

Non-Enzymatic Dynamic Kinetic Resolution of Secondary Alcohols via Enantioselective Acylation: Synthetic and Mechanistic Studies

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Supporting Information

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I. General Information

The following reagents were purchased from Aldrich and used as received: KO^t-Bu, *t*-amyl alcohol (anhydrous), toluene (anhydrous), and NEt₃ (anhydrous). (C₃Ph₃)-DMAP*¹ and Ru^{Cl₂} were synthesized as previously described. 1-Phenylethanol, 1-phenyl-1-propanol, 2-methyl-1-phenyl-1-propanol, 1-(2-methylphenyl)ethanol, and Ac₂O were purchased (Aldrich or Alfa Aesar) and purified by vacuum distillation prior to use. 1-(1-Naphthyl)ethanol was purchased from Aldrich and purified by column chromatography prior to use. The other secondary alcohols have been reported previously and were synthesized either by the addition of a Grignard reagent to an aldehyde or by the reduction of a ketone (purification: column chromatography).

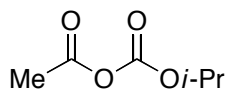
Unless otherwise specified, reactions were conducted with stirring in oven-dried glassware under an atmosphere of nitrogen.

HPLC analyses were carried out on an Agilent 1100 series system equipped with a Daicel CHIRALCEL OD column (internal diameter 4.6 mm, column length 250 mm, particle size 5 μ). GC analyses were obtained on an Agilent 6850 system equipped with a Varian CP-Chirasil-DEX CB column (internal diameter 0.25 mm, column length 25 m).

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- (1) Wurz, R. P.; Lee, E. C.; Ruble, J. C.; Fu, G. C. *Adv. Synth. Catal.* **2007**, *349*, 2345–2352.
(2) Martín-Matute, B.; Edin, M.; Bogár, K.; Kaynak, F. B.; Bäckvall, J.-E. *J. Am. Chem. Soc.* **2005**, *127*, 8817–8825.

II. Preparation of Acyl Carbonates

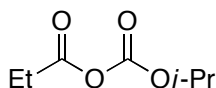
The procedures and yields have not been optimized.



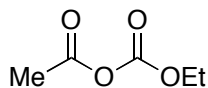
Acetyl isopropyl carbonate [60059-18-9]. Acetic acid (1.7 mL, 30 mmol) was added via syringe to a 250-mL round-bottom flask that contained anhydrous Et₂O (60 mL) and NEt₃ (4.2 mL, 30 mmol) at 0 °C. The reaction mixture was stirred for 5 min, and then isopropyl chloroformate (30 mL, 30 mmol; 1.0 M in toluene; Aldrich) was added dropwise via syringe over 10 min. The reaction mixture was stirred at 0 °C for 45 min. Next, an aqueous 10% citric acid solution (30 mL) was added. The organic layer was decanted and then washed with additional citric acid solution (20 mL), a saturated NaHCO₃ solution (30 mL), and brine (30 mL). The organic solvent was evaporated under reduced pressure, and the acyl carbonate was purified by vacuum distillation (30 °C at 500 mtorr), which provided a clear, colorless liquid (2.2 g, 50% yield).

¹H NMR (400 MHz, CDCl₃) δ 4.97 (septet, 1H, *J* = 6.4 Hz), 2.19 (s, 3H), 1.33 (d, 6H, *J* = 6.4 Hz).

¹³C NMR (100 MHz, CDCl₃) δ 165.4, 148.5, 74.5, 21.6, 21.2.



Propionyl isopropyl carbonate [176438-88-3]. Propionic acid (1.2 mL, 15 mmol) was added via syringe to a 250-mL round-bottom flask that contained anhydrous Et₂O (30 mL) and NEt₃ (2.1 mL, 15 mmol) at 0 °C. The reaction mixture was stirred for 5 min, and then isopropyl chloroformate (15 mL, 15 mmol; 1.0 M in toluene; Aldrich) was added dropwise via syringe over 10 min. The reaction mixture was stirred at 0 °C for 45 min. Next, an aqueous 10% citric acid solution (15 mL) was added. The organic layer was decanted and then washed with additional citric acid solution (10 mL), a saturated NaHCO₃ solution (15 mL), and brine (15 mL). The organic solvent was evaporated under reduced pressure, and the acyl carbonate was purified by vacuum distillation (27 °C at 450 mtorr), which provided a clear, colorless liquid (1.1 g, 46% yield).



Acetyl ethyl carbonate [15890-77-4]. Acetic acid (0.85 mL, 15 mmol) was added via syringe to a 250-mL round-bottom flask that contained anhydrous Et₂O (30 mL) and NEt₃ (2.1 mL, 15 mmol) at 0 °C. The reaction mixture was stirred for 5 min, and then ethyl chloroformate (1.4 mL, 15 mmol) was added dropwise via syringe over 10 min. The reaction mixture was stirred at 0 °C for 45 min. Next, an aqueous 10% citric acid solution (15 mL) was added. The organic layer was decanted and then washed with additional citric acid solution (10 mL), a saturated NaHCO₃ solution (15 mL), and brine (15 mL). The organic solvent was evaporated under reduced pressure, which provided a clear, colorless liquid (1.3 g, 66% yield).

¹H NMR (400 MHz, CDCl₃) δ 4.30 (quartet, 2H, *J* = 6.9 Hz), 2.19 (s, 3H), 1.34 (t, 3H, *J* = 6.9 Hz).

^{13}C NMR (100 MHz, CDCl_3) δ 165.1, 148.9, 65.6, 21.0, 13.9.

III. Dynamic Kinetic Resolutions of Secondary Alcohols

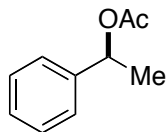
General procedure. In a nitrogen-filled glovebox (for a glovebox-free procedure, see below), Ru^{Cl} (16.6 mg, 0.026 mmol), $\text{KO}t\text{-Bu}$ (2.5 mg, 0.022 mmol), toluene (100 μL), and *t*-amyl alcohol (150 μL) were combined in an oven-dried 4-mL vial equipped with a stir bar. The resulting mixture was stirred at r.t. for 6 min, and then the alcohol (0.50 mmol) and (+)- $(\text{C}_5\text{Ph}_5)\text{-DMAP}^*$ (3.3 mg in 50 μL toluene, 0.0050 mmol) were added to the vial. The vial was then capped with a septum-lined cap, removed from the glovebox, and cooled to 10 $^\circ\text{C}$. After cooling the solution for 10 min, acetyl isopropyl carbonate (108 μL , 0.750 mmol) was added dropwise to the vial via syringe pump over 20 h (Notes: For optimal results, the tip of the needle should be aligned such that the acetyl isopropyl carbonate drips down the wall of the vial. Some grease was applied to the septum around the needle, to discourage moisture/air from entering the vial during the addition.). The reaction mixture was stirred at 10 $^\circ\text{C}$ for an additional 28 h. To remove the (+)- $(\text{C}_5\text{Ph}_5)\text{-DMAP}^*$, the mixture was then filtered through a plug of silica gel using Et_2O (10 mL) as the eluant, and the volatiles were removed under reduced pressure with minimal heat (to minimize evaporation of the products, since some are volatile). To remove a trace of a colored impurity, the residue was dissolved in CH_2Cl_2 (20 mL), H_2O (20 mL), and a 70% aqueous solution of *t*-BuOOH (2 mL). This mixture was stirred at r.t. for 2 h, and the product was isolated by extraction into CH_2Cl_2 (3 x 50 mL). The CH_2Cl_2 layers were combined and dried over MgSO_4 , and the solvent was removed under reduced pressure. The product was purified by column chromatography (hexanes \rightarrow 20% Et_2O in hexanes).

Notes: For the sake of convenience, the DKRs were set up in a glovebox. However, this method does *not* require the use of a glovebox (see the next procedure).

For the alcohol illustrated in entry 1 of Table 1, the DKR proceeded in 81% ee and 88% yield when the acetyl isopropyl carbonate was added dropwise over 1 min, rather than by syringe pump over 20 h.

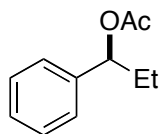
Glovebox-free procedure. Ru^{Cl} (16.6 mg, 0.026 mmol) and $\text{KO}t\text{-Bu}$ (2.5 mg, 0.022 mmol) were added to an oven-dried 4-mL vial. The vial was capped, and then it was evacuated and backfilled with nitrogen (three cycles). Toluene (100 μL) and *t*-amyl alcohol (150 μL) were added via syringe, and the resulting solution was stirred at r.t. for 6 min. (+)- $(\text{C}_5\text{Ph}_5)\text{-DMAP}^*$ (3.3 mg, 0.0050 mmol) was added to another oven-dried 4-mL vial, and this vial was capped, and then it was evacuated and backfilled with nitrogen (three cycles); next, toluene (50 μL) was added. The alcohol (0.50 mmol) and the solution of (+)- $(\text{C}_5\text{Ph}_5)\text{-DMAP}^*$ were added in turn by syringe to the vial containing Ru^{Cl} . The resulting mixture was cooled to 10 $^\circ\text{C}$. The remainder of the procedure (addition of acetyl isopropyl carbonate onward) follows the general procedure.

This procedure provided 86% ee and 95% yield for the DKR illustrated in entry 1 of Table 1, and it furnished 90% ee and 97% yield for the DKR depicted in entry 2 of Table 1.



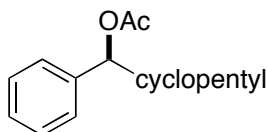
(S)-1-Phenylethyl acetate (Table 1, entry 1) [16197-93-6]. The title compound was prepared according to the general procedure, using 1-phenylethanol (61 mg, 0.50 mmol). After purification by column chromatography (hexanes → 20% Et₂O in hexanes), the title compound was isolated as a clear, colorless oil (70 mg, 85% yield; 95% calibrated GC yield) with 86% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 105 °C → 115 °C @ 1 °C/min, followed by 115 → 175 °C @ 5 °C/min; He flow rate: 1.0 ml/min; retention times: 6.5 min (major), 7.2 min (minor)).

The second run was performed with (–)-(C₃Ph₃)-DMAP*. The product was isolated as a clear, colorless oil (69 mg, 84% yield; 95% calibrated GC yield) with 87% ee.



(S)-1-Phenylpropyl acetate (Table 1, entry 2) [83860-48-4]. The title compound was prepared according to the general procedure, using 1-phenylpropanol (68 mg, 0.50 mmol). After purification by column chromatography (hexanes → 20% Et₂O in hexanes), the title compound was isolated as a clear, colorless oil (82 mg, 92% yield; 95% calibrated GC yield) with 90% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 105 °C → 115 °C @ 0.5 °C/min, followed by 115 → 175 °C @ 5 °C/min; He flow rate: 1.0 ml/min; retention times: 9.3 min (major), 9.9 min (minor)).

The second run was performed with (–)-(C₃Ph₃)-DMAP*. The product was isolated as a clear, colorless oil (86 mg, 97% yield; 97% calibrated GC yield) with 90% ee.



(S)-Cyclopentyl(phenyl) acetate (Table 1, entry 3). The title compound was prepared according to the general procedure, using cyclopentyl(phenyl)methanol (88 mg, 0.50 mmol). After purification by column chromatography (hexanes → 20% Et₂O in hexanes), the title compound was isolated as a clear, colorless oil (94 mg, 86% yield; 91% calibrated GC yield) with 82% ee (HPLC analysis of the product: Daicel CHIRALCEL OD-H column; solvent system: hexanes; 1.0 mL/min; retention times: 20.2 min (minor), 41.2 min (major)).

The second run was performed with (–)-(C₃Ph₃)-DMAP*. The product was isolated as a clear, colorless oil (94 mg, 86% yield; 88% calibrated GC yield) with 81% ee.

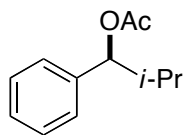
¹H NMR (400 MHz, CDCl₃) δ 7.33–7.29 (m, 4H), 7.29–7.25 (m, 1H), 5.52 (d, 1H, *J* = 9.2 Hz), 2.34 (sextet, 1H, *J* = 8.4 Hz), 2.03 (s, 3H), 1.84–1.77 (m, 1H), 1.67–1.52 (m, 3H), 1.50–1.44 (m, 1H), 1.42–1.36 (m, 2H), 1.18–1.09 (m, 1H).

¹³C NMR (100 MHz, CDCl₃) δ 170.6, 140.8, 128.4, 127.9, 127.2, 80.1, 45.6, 29.8, 29.3, 25.37, 25.35, 21.5.

IR (film) 3033, 2956, 2870, 1739, 1496, 1454, 1371, 1237, 1022, 965, 903, 761, 700 cm⁻¹.

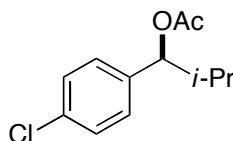
MS (EI) *m/z* (M⁺) calcd for C₁₄H₁₈O₂: 218, found: 218.

$[\alpha]_D^{24} = -43^\circ$ ($c = 1.0$, CH_2Cl_2 ; obtained with (+)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$).



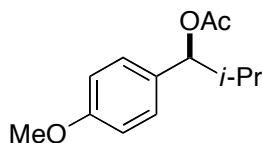
(S)-2-Methyl-1-phenylpropyl acetate (Table 1, entry 4) [84194-67-2]. The title compound was prepared according to the general procedure, using 2-methyl-1-phenylpropanol (75 mg, 0.50 mmol). After purification by column chromatography (hexanes \rightarrow 20% Et_2O in hexanes), the title compound was isolated as a clear, colorless oil (91 mg, 95% yield; 98% calibrated GC yield) with 90% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 105 $^\circ\text{C}$ \rightarrow 115 $^\circ\text{C}$ @ 0.5 $^\circ\text{C}/\text{min}$, followed by 115 \rightarrow 175 $^\circ\text{C}$ @ 5 $^\circ\text{C}/\text{min}$; He flow rate: 1.0 ml/min; retention times: 10.7 min (major), 11.3 min (minor)).

The second run was performed with (-)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$. The product was isolated as a clear, colorless oil (90 mg, 94% yield; 97% calibrated GC yield) with 91% ee.



(S)-2-Methyl-1-(4-chlorophenyl)propyl acetate (Table 1, entry 5) [137408-30-1]. The title compound was prepared according to the general procedure, using 2-methyl-1-(4-chlorophenyl)propanol (92 mg, 0.50 mmol). After purification by column chromatography (hexanes \rightarrow 20% Et_2O in hexanes), the title compound was isolated as a clear, colorless oil (100 mg, 88% yield; 87% calibrated GC yield) with 85% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 105 $^\circ\text{C}$ \rightarrow 115 $^\circ\text{C}$ @ 1 $^\circ\text{C}/\text{min}$, followed by 115 \rightarrow 175 $^\circ\text{C}$ @ 5 $^\circ\text{C}/\text{min}$; He flow rate: 1.0 ml/min; retention times: 17.2 min (major), 17.5 min (minor)).

The second run was performed with (-)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$. The product was isolated as a clear, colorless oil (100 mg, 88% yield; 90% calibrated GC yield) with 85% ee.



(S)-2-Methyl-1-(4-methoxyphenyl)propyl acetate (Table 1, entry 6). The title compound was prepared according to the general procedure, using 2-methyl-1-(4-methoxyphenyl)propanol (90 mg, 0.50 mmol). After purification by column chromatography (hexanes \rightarrow 20% Et_2O in hexanes), the title compound was isolated as a white solid (103 mg, 93% yield; 95% calibrated GC yield) with 86% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 105 $^\circ\text{C}$ \rightarrow 115 $^\circ\text{C}$ @ 0.5 $^\circ\text{C}/\text{min}$, followed by 115 \rightarrow 175 $^\circ\text{C}$ @ 5 $^\circ\text{C}/\text{min}$; He flow rate: 1.0 ml/min; retention times: 27.5 min (major), 27.8 min (minor)).

The second run was performed with (-)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$. The product was isolated as a white solid (100 mg, 90% yield; 90% calibrated GC yield) with 89% ee.

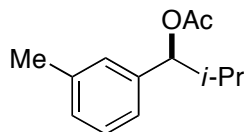
^1H NMR (400 MHz, CDCl_3) δ 7.22 (d, 2H, $J = 8.8$ Hz), 6.85 (d, 2H, $J = 8.4$ Hz), 5.39 (d, 1H, $J = 8.0$ Hz), 3.79 (s, 3H), 2.09–2.02 (m, 4H), 0.96 (d, 3H, $J = 6.8$ Hz), 0.76 (d, 3H, $J = 6.8$ Hz).

^{13}C NMR (100 MHz, CDCl_3) δ 170.5, 159.2, 132.0, 128.5, 113.7, 80.9, 55.3, 33.5, 21.4, 19.4, 18.8.

IR (film) 2965, 2873, 1724, 1676, 1606, 1515, 1474, 1456, 1377, 1298, 1244, 1177, 1104, 1032, 1016, 975, 903, 832, 812 cm^{-1} .

MS (EI) m/z (M^+) calcd for $\text{C}_{13}\text{H}_{18}\text{O}_3$: 222, found: 222.

$[\alpha]_D^{24} = -43^\circ$ ($c = 1.0$, CH_2Cl_2 ; obtained with (+)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$).



(S)-2-Methyl-1-(3-Methylphenyl)propyl acetate (Table 1, entry 7). The title compound was prepared according to the general procedure, using 2-methyl-1-(3-methyl)phenylpropanol (82 mg, 0.50 mmol). After purification by column chromatography (hexanes \rightarrow 20% Et_2O in hexanes), the title compound was isolated as a clear, colorless oil (99 mg, 96% yield; 96% calibrated GC yield) with 90% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 105 $^\circ\text{C}$ \rightarrow 115 $^\circ\text{C}$ @ 0.5 $^\circ\text{C}/\text{min}$, followed by 115 \rightarrow 175 $^\circ\text{C}$ @ 5 $^\circ\text{C}/\text{min}$; He flow rate: 1.0 ml/min; retention times: 14.9 min (major), 15.8 min (minor)).

The second run was performed with (–)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$. The product was isolated as a clear, colorless oil (99 mg, 96% yield; 97% calibrated GC yield) with 91% ee.

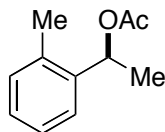
^1H NMR (400 MHz, CDCl_3) δ 7.24–7.20 (m, 1H), 7.10–7.08 (m, 3H), 5.43 (d, 1H, $J = 7.6$ Hz), 2.35 (s, 3H), 2.13–2.05 (m, 4H), 0.97 (d, 3H, $J = 6.8$ Hz), 0.80 (d, 3H, $J = 6.8$ Hz).

^{13}C NMR (100 MHz, CDCl_3) δ 170.5, 139.8, 137.8, 128.5, 128.1, 127.9, 124.2, 81.1, 33.6, 21.6, 21.3, 18.9, 18.6.

IR (film) 2964, 2875, 1736, 1686, 1610, 1450, 1371, 1237, 1160, 1023, 980, 908, 785, 705 cm^{-1} .

MS (EI) m/z (M^+) calcd for $\text{C}_{13}\text{H}_{18}\text{O}_2$: 206, found: 206.

$[\alpha]_D^{24} = -47^\circ$ ($c = 1.0$, CH_2Cl_2 ; obtained with (+)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$).



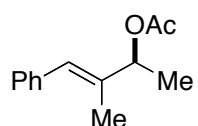
(S)-1-(2-Methylphenyl)ethyl acetate (Table 1, entry 8) [501659-37-6]. The title compound was prepared according to the general procedure, using 1-(2-methyl)phenylethanol (68 mg, 0.50 mmol). After purification by column chromatography (hexanes \rightarrow 20% Et_2O in hexanes), the title compound was isolated as a clear, colorless oil (86 mg, 96% yield; 99% calibrated GC yield) with 93% ee (GC analysis of the product: CP-Chirasil-DEX CB; heating program: 90 $^\circ\text{C}$ \rightarrow 115 $^\circ\text{C}$ @ 0.5 $^\circ\text{C}/\text{min}$, followed by 115 \rightarrow 175 $^\circ\text{C}$ @ 5 $^\circ\text{C}/\text{min}$; He flow rate: 0.7 ml/min; retention times: 20.7 min (major), 21.5 min (minor)).

The second run was performed with (–)- $(\text{C}_3\text{Ph}_3)\text{-DMAP}^*$. The product was isolated as a clear, colorless oil (78 mg, 87% yield; 94% calibrated GC yield) with 92% ee.



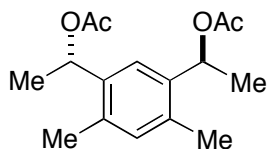
(S)-1-(1-naphthyl)ethyl acetate (Table 1, entry 9) [16197-95-8]. The title compound was prepared according to the general procedure, using 1-(1-naphthyl)ethanol (86 mg, 0.50 mmol). After purification by column chromatography (hexanes → 20% Et₂O in hexanes), the title compound was isolated as a clear, colorless oil (103 mg, 96% yield; 99% calibrated GC yield) with 89% ee (HPLC analysis of the product: Daicel CHIRALCEL OD-H column; solvent system: 1.0% *i*-PrOH in hexanes; 1.0 mL/min; retention times: 9.4 min (minor), 14.4 min (major)).

The second run was performed with (–)-(C₅Ph₅)-DMAP*. The product was isolated as a clear, colorless oil (98 mg, 91% yield; 99% calibrated GC yield) with 91% ee.



(S,E)-3-Methyl-4-phenylbut-3-en-2-yl acetate (Table 1, entry 10) [187736-05-6]. The title compound was prepared according to the general procedure, using (*E*)-3-methyl-4-phenylbut-3-en-2-ol (81 mg, 0.50 mmol). After purification by column chromatography (hexanes → 20% Et₂O in hexanes), the title compound was isolated as a clear, colorless oil (98 mg, 96% yield; 95% calibrated GC yield) with 89% ee (HPLC analysis of the product: Daicel CHIRALCEL OD-H column; solvent system: 1.0% *i*-PrOH in hexanes; 1.0 mL/min; retention times: 5.9 min (major), 7.5 min (minor)).

The second run was performed with (–)-(C₅Ph₅)-DMAP*. The product was isolated as a clear, colorless oil (85 mg, 83% yield; 99% calibrated GC yield) with 88% ee.



(1S,1'S)-1,1'-(4,6-Dimethyl-1,3-phenylene)bis(ethane-1,1-diyl) diacetate (eq 10) [205104-01-4 for the (1R,1'R) enantiomer]. In a nitrogen-filled glovebox, Ru^{Cl} (33.2 mg, 0.052 mmol), KO*t*-Bu (4.9 mg, 0.044 mmol), toluene (200 μL), and *t*-amyl alcohol (300 μL) were combined in an oven-dried 4-mL vial equipped with a stir bar. The resulting mixture was stirred at r.t. for 6 min, and then the alcohol (0.50 mmol) and (+)-(C₅Ph₅)-DMAP* (6.6 mg in 100 μL toluene, 0.010 mmol) were added to the vial. The vial was then capped with a septum-lined cap, removed from the glovebox, and cooled to 10 °C. After cooling the solution for 10 min, acetyl isopropyl carbonate (216 μL, 1.50 mmol) was added dropwise to the vial via syringe pump over 20 h (Notes: For optimal results, the tip of the needle should be aligned such that the acetyl isopropyl carbonate drips down the wall of the vial. Some grease was applied to the septum around the needle, to discourage moisture/air from entering the vial during the addition.). The reaction mixture was stirred at 10 °C for an additional 28 h. To remove the (+)-(C₅Ph₅)-DMAP*, the mixture was then filtered through a plug of silica gel using Et₂O (15 mL) as the eluant, and the volatiles were removed under reduced pressure. To remove a trace of a colored impurity, the residue was dissolved in CH₂Cl₂ (40 mL), H₂O (40 mL), and a 70% aqueous

solution of *t*-BuOOH (4 mL). This mixture was stirred at r.t. for 2 h, and the product was isolated by extraction into CH₂Cl₂ (3 x 50 mL). The CH₂Cl₂ layers were combined and dried over MgSO₄, and the solvent was removed under reduced pressure. The product was purified by column chromatography (hexanes → 20% Et₂O in hexanes), which furnished a mixture of the *d,l* and the meso diacetates (126 mg, 91% yield; dr: 7:1) as a clear, colorless oil. The diastereomers were separated by column chromatography (hexanes → 10% Et₂O in hexanes), which afforded the pure *d,l* diacetate (111 mg, 80% yield) with 99% ee.

The ee of the product was determined after deacetylation to the diol (HPLC analysis of the diol: Daicel CHIRALCEL OD-H column; solvent system: 5.0% *i*-PrOH in hexanes; 1.0 mL/min; retention times: 17.0 min (major), 28.0 min (minor)).

The second run was performed with (–)-(C₅Ph₅)-DMAP*. The mixture of the *d,l* and the meso diacetates was isolated as a clear, colorless oil (128 mg, 92% yield; dr: 7:1). An additional column chromatography afforded the pure *d,l* diacetate (110 mg, 79% yield) with 99% ee.

The spectral data match those described in the literature.³

[α]_D²⁴ = –94° (c = 1.0, CH₂Cl₂; obtained with (+)-(C₅Ph₅)-DMAP*).

IV. Preparation of Ru^{OAc}

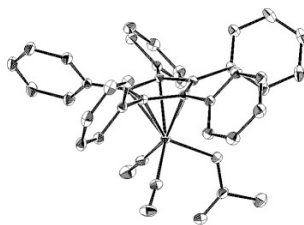
Synthesis of Ru^{OAc}. In a nitrogen-filled glovebox, Ru^{Cl} (100 mg, 0.157 mmol), KO^{*t*}-Bu (17.6 mg, 0.157 mmol), and toluene (2.5 mL) were combined in an oven-dried 20-mL vial equipped with a stir bar. The mixture was stirred at r.t. for 6 min, and then Ac₂O (151 μL, 1.60 mmol) was added to the vial via syringe. The resulting solution was stirred at r.t. for 12 h. Next, the volatiles were removed under reduced pressure, and the residue was purified by column chromatography (20% Et₂O in hexanes → 50% Et₂O in hexanes), which furnished the title compound as a yellow solid (74 mg, 71% yield; not optimized).

¹H NMR (400 MHz, CDCl₃) δ 7.21–7.17 (m, 5H), 7.10–7.06 (m, 10H), 6.99–6.97 (m, 10H), 2.00 (s, 3H).

¹³C NMR (100 MHz, CDCl₃) δ 198.2, 177.5, 132.2, 130.0, 128.3, 127.9, 106.0, 23.1.

IR (film) 3060, 2039, 1989, 1623, 1601, 1503, 1445, 1361, 1309, 1183, 1075, 1028, 803, 785, 743, 699, 675, 561 cm^{–1}.

Structure determination of Ru^{OAc} by X-ray crystallography. X-ray quality crystals were obtained by slowly evaporating CH₂Cl₂ from a saturated solution of Ru^{OAc}.



The crystal contained ca. 5% (η⁵-C₅Ph₅)Ru(CO)₂Cl, which is omitted for clarity.

(3) Ruble, J. C.; Tweddell, J.; Fu, G. C. *J. Org. Chem.* **1998**, *63*, 2794–2795.

Table 1. Crystal data and structure refinement for X11159.

Identification code	x11159	
Empirical formula	C38.91 H27.86 Cl0.05 O3.91 Ru	
Formula weight	660.56	
Temperature	100(2) K	
Wavelength	0.71073 Å	
Crystal system	Monoclinic	
Space group	P2(1)/c	
Unit cell dimensions	a = 19.5003(10) Å	a = 90°.
	b = 10.5810(5) Å	b = 104.8110(10)°.
	c = 14.9192(8) Å	g = 90°.
Volume	2976.0(3) Å ³	
Z	4	
Density (calculated)	1.474 Mg/m ³	
Absorption coefficient	0.573 mm ⁻¹	
F(000)	1349	
Crystal size	0.45 x 0.20 x 0.08 mm ³	
Theta range for data collection	2.16 to 29.94°.	
Index ranges	-27<=h<=27, -14<=k<=14, -20<=l<=20	
Reflections collected	116985	
Independent reflections	8640 [R(int) = 0.0289]	
Completeness to theta = 29.94°	100.0 %	
Absorption correction	Semi-empirical from equivalents	
Max. and min. transmission	0.9556 and 0.7825	
Refinement method	Full-matrix least-squares on F ²	
Data / restraints / parameters	8640 / 110 / 426	
Goodness-of-fit on F ²	1.057	
Final R indices [I>2sigma(I)]	R1 = 0.0212, wR2 = 0.0548	
R indices (all data)	R1 = 0.0234, wR2 = 0.0564	
Largest diff. peak and hole	0.455 and -0.642 e. Å ⁻³	

Table 2. Atomic coordinates (x 10⁴) and equivalent isotropic displacement parameters (Å²x 10³) for X11159. U(eq) is defined as one third of the trace of the orthogonalized U^{ij} tensor.

	x	y	z	U(eq)
Ru(1)	2599(1)	6419(1)	4071(1)	12(1)
C(1)	2032(1)	6959(1)	5182(1)	13(1)
C(11)	1295(1)	6702(1)	5227(1)	14(1)

C(12)	1184(1)	6442(1)	6096(1)	18(1)
C(13)	498(1)	6319(1)	6203(1)	23(1)
C(14)	-79(1)	6446(1)	5441(1)	24(1)
C(15)	27(1)	6702(1)	4574(1)	22(1)
C(16)	711(1)	6835(1)	4462(1)	18(1)
C(2)	2637(1)	6192(1)	5573(1)	12(1)
C(21)	2624(1)	4928(1)	5997(1)	13(1)
C(22)	3065(1)	4642(1)	6872(1)	16(1)
C(23)	3045(1)	3444(1)	7254(1)	18(1)
C(24)	2590(1)	2525(1)	6774(1)	19(1)
C(25)	2148(1)	2801(1)	5904(1)	19(1)
C(26)	2166(1)	3992(1)	5520(1)	16(1)
C(3)	3266(1)	6879(1)	5477(1)	13(1)
C(31)	4013(1)	6568(1)	5960(1)	14(1)
C(32)	4391(1)	7425(1)	6614(1)	18(1)
C(33)	5066(1)	7131(1)	7154(1)	23(1)
C(34)	5375(1)	5981(2)	7044(1)	25(1)
C(35)	5013(1)	5133(1)	6382(1)	25(1)
C(36)	4333(1)	5422(1)	5845(1)	20(1)
C(4)	3026(1)	8042(1)	5007(1)	13(1)
C(41)	3462(1)	9143(1)	4874(1)	14(1)
C(42)	3363(1)	10269(1)	5316(1)	18(1)
C(43)	3737(1)	11353(1)	5205(1)	22(1)
C(44)	4208(1)	11324(1)	4646(1)	22(1)
C(45)	4311(1)	10208(1)	4210(1)	20(1)
C(46)	3944(1)	9115(1)	4328(1)	17(1)
C(5)	2260(1)	8081(1)	4794(1)	13(1)
C(51)	1809(1)	9156(1)	4354(1)	14(1)
C(52)	1898(1)	9718(1)	3544(1)	18(1)
C(53)	1474(1)	10728(1)	3139(1)	22(1)
C(54)	954(1)	11184(1)	3538(1)	22(1)
C(55)	868(1)	10645(1)	4352(1)	20(1)
C(56)	1296(1)	9642(1)	4763(1)	16(1)
C(8)	3019(1)	4851(1)	3902(1)	19(1)
O(3)	3271(1)	3900(1)	3830(1)	29(1)
O(1)	1638(1)	5699(1)	3315(1)	22(1)
C(6)	1556(1)	5270(1)	2481(1)	20(1)
O(2)	2007(1)	5300(1)	2041(1)	23(1)
C(7)	839(1)	4676(2)	2083(1)	40(1)

C(9A)	1735(14)	6050(30)	3110(20)	22(5)
O(4A)	1230(11)	5740(30)	2537(16)	42(6)
C(9)	2849(1)	7030(1)	3004(1)	18(1)
O(4)	3046(1)	7430(1)	2409(1)	27(1)
Cl(1A)	3017(5)	7011(10)	2694(7)	26(2)

Table 3. Bond lengths [Å] and angles [°] for X11159.

Ru(1)-C(9)	1.894(2)
Ru(1)-C(8)	1.8955(13)
Ru(1)-C(9A)	1.953(14)
Ru(1)-O(1)	2.0689(11)
Ru(1)-C(3)	2.2213(11)
Ru(1)-C(4)	2.2360(11)
Ru(1)-C(2)	2.2363(11)
Ru(1)-C(5)	2.2487(11)
Ru(1)-C(1)	2.2880(11)
Ru(1)-Cl(1A)	2.477(11)
C(1)-C(2)	1.4290(15)
C(1)-C(5)	1.4403(16)
C(1)-C(11)	1.4802(15)
C(11)-C(12)	1.3950(17)
C(11)-C(16)	1.3984(16)
C(12)-C(13)	1.3932(17)
C(12)-H(12)	0.9500
C(13)-C(14)	1.387(2)
C(13)-H(13)	0.9500
C(14)-C(15)	1.387(2)
C(14)-H(14)	0.9500
C(15)-C(16)	1.3919(17)
C(15)-H(15)	0.9500
C(16)-H(16)	0.9500
C(2)-C(3)	1.4630(15)
C(2)-C(21)	1.4830(16)
C(21)-C(22)	1.3992(16)
C(21)-C(26)	1.4002(16)
C(22)-C(23)	1.3945(16)
C(22)-H(22)	0.9500
C(23)-C(24)	1.3864(18)

C(23)-H(23)	0.9500
C(24)-C(25)	1.3925(17)
C(24)-H(24)	0.9500
C(25)-C(26)	1.3882(17)
C(25)-H(25)	0.9500
C(26)-H(26)	0.9500
C(3)-C(4)	1.4342(16)
C(3)-C(31)	1.4882(16)
C(31)-C(36)	1.3941(17)
C(31)-C(32)	1.3968(17)
C(32)-C(33)	1.3922(17)
C(32)-H(32)	0.9500
C(33)-C(34)	1.387(2)
C(33)-H(33)	0.9500
C(34)-C(35)	1.387(2)
C(34)-H(34)	0.9500
C(35)-C(36)	1.3965(18)
C(35)-H(35)	0.9500
C(36)-H(36)	0.9500
C(4)-C(5)	1.4459(15)
C(4)-C(41)	1.4841(16)
C(41)-C(46)	1.3939(16)
C(41)-C(42)	1.3984(17)
C(42)-C(43)	1.3912(17)
C(42)-H(42)	0.9500
C(43)-C(44)	1.389(2)
C(43)-H(43)	0.9500
C(44)-C(45)	1.387(2)
C(44)-H(44)	0.9500
C(45)-C(46)	1.3936(17)
C(45)-H(45)	0.9500
C(46)-H(46)	0.9500
C(5)-C(51)	1.4839(16)
C(51)-C(52)	1.3953(16)
C(51)-C(56)	1.3977(16)
C(52)-C(53)	1.3910(17)
C(52)-H(52)	0.9500
C(53)-C(54)	1.3871(19)
C(53)-H(53)	0.9500

C(54)-C(55)	1.3900(19)
C(54)-H(54)	0.9500
C(55)-C(56)	1.3913(17)
C(55)-H(55)	0.9500
C(56)-H(56)	0.9500
C(8)-O(3)	1.1362(16)
O(1)-C(6)	1.2951(17)
C(6)-O(2)	1.2237(18)
C(6)-C(7)	1.510(2)
C(7)-H(7A)	0.9800
C(7)-H(7B)	0.9800
C(7)-H(7C)	0.9800
C(9A)-O(4A)	1.171(15)
C(9)-O(4)	1.134(3)
C(9)-Ru(1)-C(8)	88.49(6)
C(9)-Ru(1)-C(9A)	79.0(12)
C(8)-Ru(1)-C(9A)	92.8(12)
C(9)-Ru(1)-O(1)	93.43(6)
C(8)-Ru(1)-O(1)	88.15(5)
C(9A)-Ru(1)-O(1)	15.3(11)
C(9)-Ru(1)-C(3)	120.25(5)
C(8)-Ru(1)-C(3)	98.56(5)
C(9A)-Ru(1)-C(3)	157.6(13)
O(1)-Ru(1)-C(3)	145.65(5)
C(9)-Ru(1)-C(4)	97.30(5)
C(8)-Ru(1)-C(4)	130.58(5)
C(9A)-Ru(1)-C(4)	136.5(11)
O(1)-Ru(1)-C(4)	139.83(5)
C(3)-Ru(1)-C(4)	37.54(4)
C(9)-Ru(1)-C(2)	158.26(5)
C(8)-Ru(1)-C(2)	97.74(5)
C(9A)-Ru(1)-C(2)	121.2(12)
O(1)-Ru(1)-C(2)	107.52(5)
C(3)-Ru(1)-C(2)	38.32(4)
C(4)-Ru(1)-C(2)	62.92(4)
C(9)-Ru(1)-C(5)	107.84(5)
C(8)-Ru(1)-C(5)	159.77(5)
C(9A)-Ru(1)-C(5)	101.8(12)

O(1)-Ru(1)-C(5)	102.32(5)
C(3)-Ru(1)-C(5)	63.15(4)
C(4)-Ru(1)-C(5)	37.61(4)
C(2)-Ru(1)-C(5)	62.74(4)
C(9)-Ru(1)-C(1)	143.10(5)
C(8)-Ru(1)-C(1)	128.38(5)
C(9A)-Ru(1)-C(1)	95.6(13)
O(1)-Ru(1)-C(1)	87.10(4)
C(3)-Ru(1)-C(1)	62.33(4)
C(4)-Ru(1)-C(1)	61.80(4)
C(2)-Ru(1)-C(1)	36.80(4)
C(5)-Ru(1)-C(1)	37.01(4)
C(9)-Ru(1)-Cl(1A)	6.4(3)
C(8)-Ru(1)-Cl(1A)	82.3(3)
C(9A)-Ru(1)-Cl(1A)	81.1(12)
O(1)-Ru(1)-Cl(1A)	94.9(2)
C(3)-Ru(1)-Cl(1A)	119.4(2)
C(4)-Ru(1)-Cl(1A)	100.0(2)
C(2)-Ru(1)-Cl(1A)	157.6(2)
C(5)-Ru(1)-Cl(1A)	113.5(3)
C(1)-Ru(1)-Cl(1A)	149.4(3)
C(2)-C(1)-C(5)	108.92(10)
C(2)-C(1)-C(11)	125.99(10)
C(5)-C(1)-C(11)	124.95(10)
C(2)-C(1)-Ru(1)	69.63(6)
C(5)-C(1)-Ru(1)	70.01(6)
C(11)-C(1)-Ru(1)	129.88(8)
C(12)-C(11)-C(16)	119.35(11)
C(12)-C(11)-C(1)	117.45(11)
C(16)-C(11)-C(1)	122.88(11)
C(13)-C(12)-C(11)	120.40(12)
C(13)-C(12)-H(12)	119.8
C(11)-C(12)-H(12)	119.8
C(14)-C(13)-C(12)	119.93(13)
C(14)-C(13)-H(13)	120.0
C(12)-C(13)-H(13)	120.0
C(13)-C(14)-C(15)	120.03(12)
C(13)-C(14)-H(14)	120.0
C(15)-C(14)-H(14)	120.0

C(14)-C(15)-C(16)	120.42(12)
C(14)-C(15)-H(15)	119.8
C(16)-C(15)-H(15)	119.8
C(15)-C(16)-C(11)	119.88(12)
C(15)-C(16)-H(16)	120.1
C(11)-C(16)-H(16)	120.1
C(1)-C(2)-C(3)	107.63(10)
C(1)-C(2)-C(21)	125.73(10)
C(3)-C(2)-C(21)	126.64(10)
C(1)-C(2)-Ru(1)	73.57(6)
C(3)-C(2)-Ru(1)	70.28(6)
C(21)-C(2)-Ru(1)	121.64(8)
C(22)-C(21)-C(26)	118.79(11)
C(22)-C(21)-C(2)	121.26(10)
C(26)-C(21)-C(2)	119.95(10)
C(23)-C(22)-C(21)	120.18(11)
C(23)-C(22)-H(22)	119.9
C(21)-C(22)-H(22)	119.9
C(24)-C(23)-C(22)	120.52(11)
C(24)-C(23)-H(23)	119.7
C(22)-C(23)-H(23)	119.7
C(23)-C(24)-C(25)	119.68(11)
C(23)-C(24)-H(24)	120.2
C(25)-C(24)-H(24)	120.2
C(26)-C(25)-C(24)	120.08(11)
C(26)-C(25)-H(25)	120.0
C(24)-C(25)-H(25)	120.0
C(25)-C(26)-C(21)	120.75(11)
C(25)-C(26)-H(26)	119.6
C(21)-C(26)-H(26)	119.6
C(4)-C(3)-C(2)	107.35(9)
C(4)-C(3)-C(31)	125.67(10)
C(2)-C(3)-C(31)	125.69(10)
C(4)-C(3)-Ru(1)	71.79(6)
C(2)-C(3)-Ru(1)	71.40(6)
C(31)-C(3)-Ru(1)	132.30(8)
C(36)-C(31)-C(32)	118.56(11)
C(36)-C(31)-C(3)	123.13(11)
C(32)-C(31)-C(3)	118.10(11)

C(33)-C(32)-C(31)	120.80(12)
C(33)-C(32)-H(32)	119.6
C(31)-C(32)-H(32)	119.6
C(34)-C(33)-C(32)	120.17(13)
C(34)-C(33)-H(33)	119.9
C(32)-C(33)-H(33)	119.9
C(33)-C(34)-C(35)	119.63(12)
C(33)-C(34)-H(34)	120.2
C(35)-C(34)-H(34)	120.2
C(34)-C(35)-C(36)	120.26(13)
C(34)-C(35)-H(35)	119.9
C(36)-C(35)-H(35)	119.9
C(31)-C(36)-C(35)	120.56(12)
C(31)-C(36)-H(36)	119.7
C(35)-C(36)-H(36)	119.7
C(3)-C(4)-C(5)	108.72(10)
C(3)-C(4)-C(41)	127.75(10)
C(5)-C(4)-C(41)	122.87(10)
C(3)-C(4)-Ru(1)	70.67(6)
C(5)-C(4)-Ru(1)	71.67(6)
C(41)-C(4)-Ru(1)	130.88(8)
C(46)-C(41)-C(42)	119.20(11)
C(46)-C(41)-C(4)	123.78(11)
C(42)-C(41)-C(4)	117.00(10)
C(43)-C(42)-C(41)	120.46(12)
C(43)-C(42)-H(42)	119.8
C(41)-C(42)-H(42)	119.8
C(44)-C(43)-C(42)	120.01(12)
C(44)-C(43)-H(43)	120.0
C(42)-C(43)-H(43)	120.0
C(45)-C(44)-C(43)	119.83(12)
C(45)-C(44)-H(44)	120.1
C(43)-C(44)-H(44)	120.1
C(44)-C(45)-C(46)	120.43(12)
C(44)-C(45)-H(45)	119.8
C(46)-C(45)-H(45)	119.8
C(45)-C(46)-C(41)	120.06(12)
C(45)-C(46)-H(46)	120.0
C(41)-C(46)-H(46)	120.0

C(1)-C(5)-C(4)	107.24(10)
C(1)-C(5)-C(51)	126.91(10)
C(4)-C(5)-C(51)	125.50(10)
C(1)-C(5)-Ru(1)	72.98(6)
C(4)-C(5)-Ru(1)	70.72(6)
C(51)-C(5)-Ru(1)	126.90(8)
C(52)-C(51)-C(56)	118.79(11)
C(52)-C(51)-C(5)	121.30(10)
C(56)-C(51)-C(5)	119.90(10)
C(53)-C(52)-C(51)	120.86(12)
C(53)-C(52)-H(52)	119.6
C(51)-C(52)-H(52)	119.6
C(54)-C(53)-C(52)	119.89(12)
C(54)-C(53)-H(53)	120.1
C(52)-C(53)-H(53)	120.1
C(53)-C(54)-C(55)	119.83(12)
C(53)-C(54)-H(54)	120.1
C(55)-C(54)-H(54)	120.1
C(54)-C(55)-C(56)	120.31(12)
C(54)-C(55)-H(55)	119.8
C(56)-C(55)-H(55)	119.8
C(55)-C(56)-C(51)	120.30(11)
C(55)-C(56)-H(56)	119.8
C(51)-C(56)-H(56)	119.8
O(3)-C(8)-Ru(1)	177.64(11)
C(6)-O(1)-Ru(1)	121.65(10)
O(2)-C(6)-O(1)	125.23(14)
O(2)-C(6)-C(7)	121.31(13)
O(1)-C(6)-C(7)	113.45(14)
O(4A)-C(9A)-Ru(1)	175(3)
O(4)-C(9)-Ru(1)	174.81(14)

Symmetry transformations used to generate equivalent atoms:

Table 4. Anisotropic displacement parameters ($\text{\AA}^2 \times 10^3$) for X11159. The anisotropic displacement factor exponent takes the form: $-2p^2[h^2 a^{*2}U^{11} + \dots + 2 h k a^* b^* U^{12}]$

$U^{11}U^{22}$	U^{33}	U^{23}	U^{13}	U^{12}		
Ru(1)	13(1)	12(1)	10(1)	1(1)	4(1)	1(1)
C(1)13(1)	14(1)	12(1)	0(1)	4(1)	1(1)	
C(11)13(1)	12(1)	18(1)	0(1)	5(1)	1(1)	
C(12)18(1)	18(1)	21(1)	2(1)	8(1)	1(1)	
C(13)22(1)	20(1)	32(1)	1(1)	16(1)	1(1)	
C(14)16(1)	17(1)	43(1)	-3(1)	13(1)	0(1)	
C(15)13(1)	16(1)	34(1)	-4(1)	2(1)	2(1)	
C(16)16(1)	15(1)	21(1)	-1(1)	2(1)	2(1)	
C(2)13(1)	13(1)	10(1)	1(1)	3(1)	0(1)	
C(21)13(1)	12(1)	13(1)	1(1)	4(1)	1(1)	
C(22)17(1)	14(1)	15(1)	0(1)	2(1)	-1(1)	
C(23)21(1)	17(1)	16(1)	4(1)	2(1)	1(1)	
C(24)23(1)	14(1)	21(1)	3(1)	7(1)	0(1)	
C(25)20(1)	15(1)	21(1)	-1(1)	4(1)	-4(1)	
C(26)16(1)	17(1)	15(1)	1(1)	2(1)	-1(1)	
C(3)13(1)	13(1)	12(1)	0(1)	3(1)	0(1)	
C(31)12(1)	17(1)	13(1)	2(1)	4(1)	1(1)	
C(32)16(1)	20(1)	18(1)	0(1)	4(1)	-1(1)	
C(33)17(1)	30(1)	20(1)	0(1)	0(1)	-4(1)	
C(34)14(1)	36(1)	23(1)	7(1)	2(1)	3(1)	
C(35)19(1)	27(1)	27(1)	4(1)	4(1)	8(1)	
C(36)18(1)	20(1)	22(1)	-1(1)	2(1)	3(1)	
C(4)14(1)	13(1)	11(1)	0(1)	4(1)	0(1)	
C(41)14(1)	14(1)	14(1)	3(1)	2(1)	-1(1)	
C(42)20(1)	16(1)	19(1)	0(1)	4(1)	-1(1)	
C(43)22(1)	15(1)	26(1)	1(1)	1(1)	-2(1)	
C(44)18(1)	19(1)	27(1)	8(1)	-1(1)	-4(1)	
C(45)14(1)	25(1)	21(1)	7(1)	2(1)	-2(1)	
C(46)14(1)	19(1)	17(1)	2(1)	3(1)	0(1)	
C(5)14(1)	12(1)	12(1)	0(1)	4(1)	1(1)	
C(51)14(1)	12(1)	14(1)	0(1)	2(1)	1(1)	
C(52)20(1)	19(1)	17(1)	3(1)	6(1)	4(1)	
C(53)26(1)	20(1)	20(1)	7(1)	6(1)	5(1)	
C(54)24(1)	17(1)	24(1)	4(1)	2(1)	7(1)	

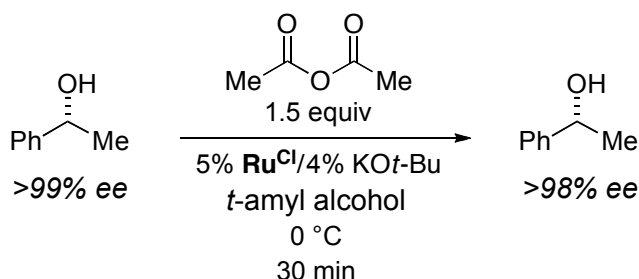
C(55)19(1)	17(1)	22(1)	-1(1)	4(1)	5(1)	
C(56)18(1)	14(1)	16(1)	-1(1)	4(1)	1(1)	
C(8)23(1)	20(1)	12(1)	0(1)	4(1)	2(1)	
O(3)39(1)	22(1)	24(1)	-3(1)	4(1)	12(1)	
O(1)16(1)	33(1)	18(1)	-8(1)	4(1)	-4(1)	
C(6)19(1)	21(1)	16(1)	-4(1)	0(1)	0(1)	
O(2)24(1)	28(1)	17(1)	-6(1)	6(1)	-1(1)	
C(7)25(1)	66(1)	27(1)	-18(1)	2(1)	-15(1)	
C(9A)	22(7)	14(11)	28(12)	1(9)	2(5)	-6(8)
O(4A)	16(9)	62(14)	41(11)	4(11)	-4(6)	-6(10)
C(9)23(1)	14(1)	16(1)	-2(1)	4(1)	0(1)	
O(4)39(1)	26(1)	20(1)	0(1)	14(1)	-7(1)	
Cl(1A)	42(5)	25(5)	17(4)	3(3)	20(3)	-7(3)

Table 5. Hydrogen coordinates ($\times 10^4$) and isotropic displacement parameters ($\text{\AA}^2 \times 10^{-3}$) for X11159.

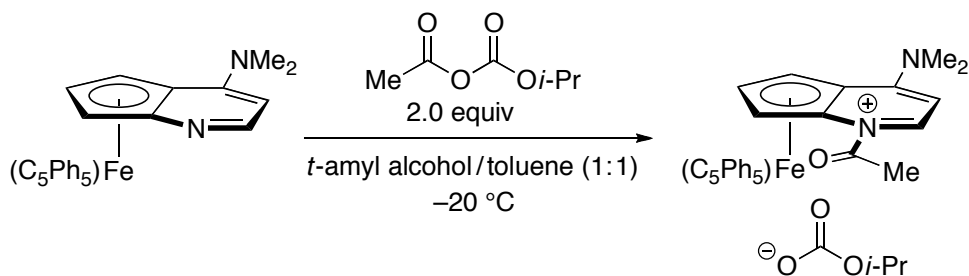
x	y	z	U(eq)	
H(12)	1579	6347	6618	22
H(13)	426	6147	6797	28
H(14)	-547	6359	5512	29
H(15)	-370	6787	4053	27
H(16)	780	7015	3868	21
H(22)	3378	5266	7206	19
H(23)	3345	3257	7848	22
H(24)	2580	1709	7037	23
H(25)	1835	2175	5574	23
H(26)	1864	4174	4926	19
H(32)	4185	8218	6691	22
H(33)	5316	7720	7600	28
H(34)	5833	5775	7419	29
H(35)	5227	4352	6294	30
H(36)	4086	4832	5397	24
H(42)	3039	10295	5695	22
H(43)	3671	12113	5511	26
H(44)	4458	12066	4563	27
H(45)	4633	10188	3829	25

H(46)	4023	8351	4036	20
H(52)	2251	9406	3267	22
H(53)	1541	11105	2590	27
H(54)	658	11863	3256	27
H(55)	516	10964	4630	24
H(56)	1239	9287	5324	19
H(7A)	616	5068	1484	60
H(7B)	538	4808	2510	60
H(7C)	897	3768	1996	60

V. Mechanistic/Reactivity Studies



Eq 5. In a nitrogen-filled glovebox, a solution of Ru^{Cl} (6.4 mg, 0.010 mmol) and KO^t-Bu (0.90 mg, 0.0080 mmol) in *t*-amyl alcohol (400 μL) in an oven-dried 5-mL vial equipped with a stir bar was stirred at room temperature for 6 min. The vial was then capped with a septum-lined cap, removed from the glovebox, and cooled to 0 °C. Next, Ac₂O (28 μL, 0.30 mmol) was added dropwise to the reaction vial over 1 min. Then, (*R*)-1-phenylethanol (24 μL, 0.20 mmol) was added, and the reaction mixture was stirred at 0 °C. After 30 min, an aliquot (40 μL) of the reaction mixture was removed and diluted with Et₂O (1.0 mL). The ee of 1-phenylethanol was determined by chiral GC analysis.



Eq 11. In a nitrogen-filled glovebox, C₅Ph₅-DMAP* (20 mg, 0.030 mmol), toluene-*d*₈ (0.25 mL), and *t*-amyl alcohol (0.25 mL) were combined in a dry NMR tube. The NMR tube was then capped with a screw-cap and removed from the glovebox. The NMR tube was cooled to -20 °C, and then acetyl isopropyl carbonate (8.6 μL, 0.60 mmol) was added to the NMR tube via syringe. The reaction was monitored by ¹H NMR as a function of time. After 1.5 h at -20 °C, the acylpyridinium salt had

formed almost quantitatively. The ^1H and ^{13}C NMR spectra of the acylpyridinium salt are included in Section VI.

Determination of the rate law: DKR of 1-phenylethanol. In a nitrogen-filled glovebox, Ru^{Cl} , $\text{KO}t\text{-Bu}$, tetradecane, toluene (250 μL), and t -amyl alcohol (250 μL) were combined in an oven-dried 4-mL vial equipped with a stir bar. The mixture was stirred at room temperature for 6 min, and then 1-phenylethanol and $(\text{C}_5\text{Ph}_5)\text{-DMAP}^*$ were added to the vial. The vial was then capped with a septum-lined cap, removed from the glovebox, and cooled to 10 $^\circ\text{C}$. After cooling the solution for 10 min, acetyl isopropyl carbonate was added dropwise to the vial via syringe over 1 min. An aliquot (40 μL) of the reaction mixture was removed after 10 min, 30 min, 50 min, 80 min, and 120 min, and then filtered through a short pad of silica. The amount of the product was determined by GC analysis (calibrated with tetradecane as the internal standard).

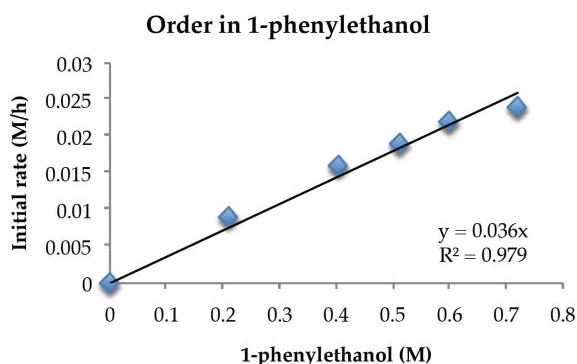
Order in 1-phenylethanol:

Table S1. Observed Initial Rates

$[\text{1-phenylethanol}]_{\text{initial}}$ (M) ^a	k_{obs} (M/h)
0.00	0.00
0.21	0.0092
0.40	0.016
0.51	0.019
0.60	0.022
0.72	0.024

^a Reaction conditions: $[\text{acetyl isopropyl carbonate}]_{\text{initial}} = 0.75 \text{ M}$, $[(\text{C}_5\text{Ph}_5)\text{-DMAP}^*]_{\text{initial}} = 5.0 \text{ mM}$, $[\text{Ru}^{\text{Cl}}]_{\text{initial}} = 0.026 \text{ M}$, and $[\text{KO}t\text{-Bu}]_{\text{initial}} = 0.022 \text{ M}$.

Figure S1.



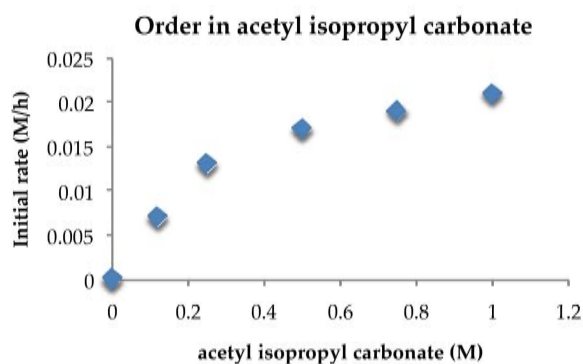
Order in acetyl isopropyl carbonate:

Table S2. Observed Initial Rates

[acetyl isopropyl carbonate] _{initial} (M) ^a	k _{obs} (M/h)
0.00	0.00
0.12	0.0071
0.25	0.013
0.50	0.017
0.75	0.019
1.00	0.021

^a Reaction conditions: [1-phenylethanol]_{initial} = 0.50 M, [(C₅Ph₅)-DMAP*]_{initial} = 5.0 mM, [Ru^{Cl}]_{initial} = 0.026 M, and [KO*t*-Bu]_{initial} = 0.022 M.

Figure S2.



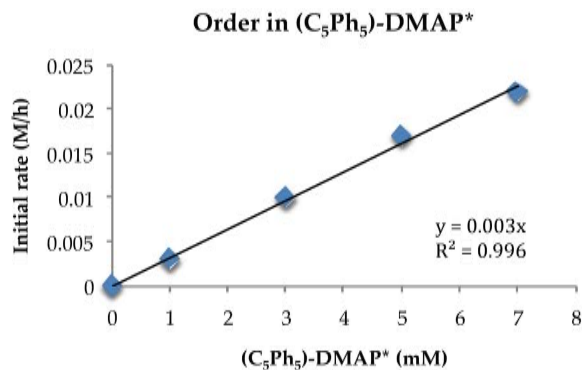
Order in (C₅Ph₅)-DMAP*:

Table S3. Observed Initial Rates

[(C ₅ Ph ₅)-DMAP*] _{initial} (mM) ^a	k _{obs} (M/h)
0.0	0.00
1.0	0.0034
3.0	0.010
5.0	0.017
7.0	0.022

^a Reaction conditions: [1-phenylethanol]_{initial} = 0.50 M, [acetyl isopropyl carbonate]_{initial} = 0.75 M, [Ru^{Cl}]_{initial} = 0.026 M, and [KO*t*-Bu]_{initial} = 0.022 M.

Figure S3.



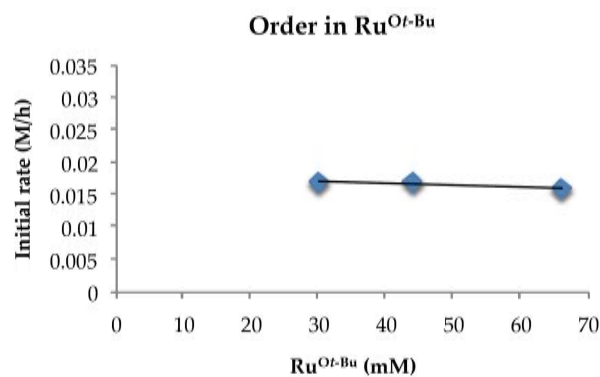
Order in Ru^{O-*t*Bu}

Table S4. Observed Initial Rates

[Ru ^{O-<i>t</i>Bu}] _{initial} (M) ^a	k _{obs} (M/h)
0.030	0.017
0.044	0.017
0.066	0.016

^a Reaction conditions: [1-phenylethanol]_{initial} = 0.50 M, [acetyl isopropyl carbonate]_{initial} = 0.75 M, and [(C₅Ph₅)-DMAP*]_{initial} = 5.0 mM.

Figure S4.



VI. ^1H NMR Spectra and GC/HPLC traces

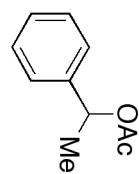
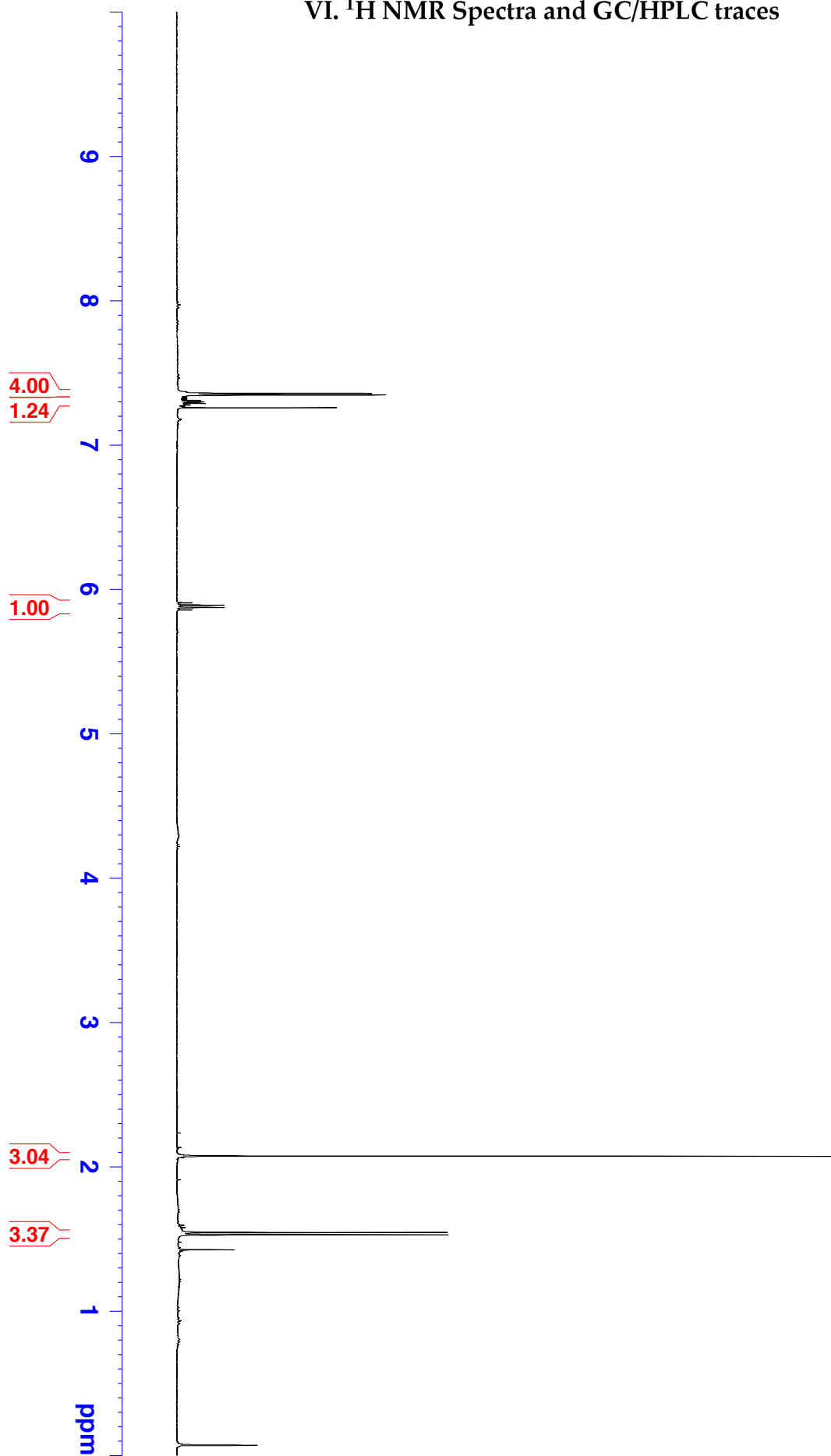


Table 1, entry 1



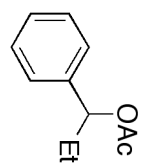
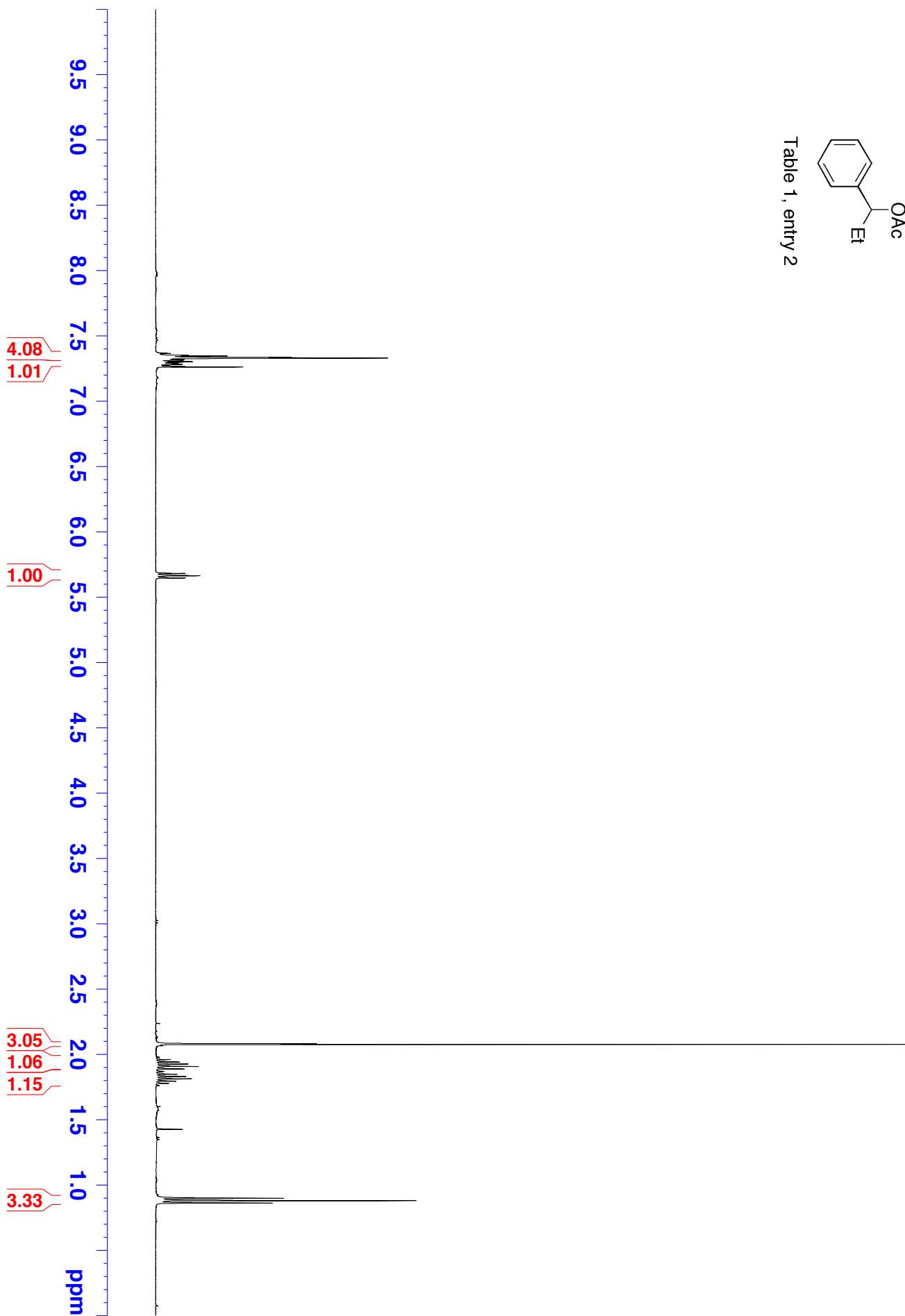


Table 1, entry 2



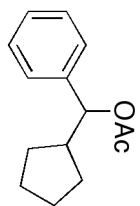
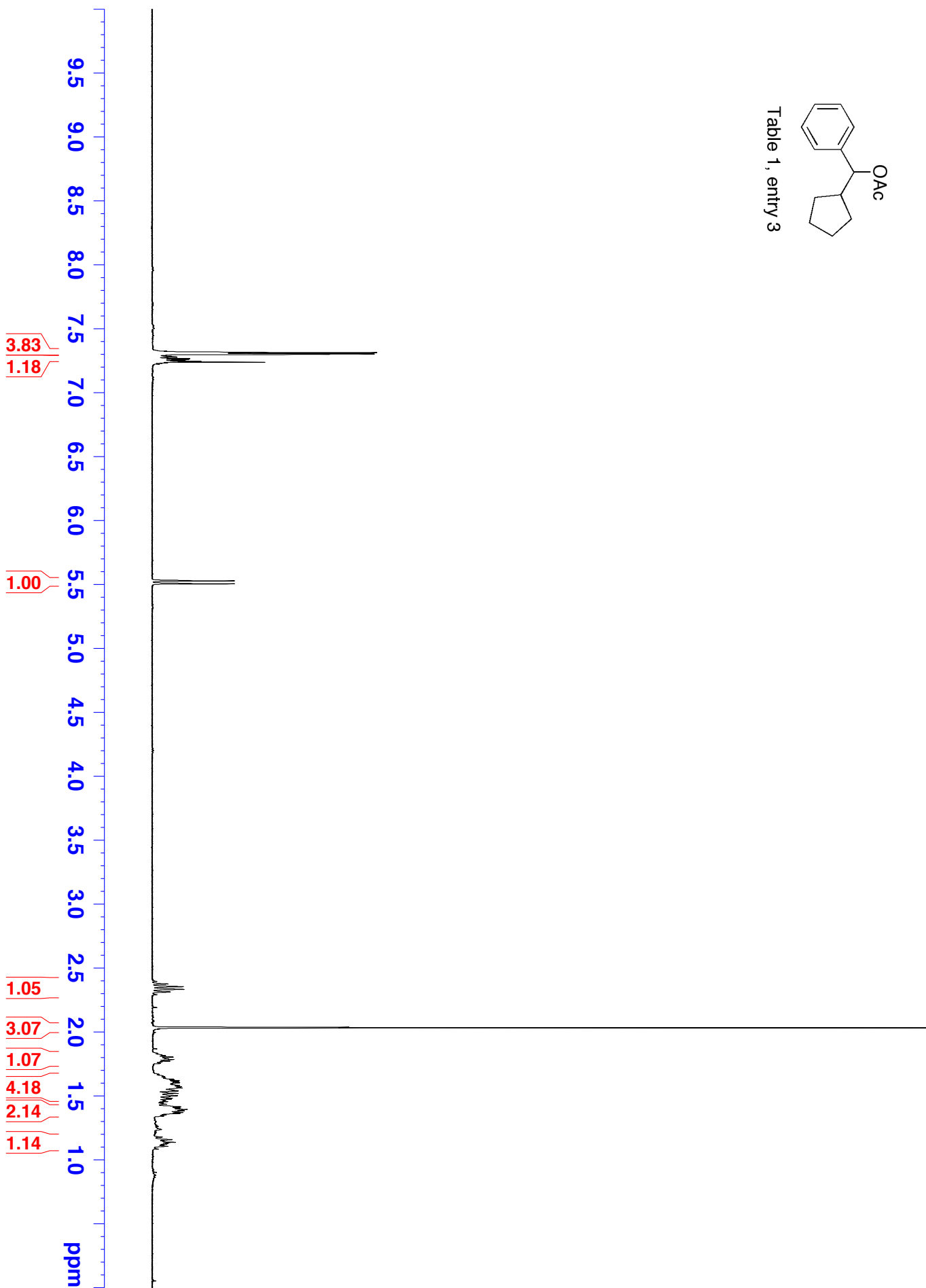


Table 1, entry 3



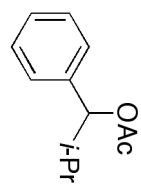
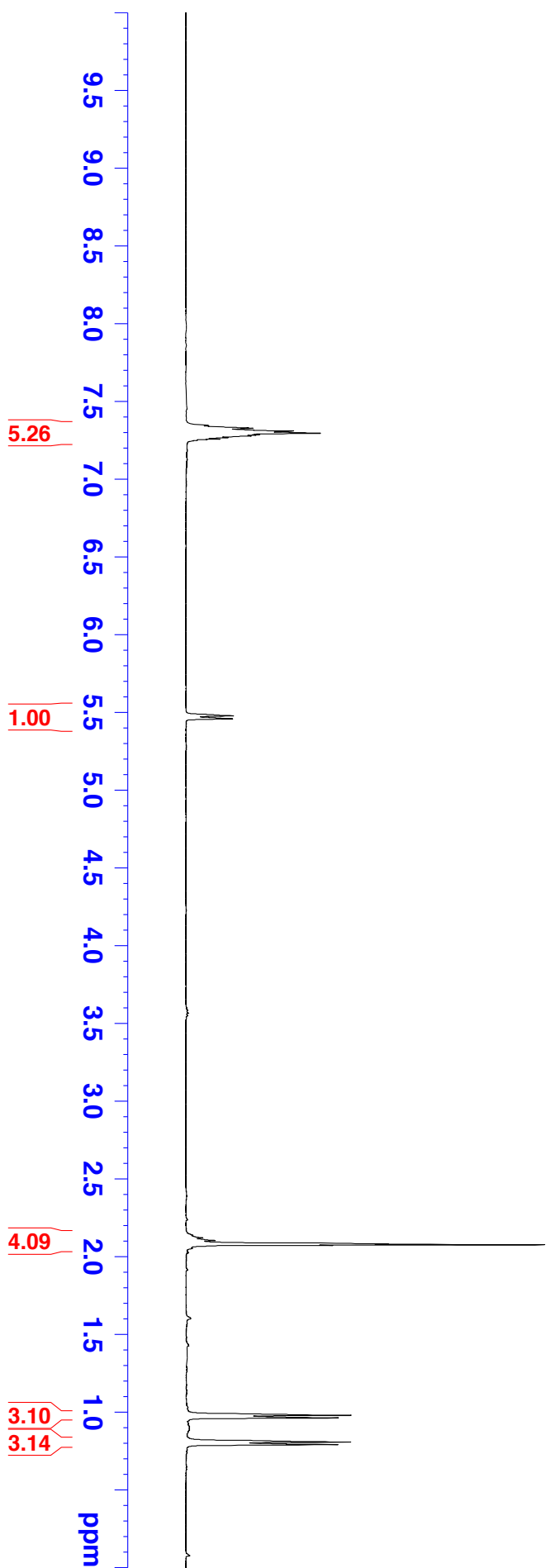


Table 1, entry 4



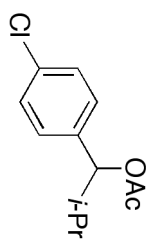
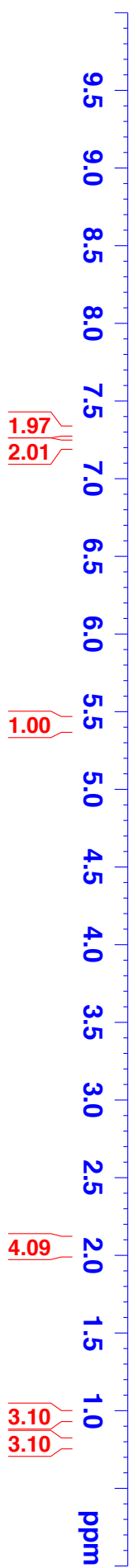


Table 1, entry 5



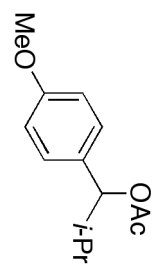
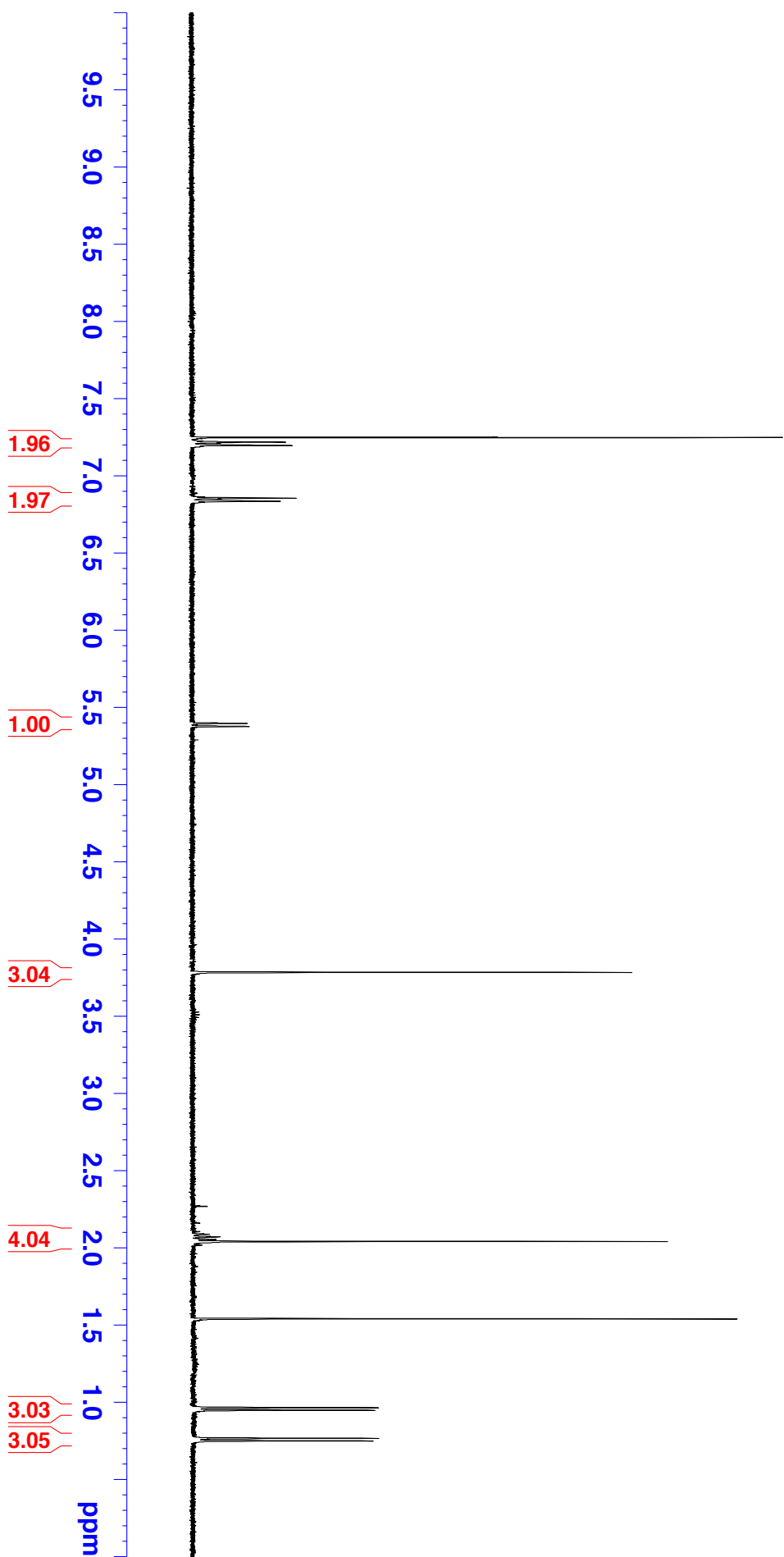


Table 1, entry 6



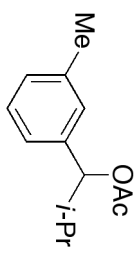
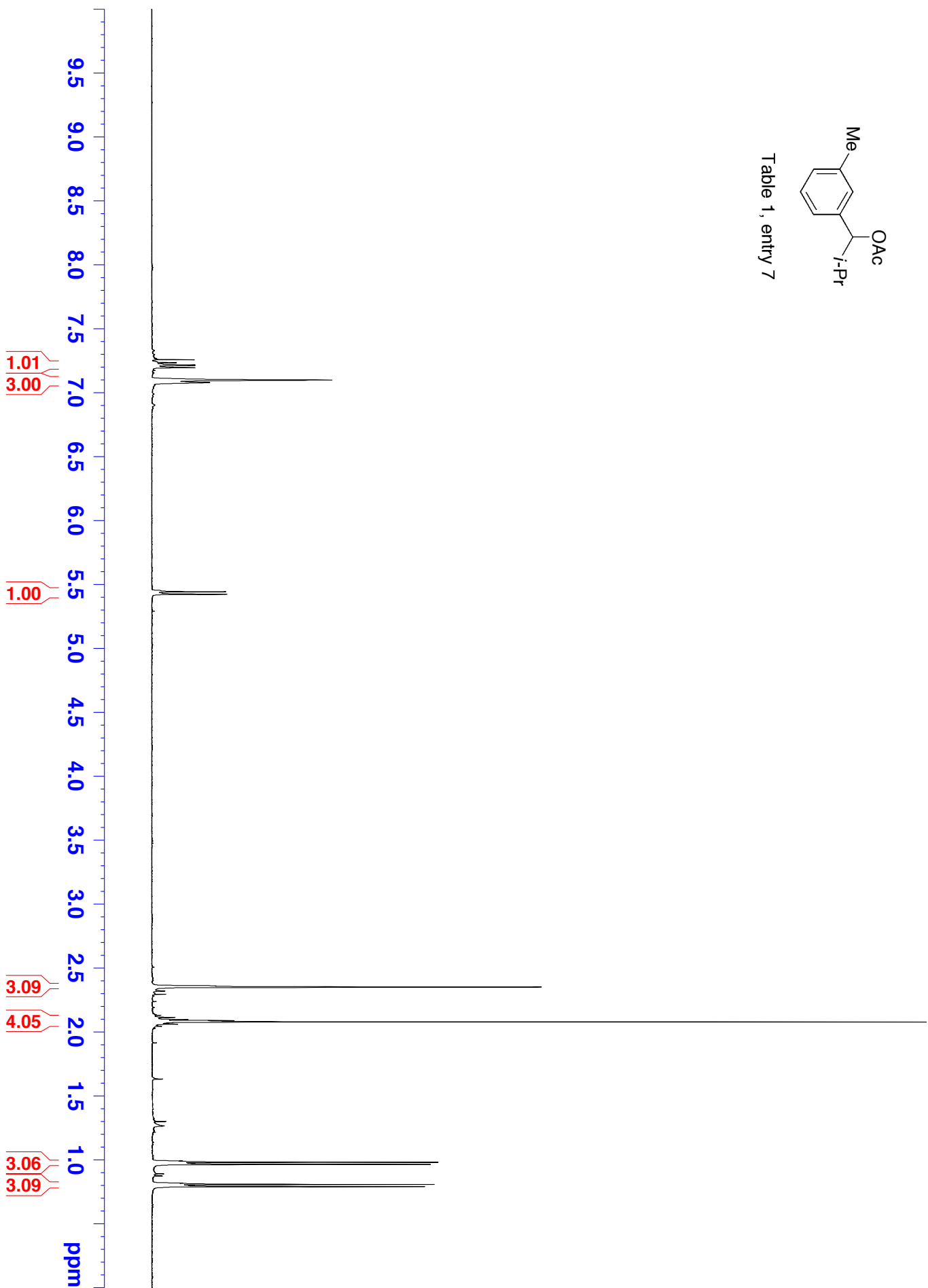


Table 1, entry 7



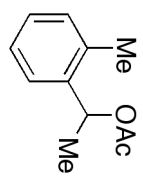
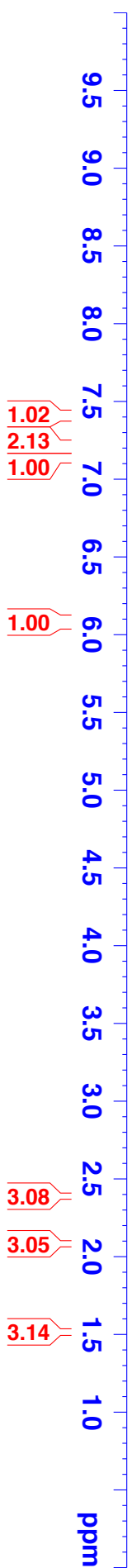


Table 1, entry 8



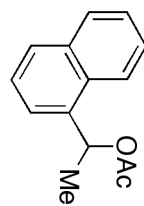
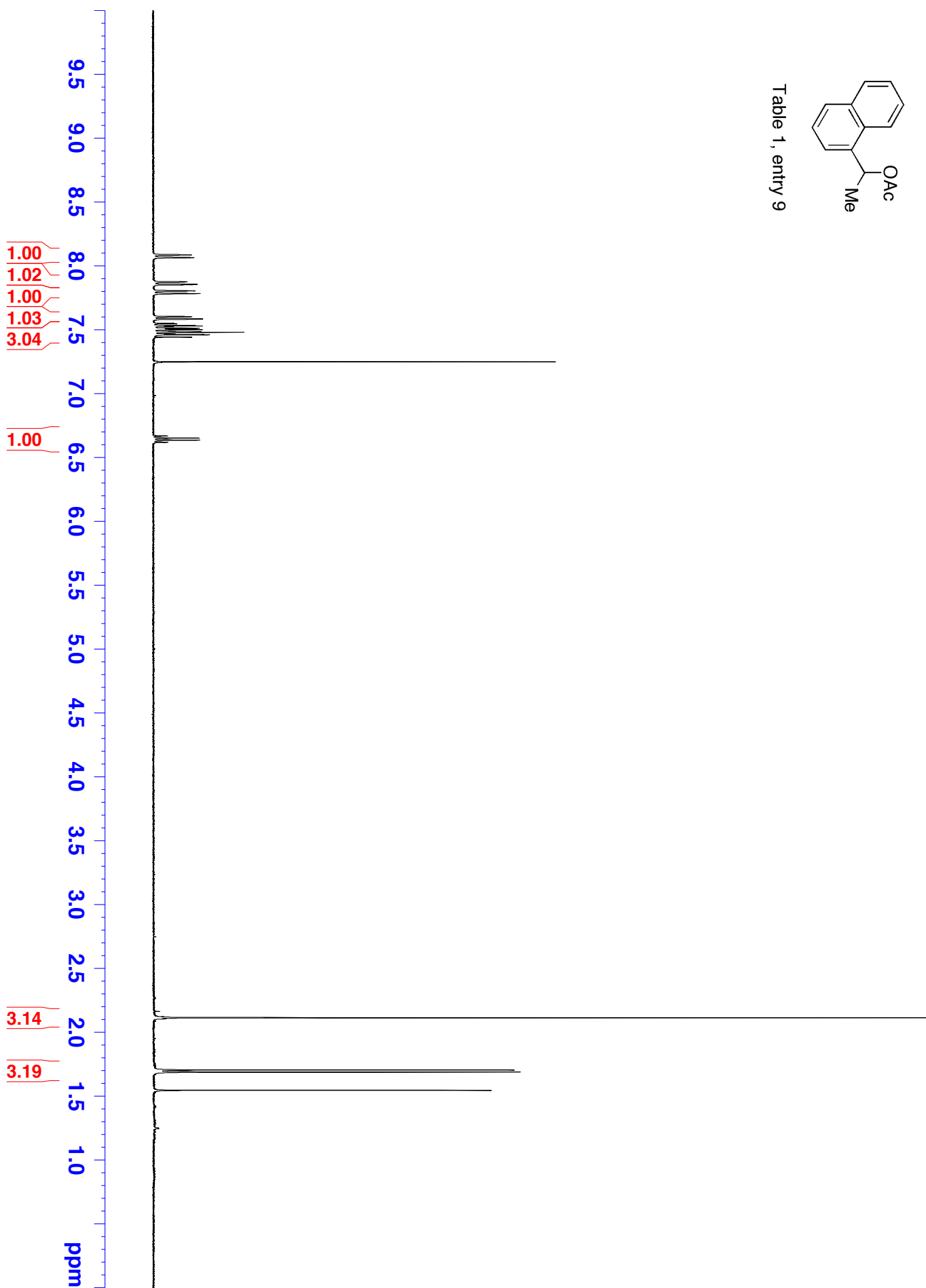


Table 1, entry 9



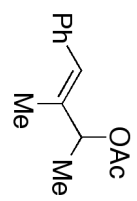
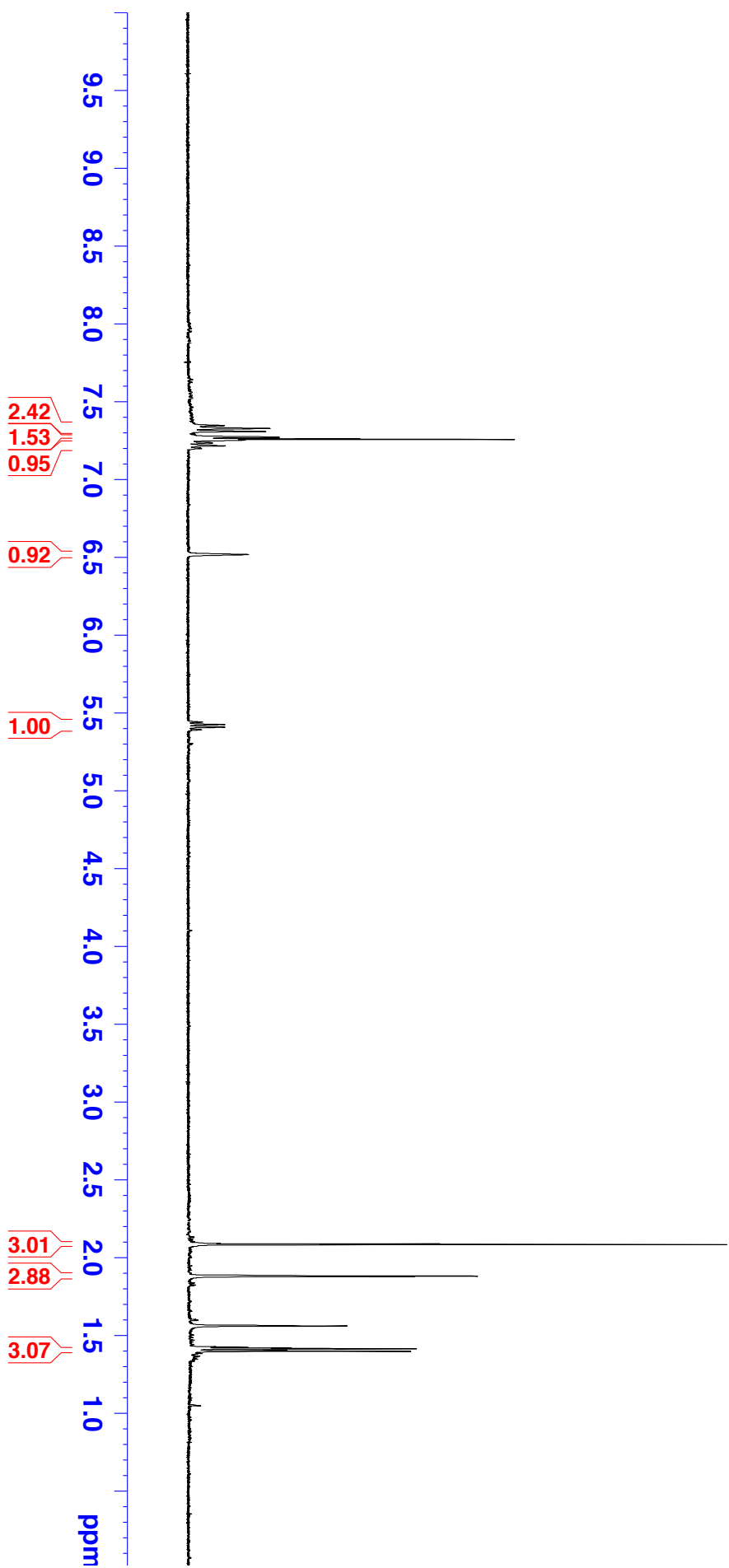
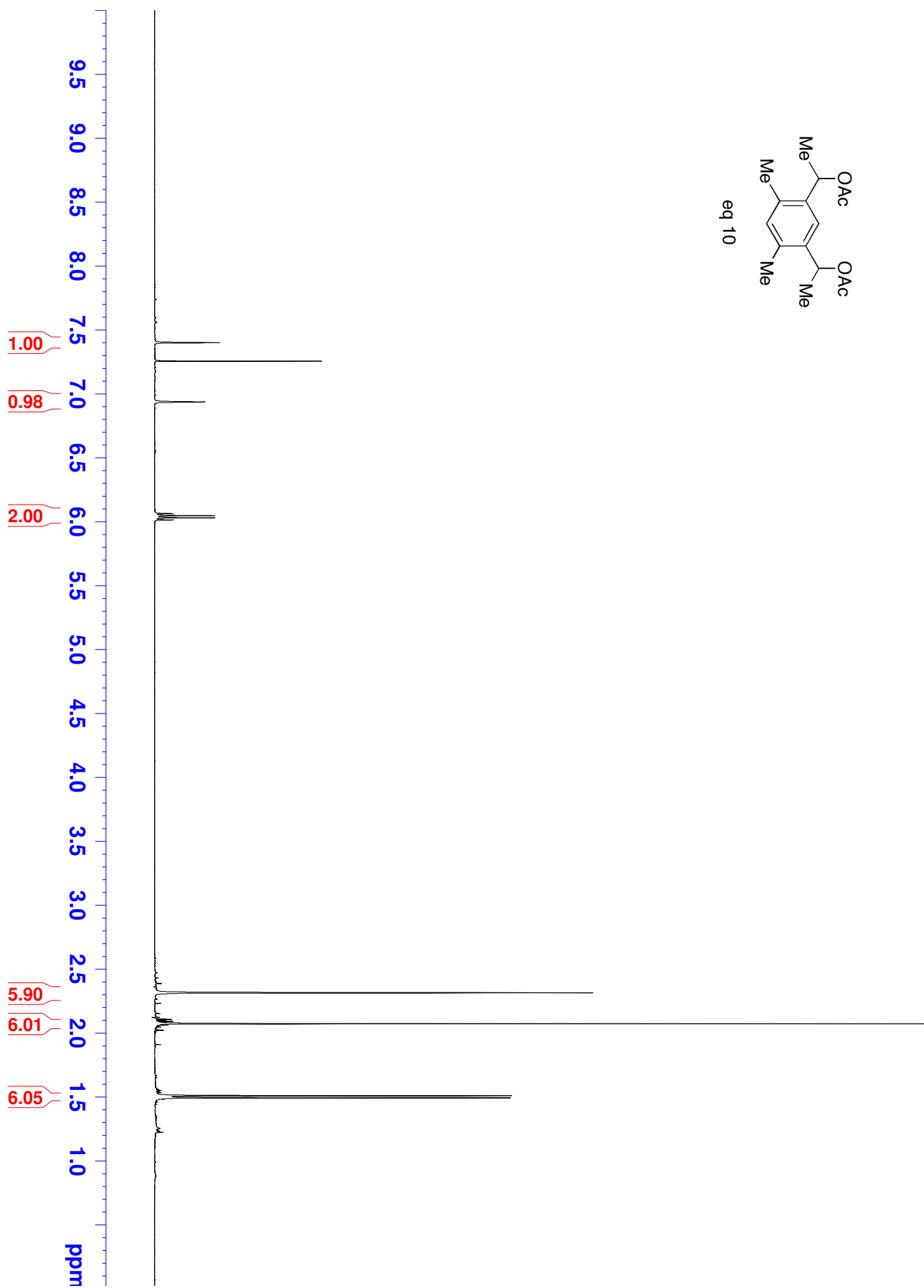
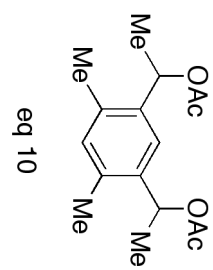
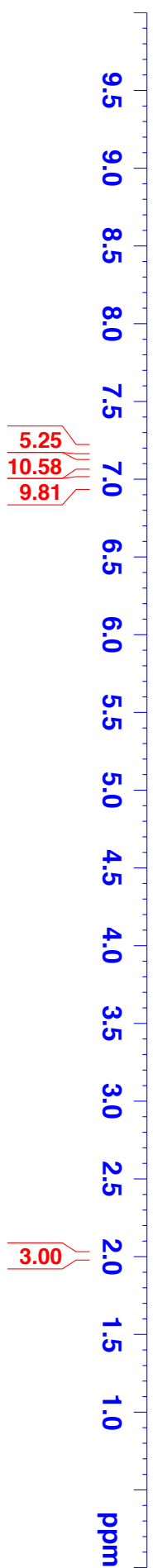
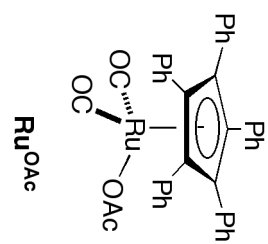
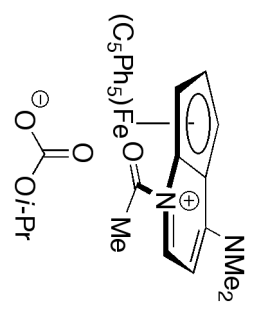


Table 1, entry 10

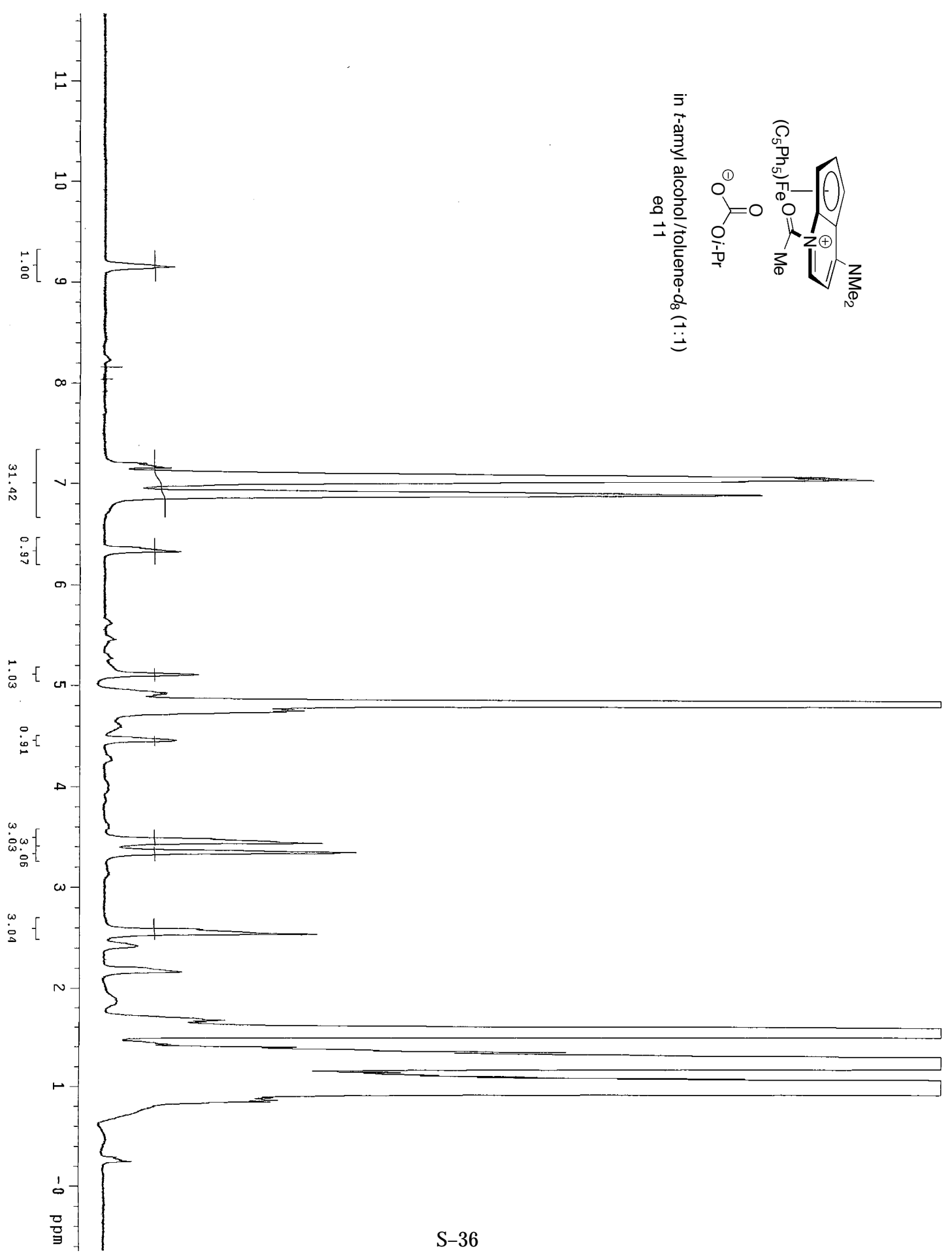




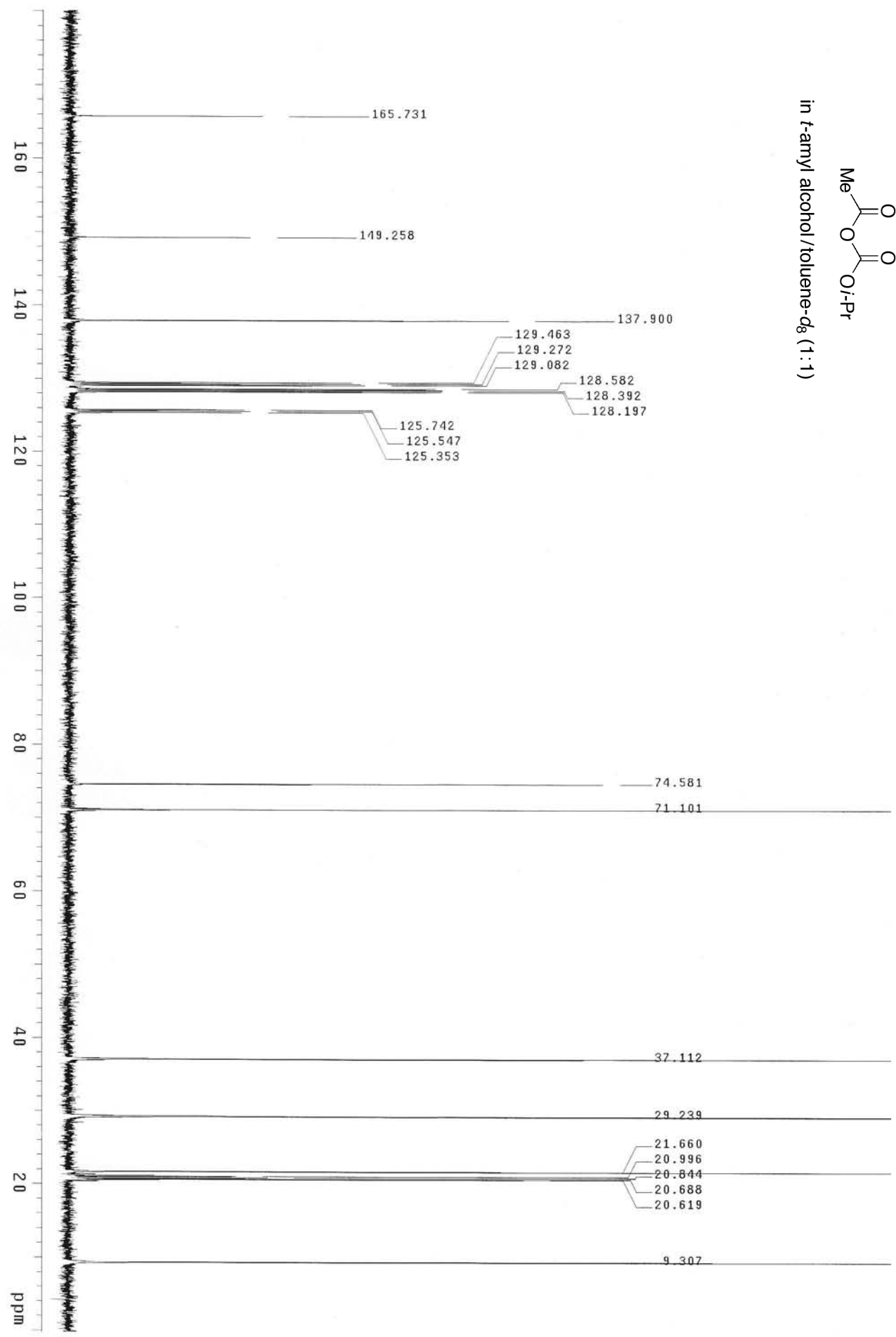
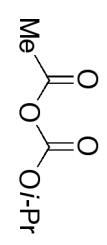


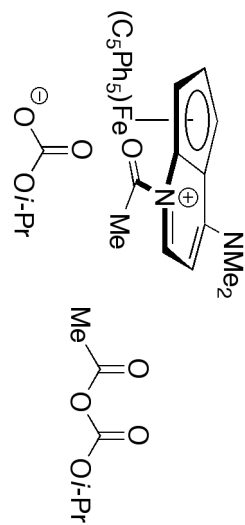


in *t*-amyl alcohol/toluene- d_8 (1:1)
 eq 11

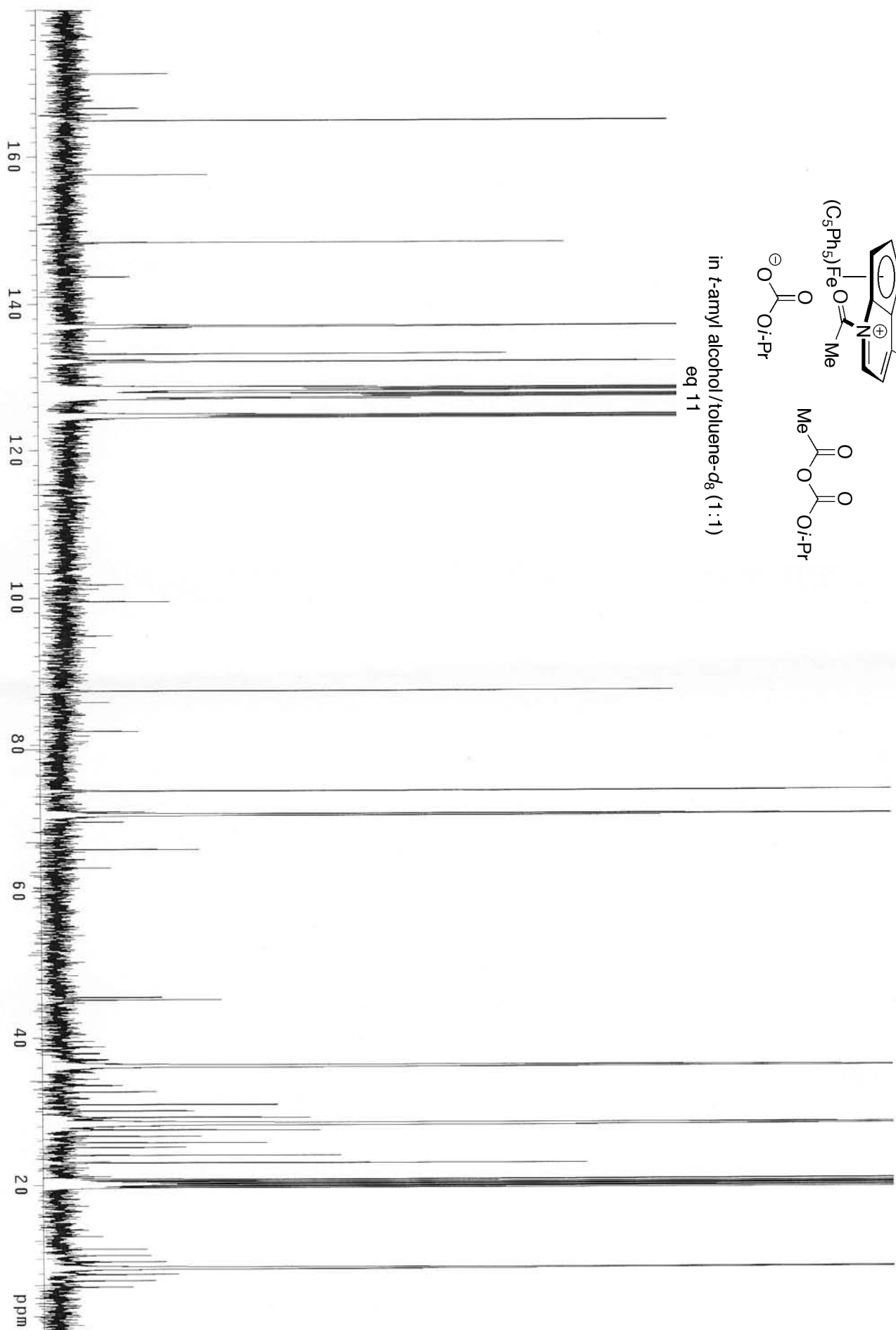


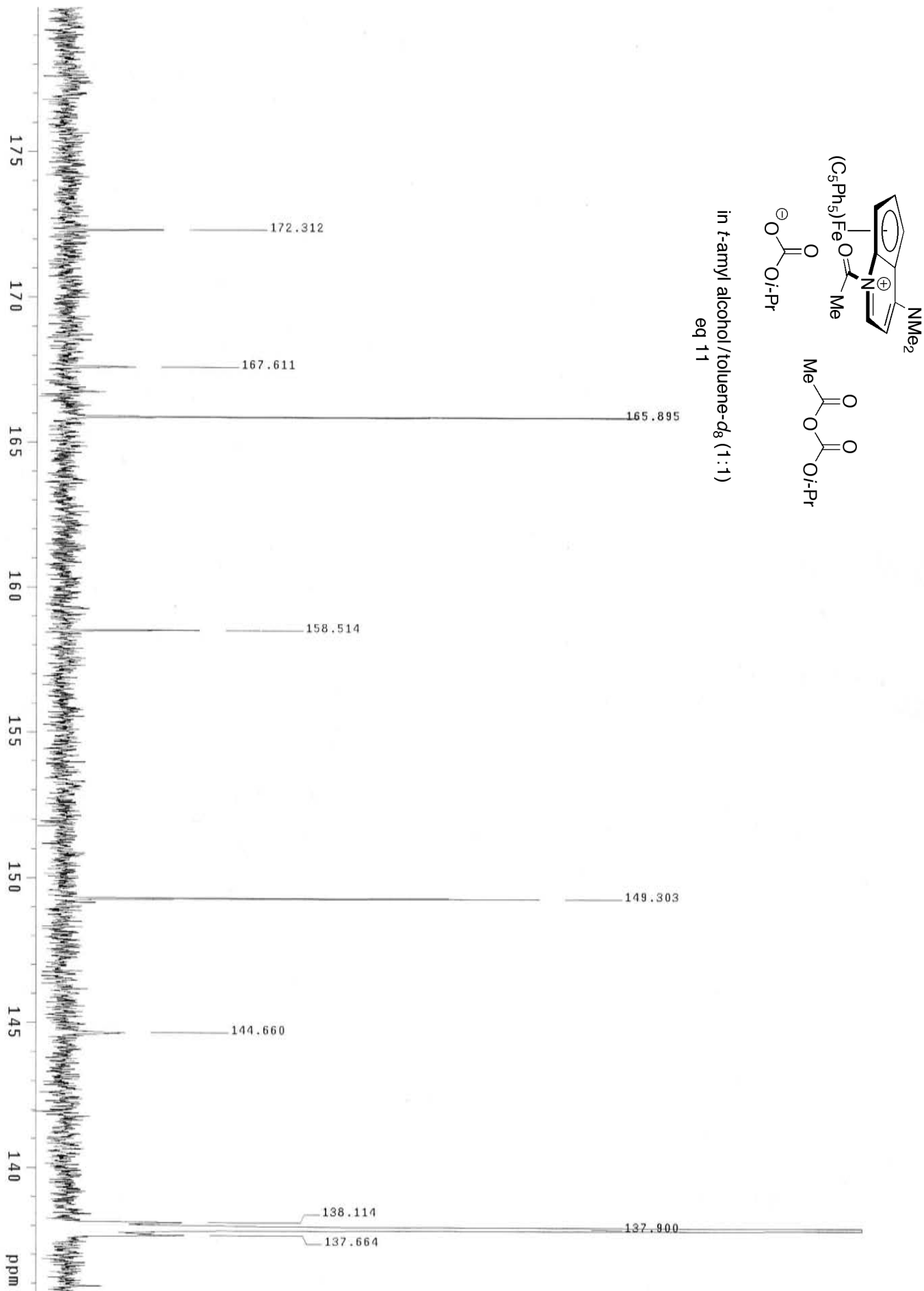
in *t*-amyl alcohol/toluene- d_6 (1:1)





in *t*-amyl alcohol/toluene- d_8 (1:1)
eq 11





```

=====
Injection Date   : 11/7/2011 9:41:11 PM      Seq. Line :    1
Sample Name     :                          Location  : Vial 2
Acq. Operator   : HD                       Inj       :    1
Acq. Instrument : Instrument 3              Inj Volume: 1 µl
Different Inj Volume from Sequence !      Actual Inj Volume : 3 µl
Acq. Method     : C:\HPCHEM\3\METHODS\AU105115.M
Last changed    : 6/10/2008 10:06:18 AM by BJ
Analysis Method : C:\HPCHEM\3\METHODS\SY90115.M
Last changed    : 12/27/2011 5:54:50 PM by HD
                  (modified after loading)
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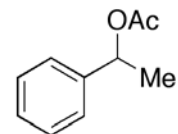
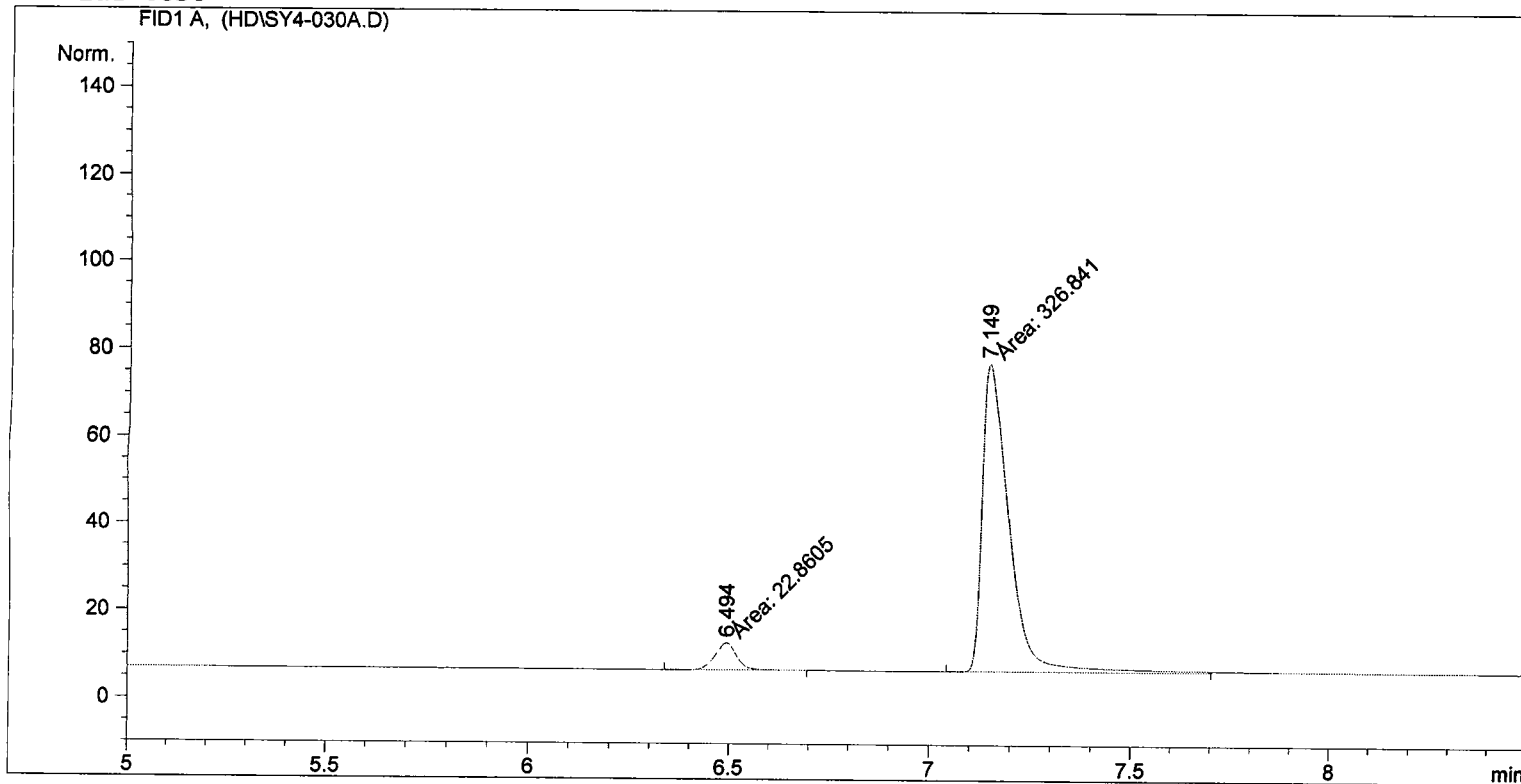


Table 1, entry 1
with (-)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.494	MM	0.0623	22.86055	6.11847	6.53716
2	7.149	MM	0.0771	326.84085	70.63768	93.46284

Totals : 349.70140 76.75615

Results obtained with enhanced integrator!

=====
*** End of Report ***


```

=====
Injection Date   : 11/25/2011 7:08:08 PM          Seq. Line :    3
Sample Name     :                               Location  : Vial 3
Acq. Operator   : SN                           Inj       :    1
Acq. Instrument : Instrument 3                  Inj Volume: 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 3 µl
Acq. Method    : C:\HPCHEM\3\METHODS\AU105115.M
Last changed   : 6/10/2008 10:06:18 AM by BJ
Analysis Method: C:\HPCHEM\3\METHODS\SY90115.M
Last changed   : 12/27/2011 5:51:06 PM by HD
                (modified after loading)
=====

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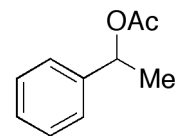
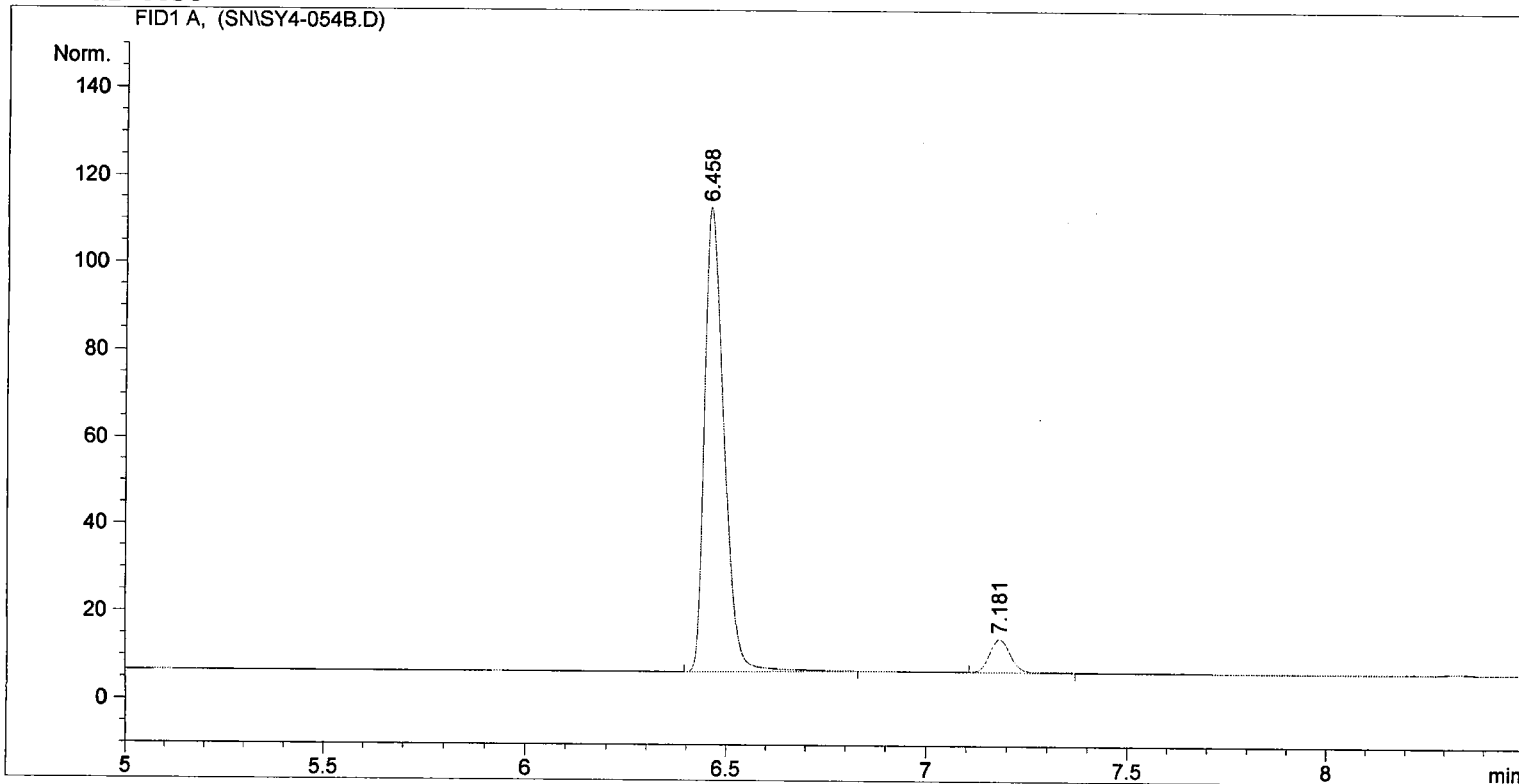


Table 1, entry 1
with (+)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	6.458	PB	0.0513	363.56866	106.53935	93.02377
2	7.181	BB	0.0570	27.26548	7.49099	6.97623

Totals : 390.83414 114.03034

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 12/28/2011 12:09:26 AM      Seq. Line   :    7
Sample Name     :                               Location    : Vial 4
Acq. Operator   : HD                           Inj         :    1
Acq. Instrument : Instrument 3                  Inj Volume  : 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 2 µl
Acq. Method     : C:\HPCHEM\3\METHODS\SY105115.M
Last changed    : 12/27/2011 7:22:35 PM by HD
Analysis Method : C:\HPCHEM\3\METHODS\SY100ISA.M
Last changed    : 12/28/2011 1:37:54 PM by HD
                  (modified after loading)
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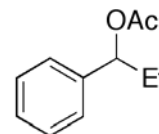
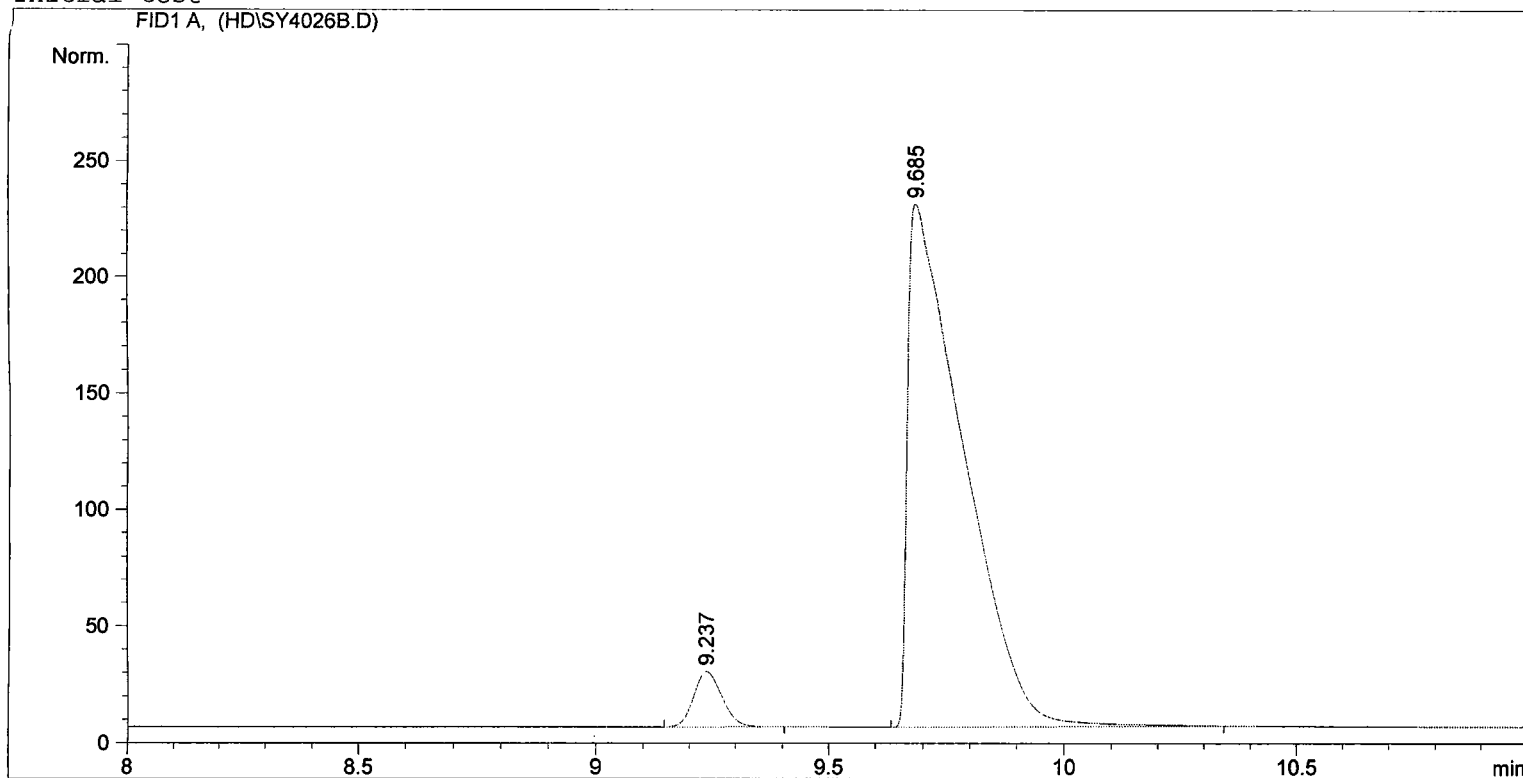


Table 1, entry 2
with (-)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By       :      Signal
Multiplier      :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	9.237	BP	0.0667	100.69203	23.61509	5.23181
2	9.685	PB	0.1042	1823.91992	224.75095	94.76819

Totals : 1924.61195 248.36604

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 11/24/2011 3:48:47 PM      Seq. Line :    8
Sample Name     :                             Location  : Vial 1
Acq. Operator  : HD                          Inj       :    1
Acq. Instrument : Instrument 3                 Inj Volume: 1 µl
Different Inj Volume from Sequence !           Actual Inj Volume : 3 µl
Acq. Method    : C:\HPCHEM\3\METHODS\SY105115.M
Last changed   : 11/24/2011 12:17:48 PM by jc
Analysis Method : C:\HPCHEM\3\METHODS\SY90115.M
Last changed   : 12/27/2011 5:47:31 PM by HD
                (modified after loading)
    
```

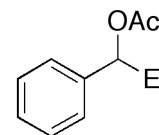
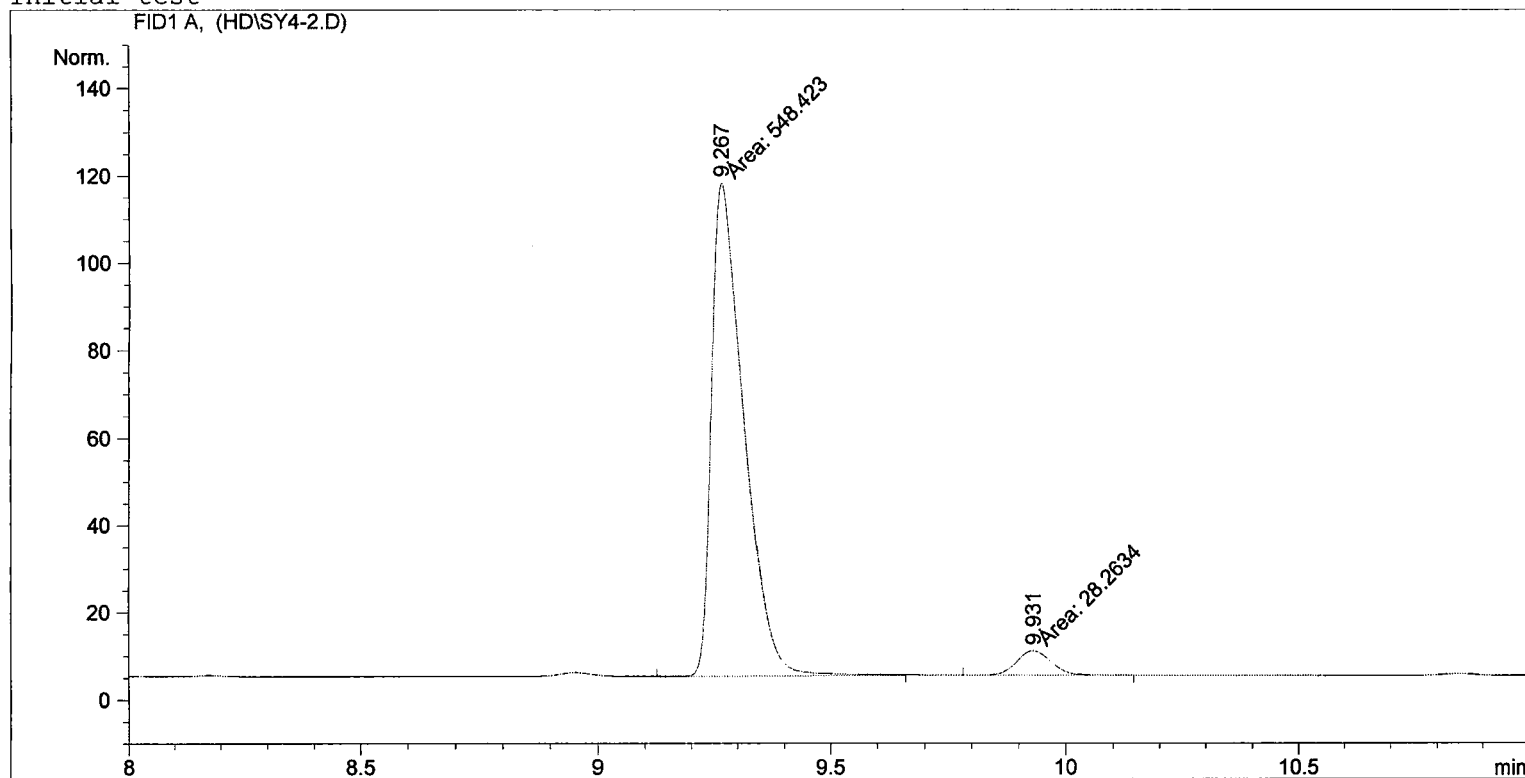


Table 1, entry 2
with (+)-C₅Ph₅-DMAP*

Initial test



Area Percent Report

```

=====
Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	9.267	MM	0.0809	548.42279	112.92822	95.09900
2	9.931	MM	0.0841	28.26339	5.60311	4.90100

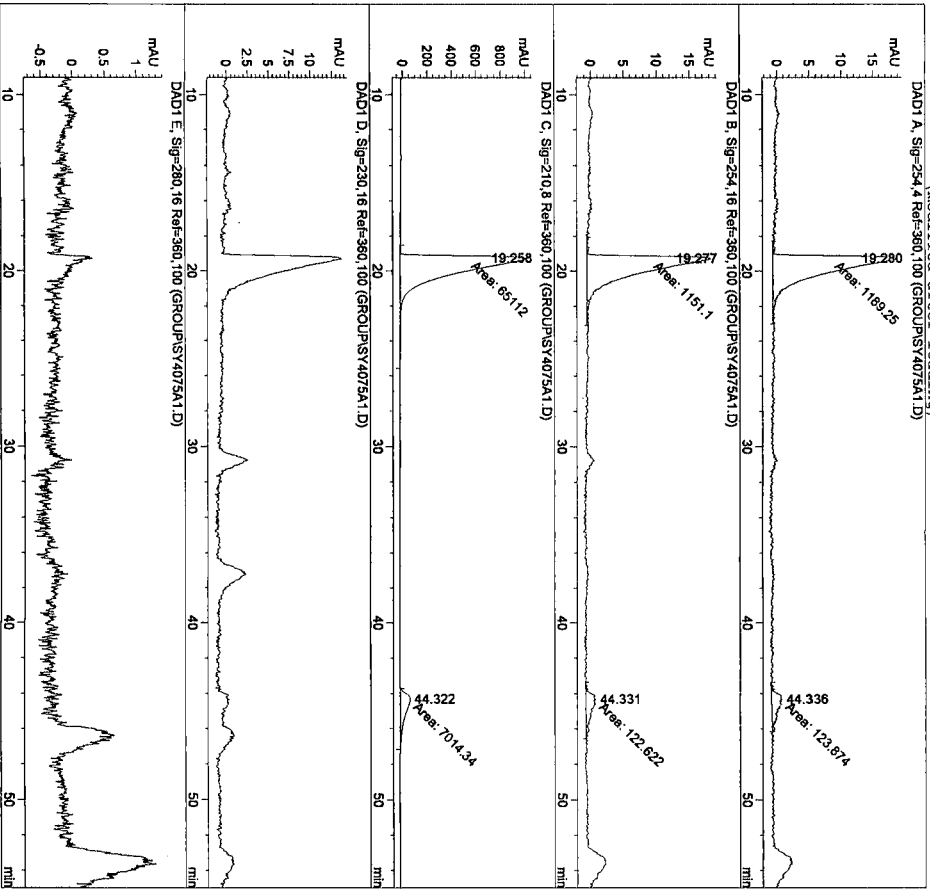
Totals : 576.68618 118.53133

Results obtained with enhanced integrator!

*** End of Report ***

Injection Date : 3/1/2012 1:22:20 AM
 Sample Name : JTM
 Acq. Operator : JTM
 Acq. Instrument : Instrument 1
 Acq. Method : C:\HPCHEM\1\METHODS\ODH-0060.M
 Last changed : 1/19/2012 5:01:32 PM by SN
 Analysis Method : C:\HPCHEM\1\METHODS\YL-AD02.M
 Last changed : 3/1/2012 11:00:43 AM by JTM

Seq. Line : 23
 Location : Vial 73
 Inj Volume : 5 µl



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.280	NM	1.0234	1189.24658	19.36803	90.5665
2	44.336	NM	1.2010	123.87383	1.71897	9.4335
Totals :				1313.12041	21.08699	

Signal 2: DAD1 B, Sig=254,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.277	NM	1.0216	1151.10132	18.77863	90.3730
2	44.331	NM	1.2736	122.62193	1.60466	9.6270
Totals :				1273.72325	20.38329	

Signal 3: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	19.258	NM	1.0545	6.5112064	1029.08911	90.2749
2	44.322	NM	1.3988	7014.33838	83.57714	9.7251
Totals :				7.2126464	1112.66625	

Results obtained with enhanced integrator!
 Signal 4: DAD1 D, Sig=230,16 Ref=360,100
 Signal 5: DAD1 E, Sig=280,16 Ref=360,100

*** End of Report ***

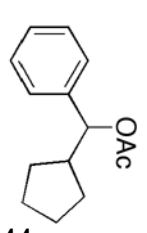
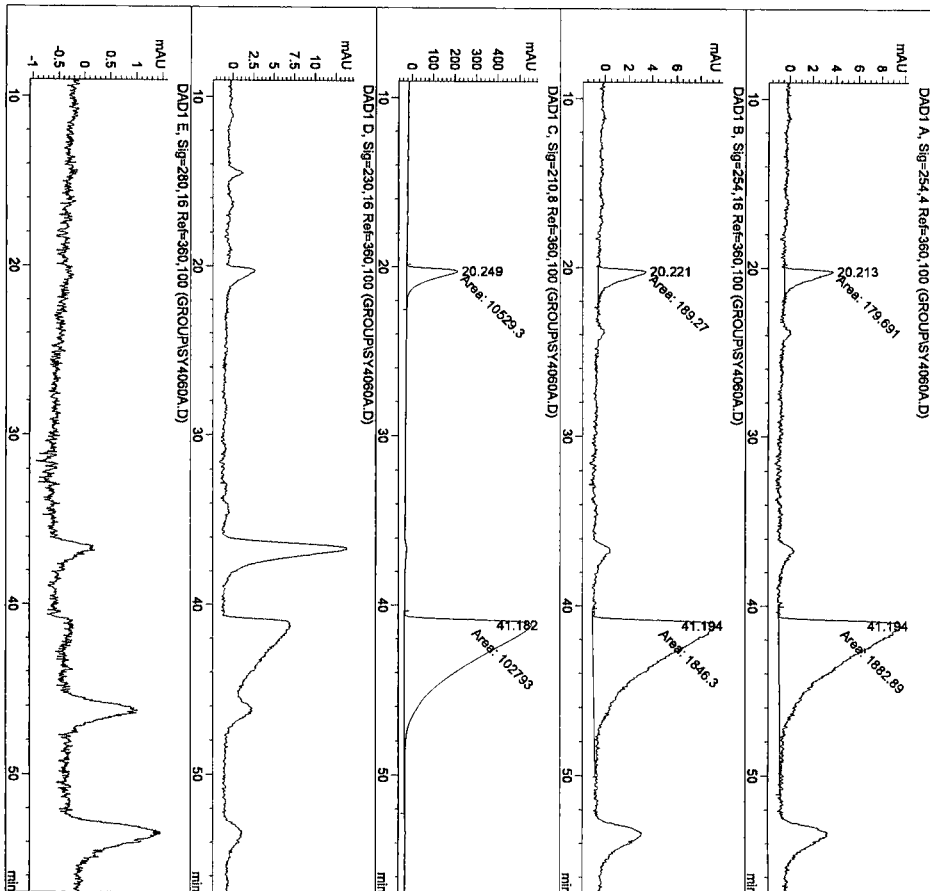


Table 1, entry 3
 with (-)-C₅Ph₅-DMAP*

Injection Date : 2/28/2012 7:45:00 PM
 Sample Name :
 Acq. Operator : JTM
 Acq. Instrument : Instrument 1
 Acq. Method : C:\HPCHEM\1\METHODS\ODH-0060.M
 Last changed : 1/19/2012 5:01:32 PM by SN
 Analysis Method : C:\HPCHEM\1\METHODS\YI-AD02.M
 Last changed : 2/28/2012 8:45:25 PM by JTM

Seq. Line : 6
 Location : Vial 71
 Inj : 1
 Inj Volume : 5 µl



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.213	MM	0.7092	179.69051	4.22263	8.7119
2	41.194	MM	2.9283	1882.89256	10.71659	91.2881
Totals :				2062.58308	14.93922	

Signal 2: DAD1 B, Sig=254,16 Ref=360,100

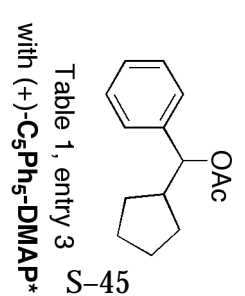
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.221	MM	0.7715	189.27020	4.08862	9.2981
2	41.194	MM	2.9698	1846.30212	10.36168	90.7019
Totals :				2035.57233	14.45030	

Signal 3: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	20.249	MM	0.7388	1.0529364	237.52646	9.2915
2	41.182	MM	2.9146	1.0279365	587.81067	90.7085
Totals :				1.1332265	825.33713	

Results obtained with enhanced integrator!
 Signal 4: DAD1 D, Sig=230,16 Ref=360,100
 Signal 5: DAD1 E, Sig=280,16 Ref=360,100

*** End of Report ***



```

=====
Injection Date   : 12/28/2011 12:45:56 AM      Seq. Line   :    8
Sample Name     :                               Location    : Vial 5
Acq. Operator   : HD                           Inj         :    1
Acq. Instrument : Instrument 3                  Inj Volume  : 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 2 µl
Acq. Method     : C:\HPCHEM\3\METHODS\SY105115.M
Last changed    : 12/27/2011 7:22:35 PM by HD
Analysis Method : C:\HPCHEM\3\METHODS\SY100ISA.M
Last changed    : 12/28/2011 1:39:13 PM by HD
                  (modified after loading)
    
```

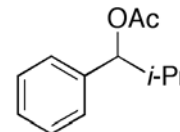
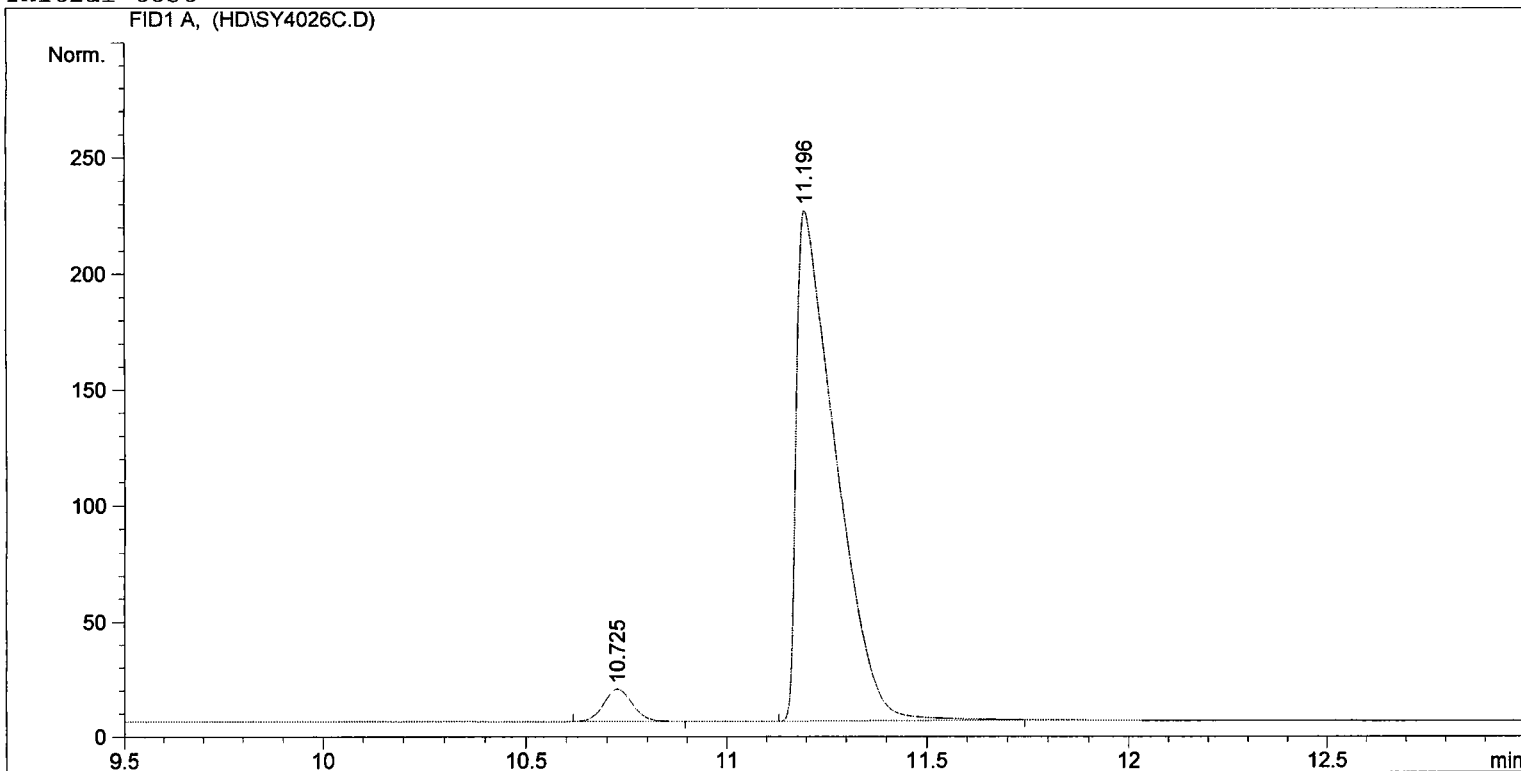


Table 1, entry 4
with (-)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution      :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	10.725	BB	0.0768	69.37827	14.00787	4.39299
2	11.196	PB	0.0933	1509.91589	220.63409	95.60701

Totals : 1579.29417 234.64197

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 12/10/2011 7:18:42 PM      Seq. Line   : 11
Sample Name     :                               Location    : Vial 2
Acq. Operator  : hd                            Inj         : 1
Acq. Instrument : Instrument 3                  Inj Volume  : 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 3 µl
Acq. Method    : C:\HPCHEM\3\METHODS\SY105115.M
Last changed   : 11/24/2011 12:17:48 PM by jc
Analysis Method : C:\HPCHEM\3\METHODS\SY90115.M
Last changed   : 12/27/2011 6:04:59 PM by HD
                (modified after loading)
    
```

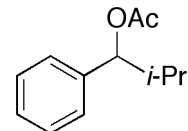
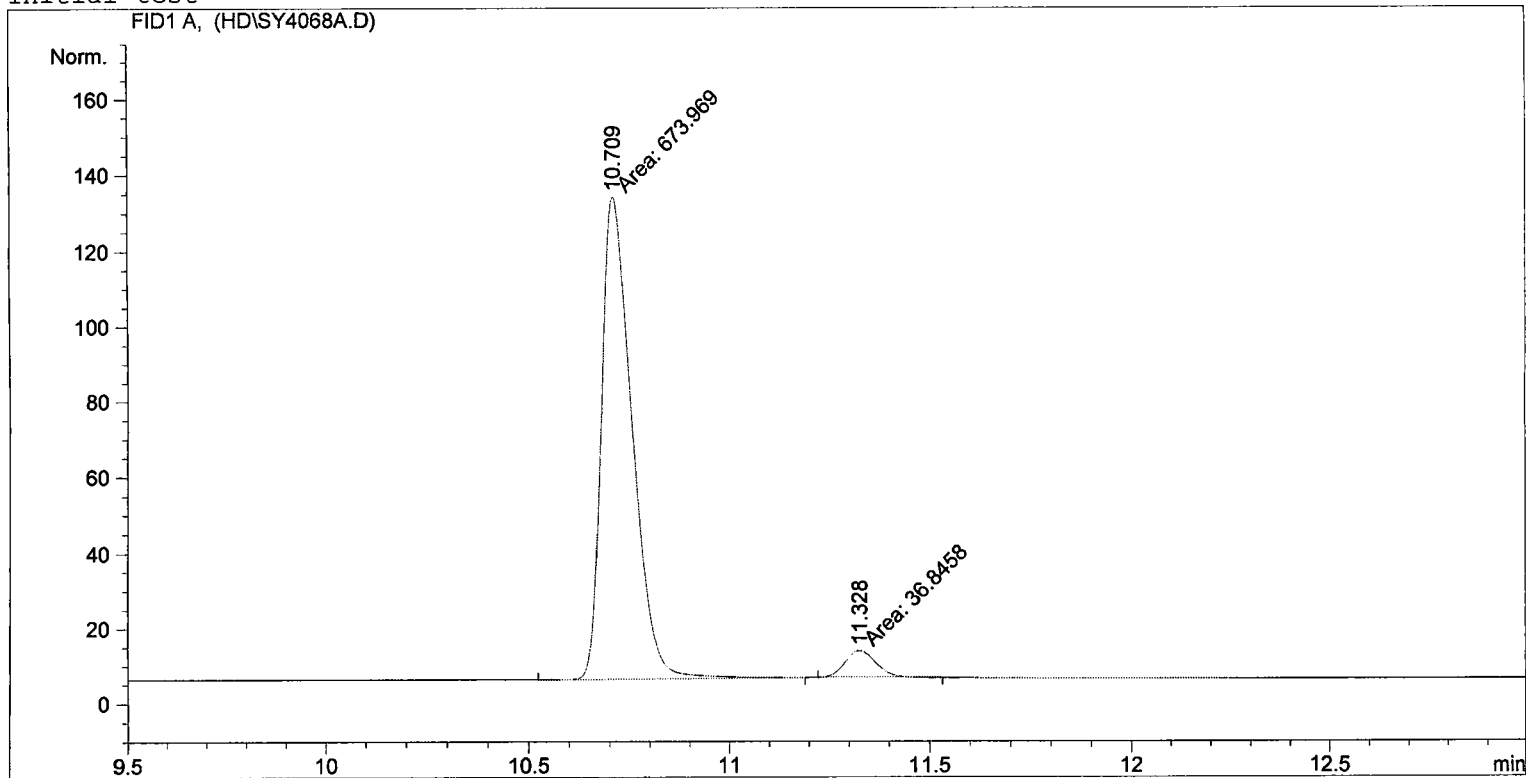


Table 1, entry 4
with (+)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	10.709	MM	0.0879	673.96893	127.77182	94.81639
2	11.328	MM	0.0871	36.84584	7.04818	5.18361

Totals : 710.81477 134.82000

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date : 12/28/2011 4:09:40 PM      Seq. Line : 11
Sample Name   :                               Location  : Vial 3
Acq. Operator : HD                           Inj       : 1
Acq. Instrument : Instrument 3                Inj Volume: 1 µl
Different Inj Volume from Sequence !      Actual Inj Volume : 2 µl
Acq. Method   : C:\HPCHEM\3\METHODS\AU105115.M
Last changed  : 6/10/2008 10:06:18 AM by BJ
Analysis Method : C:\HPCHEM\3\METHODS\SY100ISA.M
Last changed  : 12/28/2011 6:10:33 PM by HD
                (modified after loading)
=====
    
```

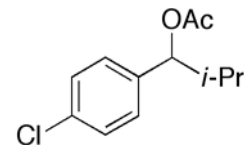
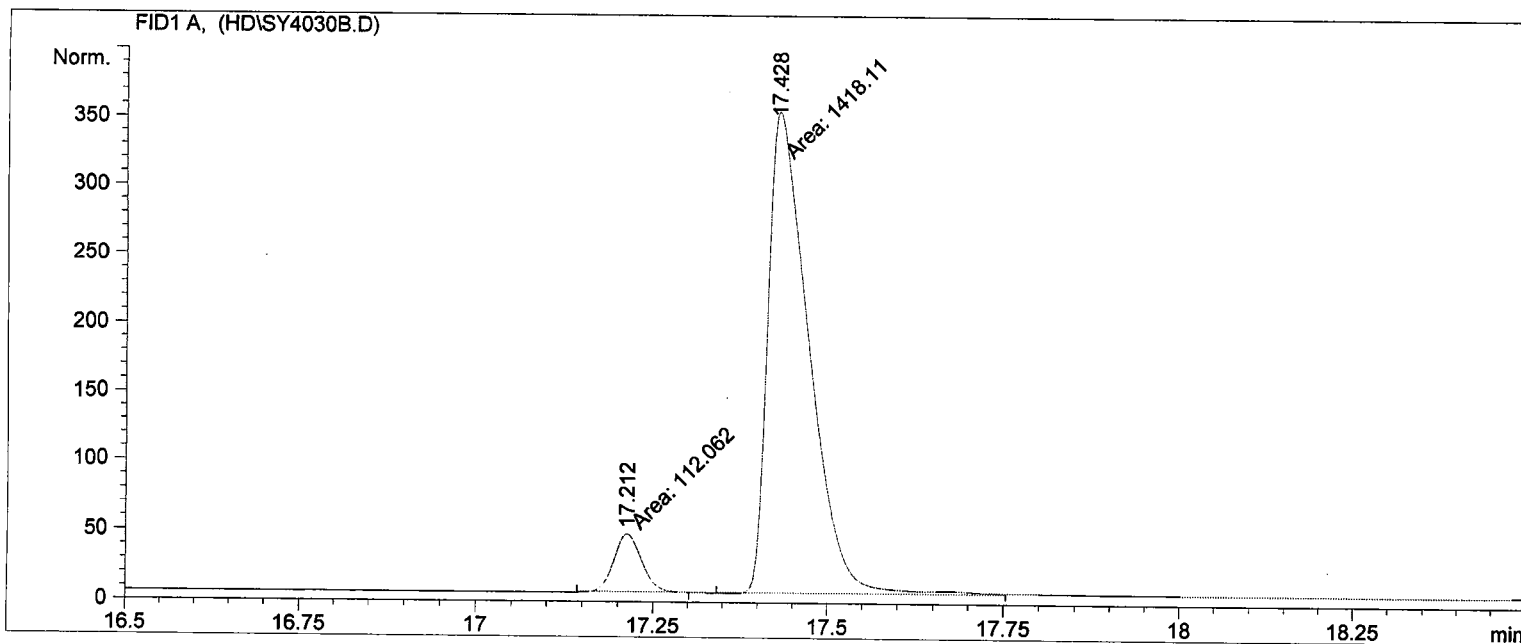


Table 1, entry 5
with (-)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	17.212	MM	0.0453	112.06217	41.26473	7.32348
2	17.428	MM	0.0678	1418.11487	348.77933	92.67652

Totals : 1530.17704 390.04406

Results obtained with enhanced integrator!

=====
Summed Peaks Report
=====

Signal 1: FID1 A,

=====
Final Summed Peaks Report
=====

Signal 1: FID1 A,


```

=====
Injection Date   : 12/2/2011 2:16:51 PM      Seq. Line   :    4
Sample Name     :                          Location    : Vial 2
Acq. Operator   : CC                        Inj         :    1
Acq. Instrument : Instrument 3               Inj Volume  : 1 µl
Different Inj Volume from Sequence !      Actual Inj Volume : 3 µl
Acq. Method     : C:\HPCHEM\3\METHODS\AU105115.M
Last changed    : 6/10/2008 10:06:18 AM by BJ
Analysis Method : C:\HPCHEM\3\METHODS\SY90115.M
Last changed    : 12/27/2011 6:15:42 PM by HD
                  (modified after loading)
=====
    
```

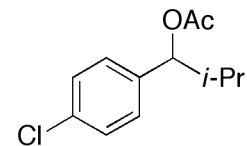
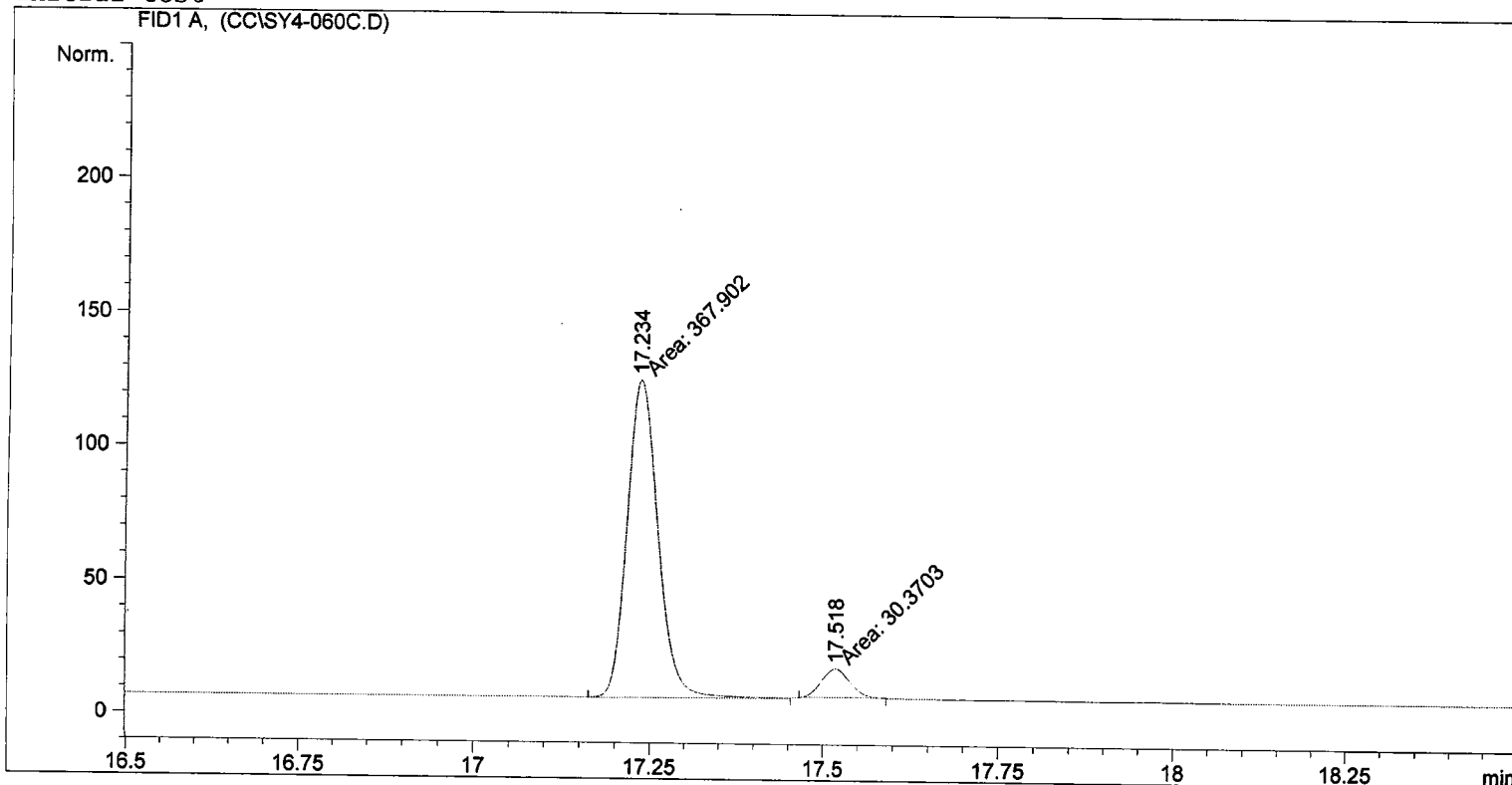


Table 1, entry 5
with (+)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	17.234	MM	0.0516	367.90179	118.76257	92.37450
2	17.518	MM	0.0468	30.37025	10.82276	7.62550

Totals : 398.27205 129.58533

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 11/25/2011 8:01:16 PM      Seq. Line   :    5
Sample Name     :                               Location    : Vial 2
Acq. Operator   : SN                          Inj         :    1
Acq. Instrument : Instrument 3                 Inj Volume  : 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 3 µl
Acq. Method     : C:\HPCHEM\3\METHODS\SY105115.M
Last changed    : 11/24/2011 12:17:48 PM by jc
Analysis Method : C:\HPCHEM\3\METHODS\COOLDOWN.M
Last changed    : 4/19/2012 3:17:34 PM by hc
                  (modified after loading)
=====
    
```

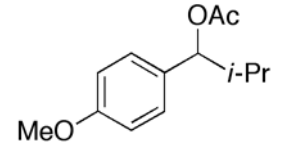
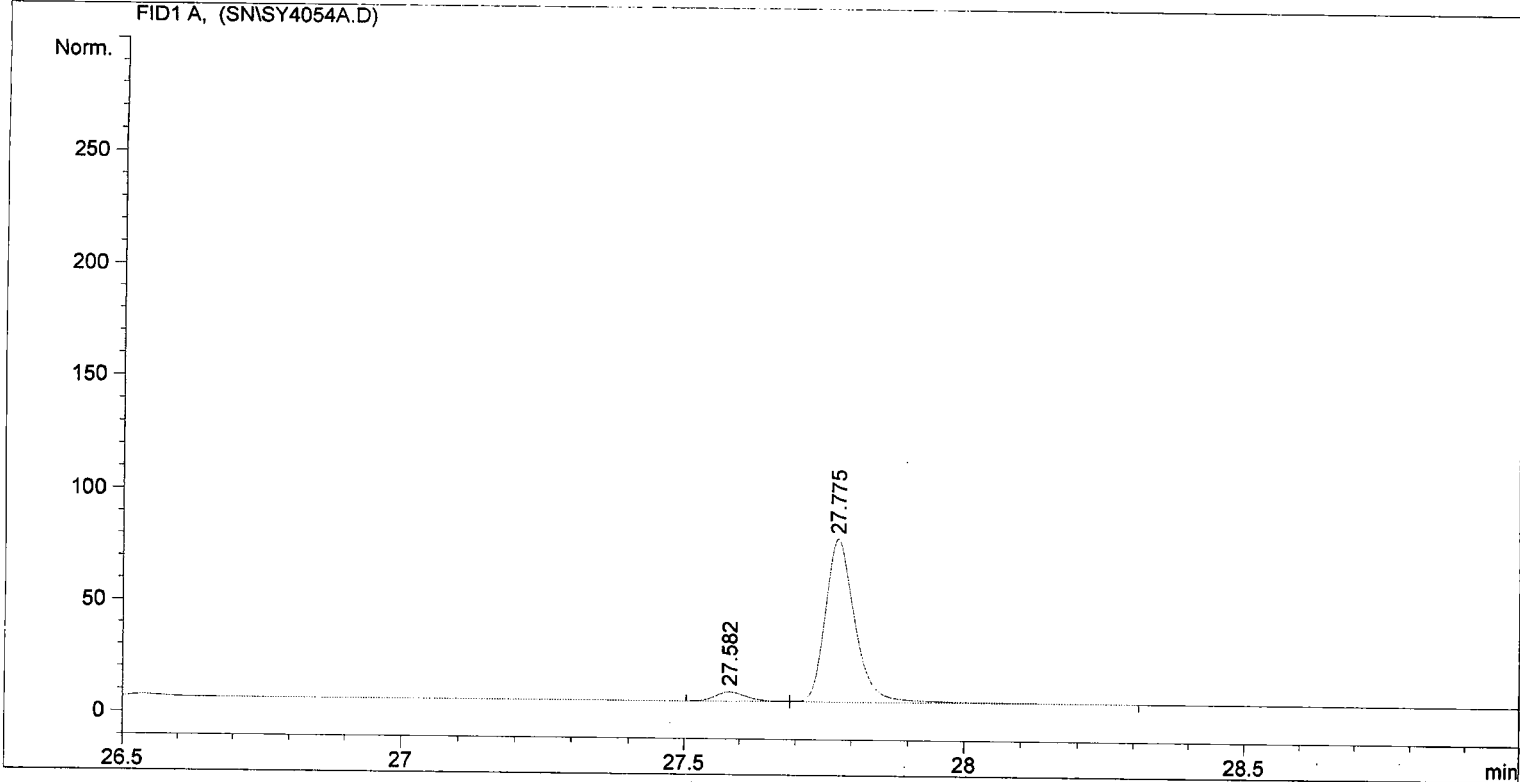


Table 1, entry 6
with (-)-C₅Ph₅-DMAP*

standby



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	27.582	PV	0.0603	16.20505	4.08384	5.61661
2	27.775	VB	0.0557	272.31488	72.77936	94.38339

Totals : 288.51993 76.86319

Results obtained with enhanced integrator!

=====
Summed Peaks Report
=====

Signal 1: FID1 A,

```

=====
Injection Date   : 12/3/2011 12:36:06 PM      Seq. Line   :    9
Sample Name     :                               Location    : Vial 2
Acq. Operator   : CC                          Inj         :    1
Acq. Instrument : Instrument 3                 Inj Volume  : 1 µl
Different Inj Volume from Sequence !         Actual Inj Volume : 3 µl
Acq. Method     : C:\HPCHEM\3\METHODS\SY105115.M
Last changed    : 11/24/2011 12:17:48 PM by jc
Analysis Method : C:\HPCHEM\3\METHODS\SY90115.M
Last changed    : 12/27/2011 6:23:02 PM by HD
                  (modified after loading)
    
```

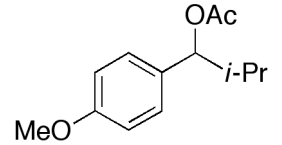
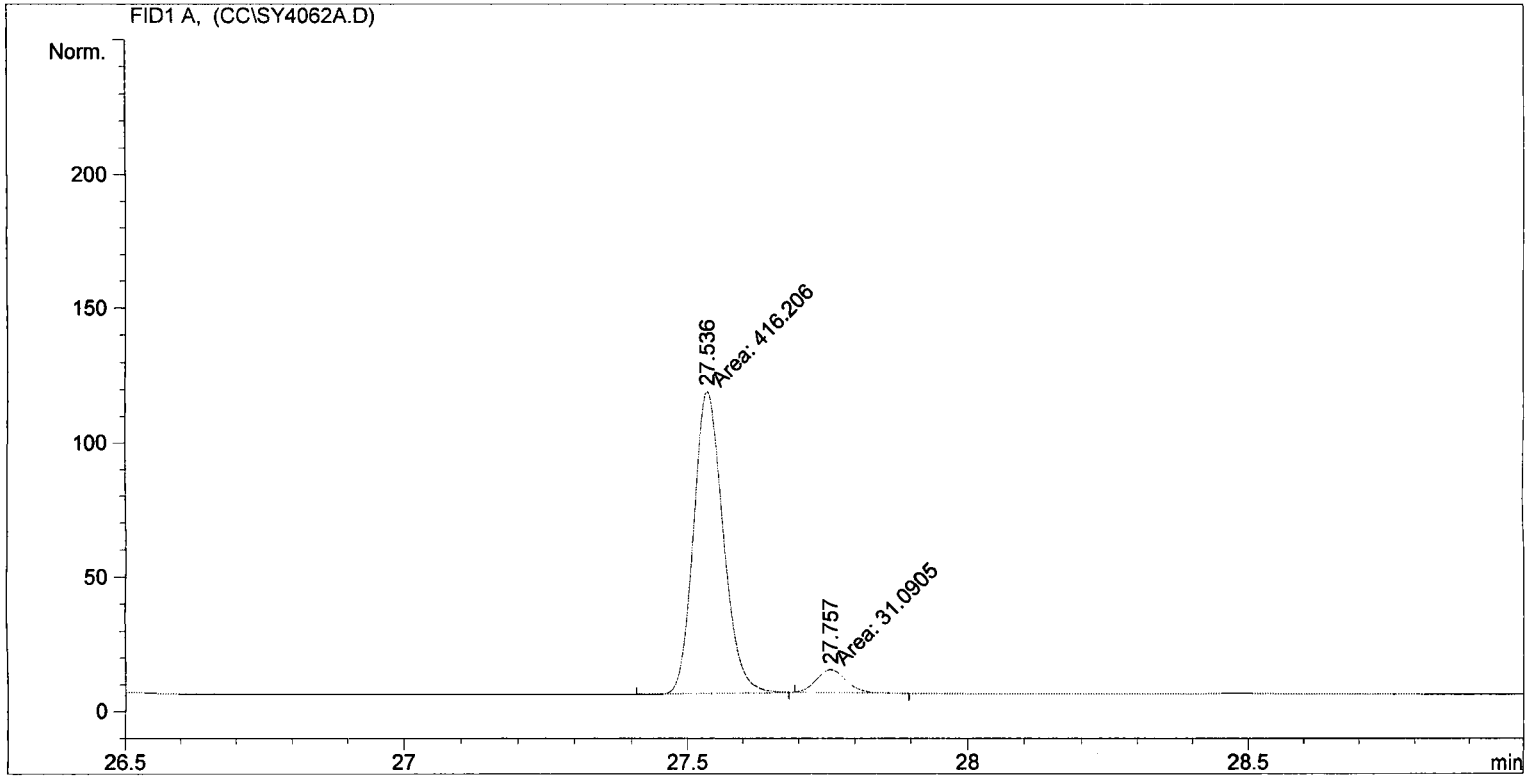


Table 1, entry 6
with (+)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By       :      Signal
Multiplier      :      1.0000
Dilution        :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	27.536	MM	0.0617	416.20593	112.49719	93.04924
2	27.757	MM	0.0609	31.09052	8.51014	6.95076

Totals : 447.29645 121.00733

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 12/28/2011 4:36:13 PM      Seq. Line   : 12
Sample Name     :                               Location    : Vial 4
Acq. Operator  : HD                           Inj         : 1
Acq. Instrument : Instrument 3                 Inj Volume  : 1 µl
Different Inj Volume from Sequence !         Actual Inj Volume : 2 µl
Acq. Method    : C:\HPCHEM\3\METHODS\SY105115.M
Last changed   : 12/27/2011 7:22:35 PM by HD
Analysis Method : C:\HPCHEM\3\METHODS\SY100ISA.M
Last changed   : 12/28/2011 6:12:44 PM by HD
                (modified after loading)
    
```

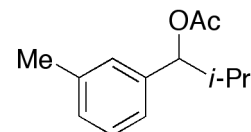
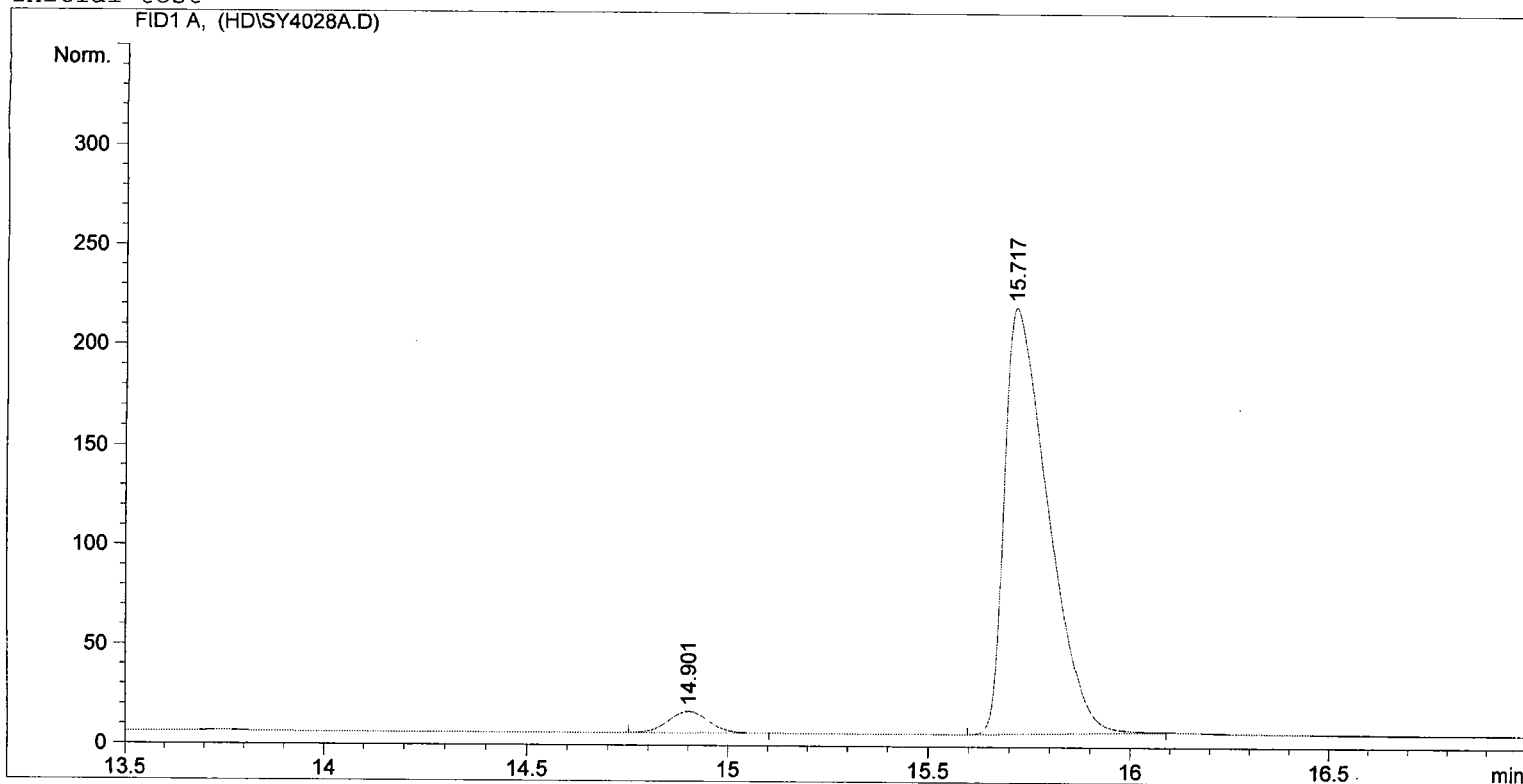


Table 1, entry 7
with (-)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      : Signal
Multiplier    : 1.0000
Dilution      : 1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	14.901	BB	0.1087	75.04472	10.64902	4.48550
2	15.717	PB	0.1104	1598.00769	213.32225	95.51450

Totals : 1673.05241 223.97127

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date : 12/28/2011 5:12:40 PM      Seq. Line : 13
Sample Name    :                               Location  : Vial 5
Acq. Operator  : HD                           Inj       : 1
Acq. Instrument : Instrument 3                 Inj Volume: 1 µl
Different Inj Volume from Sequence !         Actual Inj Volume : 2 µl
Acq. Method    : C:\HPCHEM\3\METHODS\SY105115.M
Last changed   : 12/27/2011 7:22:35 PM by HD
Analysis Method : C:\HPCHEM\3\METHODS\SY100ISA.M
Last changed   : 12/28/2011 6:15:38 PM by HD
                (modified after loading)
    
```

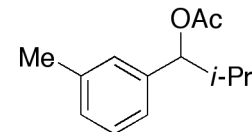
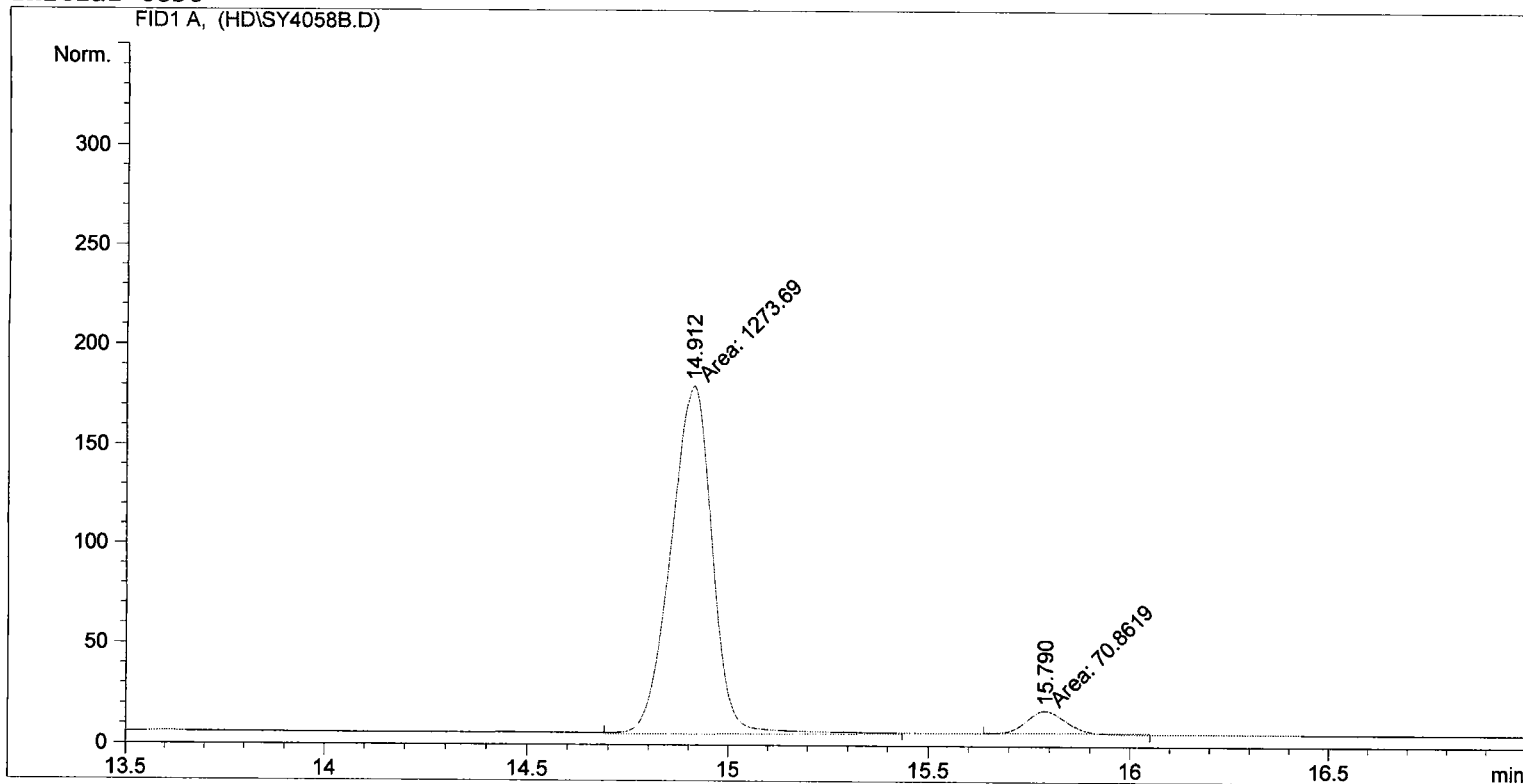


Table 1, entry 7
with (+)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      : Signal
Multiplier     : 1.0000
Dilution       : 1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	14.912	MM	0.1215	1273.69214	174.75676	94.72971
2	15.790	MM	0.1074	70.86195	10.99810	5.27029

Totals : 1344.55408 185.75486

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 12/27/2011 6:22:37 PM      Seq. Line   :    3
Sample Name     :                               Location    : Vial 5
Acq. Operator   : HD                           Inj         :    1
Acq. Instrument : Instrument 3                  Inj Volume  : 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 3 µl
Acq. Method     : C:\HPCHEM\3\METHODS\SY90115.M
Last changed    : 12/27/2011 3:02:56 PM by HD
Analysis Method : C:\HPCHEM\3\METHODS\SY105130.M
Last changed    : 12/27/2011 7:18:21 PM by HD
                  (modified after loading)
=====

```

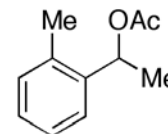
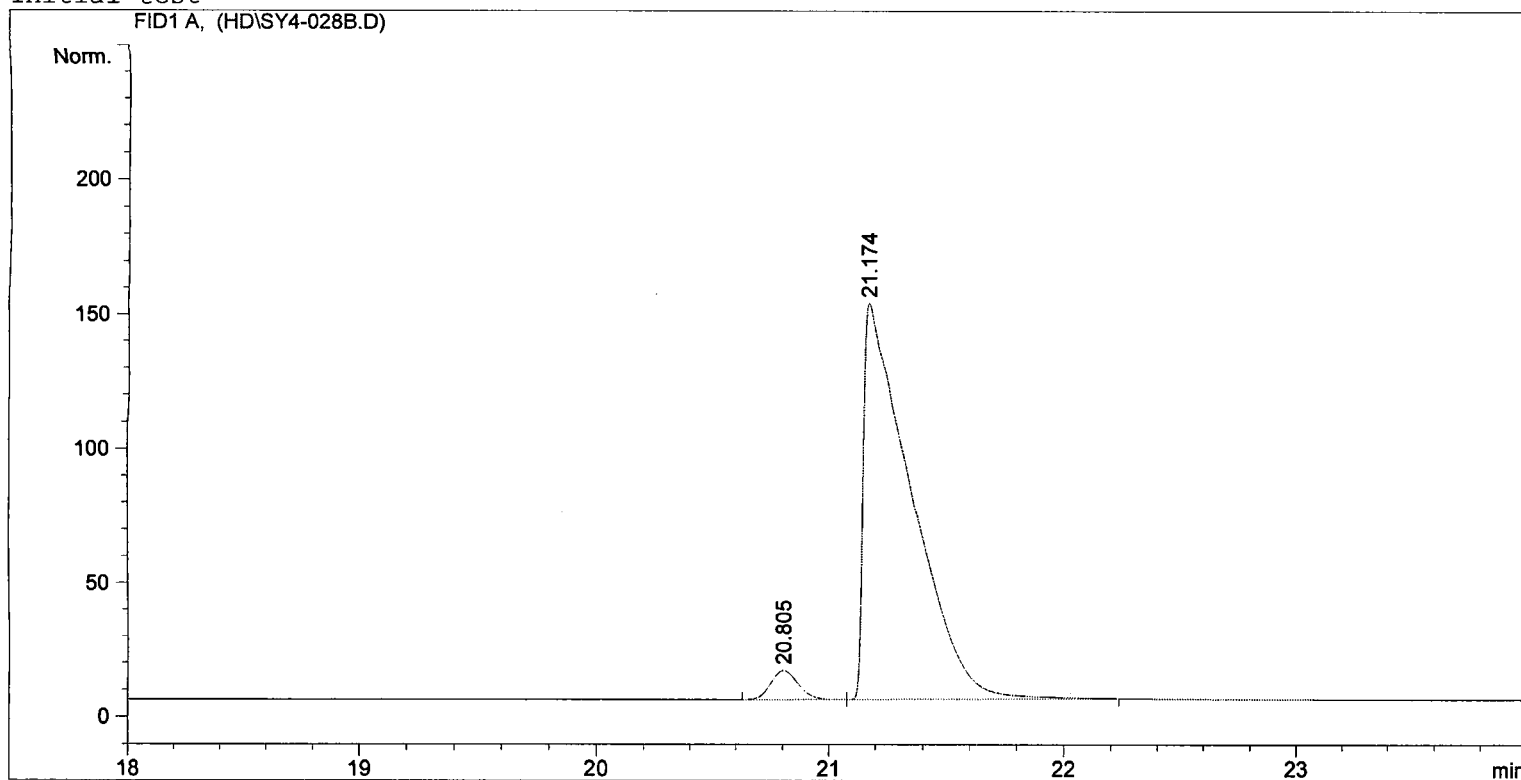


Table 1, entry 8
with (-)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By       :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs

```

Signal 1: FID1 A,

Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	20.805	PP	0.1294	88.95095	10.76215	4.09658
2	21.174	VB	0.1783	2082.39575	147.86237	95.90342

Totals : 2171.34670 158.62451

Results obtained with enhanced integrator!

=====
*** End of Report ***

```

=====
Injection Date   : 12/27/2011 8:26:19 PM      Seq. Line   :    1
Sample Name     :                               Location    : Vial 3
Acq. Operator   : HD                          Inj         :    1
Acq. Instrument : Instrument 3                 Inj Volume  : 1 µl
Different Inj Volume from Sequence !          Actual Inj Volume : 3 µl
Acq. Method     : C:\HPCHEM\3\METHODS\SY90115.M
Last changed    : 12/27/2011 3:02:56 PM by HD
Analysis Method : C:\HPCHEM\3\METHODS\SY100ISA.M
Last changed    : 12/28/2011 1:42:35 PM by HD
                  (modified after loading)
=====
    
```

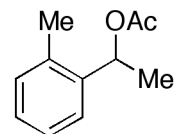
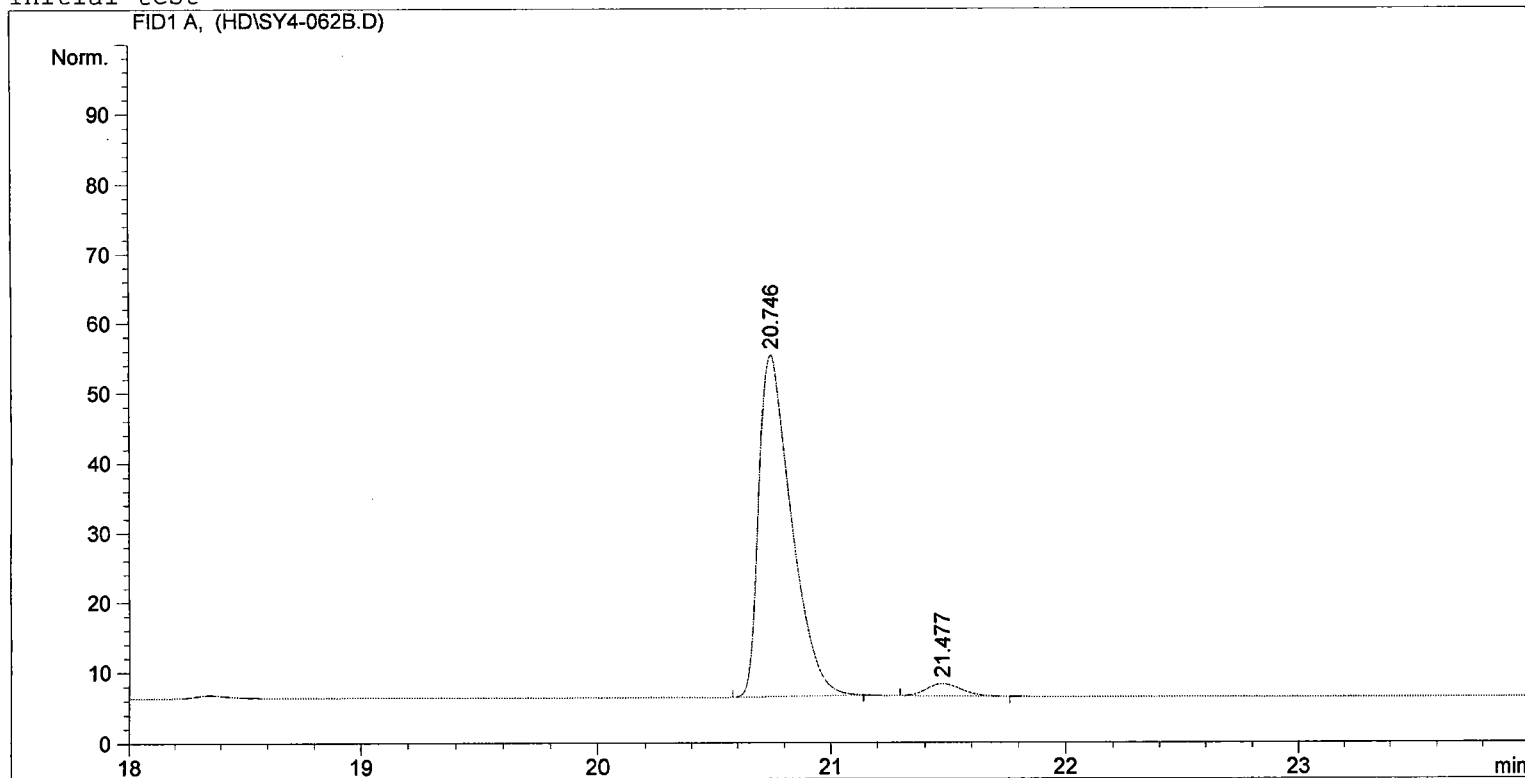


Table 1, entry 8
with (+)-C₅Ph₅-DMAP*

Initial test



=====
Area Percent Report
=====

```

Sorted By      :      Signal
Multiplier     :      1.0000
Dilution       :      1.0000
Use Multiplier & Dilution Factor with ISTDs
    
```

Signal 1: FID1 A,

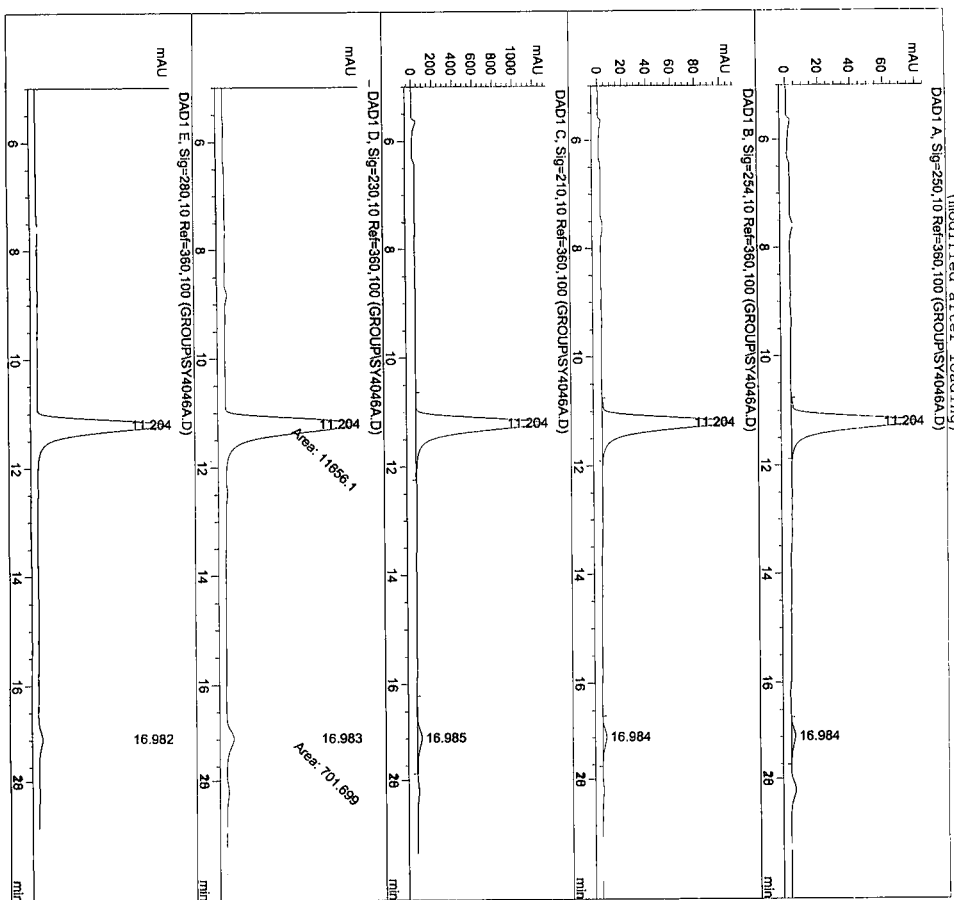
Peak #	RetTime [min]	Type	Width [min]	Area [pA*s]	Height [pA]	Area %
1	20.746	PB	0.1371	471.79797	48.86399	96.36722
2	21.477	BP	0.1221	17.78549	1.73504	3.63278

Totals : 489.58346 50.59904

Results obtained with enhanced integrator!

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*** End of Report ***

Injection Date : 11/25/2011 7:03:54 PM
 Sample Name : JTM
 Acq. Operator : JTM
 Acq. Instrument : Instrument 1
 Different Inj Volume from Sequence : Actual Inj Volume : 15 µl
 Inj Volume : 1 µl
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01-60.M
 Last changed : 11/25/2011 4:57:32 PM by NB
 Analysis Method : C:\HPCHEM\1\METHODS\VL-AD08.M
 Last changed : 12/27/2011 7:49:48 PM by JTM
 (modified after loading)



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.204	BB	0.2482	1293.14844	78.50520	95.8974
2	16.984	PB	0.3429	55.32301	2.41821	4.1026
Totals :				1348.47145	80.92342	

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.204	BB	0.2474	1709.65356	104.23670	95.7928
2	16.984	PB	0.3439	75.08683	3.19879	4.2072
Totals :				1784.74039	107.43549	

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.204	VB	0.2630	2.1460164	1245.90479	94.7669
2	16.985	VB	0.3941	1185.03430	45.16370	5.2331
Totals :				2.26451e4	1291.06848	

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.204	NM	0.3314	1.16561e4	586.18079	94.3218
2	16.983	NM	0.3784	701.69916	30.90495	5.6782
Totals :				1.23578e4	617.08573	

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	11.204	BB	0.2462	4694.26367	287.88928	95.8319
2	16.982	BB	0.3586	204.17297	8.73924	4.1681
Totals :				4898.43665	296.62852	

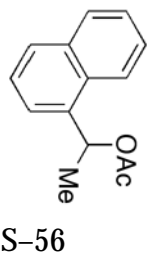
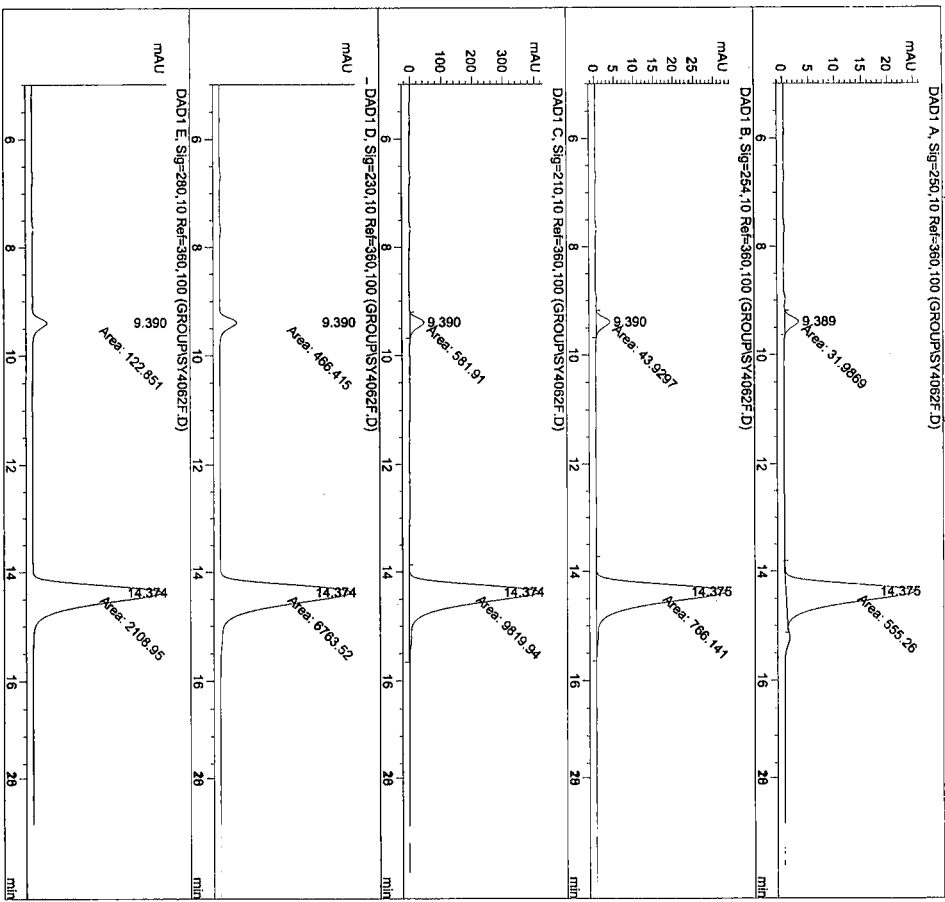


Table 1, entry 9
 with (-)-C₉Ph₅-DMAP*

Injection Date : 1/13/2012 2:31:26 PM
 Sample Name : JTM
 Acq. Operator : Instrument 1
 Acq. Instrument : Instrument 1
 Different Inj Volume from Sequence : Actual Inj Volume : 15 µl
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01-40.M
 Last Changed : 8/17/2010 4:39:48 PM by JTM
 Analysis Method : C:\HPCHEM\1\METHODS\OD-10-40.M
 Last Changed : 4/16/2012 1:52:26 PM by CE
 (modified after loading)

Seq. Line : 2
 Location : Vial 1
 Inj : 1
 Inj Volume : 15 µl



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTRS

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.389	MM	0.2057	31.98688	2.59194	5.4469
2	14.375	MM	0.3774	555.26025	24.52152	94.5531
Totals :				587.24713	27.11346	

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.390	MM	0.2120	43.92972	3.45371	5.4229
2	14.375	MM	0.3905	766.14087	32.69531	94.5771
Totals :				810.07059	36.14901	

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.390	MM	0.2145	581.91028	45.20829	5.5943
2	14.374	MM	0.3936	9819.93750	415.82935	94.4057
Totals :				1.04018e4	461.03764	

Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.390	MM	0.2201	466.41537	35.31659	6.4512
2	14.374	MM	0.4124	6763.52100	273.33804	93.5488
Totals :				7229.93637	308.65463	

Signal 5: DAD1 E, Sig=280,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	9.390	MM	0.2165	122.85142	9.45809	5.5046
2	14.374	MM	0.3907	2108.95215	89.95865	94.4954
Totals :				2231.80357	99.41774	

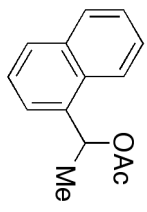
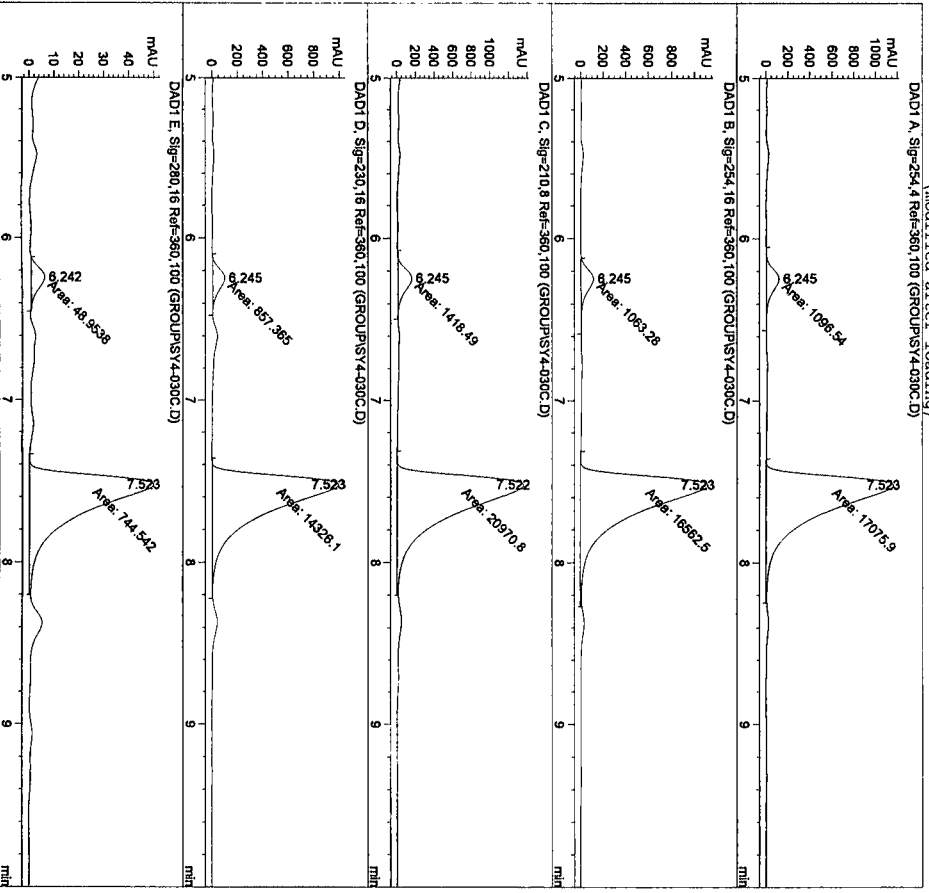


Table 1, entry 9
 with (+)-C₉Ph₅-DMAP⁺

Injection Date : 11/12/2011 12:04:53 PM
Sample Name : SN
Acq. Operator : SN
Acq. Instrument : Instrument 1
Different Inj Volume from Sequence : Actual Inj Volume : 5 µl
Acq. Method : C:\HPCHEM\1\METHODS\ODH-0130.M
Last changed : 8/3/2011 6:13:51 PM by JTM
Analysis Method : C:\HPCHEM\1\METHODS\SNIA0125.M
Last changed : 12/27/2011 7:59:40 PM by SN

Seq. Line : 2
Location : Vial 63



Area Percent Report

Sorted By : Signal
Multiplier : 1.0000
Dilution : 1.0000
Use Multiplier & Dilution Factor with ISTDs

Table with 5 columns: Peak RetTime [min], Width [min], Area [mAU*s], Height [mAU], Area %. Data for Signal 1: DAD1 A, Sig=254,4 Ref=360,100.

Table with 5 columns: Peak RetTime [min], Width [min], Area [mAU*s], Height [mAU], Area %. Data for Signal 2: DAD1 B, Sig=254,16 Ref=360,100.

Table with 5 columns: Peak RetTime [min], Width [min], Area [mAU*s], Height [mAU], Area %. Data for Signal 3: DAD1 C, Sig=210,8 Ref=360,100.

Table with 5 columns: Peak RetTime [min], Width [min], Area [mAU*s], Height [mAU], Area %. Data for Signal 4: DAD1 D, Sig=230,16 Ref=360,100.

Table with 5 columns: Peak RetTime [min], Width [min], Area [mAU*s], Height [mAU], Area %. Data for Signal 5: DAD1 E, Sig=280,16 Ref=360,100.

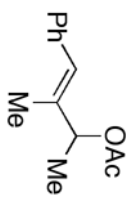
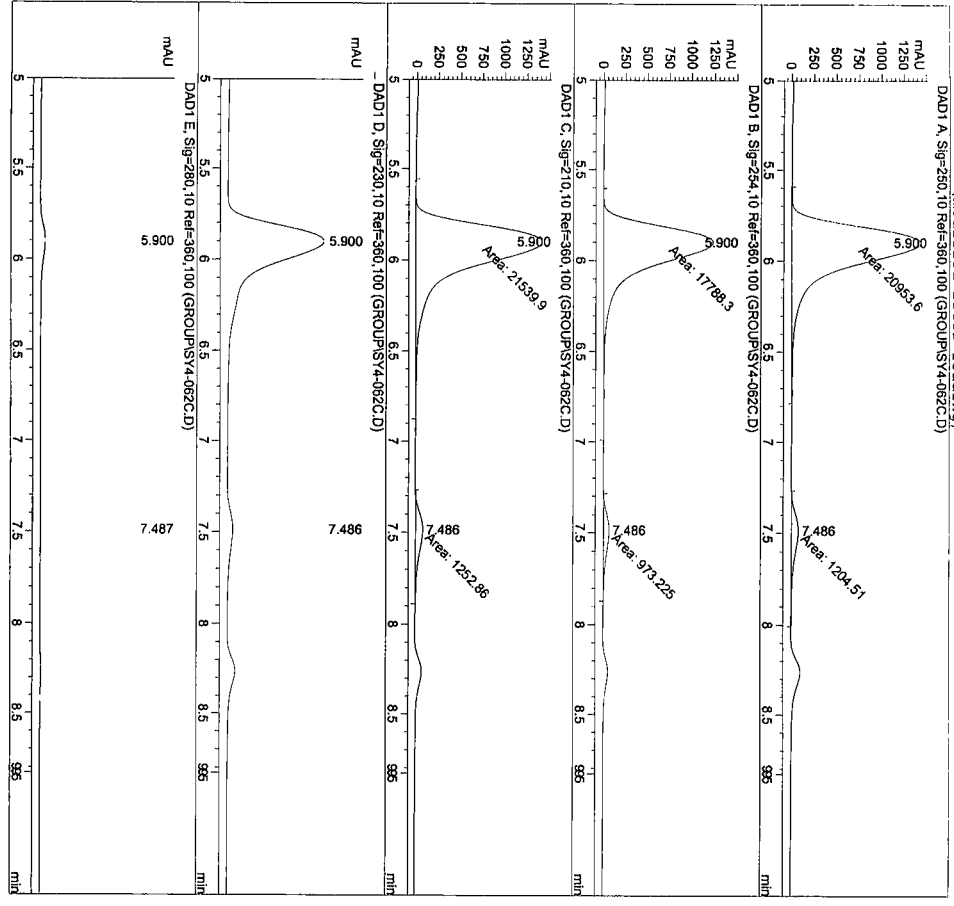


Table 1, entry 10
with (-)-C5Ph5-DMAP*

Injection Date : 12/8/2011 12:22:44 AM
 Sample Name : JTM
 Acq. Operator : JTM
 Acq. Instrument : Instrument 1
 Different Inj Volume from Sequence : Actual Inj Volume : 15 µl
 Acq. Method : C:\HPCHEM\1\METHODS\OD-01-30.M
 Last changed : 4/7/2011 2:40:51 AM by CC
 Analysis Method : C:\HPCHEM\1\METHODS\YL-AD07.M
 Last changed : 12/27/2011 8:21:33 PM by JTM



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=250,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.900	MM	0.2418	2.0953664	1444.29919	94.5640
2	7.486	MM	0.2629	1204.50964	76.36211	5.4360
Totals : 2.21581e4 1520.66131						

Signal 2: DAD1 B, Sig=254,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.900	MM	0.2407	1.7788364	1231.64832	94.8127
2	7.486	MM	0.2537	973.22461	63.92531	5.1873
Totals : 1.87616e4 1295.57362						

Signal 3: DAD1 C, Sig=210,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.900	MM	0.2526	2.153964	1420.97913	94.5033
2	7.486	MM	0.2553	1252.85693	81.78120	5.4967
Totals : 2.27927e4 1502.76032						

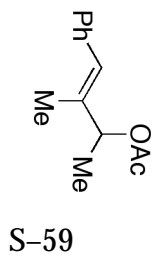
Signal 4: DAD1 D, Sig=230,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.900	BB	0.2265	1.5909664	1076.05347	94.1519
2	7.486	PP	0.2571	988.19049	59.09837	5.8481
Totals : 1.68978e4 1135.15183						

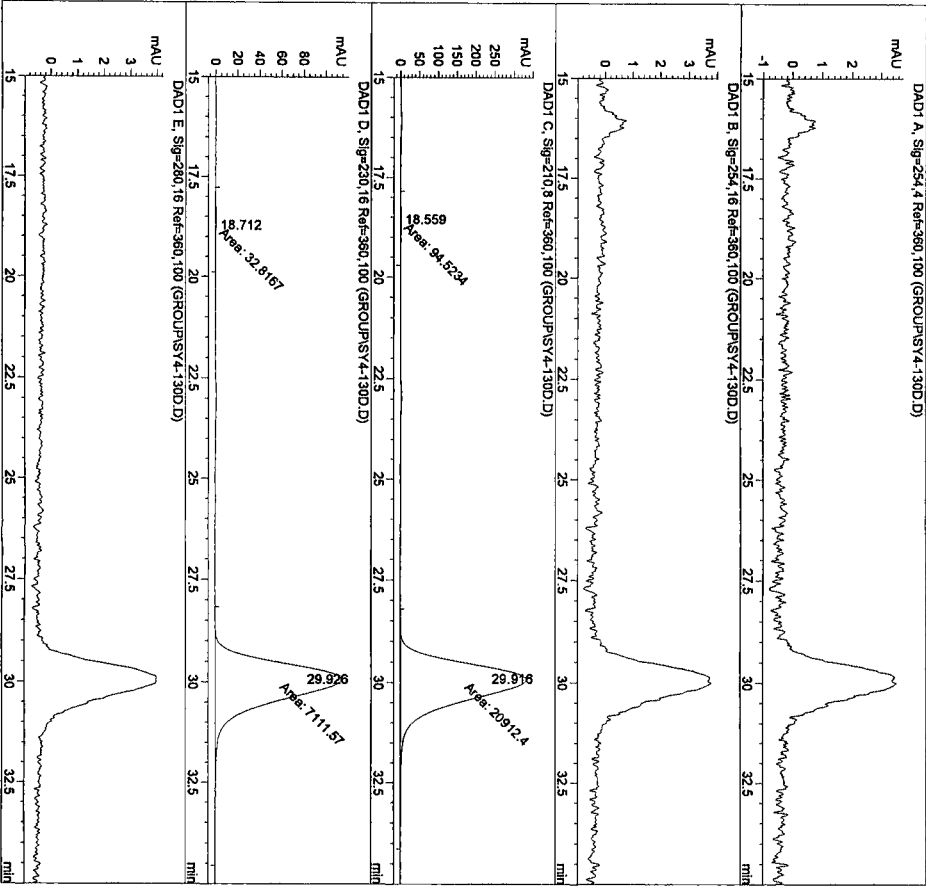
Signal 5: DAD1 E, Sig=280,10 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	5.900	BB	0.2255	767.51825	52.22518	94.1434
2	7.487	BP	0.2459	47.74647	3.02838	5.8566
Totals : 815.26472 55.25356						

Table 1, entry 10
 with (+)-C₅Ph₅-DMAP*



Injection Date : 1/22/2012 7:29:46 PM Seq. Line : 1
 Sample Name : Location : Vial 11
 Acq. Operator : JTM Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 µl
 Different Inj Volume from Sequence : Dilution : 1.000
 Acq. Method : C:\HPCHEM\1\METHODS\ODH-0540.M Use Multiplier & Dilution Factor with ISTDs
 Last changed : 9/7/2011 9:23:44 AM by SN
 Analysis Method : C:\HPCHEM\1\METHODS\VI-ADD08.M
 Last changed : 4/19/2012 1:41:14 PM by JTM



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Signal 2: DAD1 B, Sig=254,16 Ref=360,100

Signal 3: DAD1 C, Sig=210,8 Ref=360,100

Signal 4: DAD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.559	MM	0.8222	94.52335	1.91597	0.4500
2	29.916	MM	1.0443	2.09124e4	333.75858	99.5500
Totals :				2.10069e4	335.67454	

Results obtained with enhanced integrator!

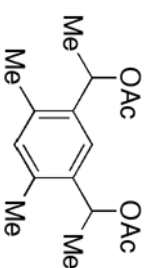
Signal 4: DAD1 D, Sig=230,16 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	18.712	MM	0.8052	32.81665	6.79271e-1	0.4593
2	29.926	MM	1.0365	7111.57129	114.34914	99.5407
Totals :				7144.38794	115.02841	

Results obtained with enhanced integrator!

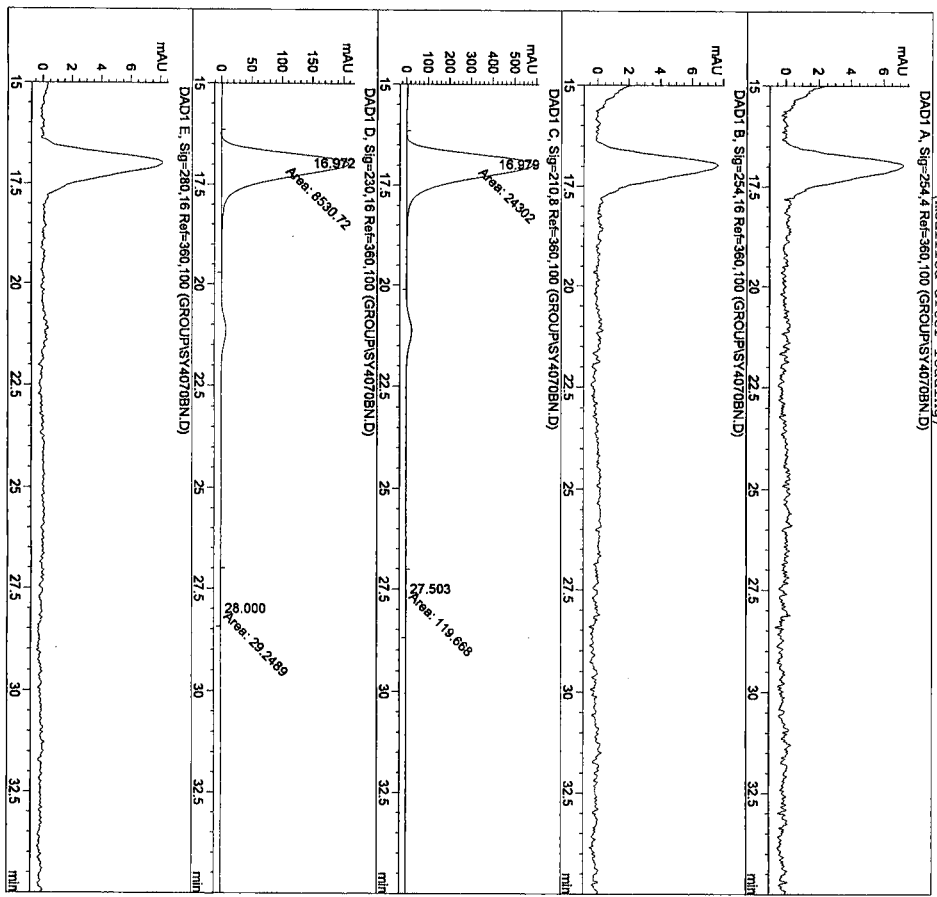
Signal 5: DAD1 E, Sig=280,16 Ref=360,100

*** End of Report ***



with (-)-C₆Ph₅-DMAP⁺

Injection Date : 4/19/2012 10:40:12 AM Seq. Line : 46
 Sample Name : JTM Location : Vial 22
 Acq. Operator : JTM Inj : 1
 Acq. Instrument : Instrument 1 Inj Volume : 5 µl
 Acq. Method : C:\HPCHEM\1\METHODS\OPH-0540.M
 Last changed : 9/7/2011 9:23:44 AM by SN
 Analysis Method : C:\HPCHEM\1\METHODS\YI-AD08.M
 Last changed : 4/19/2012 1:40:50 PM by JTM
 (modified after loading)



Area Percent Report
 Sorted By : Signal
 Multiplier : 1.0000
 Dilution : 1.0000
 Use Multiplier & Dilution Factor with ISTDs

Signal 1: DAD1 A, Sig=254,4 Ref=360,100

Signal 2: DAD1 B, Sig=254,16 Ref=360,100

Signal 3: DAD1 C, Sig=210,8 Ref=360,100

Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.979	MM	0.6942	2.43020e4	583.45728	99.5100
2	27.503	MM	0.9062	119.66791	2.20082	0.4900

Totals : 2.4421e4 585.65809

Results obtained with enhanced integrator!

Signal 4: DAD1 D, Sig=230,16 Ref=360,100

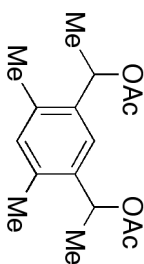
Peak #	RetTime [min]	Type	Width [min]	Area [mAU*s]	Height [mAU]	Area %
1	16.972	MM	0.6832	8530.72363	208.09496	99.6583
2	28.000	MM	0.8272	29.24886	5.89296e-1	0.3417

Totals : 8559.97249 208.68425

Results obtained with enhanced integrator!

Signal 5: DAD1 E, Sig=280,16 Ref=360,100

*** End of Report ***



eq 10

With (+)-C₉Ph₅-DMAP*