Table S1 List of primers

Primer	Sequence	Note
pbfBamH1	5′-AATTGGATCCACTAACCTTATTGTCCCTTG-3′	*Pbf* RNAi construct
pbfBspE1	5′-TATATCCGGAATGGACATGATCTCCGGCAG-3′	
pbfBblII	5′-AATAAGATCTATGGACATGATCTCCGGCAG-3′	
pbfSac1	5′-AATTGAGCTCACTAACCTTATTGTCCCTTG-3′	
Umc1066F	5′-ATGGAGCACGTCATCTCAATGG-3′	*O2* polymorphic markers
Umc1066R	5′-AGCAGCAGCAACGTCTATGACACT-3′	
P1	5′- TATATGAATTCAAGGATCCGCATGCCGGTCAACATGGTG -3′	P35S forward, *EcoR*1 and *BamH*1 introduced
P2	5′- AAATCCATGGTAGATCCCCCGTTCGTAAATG -3′	P35S reverse, *Nco*1 introduced
P3	5′- ATTACCATGGACATGATCTCCGGCAG -3′	*Pbf* CDS forward, *Nco*1 introduced
P4	5′- AATTGAGCTCACTAACCTTATTGTCCCTTG -3′	*Pbf* CDS reverse, *Sac*1 introduced
P5	5′- TATATGAATTCAACCTAGGGCATGCCGGTCAACATGGTG -3′	P35S forward, *EcoR*1 and *Avr*II introduced
P6	5′- AAATTCTAGAGGTAGATCCCCCGTTCGTAAATG -3′	P35S reverse, *Xba*1 introduced
P7	5′- ATTATCTAGACCATGGAGCACGTCATCTCAATG -3′	*O2* CDS forward, *Xba*1 introduced
P8	5′- TAATGAGCTCCTTATTCAGCGACGCCTG -3′	*O2* CDS reverse, *Sac*1 introduced
P9	5′-TATAGAGCTCGGCGCAAAAATCACCAGTC-3′	T35S forward, *Sac*1 introduced
P10	5′- ATTAAGCTTAATTCCTAGGTGCAGGTCACTGGATTTTGG -3′	T35S reverse, *Hind*III and *Avr*II introduced
P11	5′- AAAGCTGTACAAGTAAATAGAAATATTTGTGTTGTATCG -3′	Terminator of 10-kDa δ-zein gene, forward
P12	5′- ATCAAGCTTACTAGTGGATCCTACCAGCTGAGAATTAGGAG -3′	Terminator of 10-kDa δ-zein gene, reverse, *Hind*III and *BamH*1 introduced
P13	5′- ATTTGAATTCGTCCACGCGCAAATAGACC -3′	22-kDa zein promoter forward, *EcoR*1 introduced
P14	5′- TAATCCATGGTTGTTTGGTCGTTGCTAGTG -3′	22-kDa zein promoter

		reverse, *Nco*1 introduced
γ50F	5′-ATGAAGCTGGTGCTTGTGGTTC-3′	For γ50 RT-PCR
γ50R	5′-TAATGTCATTGCTGCTGCATGG-3′	
γ27F	5′-ATGAGGGTGTTGCTCGTTGC-3′	For γ27 RT-PCR
γ27R	5′-ACTCAACTAGCTAGCTAGCC-3′	
α22F1	5′-ACACCATATGTTCATTATTCCACAATGCTCA-3′	For α22 RT-PCR
α22R1	5′-TTAAGGATCCTATATAATCTAAAAGATGGCA-3′	
α19AF	CTCTTAa/gATTAGTAGCTAATAt/cATC	For α19A RT-PCR
α19AR	CTGGGAAGCCACAAACATCA	
α19BDF	ATTAGTCGGTAATCCATCAACC	For α19B and D RT-PCR
α19BDR	CTAGAAGATGGCACCACCAATG	
δ18F	5′-CGCCATGGCAGCCAAGATG-3′	For δ18 RT-PCR
δ18R	5′-ATGCCGACTTCATTATTGGG-3′	
γ16F	5′-TCGACACCATGAAGGTGCTG-3′	For γ16 RT-PCR
γ16R	5′-TGGTGATGGGTGACACTACG-3′	
β15F	5′-AGGATCGTCGAACAGAACAGC-3′	For β15 RT-PCR
β15R	5′-AGATGGATAGAGGAGATTTCCC-3′	
δ10F	5′-ATACTCTAGGAAGCAAGGAC-3′	For δ10 RT-PCR
δ10R	5′-TAAGAACATGGGTGGAATCG-3′	
δ18-10F	5′-CGCCATGGCAGCCAAGATG-3′	Common primer pair for
δ18-10R	5′-TATCTAGAATGCAGCACCAAC-3′	δ18 and δ10 RT-PCR
PbfF	5′-ATGGACATGATCTCCGGCAG-3′	For *Pbf* RT-PCR
PbfR	5′-ACTAACCTTATTGTCCCTTG-3′	
O2F	5′-ATGGAGCACGTCATCTCAATG-3′	For *O2* RT-PCR
O2R	5′-CCTTATTCAGCGACGCCTG-3′	
GFPF1	5′-ATGGTGAGCAAGGGCGAGG-3′	For *Gfp* RT-PCR
GFPR1	5′-TTACTTGTACAGCTCGTCC-3′	
P22F1	5′-ACATGTGTAAAGGTGAAGAG-3′	
GFPR2	5′- GTGAGCAAGGGCGAGGAGCT-3′	
P22bisulF	5′-ATATGTGTAAAGGTGAAGAG-3′	
α22bisulR	5′-CTAATATCTTAATAACCATT -3′	
GFPbisulR	5′-AACTCCTCACCCTTACTCAC-3′	
P27bisulF	5′-GAAATATGGTGAGTTATGTTGAG-3′	
γ27bisulR	5′-AAAACAACAAACAACACCCTC-3′	
P35SbisulF	5′-ATGTTGGTTAATATGGTGGAG-3′	
BarbisulR	5′-ACCACCAACATATCCACCTC-3′	
PbfbisulR	5′-CTTTAAATCCCTAACTTCCTCC-3′	

β15bisulF	5’- GAGTAATGTATTGGTTGTGAG -3’
β15bisulR	5’- CTATTCAACAATCCTCTCTAC -3’

Restriction enzyme cutting sites are colored in red.