

1 TABLE S1. Bacterial strains and plasmids

Strain or plasmid	Relevant characteristics	Source or Reference
<i>A. tumefaciens</i> strains		
C58	Wild-type virulent strain containing nopaline-type Ti plasmid pTiC58	Eugene Nester
Ach5	Wild-type virulent strain containing octopine-type Ti plasmid pTiAch5	[1]
1D1609	Wild-type virulent strain containing octopine-type Ti plasmid pTi1D1609	[2]
EML1226	<i>chvG</i> in-frame deletion mutant, C58 Δ <i>chvG</i>	This study
EML3682	<i>chvI</i> in-frame deletion mutant, C58 Δ <i>chvI</i>	This study
EML1654	<i>exoR</i> in-frame deletion mutant, C58 Δ <i>exoR</i>	This study
EML2104	<i>exoR chvG</i> in-frame deletion mutant, C58 Δ <i>chvG</i> Δ <i>exoR</i>	This study
<i>E. coli</i> strains		
DH10B	Host for DNA cloning	Invitrogen
BL21(DE3)	Host for overexpressing proteins driven by T7 promoter	[3]
<i>S. cerevisiae</i> AH109	Host for yeast two-hybrid analysis	Clontech
Plasmids		
pRL662	Gm ^r , broad-host-range vector derived from pBBR1MCS-2	[4]
pTrc200	Sm ^r Sp ^r , pSV1 origin <i>lacI^r</i> , <i>trc</i> promoter expression vector	[5]
pET22b(+)	Ap ^R , overexpression vector to generate C-terminal His-tagged protein driven by T7 promoter	Novagen
pET28a	Km ^r , overexpression vector to generate N-terminal His-tagged protein driven by T7 promoter	Novagen
pJQ200SK	Gm ^r , suicide plasmid containing Gm ^r and <i>sacB</i> gene for selection of double crossover	[6]
pBluescript SK(+)-HA	pBluescriptsk+::HA for HA epitope tagging vector	This study
pGADT7	Ap ^r , AD vector used in yeast two-hybrid assay	Clontech
pGBKT7	Km ^r , DNA-BD vector used in yeast-two hybrid assay	Clontech
pJQ200SK- Δ <i>chvG</i>	Gm ^r , used in generating entire <i>chvG</i> deletion mutant of <i>A. tumefaciens</i> C58	This study
pJQ200SK- Δ <i>chvI</i>	Gm ^r , used in generating entire <i>chvI</i> deletion mutant of <i>A. tumefaciens</i> C58	This study
pJQ200SK- Δ <i>exoR</i>	Gm ^r , used in generating entire <i>exoR</i> deletion mutant of <i>A. tumefaciens</i> C58	This study
pJQ200SK- Δ <i>exoR-strep</i>	Gm ^r , used in generating entire <i>exoR</i> replacement with <i>exoR-strep</i> of <i>A.</i>	This study

<i>tumefaciens</i> C58		
pET28a-ChvI	Km ^r , pET28a expressing His-ChvI	This study
pET28a-ChvI D52E	Km ^r , pET28a expressing His-ChvI with D52E substitution	This study
pChvG	Gm ^r , pRL662 expressing ChvG driven by <i>lacZp</i>	This study
pChvI	Gm ^r , pRL662 expressing ChvI driven by <i>lacZp</i>	This study
pChvI D52E	Gm ^r , pRL662 expressing ChvI with D52E substitution	This study
pChvI D52A	Gm ^r , pRL662 expressing ChvI with D52A substitution	This study
pChvG-HA	Gm ^r , pRL662 expressing ChvG-HA driven by <i>lacZp</i>	This study
pChvI-ChvG-HA	Gm ^r , pRL662 expressing ChvI-ChvG-HA driven by native protomotor	This study
pExoR	Sm ^r Sp ^r , pTrc200 expressing ExoR driven by <i>trc</i> promoter	This study
pExoR G73C	Sm ^r Sp ^r , pTrc200 expressing ExoR with G73C substitution	This study
pExoR S153Y	Sm ^r Sp ^r , pTrc200 expressing ExoR with S153Y substitution	This study
pExoR G73C S153Y	Sm ^r Sp ^r , pTrc200 expressing ExoR with G73C and S153Y substitutions	This study
pGBKT7-ChvG-peri	Km ^r , DNA-BD vector expressing ChvG periplasmic domain	This study
pGBKT7-53	Km ^r , DNA-BD vector expressing murine p53	Clontech
pGADT7-ExoR	Ap ^r , AD vector expressing ExoR	This study
pGADT7-ExoR G73C	Ap ^r , AD vector expressing ExoR G73C	This study
pGADT7-ExoR S153Y	Ap ^r , AD vector expressing ExoR S153Y	This study
pGADT7-ExoR G73C S153Y	AP ^r , AD vector expressing ExoR G73C S153Y	This study
pGADT7-T	Ap ^r , AD vector expressing SV40 large T-antigen	Clontech

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1 **References**

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