

Table S1. List of nematode and bacteria strains used in this study

Strain		Genotype or description	Source or reference
Nematode			
<i>C. elegans</i>	N2	Wild-type variant Bristol	CGC ^a
	SU159	<i>ajm-1(ok160) X; jcEx44 [pJS191 (<i>ajm-1::gfp</i>) + pRF4 (<i>rol-6(su1006)</i>)]</i>	CGC ^a
Bacteria			
<i>S. Typhimurium</i>	ATCC 14028	Wild-type 14028 strain	ATCC ^b
	ATCC 14028::GFP	Wild-type 14028 <i>proU::gfp</i> , chloramphenicol ^r	MR Lab, [1]
	ATCC 14028::RFP	Wild-type 14028 + pMW635, <i>araC</i> , ampicillin ^r	MR Lab, Oliver Pabst
	<i>hilA</i>	ATCC 14028 <i>hilA</i> , kanamycin ^r	[2]
	<i>ssaV</i>	ATCC 14028 non-polar <i>ssaV</i> , chloramphenicol ^r	[3]
	<i>rfaL</i>	ATCC 14028 <i>rfaL</i> , chloramphenicol ^r	[4]
	<i>trxA</i>	ATCC 14028 non-polar <i>trxA</i>	[4]
	<i>trxA/pFA3</i>	<i>trxA</i> + wild-type TrxA, E _o ^r -270 mV, <i>araC</i> , chloramphenicol ^r	[4,5]
	<i>trxA/pFA5</i>	<i>trxA</i> + Grx-type TrxA, E _o ^r -195 mV, <i>araC</i> , chloramphenicol ^r	[4,5]
	<i>trxA/pFA6</i>	<i>trxA</i> + DsbA-type TrxA, E _o ^r -204mV, <i>araC</i> , chloramphenicol ^r	[4,5]
	<i>trxA/pFA7</i>	<i>trxA</i> + PDI-type TrxA, E _o ^r -221 mV, <i>araC</i> , chloramphenicol ^r	[4,5]
	<i>trxA/pFA8</i>	<i>trxA</i> + non-catalytic TrxA, <i>araC</i> , chloramphenicol ^r	[4,5]
	<i>trxA/pBAD33</i>	<i>trxA</i> + empty vector, <i>araC</i> , chloramphenicol ^r	[4,5]
	<i>trxB</i>	ATCC 14028 <i>trxB</i> , kanamycin ^r	[4]
	<i>trxC</i>	ATCC 14028 <i>trxC</i> , kanamycin ^r	[4]
	<i>dsbC</i>	ATCC 14028 <i>dsbC</i> , kanamycin ^r	[4]
<i>S. Dublin</i>	STM75	Wild-type STM75 strain	[6]
	STM75::RFP	Wild-type STM75 + pMW635, <i>araC</i> , ampicillin ^r	MR Lab, Oliver Pabst
<i>E. coli</i>	OP50	Laboratory food source	CGC ^a
	OP50::GFP	OP50 + pFVP25.1, ampicillin ^r	CGC ^a
<i>B. thailandensis</i>	ATCC 700388	Wild-type reference strain isolated from Thailand	ATCC ^b

^a Caenorhabditis Genetics Center, University of Minnesota, Minneapolis, U.S.A.^b American Type Cell Collection, Virginia, U.S.A.

REFERENCES

1. Hautefort I, Proença MJ, Hinton JCD (2003) Single-copy green fluorescent protein gene fusions allow accurate measurement of *Salmonella* gene expression in vitro and during infection of mammalian cells. *Appl Environ Microbiol* 69: 7480-7491.
2. Negrea A, Bjur E, Ygberg SE, Elofsson M, Wolf-Watz H, et al. (2007) Salicylidene acylhydrazides that affect type III protein secretion in *Salmonella enterica* serovar Typhimurium. *Antimicrob Agents Chemother* 51: 2867-2876.
3. Shea JE, Beuzon CR, Gleeson C, Mundy R, Holden DW (1999) Influence of the *Salmonella* Typhimurium pathogenicity island 2 type III secretion system on bacterial growth in the mouse. *Infect Immun* 67: 213-219.
4. Bjur E, Eriksson-Ygberg S, Aslund F, Rhen M (2006) Thioredoxin 1 promotes intracellular replication and virulence of *Salmonella enterica* serovar Typhimurium. *Infect Immun* 74: 5140-5151.
5. Mössner E, Huber-Wunderlich M, Rietsch A, Beckwith J, Glockshuber R, et al. (1999) Importance of redox potential for the *in vivo* function of the cytoplasmic disulfide reductant thioredoxin from *Escherichia coli*. *J Biol Chem* 274.
6. Tezcan-Merdol D, Eriksson Ygberg S, Rhen M (2007) The *Salmonella enterica* virulence plasmid and the spv gene cluster. In: Rhen M, Maskell D, Mastroeni P, Therfall J, editors. *Salmonella: molecular biology and pathogenesis*. Norfolk: Horizon Bioscience. pp. 89-103.