Supporting Information

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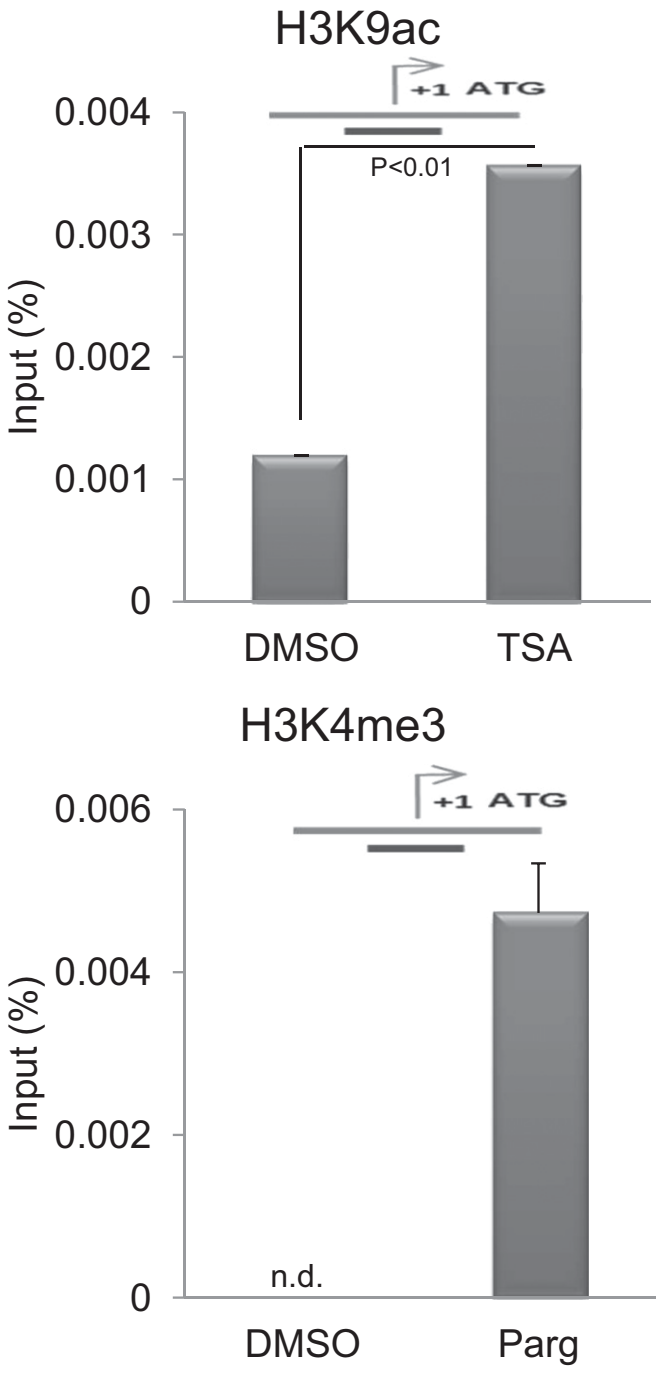

Fig. S1. Trichostatin A (TSA) and pargyline regulate chromatin modifications at the IL-1β promoter. Zebrafish eggs microinjected at the one-cell stage with 5–10 ng *Vibrio anguillarum* DNA (vDNA) per egg were dechorionated at 24 h post fertilization (hpf) and treated manually with 100 nM TSA or 3 μM pargyline for 5 h. Larvae then were processed and analyzed by ChIP (H3K9ac and H3K4me3). The amplicon used for each promoter is indicated by a diagram above the bars. Error bars indicate the SD of triplicate samples using 30 pooled larvae per treatment. n.d., not detected.

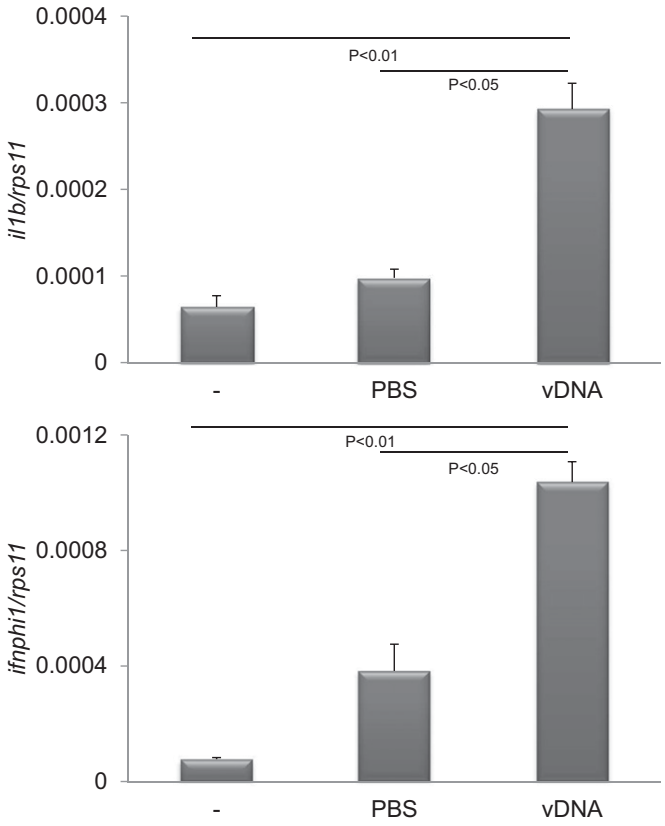

Fig. S2. vDNA induces IL-1β and IFNφ1 expression. Zebrafish eggs microinjected at the one-cell stage with vDNA or vehicle alone (PBS) (5–10 ng per egg) were processed at 30 hpf. IL-1β and IFNφ1 transcript levels then were analyzed by quantitative PCR. Error bars indicate the SD of triplicate samples using 30 pooled larvae per treatment.

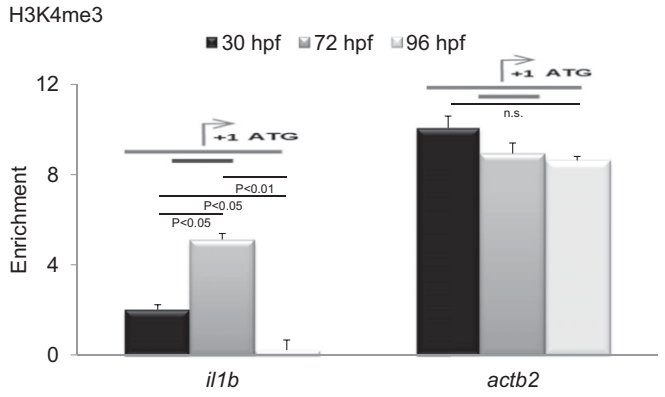

Fig. S3. Dynamics of chromatin modifications at the IL-1β gene promoter. Zebrafish embryos or naturally hatched larvae were processed and analyzed by ChIP (H3K4me3) at the indicated times. The amplicon used for each promoter is indicated by a diagram above the bars. Error bars indicate the SD of triplicate samples using 30 pooled larvae per sample point. n.s., nonsignificant.

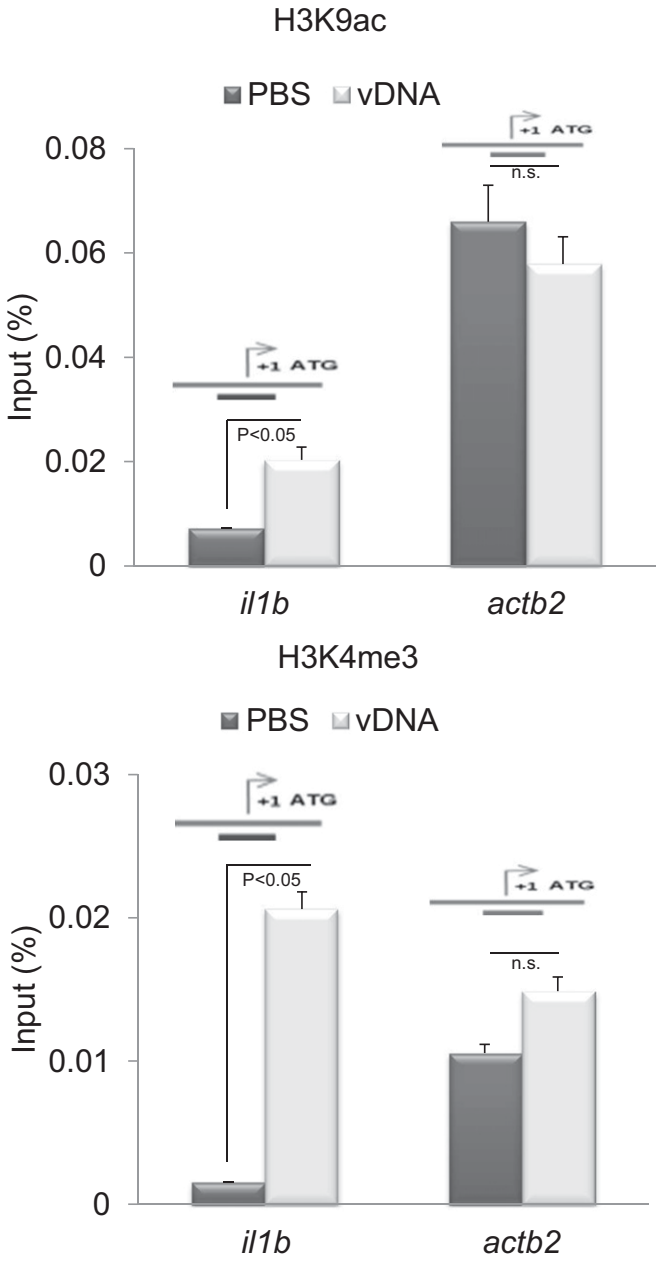

Fig. S4. vDNA injection increases chromatin modifications at the IL-1β gene promoter. Zebrafish eggs microinjected at the one-cell stage with 5–10 ng vDNA per egg were processed at 30 hpf and analyzed by ChIP (H3K9ac and H3K4me3). The amplicon used for each promoter is indicated by a diagram above the bars. Error bars indicate the SD of triplicate samples using 30 pooled larvae per sample. n.s., nonsignificant.

Table S1. Primers used to analyze gene expression in this study

Gene symbol	GenBank accession no.	Gene name	Primers	Sequence (5′ to 3′)
c3a	BC055564	*Complement component c3a*	F	ATGAGCTCCTGCAGAGGTGT
			R	AGTGGTTGTTGGAGGTCTGG
cclc25ab	NM_001129894	*C-C motif chemokine c25 ab*	F	AGCACCTCTCGCTTTGTGTT
			R	TGTTTGAAAGGCACTTGACG
defbl1	NM_001081553	*Defensin, beta-like 1*	F	CAGGACTGCCATCATCTGAA
			R	CTCCTTGTCTGCAAACACCA
ifnphi1	NM_207640	*IFN phi 1*	F3	GAGCACATGAACTCGGTGAA
			R3	TGCGTATCTTGCCACACATT
ifnphi2	NM_001111082	*IFN phi 2*	F1	CCTCTTTGCCAACGACAGTT
			R1	CGGTTCCTTGAGCTCTCATC
ifnphi3	NM_001111083	*IFN phi 3*	F1	TTCTGCTTTGTGCAGGTTTG
			R1	GGTATAGAAACGCGGTCGTC
il1b	NM_212844	*Interleukin 1 beta*	F5	GGCTGTGTGTTTGGGAATCT
			R5	TGATAAACCAACCGGGACA
il8	CT826376	*Interleukin 8*	F	GTCGCTGCATTGAAACAGAA
			R	CTTAACCCATGGAGCAGAGG
il8l2	EH441857	*Interleukin 8-like 2*	F	GCTGGATCACACTGCAGAAA
			R	TGCTGCAAACTTTTCCTTGA
il10	NM_001020785	*Interleukin 10*	F2	ATTTGTGGAGGGCTTTCCTT
			R2	AGAGCTGTTGGCAGAATGGT
il12a	AB183001	*Interleukin 12a*	F1	AGCAGGACTTGTTTGCTGGT
			R1	TCCACTGCGCTGAAGTTAGA
lta	NM_001024821.1	*Lymphotoxin alpha (TNF superfamily, member 1)*	F2	AAGCCAAACGAAGGTCA
			R2	AACCCATTTCAGCGATTGTC
lyz	NM_139180	*Lysozyme*	F	TGGCAGTGGTGTTTTTGTGT
			R	TCAAATCCATCAAGCCCTTC
mxb	NM_001128672	*Myxovirus (influenza) resistance B*	F1	AATGGTGATCCGCTATCTGC
			R	TCTGGCGGCTCAGTAAGTTT
mxc	NM_001007284	*Myxovirus (influenza virus) resistance C*	F	GAGGCTTCACTTGGCAACTC
			R	TTGTTCCAATAAGGCCAAGC
nos2b	NM_001113501	*Nitric oxide synthase 2b, inducible*	F1	GGCTTGCACTGCTTTTAAGG
			R1	TCCAGAGTGCAACTGTCCTG
ptgs1	NM_153656	*Prostaglandin-endoperoxide synthase 1*	F	TTTTGCTGCTGAGTGTGTCC
			R	CGAACACAGATCCCTTGGTT
ptgs2b	NM_001025504	*Prostaglandin-endoperoxide synthase 2b*	F1	TGGATCTTTCCTGGTGAAGG
			R1	GAAGCTCAGGGGTAGTGCAG
ticam1	NM_001044759	*Toll-like receptor adaptor molecule 1*	F1	ATGGAGAGCGCTTGAACTGT
			R1	TTGTCGCACAAACTCTCCTG
tlr3	NM_001013269	*Toll-like receptor 3*	F1	AAAGGGCTACGTTTGGTGTG
			R1	GTTGGTGGAGTTCAGCATT
tlr4ba	NM_001131051	*Toll-like receptor 4b, duplicate a*	F	CAATGGCTTGGGTACTTTGC
			R	GATTTGAGGAGTGCCGGATA
tlr22	NM_001128675	*Toll-like receptor 22*	F1	TGGGCCAAGAAGAATGAATC
			R1	ATGACAACAGGAGGGTGAGG
tnfa	NM_212859	*Tumor necrosis factor a (TNF superfamily, member 2)*	F2	GCGCTTTTCTGAATCCTACG
			R2	TGCCCAGTCTGTCTCCTTCT
rps11	NM_213377	*Ribosomal protein S11*	F	ACAGAAATGCCCCTTCACTG
			R	GCCTCTTCTCAAAACGGTTG

The gene symbols follow the Zebrafish Nomenclature Guidelines (https://wiki.zfin.org/display/general/ZFIN+Zebrafish+Nomenclature+Guidelines).

Table S2. ChIP-quantitative PCR primer sequences and position of amplicons of each gene used in this study

Gene symbol	Ensembl accession no.	Gene name	Primers	Sequence (5′ to 3′)	Position relative to transcription start site (nt)
actb2	ENSDARG00000037870	*Actin beta2*	F	ACTATGAACTGAACCGACTG	+919/+1072
			R	CTGCGATCAATTACACAACC	
ifnphi1	ENSDARG00000025607	*IFN phi 1*	F	CCCGAATAAATCCAGCACTC	−84/+100
			R	TTATCCTGTATCGGCCAAGC	
il1b	ENSDARG0000005419	*Interleukin 1 beta*	F	ATCAAGGAATTTCCCGCTTC	−62/+64
			R	TGCACTCCACATACCAGAGC	
il12a	ENSDARG00000038878	*Interleukin 12a*	F	GCGTGTGAGTGTGTGTGTGT	−91/+55
			R	GAGCCAGAGCCATGCTAATC	
tert	ENSDARG00000042637	*Telomerase reverse transcriptase*	F	GCGTGTCGTTATCTGGGAGT	−69/-10
			R	CGTGCTTGGGGAGTTCTATT	

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