Student t test. *B*, *C*, CD5-CK2 signaling does not affect parameters of early T-cell activation. *B*, Measurement of CD25 and CD69 upregulation on CD4⁺ T-cell development following 0-24h stimulation with anti-CD3. Data are from one mouse per group, $n=2$ experiments with 2-3 mice per experiment. *C*, Calcium mobilization in CD4⁺ T-cells following stimulation with anti-CD3. Data are from one representative mouse per group from a total of 2-3 mice/experiment/group, $n=2$ experiments.

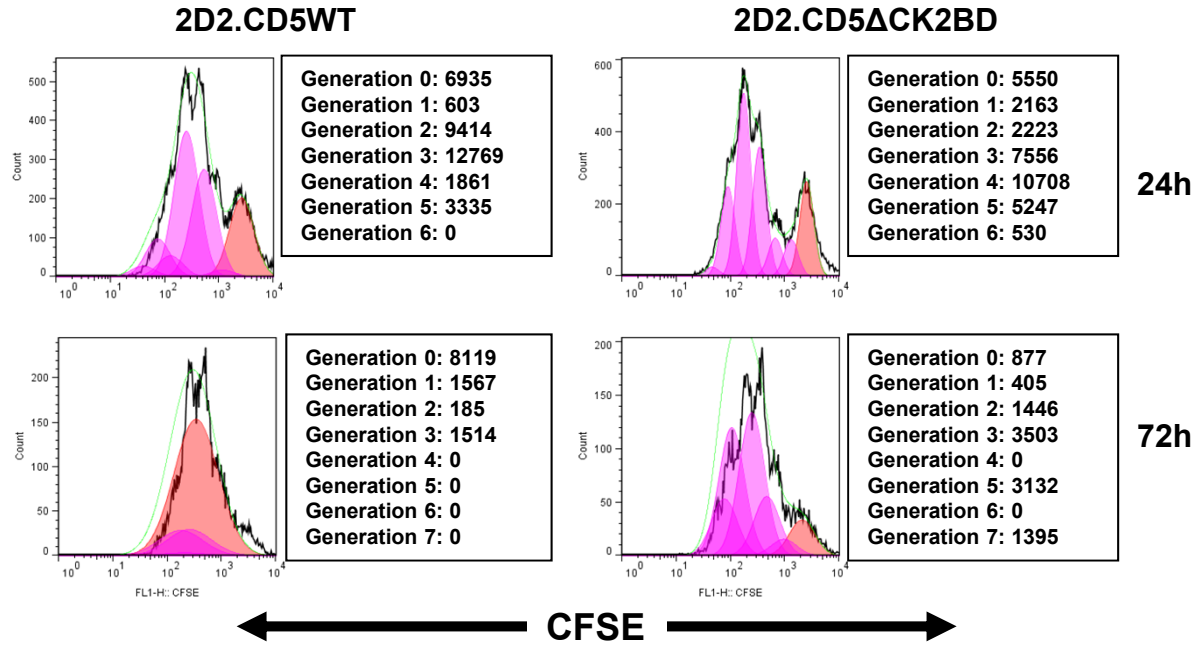
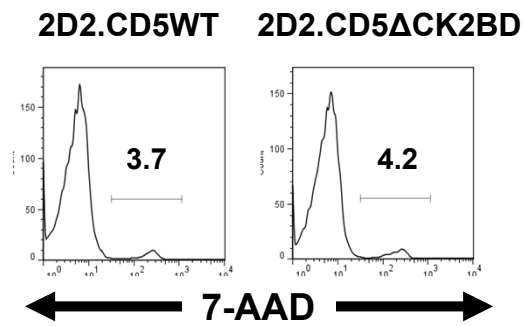
SUPPLEMENTAL FIGURE 2. *A*, Th1, Th17 and Th2 cell differentiation from naïve CD4⁺ T-cells obtained from 2D2.CD5WT, 2D2.CD5KO or 2D2.CD5 Δ CK2BD mice. Sorted CD4⁺ T cells were co-cultured with irradiated APCs and stimulated with anti-CD3 under Th1-, Th17-, or Th2- polarizing conditions for 5 days (*A*). Numbers in each quadrant represent the average frequency of CD4⁺ T-cells expressing IFN γ , IL-17a and/or IL-4 \pm SEM from four independent experiments performed with at least three mice per experiment. *B*, IL-23R expression on CD4⁺ T-cells on day 5 after stimulation with 10 μ g/ml MOG₃₅₋₅₅ peptide under Th17 polarizing conditions. *C*, *Rorc* and *Il23r* RNA expression in 2D2.CD5WT and 2D2.CD5 Δ CK2BD CD4⁺ T-cells measured on day 5 of stimulation with MOG₃₅₋₅₅ peptide under Th17 polarizing conditions. The fold difference was determined by normalizing to the expression of *Gapdh*. *D*, EdU incorporation determined on day 5 in Th17 polarizing cultures. Data are representative of three independent experiments performed with at least two mice per experiment. * p <0.05, ** p <0.01, *** p <0.001 with Student t test.

SUPPLEMENTAL FIGURE 3. T-cell proliferation and cell death following restimulation in co-cultures of previously stimulated CD4⁺ T-cells from 2D2.CD5WT and 2D2.ΔCK2BD mice. *A*, 2D2.CD5WT CD4⁺ T-cells and 2D2.CD4ΔCK2 CD4⁺ T-cells were stimulated for 24h with MOG₃₅₋₅₅ peptide and rested for 3d separately. On day 3, dead cells were removed by Ficoll-Paque density gradient centrifugation and co-cultured in the presence of 10 μg/ml MOG₃₅₋₅₅ after labeling one of the populations with CFSE. The dilution of CFSE was determined by flow cytometry at the indicated times and analyzed with FlowJo software *B*, 7-AAD incorporation in cells from above following 24h restimulation. *C*. 7-AAD incorporation in T-cells primed in vivo and restimulated in vitro with MOG₃₅₋₅₅ peptide. Data are from one representative mouse per group from a total of 2-3 mice/experiment/group, *n*= 2 experiments.

SUPPLEMENTAL FIGURE 4. The CD5-CK2 signaling pathway regulates cognate peptide or anti-CD3 mAb stimulated cytokine production from CD4⁺ T-cells. *A-E*, Naïve 2D2.CD5WT, 2D2.CD5KO, and 2D2.CD5ΔCK2BD CD4⁺ T-cells were co-cultured with irradiated APCs and stimulated with 10 μg/ml MOG₃₅₋₅₅ peptide (vs. 100 μg/ml MOG₃₅₋₅₅ in Figure 6) for 1-5 d. *F-J*, Naïve CD5WT, CD5KO, and CD5ΔCK2BD CD4⁺ T-cells were co-cultured with irradiated APCs and stimulated with 1 μg/ml anti-CD3 for 1-5 d. Supernates were collected on days 1, 2, 3, and 5 to quantify secreted levels of IFNγ (*A* and *F*), IL-17a (*B* and *G*), IL-6 (*C* and *H*), IL-2 (*D* and *I*), and IL-10 (*E* and *J*) by ELISA. Data ± SEM represent values 2-3 independent mice/group/ experiment, *n*=4 experiments.

A**B****C**



A**B****C**