

# Synthesis and NMR Characterization of (Z,Z,Z,Z,E,E, $\omega$ )-Heptaprenol

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## Experimental Procedures.

**General Information.** All reactions were performed under an atmosphere of nitrogen, unless noted otherwise. Thin-layer chromatography was performed with Whatman reagents 0.25-mm silica gel 60-F plates. Column chromatography was carried out with silica gel 60, 230–400 mesh (0.040–0.063 mm particle size) purchased from EM Science. High-resolution mass spectra were obtained at the Department of Chemistry and Biochemistry, University of Notre Dame by ESI ionization, using Bruker micrOTOF/Q2 mass spectrometer.

**NMR Spectroscopy.** 1D  $^1\text{H}$ ,  $^{13}\text{C}\{^1\text{H}\}$ , Dept and 2D homo-COSY and heteronuclear  $^1\text{H}$ - $^{13}\text{C}$  HETCOR spectra were recorded on a Varian INOVA-500 or Varian DirectDrive 600 spectrometer. For compounds **18-OH**, **20-OH**, **7b**, **4**, and **4B2**, 1D  $^1\text{H}$  and  $^{13}\text{C}\{^1\text{H}\}$ , and 2D homo- DQF-COSY, TOCSY, ROESY and heteronuclear  $^1\text{H}$ - $^{13}\text{C}$  HSQC, HSQC-TOCSY, HMBC spectra were recorded on a Bruker AVANCE II 800. The structures of prenil **4** and **4B2** [10 mg of each was dissolved in 600  $\mu\text{L}$   $\text{CDCl}_3$  containing 3 mg  $\text{Eu}(\text{hfc})_3$ ] were determined by interpretation of the homonuclear DQF-COSY, TOCSY, ROESY and heteronuclear  $^1\text{H}$ - $^{13}\text{C}$  HSQC, HSQC-TOCSY, HMBC NMR spectra. Proton connectivities were derived from the DQF-COSY and TOCSY spectra.  $^{13}\text{C}$  resonances corresponding to carbons with directly attached protons were assigned using HSQC and HSQC-TOCSY spectra. HMBC spectra were used to assign resonances of the quaternary carbons and to validate the connectivities established by the other spectra. ROESY spectra were utilized to establish the *E* or *Z* configurations for each C=C double bond. Experiments were performed at 25  $^\circ\text{C}$  using a Bruker AVANCE II spectrometer equipped with a TCI cryoprobe and operating at a  $^1\text{H}$  resonance frequency of 800.13 MHz. Standard pulse sequences were used.<sup>1-6</sup> Time domain data ( $t_2$  and  $t_1$ ) for 2D experiments were recorded as 2048  $\times$  1024 complex matrices with 16 and 32 scans per  $t_1$  increment for homonuclear and heteronuclear spectra, respectively. Relaxation delay between individual scans and spin-lock time for TOCSY experiments was 1.4 s and 60 ms, respectively. For ROESY experiments, a relaxation delay of 4 s and a spin-lock time of 400 ms gave 2048  $\times$  512 complex time domain points. Linear prediction to 1024 complex points was applied in

the  $t_1$  domain. The data were zero filled to obtain final  $2048 \times 2048$  complex matrices. In all other homonuclear 2D experiments, zero filling was used only in the  $t_1$  domain to obtain final  $2048 \times 2048$  complex time domain data. In heteronuclear 2D experiments, linear prediction to 2048 complex data points was employed in the  $t_1$  domain, which was zero filled to 4096 to get final  $2048 \times 4096$  complex time domain data. Shifted sine bell weighting functions were applied in both domains prior to double Fourier transformation. In the 3D  $^1\text{H}$ - $^{13}\text{C}$ -HSQC-TOCSY experiments,  $1024 \times 48 \times 128$  complex time domain points in  $t_3$ ,  $t_2$ , and  $t_1$  domains were collected with 16 scans per time increment, relaxation delay of 1.3 s, and 60 ms spin-lock. Linear predictions to 128 and 256 complex points were utilized in the  $t_2$  and  $t_1$  domains. Zero filling gave  $2048 \times 256 \times 512$  complex matrices. Shifted sine bell weighting functions were applied in all three domains prior to triple Fourier transformation. Spectra were processed using Bruker TopSpin 2.1 software.  $^1\text{H}$  spectra, the  $^1\text{H}$  dimension in 2D heteronuclear spectra, and the 1D  $^{13}\text{C}\{^1\text{H}\}$  spectra were referenced to solvent (DMSO,  $\delta_{\text{H}}$  2.5 and  $\delta_{\text{C}}$  39.5;  $\text{CDCl}_3$ ,  $\delta_{\text{H}}$  7.27 and  $\delta_{\text{C}}$  77.23). The  $^{13}\text{C}$  dimension in the 2D heteronuclear spectra was referenced indirectly.<sup>7</sup>

Six functional group transformations, used for the synthesis of compounds described in this paper, are given as general procedures A – F.

***General procedure A. Deprotection of O-THP ether of prenil.***

THP-protected ether (20 mmol) was dissolved in MeOH (100 mL) and TsOH·H<sub>2</sub>O (0.6 g) was added. The resulting mixture was stirred for 6 h at room temperature. After addition of Et<sub>3</sub>N (0.5 mL), the resulting solution was filtered through a layer of silica gel and concentrated under reduced pressure. The crude compound was used for the next step without further purification or it was purified by short-path column chromatography on silica gel.

***General procedure B. Bromination of prenil.***

B1. Triethylamine (3.3 mL, 24 mmol) was added to a solution of alcohol (20 mmol) in anhydrous THF (60 mL) and the solution was cooled down in an ice-water bath. To this solution, MsCl (1.9 mL, 24 mmol) was added dropwise. After stirring for 1 h in an ice-water bath, the reaction mixture was quickly filtered through a layer of Celite to a pre-cooled THF solution of LiBr (2.1 g, 24 mmol in 20 mL of THF) and Celite pad was quickly washed with THF ( $2 \times 10$  mL). The combined solution was stirred for 2 h in an ice-water bath. The volatiles were evaporated under reduced pressure and the resulting bromide was taken up in hexanes/ethyl acetate mixture (1:1, 200 mL). The solution was washed with water ( $50 \text{ mL} \times 2$ ) and brine (100 mL), dried over anhydrous  $\text{MgSO}_4$ , filtered, and concentrated under reduced pressure. The crude compound was used for the next step without further purification or it was purified by short-path column chromatography on silica gel.

B2. Phosphorus tribromide (0.83 mL, 8.7 mmol) in dry ether (20 mL) was added to a mixture of alcohol (20 mmol) and pyridine (0.15 mL) in dry ether (50 mL) over 1 h in an ice-water bath. After stirring for 2 h in an ice-water bath, it was poured into ice-water (100 mL) and then extracted with hexanes ( $50 \text{ mL} \times 3$ ). The organic layer was washed with water ( $50 \text{ mL} \times 2$ ), saturated  $\text{NaHCO}_3$  ( $50 \text{ mL} \times 2$ ), and brine (50 mL), dried over anhydrous  $\text{MgSO}_4$ , filtered, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel chromatography.

B3. Same as general procedure C except using LiBr, instead of LiCl

***General procedure C. Chlorination of prenol.***

A cold solution of alcohol (20 mmol) and LiCl (2.1 g, 50 mmol) in DMF (200 mL) was treated with *s*-collidine (6.6 mL, 50 mmol) and MsCl (3.9 mL, 50 mmol) in an ice-water bath. After stirring in an ice-water bath for 2 h, the reaction mixture was diluted with ethyl acetate (800 mL). The mixture was washed with saturated  $\text{NH}_4\text{Cl}$  ( $300 \text{ mL} \times 2$ ) and brine (300 mL), dried over anhydrous  $\text{MgSO}_4$ , filtered, and concentrated under reduced pressure to afford the corresponding chloride. The crude compound was

used for the next step without further purification or it was purified by short-path column chromatography on silica gel.

***General procedure D. Sulfonylation of prenyl halide.***

The halide (chloride or bromide, 20 mmol) in acetonitrile (10 mL) was added dropwise to a suspension of sodium *p*-toluenesulfinate (4.3 g, 24 mmol) in DMF (120 mL) in an ice-water bath. After the reaction mixture was stirred at room temperature for 6 h, it was diluted with ethyl acetate (500 mL). The mixture was washed with water (200 mL  $\times$  2) and brine (200 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated to afford the corresponding sulfone. The crude compound was purified by column chromatography on silica gel.

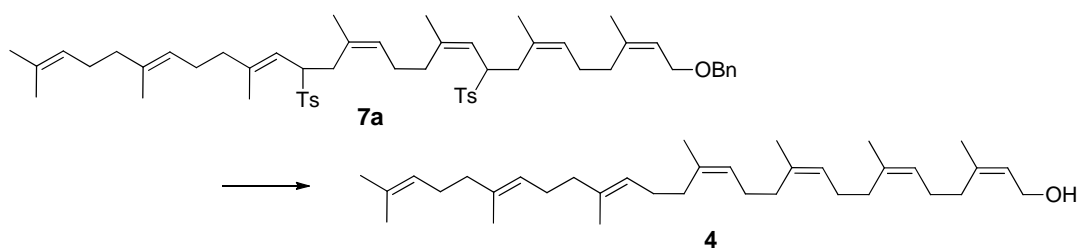
***General procedure E. Coupling of prenyl sulfone and prenyl bromide.***

Sulfone (20 mmol) was dissolved in a mixture of anhydrous THF and HMPA (4:1, 80 mL) and the resulting solution was cooled to  $-78$  °C. To this cold solution, *n*-BuLi (12.5 mL, 20 mmol, 1.6 M in hexane) was added dropwise and stirring was continued for 1.5 h. A solution of bromide (17.4 mmol) in THF-HMPA (4:1, 20 mL) was added dropwise to a solution of sulfone over 1 h. The mixture was stirred for an additional 5 h. After warming the mixture to 0 °C, it was poured into ice-water and extracted with hexanes-ether (1:1, 150 mL  $\times$  2). The extract was washed with water (150 mL) and brine (150 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The residue was purified by column chromatography on silica gel to afford the product.

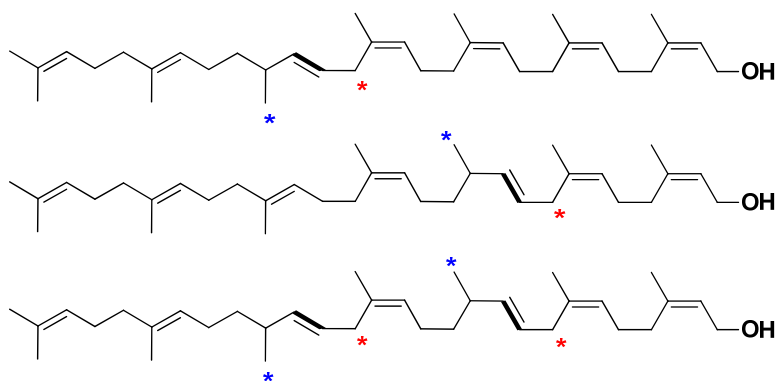
***General procedure F. Deprotection of benzyl ether and desulfonylation.***

**Transformation of 7a to 4 and 7b to 4/4B2 are given as representative examples.**

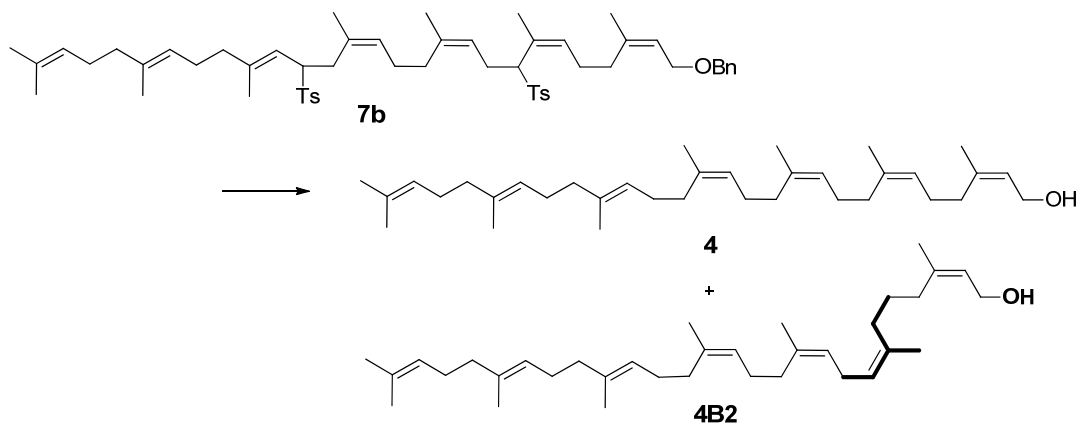
### Synthesis of compound **4** from **7a**.



Ethylamine (150 mL) was added to a flask containing lithium (1.0 g, 0.14 mol) at  $-78\text{ }^{\circ}\text{C}$  and ether (50 mL) was added. After stirring for 10 min at  $-78\text{ }^{\circ}\text{C}$ , an ethereal solution of compound **7a** (6.0 g, 6.7 mmol in 50 mL) was added dropwise to the blue solution over 20 min. After stirring for 40 min at  $-78\text{ }^{\circ}\text{C}$ , the reaction was quenched by addition of isoprene (10 mL) and MeOH (50 mL). After addition of saturated  $\text{NH}_4\text{Cl}$  (200 mL), the reaction mixture was extracted with ethyl acetate (100 mL  $\times$  3). The combined organic layer was washed with brine (150 mL), dried over anhydrous  $\text{MgSO}_4$ , filtered through a layer of silica gel, and concentrated under reduced pressure. The residue (2.8 g) was purified by column chromatography (silica gel, 35 g; i.d. of column, 3/4 inch; ethyl acetate:hexane, 1:7) to afford the desired product (2.0 g, 59%). The purity of **4** is  $>90\%$  as assessed by  $^1\text{H}$  NMR. As isolated by the silica flash chromatography **4** contains  $< 10\%$  of impurities presumed to arise as a result of double bond isomerization. Neither the number of these impurities nor their structures were determined. The structures of the three impurities anticipated as possibly present are shown below. We anticipate greater purity of **4** can be achieved by column chromatography using silver nitrate-impregnated silica gel.<sup>8-10</sup>

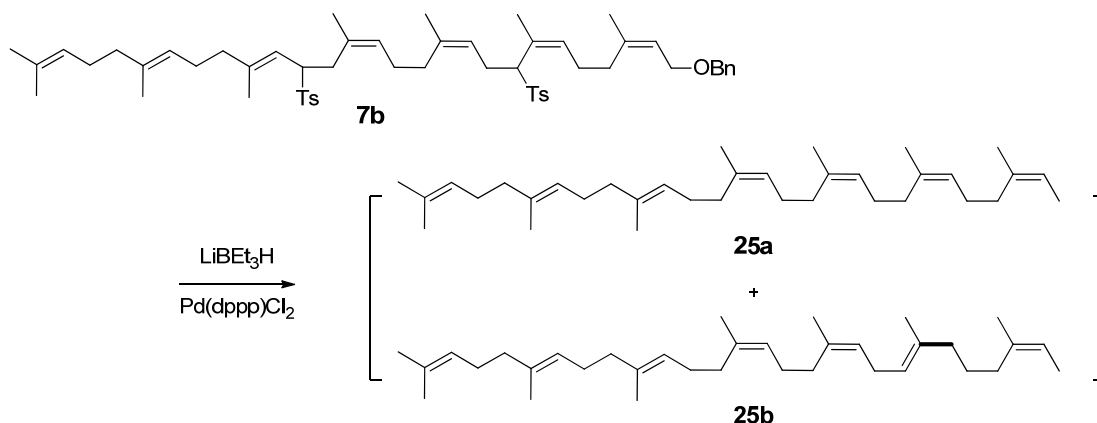


### Synthesis of compounds **4/4B2** from **7b**.



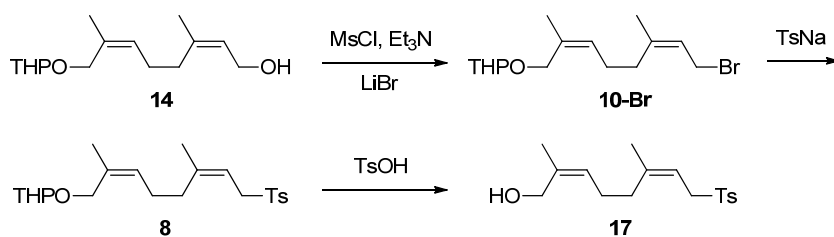
Ethylamine (200 mL) was added to a flask containing lithium (1.9 g, 0.27 mol) at  $-78\text{ }^{\circ}\text{C}$  and ether (100 mL) was added. After stirring for 10 min at  $-78\text{ }^{\circ}\text{C}$ , an ethereal solution of compound **7b** (8.0 g, 9.0 mmol in 100 mL) was added dropwise to the blue solution over 20 min. After stirring for 40 min at  $-78\text{ }^{\circ}\text{C}$ , the reaction was quenched by addition of isoprene (15 mL) and MeOH (70 mL). After addition of saturated  $\text{NH}_4\text{Cl}$  (300 mL), the reaction mixture was extracted with ethyl acetate (150 mL  $\times$  3). The combined organic layer was washed with brine (200 mL), dried over anhydrous  $\text{MgSO}_4$ , filtered through a layer of silica gel, and concentrated under reduced pressure. The residue (3.5 g) was purified by column chromatography (silica gel, 50 g; i.d. of column, 1 inch; ethyl acetate:hexanes, 1:7) to afford the mixture of **4/4B2** (2.8 g, 1:3 mixture, 63%). The mixture was further subjected to column chromatography (silica gel, 50 g; i.d. of column, 3/4 inch; ethyl acetate:hexanes, 1:15) to afford **4** (0.23 g), **4B2** (0.81 g), and **4/4B2** mixture (1.6 g).

### Deprotection of benzyl ether and desulfonation of **7b** using $\text{LiBEt}_3\text{H}/\text{Pd}(\text{dppp})\text{Cl}_2$ .



LiBEt<sub>3</sub>H (10 mL, 10 mmol, 1.0 M solution in THF) was added dropwise over 15 min to a cold solution of compound **7b** (1.8 g, 2.0 mmol) and bis(diphenylphosphino)propanepalladium(II) dichloride (Pd(dppp)Cl<sub>2</sub> (0.24 g, 0.40 mmol) in anhydrous THF (40 mL) in an ice-water bath. Stirring was continued for 4 h, and the reaction mixture was diluted with hexanes:ether (1:1, 200 mL). The mixture was washed with saturated NH<sub>4</sub>Cl (100 mL × 2) and brine (100 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. <sup>1</sup>H NMR analysis indicated that the crude material has at least two products. TLC analysis showed that the major products were considerably less polar than expected for **4** (heptaprenol). The major material obtained from column chromatography (silica gel, 15 g; i.d. of column, 1/2 inch; hexanes) of this crude product was 0.78 g of a 1:3 mixture (by <sup>1</sup>H NMR analysis) of hydrocarbons (arising from reductive desulfonylation and deoxygenation). The possible structures of the two isolated reaction products are formulated as **25a** and **25b** (major). These structures are suggested by the absence any aromatic resonances, and the further absence of the resonance anticipated for the methylene of (either a free or an *O*-Bn-protected) allyl alcohol in <sup>1</sup>H NMR spectrum. As a result, further evaluation of this method for desulfonylation was not made.

#### Synthesis of compounds **17** from **14** using general procedures B1/D/A

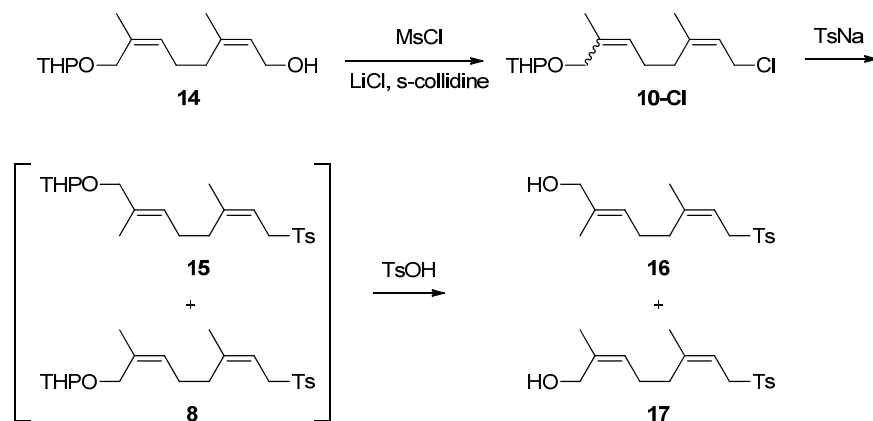


Triethylamine (17 mL, 122 mmol) was added to a solution of **14** (26 g, 102 mmol) in anhydrous THF (300 mL) and the solution was cooled down in an ice-water bath. To this solution, MsCl (9.4 mL, 122 mmol) was added dropwise. After stirring for 1 h in an ice-water bath, the reaction mixture was quickly filtered through a layer of Celite to a pre-cooled THF solution of LiBr (10.4 g, 120 mmol in 20 mL of THF) and Celite pad was quickly washed with THF (2 × 100 mL). The combined solution was stirred for 2 h in an ice-water bath. The volatiles were evaporated under reduced pressure. The crude bromide was taken up in hexanes/ethyl acetate mixture (1:1, 500 mL). The solution was washed with water (300



mL) and brine (300 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure. The crude compound (**10-Br**, ~27 g) in acetonitrile (60 mL) was added dropwise to a suspension of sodium *p*-toluenesulfinate (21.4 g, 120 mmol) in DMF (250 mL) in an ice-water bath. After the reaction mixture was stirred at room temperature for 6 h, it was diluted with ethyl acetate (800 mL). The mixture was washed with water (400 mL × 2) and brine (400 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated to afford crude compound. It was purified by column chromatography (silica gel, 350 g; i.d. of column, 3 inch; ethyl acetate:hexanes, 1:5 to 1:3) to give **8** (31 g, 78%) and **8/15** mixture (3 g). A small amount of **8** was taken for the next step. Compound **8** (1.0 g, 2.5 mmol) was dissolved in MeOH (10 mL) and TsOH·H<sub>2</sub>O (0.1 g) was added. The resulting mixture was stirred for 6 h at room temperature. After addition of Et<sub>3</sub>N (50 μL), the resulting solution was filtered through a layer of silica gel and concentrated under reduced pressure. The crude compound was purified by column chromatography (silica gel, 10 g; i.d. of column, 1/2 inch; ethyl acetate:hexanes, 1:1) to give **17** (0.63 g, 80%).

#### Synthesis of compounds **16/17** from **14** using general procedures C/D/A



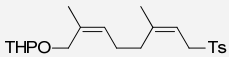
A cold solution of **14** (10 g, 39 mmol) and LiCl (4.1 g, 97 mmol) in DMF (300 mL) was treated with *s*-collidine (13 mL, 98 mmol) and MsCl (7.6 mL, 98 mmol) in an ice-water bath. After stirring at ice-water temperature for 2 h, the reaction mixture was diluted with ethyl acetate (800 mL). The mixture was washed with saturated NH<sub>4</sub>Cl (300 mL × 2) and brine (300 mL × 2), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated under reduced pressure to afford **10-Cl**. The crude compound (**10-Cl**, ~10 g)

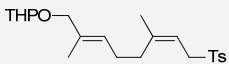
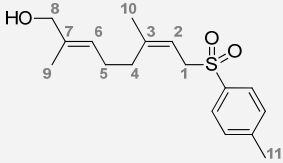
in acetonitrile (20 mL) was added dropwise to a suspension of sodium *p*-toluenesulfinate (8.2 g, 46 mmol) in DMF (200 mL) in an ice-water bath. After the reaction mixture was stirred at room temperature for 6 h, it was diluted with ethyl acetate (600 mL). The mixture was washed with water (200 mL  $\times$  2) and brine (200 mL), dried over anhydrous MgSO<sub>4</sub>, filtered, and concentrated to afford a mixture of compounds **8** and **15** (8.2 g, 53% combined yield). NMR analysis showed a 5:4 ratio of **8**:**15**. A sample of the mixture (1.0 g) was purified by column chromatography (silica gel, 15 g; i.d. of column, 1/2 inch; ethyl acetate:hexanes, 1:5) to give **8** (0.35 g) and **15** (0.30 g).

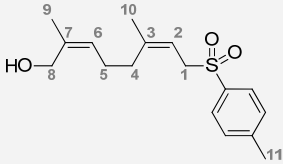
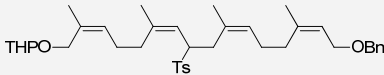
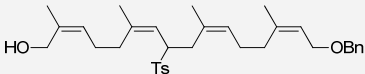
The **8/15** mixture (2.0 g, 5.1 mmol) was dissolved in MeOH (20 mL) and TsOH·H<sub>2</sub>O (0.2 g) was added. The resulting mixture was stirred for 6 h at room temperature. After addition of Et<sub>3</sub>N (100  $\mu$ L), the resulting solution was filtered through a layer of silica gel and concentrated under reduced pressure. The crude compound was purified by column chromatography (silica gel, 30 g; i.d. of column, 3/4 inch; ethyl acetate:hexanes, 1:1) to give **16/17** as a mixture (1.3 g, 81% combined yield). The mixture was subjected to column chromatography (silica gel, 20 g; i.d. of column, 1/2 inch; ethyl acetate:hexanes, 2:3) to give **16** (0.30 g), **17** (0.40 g), and a **16/17** mixture (0.50 g).

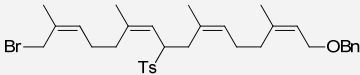
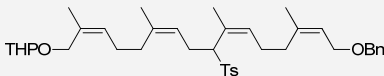
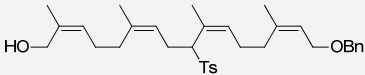
### Table S1. Characterization data of synthesized compounds.

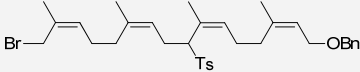
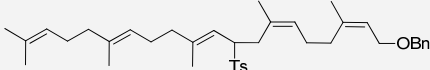
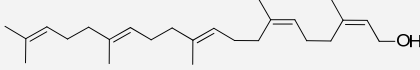
Spectral data of synthesized compounds are given in Table S1. The used preparation procedure used for each compound is indicated. When the compound was known in the literature, the literature is given.

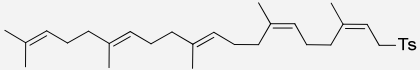
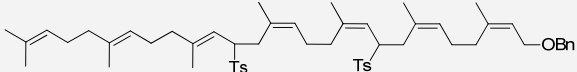
Compound No	
<b>8</b>	
Procedure used	B1 or B3 followed by D
Yield	31 g (78%, B1) and 2 g (57%, B3) from <b>14</b>
Lit	Ref <sup>1</sup> No spectroscopic characterization is given.
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) $\delta$ 1.46 - 1.61 (m, 4H), 1.64 - 1.68 (m, 1H), 1.70 (2 $\times$ s, 6H), 1.73 - 1.83 (m, 3H), 1.93 (q, <i>J</i> = 7.6 Hz, 2H), 2.41 (s, 3H), 3.45 - 3.51 (m, 1H), 3.77 (dd, <i>J</i> =

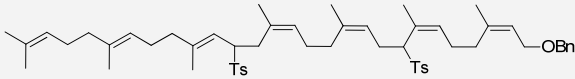
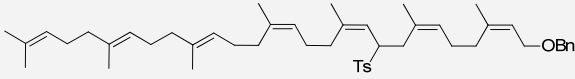
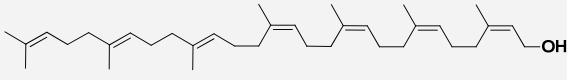
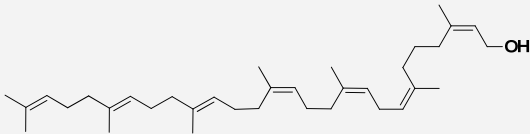
<sup>13</sup> C NMR	7.8, 3.2 Hz, 2H), 3.80 - 3.86 (m, 1H), 4.00 (dd, <i>J</i> = 20.5, 11.4 Hz, 2H), 4.53 (t, <i>J</i> = 3.4 Hz, 1H), 5.18 (q, <i>J</i> = 7.2 Hz, 2H), 7.30 (d, <i>J</i> = 8.0 Hz, 2H), 7.72 (d, <i>J</i> = 8.2 Hz, 2H) (126 MHz, CDCl <sub>3</sub> ) δ 19.3 (CH <sub>2</sub> ), 21.5 (CH <sub>3</sub> ), 21.6 (CH <sub>3</sub> ), 23.4 (CH <sub>3</sub> ), 25.4 (CH <sub>2</sub> ), 25.5 (CH <sub>2</sub> ), 30.5 (CH <sub>2</sub> ), 31.9 (CH <sub>2</sub> ), 55.7 (CH <sub>2</sub> ), 62.0 (CH <sub>2</sub> ), 65.0 (CH <sub>2</sub> ), 97.1 (CH), 111.2 (vinyl CH), 128.1 (vinyl CH), 128.3 (CH), 129.5 (CH), 132.6 (C), 135.8 (C), 144.4 (C), 145.3 (C)
HRMS (ESI)	calcd for C <sub>22</sub> H <sub>32</sub> NaO <sub>4</sub> S (M+Na <sup>+</sup> ) 415.1914, found 415.1912
<b>15</b>	
Procedure used	C followed by D
Yield	8.2 g (53%, as 5:4 mixture of <b>8</b> : <b>15</b> ) from <b>14</b> 0.3 g ( <b>15</b> , isolated from 1.0 g of <b>8</b> / <b>15</b> mixture)
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) δ 1.43 - 1.55 (m, 4H), 1.56 (s, 3H), 1.62 - 1.68 (m, 1H), 1.69 (s, 3H), 1.78 (t, <i>J</i> = 7.7 Hz, 3H), 1.84 - 1.94 (m, 2H), 2.39 (s, 3H), 3.41 - 3.51 (m, 1H), 3.71 - 3.77 (m, 2H), 3.78 - 3.86 (m, 1H), 4.02 (d, <i>J</i> = 11.6 Hz, 1H), 4.53 (t, <i>J</i> = 3.6 Hz, 1H), 5.16 (t, <i>J</i> = 7.7 Hz, 1H), 5.24 (dd, <i>J</i> = 7.0, 6.2 Hz, 1H), 7.28 (d, <i>J</i> = 8.2 Hz, 2H), 7.69 (d, <i>J</i> = 8.2 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 13.9 (CH <sub>3</sub> ), 19.3 (CH <sub>2</sub> ), 21.5 (CH <sub>3</sub> ), 23.3 (CH <sub>3</sub> ), 25.3 (CH <sub>2</sub> ), 25.5 (CH <sub>2</sub> ), 30.5 (CH <sub>2</sub> ), 31.3 (CH <sub>2</sub> ), 55.8 (CH <sub>2</sub> ), 62.0 (CH <sub>2</sub> ), 72.5 (CH <sub>2</sub> ), 97.4 (CH), 111.0 (vinyl CH), 126.3 (vinyl CH), 128.2 (CH), 129.4 (CH), 132.6 (C), 135.7 (C), 144.3 (C), 145.4 (C)
HRMS (ESI)	calcd for C <sub>22</sub> H <sub>32</sub> NaO <sub>4</sub> S (M+Na <sup>+</sup> ) 415.1914, found 415.1902
<b>16</b>	
Procedure used	A
Yield	1.3 g (81%) from <b>8</b> / <b>15</b> mixture 0.3 g ( <b>16</b> , isolated from <b>16</b> / <b>17</b> mixture)
<sup>1</sup> H NMR	(600 MHz, DMSO- <i>d</i> <sub>6</sub> ) δ 1.47 (s, 3H, H-9), 1.67 (s, 3H, H-10), 1.81 (s, 4H, H-4, H-5), 2.38 (s, 3H, H-11), 3.72 (s, 2H, H-8), 3.94 (d, <i>J</i> = 7.7 Hz, 2H, H-1), 5.05 (t, <i>J</i> = 7.8 Hz, 1H, H-2), 5.20 (br. s., 1H, H-6), 7.41 (d, <i>J</i> = 7.9 Hz, 2H), 7.71 (d, <i>J</i> = 8.3 Hz, 2H)
<sup>13</sup> C NMR	(151 MHz, DMSO- <i>d</i> <sub>6</sub> ) δ 13.4 (C-9), 21.0 (C-11), 23.2 (C-10), 25.2, 31.2 (C-4, C-5), 54.8 (C-1), 66.3 (C-8), 111.3 (C-2), 122.7 (C-6), 127.9, 129.6, 135.7, 136.1, 144.1, 144.7
HRMS (ESI)	calcd for C <sub>17</sub> H <sub>24</sub> NaO <sub>3</sub> S (M+Na <sup>+</sup> ) 331.1338, found 331.1334

<b>17</b>	
Procedure used	A
Yield	0.6 g (80%) from <b>8</b>
Lit	Ref <sup>1</sup> No spectroscopic characterization is given.
<sup>1</sup> H NMR	(600 MHz, CDCl <sub>3</sub> ) δ 1.69 (s, 3H, H-10), 1.72 (s, 3H, H-9), 1.76 - 1.86 (m, 2H, H-4), 1.89 - 2.03 (m, 2H, H-5), 2.40 (s, 3H, H-11), 3.74 (d, <i>J</i> = 7.9 Hz, 2H, H-1), 4.00 (s, 2H, H-8), 5.12 (q, <i>J</i> = 8.2 Hz, 2H, H-6, H-2), 7.29 (d, <i>J</i> = 8.1 Hz, 2H), 7.69 (d, <i>J</i> = 8.1 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 21.3 (C-9), 21.5 (C-11), 23.4 (C-10), 25.5 (C-5), 32.0 (C-4), 55.7 (C-1), 61.0 (C-8), 110.9 (C-2), 126.7 (C-6), 128.2, 129.5, 135.3, 135.7, 144.5, 145.7
HRMS (ESI)	calcd for C <sub>17</sub> H <sub>24</sub> NaO <sub>3</sub> S (M+Na <sup>+</sup> ) 331.1338, found 331.1330
<b>18</b>	
Procedure used	E
Yield	35 g (63%) from <b>8</b>
Lit	Ref <sup>1</sup> No spectroscopic characterization is given.
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) δ 1.60, 1.64, 1.72 (4 × s, 12H), 1.44 - 1.91 (m, 10H), 1.92 - 2.10 (m, 4H), 2.41 (s, 3H), 2.46 (dt, <i>J</i> = 11.4, 6.6 Hz, 1H), 2.68 (d, <i>J</i> = 13.2 Hz, 1H), 3.44 - 3.57 (m, 1H), 3.80 - 3.90 (m, 2H), 3.91 - 4.07 (m, 4H), 4.41 - 4.51 (m, 2H), 4.54 (br. s., 1H), 4.95 (d, <i>J</i> = 10.6 Hz, 1H), 5.07 - 5.20 (m, 2H), 5.41 (t, <i>J</i> = 6.4 Hz, 1H), 7.21 - 7.40 (m, 6H), 7.66 - 7.74 (m, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 19.3, 19.4, 21.5, 21.6, 23.2, 23.4, 23.5, 25.4, 25.4, 26.4, 30.1, 30.1, 30.5, 32.0, 32.1, 32.1, 62.0, 63.1, 63.2, 65.0, 65.0, 66.2, 72.0, 97.2, 97.3, 117.8, 122.1, 127.4, 127.7, 128.0, 128.1, 128.1, 128.2, 129.1, 129.3, 130.4, 132.4, 132.4, 134.8, 138.4, 139.9, 144.3, 144.6, 144.6
HRMS (ESI)	calcd for C <sub>39</sub> H <sub>54</sub> NaO <sub>5</sub> S (M+Na <sup>+</sup> ) 657.3584, found 657.3587
<b>18-OH</b>	
Procedure used	A
Yield	14 g (93%) from <b>18</b>
Lit	Ref <sup>1</sup>

<sup>1</sup> H NMR/ <sup>13</sup> C NMR HRMS (ESI)	No spectroscopic characterization is given. Full assignment is given in Table S3. calcd for C <sub>34</sub> H <sub>46</sub> NaO <sub>4</sub> S (M+Na <sup>+</sup> ) 573.3009, found 573.3010
<b>19</b>	
Procedure used	B2
Yield	10 g (74%) from <b>18-OH</b>
Lit	Ref <sup>d1</sup> No spectroscopic characterization
<sup>1</sup> H NMR	<sup>1</sup> H NMR (600 MHz, CDCl <sub>3</sub> ) δ 1.59, 1.68, 1.71, 1.78 (4 × s, 12H), 1.65 - 1.91 (m, 4H), 1.94 - 2.11 (m, 4H), 2.42 (s, 3H), 2.66 (d, <i>J</i> = 13.2 Hz, 1H), 3.79 - 3.94 (m, 3H), 3.97 (d, <i>J</i> = 6.8 Hz, 2H), 4.48 (s, 2H), 4.99 (d, <i>J</i> = 10.6 Hz, 1H), 5.12 - 5.27 (m, 2H), 5.40 (t, <i>J</i> = 6.1 Hz, 1H), 7.23 - 7.37 (m, 6H), 7.72 (d, <i>J</i> = 8.1 Hz, 2H)
<sup>13</sup> C NMR	(151 MHz, CDCl <sub>3</sub> ) δ 21.6, 21.8, 23.3, 23.4, 23.7, 26.0, 26.4, 30.4, 31.4, 31.9, 32.0, 63.3, 66.2, 72.0, 118.0, 122.1, 127.4, 127.7, 128.1, 128.2, 129.1, 129.3, 130.2, 130.4, 132.2, 134.8, 138.4, 139.8, 144.2, 144.4
HRMS (ESI)	calcd for C <sub>34</sub> H <sub>45</sub> BrNaO <sub>3</sub> S (M+Na <sup>+</sup> ) 635.2165, found 635.2170
<b>20</b>	
Procedure used	E
Yield	38 g (83%) from <b>11</b>
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) δ 1.61, 1.63, 1.77, 1.79 (4 × s, 12H), 1.47 - 1.90 (m, 10H), 1.96 - 2.06 (m, 2H), 2.11 (q, <i>J</i> = 7.3 Hz, 2H), 2.41 (s, 3H), 2.59 (m, <i>J</i> = 11.7, 6.3 Hz, 1H), 2.74 - 2.82 (m, 1H), 3.51 (dd, <i>J</i> = 11.0, 5.0 Hz, 1H), 3.85 - 3.99 (m, 3H), 3.99 - 4.13 (m, 2H), 4.43 - 4.50 (m, 2H), 4.59 (t, <i>J</i> = 3.1 Hz, 1H), 4.82 (t, <i>J</i> = 6.4 Hz, 1H), 5.29 (t, <i>J</i> = 6.9 Hz, 1H), 5.33 - 5.45 (m, 2H), 7.24 - 7.39 (m, 6H), 7.71 (d, <i>J</i> = 8.0 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 19.4 (CH <sub>2</sub> ), 21.5 (CH <sub>3</sub> ), 21.7 (CH <sub>3</sub> ), 23.2 (CH <sub>3</sub> ), 23.3 (CH <sub>2</sub> ), 25.4 (CH <sub>2</sub> ), 25.8 (CH <sub>2</sub> ), 25.9 (CH <sub>2</sub> ), 30.6 (CH <sub>2</sub> ), 31.3 (CH <sub>2</sub> ), 32.1 (CH <sub>2</sub> ), 62.0 (CH <sub>2</sub> ), 62.1 (CH <sub>2</sub> ), 65.1 (CH <sub>2</sub> ), 66.1 (CH, CH <sub>2</sub> ), 72.1 (CH <sub>2</sub> ), 97.3 (CH), 119.4, 122.4, 126.3, 127.5, 127.6, 128.3, 128.6, 128.7, 129.4, 132.3, 135.1, 135.3, 138.1, 138.3, 139.3, 144.3
HRMS (ESI)	calcd for C <sub>39</sub> H <sub>54</sub> NaO <sub>5</sub> S (M+Na <sup>+</sup> ) 657.3584, found 657.3591
<b>20-OH</b>	
Procedure used	A
Yield	16 g (80%) from <b>20</b>
<sup>1</sup> H NMR/ <sup>13</sup> C NMR	Full assignment is given in Table S3.

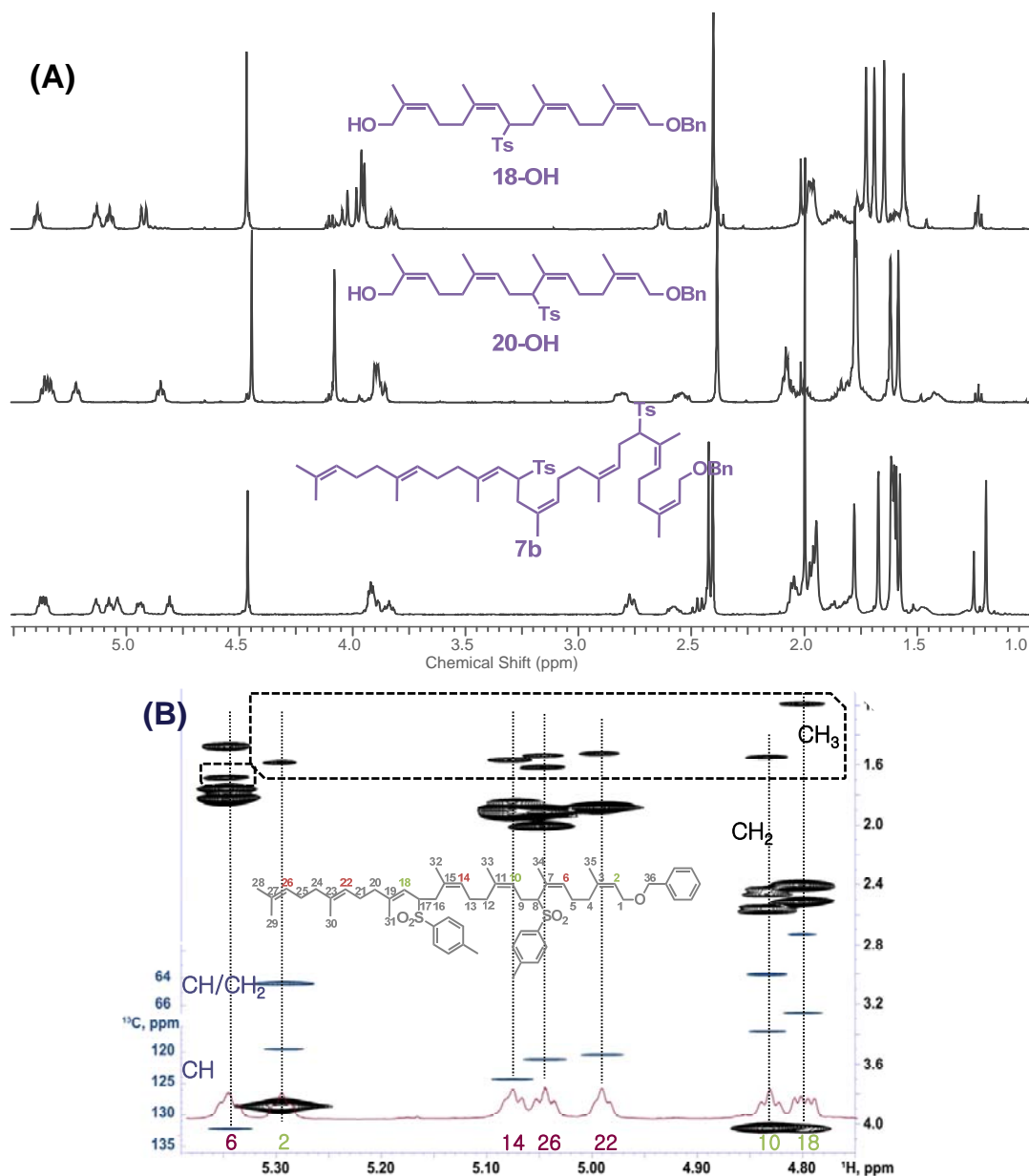
HRMS (ESI)	calcd for C <sub>34</sub> H <sub>46</sub> NaO <sub>4</sub> S (M+Na <sup>+</sup> ) 573.3009, found 573.2995
<b>21</b>	
Procedure used	B1 or B2
Yield	3.4 g (55%) or 10.8 (88%) from <b>20-OH</b>
<sup>1</sup> H NMR	<sup>1</sup> H NMR (500 MHz, CDCl <sub>3</sub> ) δ 1.40 - 1.57 (m, 1H), 1.64, 1.68, 1.82, 1.86 (4 × s, 12H), 1.73 - 1.94 (m, 3H), 2.01 - 2.19 (m, 4H), 2.44 (s, 3H), 2.56 - 2.67 (m, 1H), 2.78 - 2.88 (m, 1H), 3.90 - 4.03 (m, 5H), 4.49 (s, 2H), 4.88 (t, <i>J</i> = 6.5 Hz, 1H), 5.27 - 5.45 (m, 3H), 7.28 - 7.40 (m, 7H), 7.74 (d, <i>J</i> = 8.2 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 21.6, 21.9, 23.3, 23.4, 26.0, 26.3, 31.4, 31.4, 32.0, 66.2, 66.3, 72.1, 119.8, 122.4, 126.4, 127.5, 127.7, 128.3, 128.8, 129.4, 130.6, 132.1, 135.3, 137.7, 138.3, 139.4, 144.4
HRMS (ESI)	calcd for C <sub>34</sub> H <sub>45</sub> BrNaO <sub>3</sub> S (M+Na <sup>+</sup> ) 635.2165, found 635.2170
<b>22</b>	
Procedure used	E
Yield	9.0 g (71%) from <b>12</b>
<sup>1</sup> H NMR	(500 MHz, acetone- <i>d</i> <sub>6</sub> ) δ 1.58, 1.59, 1.61, 1.65, 1.74 (5 × s, 15H), 1.09 - 1.30 (m, 3H), 1.59 (d, <i>J</i> = 4.6 Hz, 6H), 1.61 (s, 3H), 1.65 (s, 4H), 1.74 (s, 3H), 1.92 - 2.12 (m, 14H), 2.43 (s, 3H), 2.50 (dd, <i>J</i> = 13.0, 11.4 Hz, 1H), 2.70 (dd, <i>J</i> = 13.2, 2.4 Hz, 1H), 3.98 (td, <i>J</i> = 11.0, 3.2 Hz, 1H), 4.02 (d, <i>J</i> = 6.8 Hz, 2H), 4.48 (s, 2H), 4.95 (d, <i>J</i> = 10.0 Hz, 1H), 5.06 - 5.16 (m, 2H), 5.21 (br. s., 1H), 5.33 - 5.44 (m, 1H), 7.23 - 7.29 (m, 1H), 7.31 - 7.37 (m, 3H), 7.42 (d, <i>J</i> = 8.2 Hz, 2H), 7.75 (d, <i>J</i> = 8.2 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 15.9 (CH <sub>3</sub> ), 16.3 (CH <sub>3</sub> ), 17.6 (CH <sub>3</sub> ), 21.6 (CH <sub>3</sub> ), 23.4 (CH <sub>3</sub> ), 23.4 (CH <sub>3</sub> ), 25.6 (CH <sub>3</sub> ), 26.1 (CH <sub>2</sub> ), 26.4 (CH <sub>2</sub> ), 26.6 (CH <sub>2</sub> ), 29.6 (CH <sub>2</sub> ), 32.1 (CH <sub>2</sub> ), 39.6 (CH <sub>2</sub> ), 39.7 (CH <sub>2</sub> ), 63.5 (CH), 66.3 (CH <sub>2</sub> ), 72.1 (CH <sub>2</sub> ), 117.0 (CH), 122.1 (CH), 123.3 (CH), 124.1 (CH), 127.4 (CH), 127.7 (CH), 127.7 (CH), 127.8 (CH), 128.3 (CH), 129.1 (CH), 129.3 (CH), 130.6 (CH), 131.3 (CH), 134.9, 135.5, 138.4, 139.9, 144.2, 145.1
HRMS (ESI)	calcd for C <sub>39</sub> H <sub>54</sub> NaO <sub>3</sub> S (M+Na) 625.3686, found 625.3680
<b>23</b>	
Procedure used	F
Yield	3.6 g (71%) from <b>22</b>
Lit	Ref <sup>8</sup>
<sup>1</sup> H NMR	<sup>1</sup> H, <sup>13</sup> C, HRMS (500 MHz, CDCl <sub>3</sub> ) δ 1.59, 1.60, 1.67, 1.68, 1.73 (6 × s, 18H), 1.93 - 2.11 (m, 16H),

<sup>13</sup> C NMR	4.04 - 4.19 (m, 2H), 5.06 - 5.16 (m, 4H), 5.43 (t, <i>J</i> = 6.8 Hz, 1H) (126 MHz, CDCl <sub>3</sub> ) δ 15.9 (CH <sub>3</sub> ), 17.6 (CH <sub>3</sub> ), 23.3 (CH <sub>3</sub> ), 23.4 (CH <sub>3</sub> ), 25.6 (CH <sub>3</sub> ), 26.2 (CH <sub>2</sub> ), 26.5 (CH <sub>2</sub> ), 26.7 (CH <sub>2</sub> ), 31.8 (CH <sub>2</sub> ), 32.1 (CH <sub>2</sub> ), 39.6 (CH <sub>2</sub> ), 58.8 (CH <sub>2</sub> ), 123.9 (CH), 124.1 (CH), 124.3 (CH), 124.4 (CH), 124.5 (CH), 131.1, 134.8, 135.1, 136.0, 139.5
HRMS (ESI)	calcd for C <sub>25</sub> H <sub>42</sub> NaO (M+Na) 381.3128, found 381.2975
<b>24</b>	
Procedure used	B2 followed by D
Yield	2.6 g (55%) from <b>23</b>
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) δ 1.59, 1.65, 1.67, 1.72 (6 × s, 18H), 1.74 - 1.81 (m, 2H), 1.87 (q, <i>J</i> = 7.3 Hz, 3H), 1.93 - 2.09 (m, 14H), 2.43 (s, 3H), 3.77 (d, <i>J</i> = 7.8 Hz, 2H), 4.94 (t, <i>J</i> = 6.9 Hz, 1H), 5.09 (m, 3H), 5.19 (t, <i>J</i> = 7.6 Hz, 1H), 7.31 (m, <i>J</i> = 8.0 Hz, 2H), 7.73 (m, <i>J</i> = 8.2 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 15.9, 17.6, 21.6, 23.3, 23.5, 25.6, 25.7, 26.5, 26.5, 26.7, 31.8, 32.0, 39.7, 55.9, 111.0, 123.8, 123.9, 124.1, 124.3, 128.4, 129.5, 131.2, 134.9, 135.2, 135.9, 136.0, 144.4, 145.7
HRMS (ESI)	calcd for C <sub>32</sub> H <sub>48</sub> NaO <sub>2</sub> S (M+Na) 519.3267, found 519.3261
<b>7a</b>	
Procedure used	E
Yield	14 g (77%) from <b>19</b>
Lit	Ref <sup>11</sup> No spectroscopic characterization is given.
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) δ 1.41 - 1.53 (m, 2H), 1.55, 1.56, 1.58, 1.59, 1.64, 1.67, 1.72 (8 × s, 24H), 1.72 - 1.83 (m, 3H), 1.77 - 1.81 (m, 1H), 1.87 - 2.07 (m, 18H), 2.38 - 2.48 (m, 10H), 2.61 - 2.71 (m, 1H), 2.74 (d, <i>J</i> = 13.6 Hz, 1H), 3.75 - 3.89 (m, 3H), 3.93 - 4.03 (m, 2H), 4.46 - 4.51 (m, 2H), 4.87 - 5.10 (m, 7H), 5.17 (d, <i>J</i> = 8.8 Hz, 1H), 5.41 (t, <i>J</i> = 6.8 Hz, 1H), 7.24 - 7.38 (m, 11H), 7.65 - 7.76 (m, 5H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 15.9, 15.9, 16.2, 16.2, 17.6, 21.6, 23.2, 23.2, 23.3, 23.4, 23.4, 23.5, 25.6, 25.6, 26.1, 26.1, 26.5, 26.6, 29.4, 29.5, 30.2, 31.7, 31.8, 32.1, 39.6, 39.7, 39.7, 56.0, 63.1, 63.3, 63.3, 66.3, 72.0, 117.0, 117.8, 117.8, 122.1, 123.3, 124.1, 124.1, 127.3, 127.4, 127.7, 128.0, 128.1, 128.3, 128.5, 129.1, 129.2, 129.3, 129.3, 129.3, 129.5, 130.4, 130.7, 130.8, 131.3, 134.7, 134.8, 135.5, 135.6, 138.4, 139.9, 144.3, 144.3, 144.4, 144.5, 145.0
HRMS (ESI)	calcd for C <sub>56</sub> H <sub>76</sub> NaO <sub>5</sub> S <sub>2</sub> (M+Na) 915.5026, found 915.5019

<b>7b</b>	
Procedure used	E
Yield	17 g (83%) from <b>21</b>
<sup>1</sup> H NMR/ <sup>13</sup> C NMR	Full assignment is given in Table S4.
HRMS (ESI)	calcd for C <sub>56</sub> H <sub>76</sub> O <sub>5</sub> S <sub>2</sub> (M-H <sup>-</sup> ) 891.5050, found 891.5070
<b>7c</b>	
Procedure used	E
Yield	3.6 g (93%) from <b>24</b>
<sup>1</sup> H NMR	(500 MHz, CDCl <sub>3</sub> ) δ 1.43 - 1.54 (m, 2H), 1.60, 1.64, 1.65, 1.68, 1.73 (8 × s, 24H), 1.74 - 1.82 (m, 2H), 1.87 - 2.12 (m, 16H), 2.41 (s, 3H), 2.46 (dd, <i>J</i> = 13.0, 11.6 Hz, 1H), 2.72 (dd, <i>J</i> = 13.2, 1.8 Hz, 1H), 3.85 (td, <i>J</i> = 10.9, 2.6 Hz, 1H), 3.99 (d, <i>J</i> = 6.8 Hz, 2H), 4.49 (s, 2H), 4.90 (t, <i>J</i> = 6.0 Hz, 1H), 4.97 (d, <i>J</i> = 10.6 Hz, 1H), 5.07 - 5.27 (m, 4H), 5.42 (t, <i>J</i> = 6.4 Hz, 1H), 7.24 - 7.41 (m, 7H), 7.71 (d, <i>J</i> = 8.2 Hz, 2H)
<sup>13</sup> C NMR	(126 MHz, CDCl <sub>3</sub> ) δ 15.9, 17.5, 21.4, 23.2, 23.2, 23.3, 23.5, 25.6, 26.4, 26.4, 26.5, 26.6, 30.0, 31.7, 32.0, 32.1, 39.6, 39.6, 63.1, 66.2, 72.0, 117.5, 122.0, 123.8, 123.9, 124.0, 124.2, 127.3, 127.6, 127.9, 128.2, 129.0, 129.2, 130.4, 134.8, 135.1, 135.6, 138.4, 139.8, 144.1, 144.8
HRMS (ESI)	calcd for C <sub>49</sub> H <sub>70</sub> NaO <sub>3</sub> S (M+Na) 761.4938, found 761.4942
<b>4</b>	
Procedure used	F
Yield	2.0 g, 59% from <b>7a</b> (>90% purity) 2.8 g, 63% (as 1:3 mixture of <b>4:4B2</b> ) from <b>7b</b> 0.2 g ( <b>4</b> , isolated from <b>4:4B2</b> mixture) 1.6 g, 68% from <b>7c</b> (>91% purity)
Lit	Compound <b>4</b> was isolated as colorless oil and was stored at 4 °C. Ref <sup>8</sup> <sup>1</sup> H, <sup>13</sup> C, HRMS are given.
<sup>1</sup> H NMR/ <sup>13</sup> C NMR	Full assignment is given in Table S2.
HRMS (ESI)	calcd for C <sub>35</sub> H <sub>58</sub> NaO (M+Na <sup>+</sup> ) 517.4380, found 517.4348
<b>4B2</b>	
Procedure used	F
Yield	2.8 g, 63% (as 1:3 mixture of <b>4:4B2</b> ) from <b>7b</b>



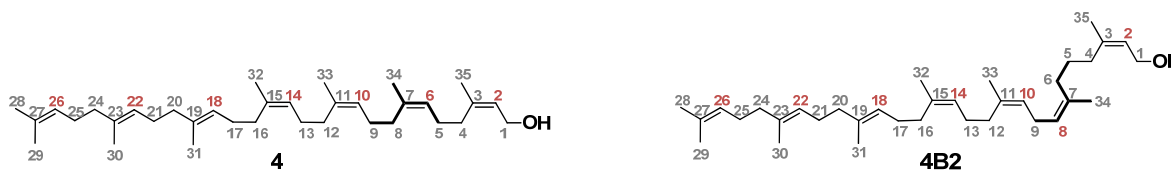
<sup>1</sup> H NMR/ <sup>13</sup> C NMR	0.8 g ( <b>4B2</b> , isolated from <b>4/4B2</b> mixture)
HRMS (ESI)	Full assignment is given in Table S2. calcd for C <sub>35</sub> H <sub>58</sub> NaO (M+Na <sup>+</sup> ) 517.4380, found 517.4340



**Figure S1.**  $^1\text{H}$  NMR spectra of intermediates (**18-OH**, **20-OH** and **7b**) (A).  $^1\text{H}$ - $^{13}\text{C}$ -HSQC-TOCSY (crosspeaks in blue) and TOCSY (crosspeak in black) of **7b** (800.13 MHz).

Stereochemical verification within each route was by NMR. Notably, intermediates **18-OH**, **20-OH** and **7b** had well-resolved  $^1\text{H}$  resonances (Figure S1A, Tables 1 and 2). Figure S1B further shows the  $^1\text{H}$ - $^{13}\text{C}$ -HSQC-TOCSY spectrum of **7b** (crosspeaks in blue with respect to the y-axis on the left) and TOCSY spectrum (crosspeaks in black with respect to the y-axis on the right). The seven vinyl resonances (of H-2, -6, -10, -14, -18, -22, and -28) in **7b** were each resolved, and each resonance showed crosspeaks to the proximal  $\text{CH}/\text{CH}_2/\text{CH}_3$  carbons. The  $^1\text{H}$  and  $^{13}\text{C}$  resonance assignments for three compounds are given in Tables S1 and S2.

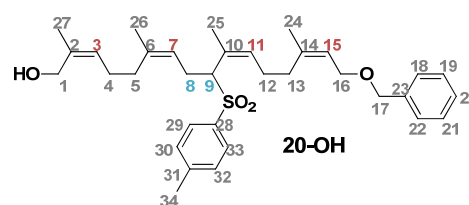
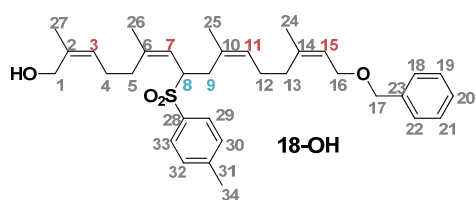
**Table S2.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data<sup>a</sup> for **4** and **4B2** in  $\text{CDCl}_3$  at 298 K (800.13 MHz, obtained using a solution of 10 mg of **4** (or **4B2**) and 3 mg of  $\text{Eu}(\text{hcf})_3$  in 600  $\mu\text{L}$  of  $\text{CDCl}_3$ ).



<b>4</b>					<b>4B2</b>			
Carbon	Type	$\delta_{\text{C}}$	$\delta_{\text{H}}$	HMBC <sup>b</sup>	Type	$\delta_{\text{C}}$	$\delta_{\text{H}}$	HMBC <sup>b</sup>
1	CH <sub>2</sub>	63.38	6.08		CH <sub>2</sub>	65.60	6.95	
2	CH	125.24	6.62		CH	125.62	7.08	
3	C	140.68			C	141.76		
4	CH <sub>2</sub>	32.81	2.70	2, 3, 5, 35	CH <sub>2</sub>	32.69	2.79	2, 3, 5, 6, 35
5	CH <sub>2</sub>	26.65	2.45	3, 4, 6, 7	CH <sub>2</sub>	27.20	1.96	3, 4, 6, 7
6	CH	124.74	5.41	5, 8, 34	CH <sub>2</sub>	40.00	2.29	4, 5, 7, 8, 34
7	C	136.20			C	135.03		
8	CH <sub>2</sub>	32.30	2.11 2.18	9, 10	CH	124.04	5.27	6, 7, 34
9	CH <sub>2</sub>	26.43	2.12 2.18	10, 11	CH <sub>2</sub>	27.19	2.77	7, 8, 10
10	CH	124.96	5.23	8, 9, 33	CH	124.27	5.18	9, 12, 33
11	C	135.33			C	135.32		
12	CH <sub>2</sub>	32.29	2.11 2.18	11, 14, 33	CH <sub>2</sub>	32.53	2.10	10, 11, 33
13	CH <sub>2</sub>	26.61	2.12 2.18	12, 14, 15	CH <sub>2</sub>	26.62	2.10	12, 14, 15
14	CH	124.93	5.19	13, 32	CH	125.21	5.17	13, 16, 32
15	C	135.40			C	135.69		
16	CH <sub>2</sub>	32.00	2.08 2.10	15, 17, 18, 32	CH <sub>2</sub>	32.26	2.07	14, 17, 18
17	CH <sub>2</sub>	26.61	2.09 2.11	16, 18, 19	CH <sub>2</sub>	26.89	2.08	15, 16, 18, 19
18	CH	124.15	5.18	16, 17, 20, 31	CH	124.43	5.16	17, 20, 31
19	C	135.13			C	135.42		
20	CH <sub>2</sub>	39.73	2.00	18, 19, 21	CH <sub>2</sub>	40.01	2.00	19, 21, 31
21	CH <sub>2</sub>	26.65	2.02 2.08	20, 22, 23	CH <sub>2</sub>	26.90	2.07	20, 22, 23
22	CH	124.23	5.14	20, 21, 24, 30	CH	124.49	5.13	18, 21, 22, 24, 30
23	C	134.87			C	135.18		
24	CH <sub>2</sub>	39.83	2.012	22, 23, 25, 26, 30	CH <sub>2</sub>	39.97	1.99	22, 23, 25, 26, 30
25	CH <sub>2</sub>	26.73	2.008 2.07	24, 26, 27	CH <sub>2</sub>	27.03	2.07	23, 26, 27
26	CH	124.38	5.12	28, 29	CH	124.66	5.11	24, 25, 28, 29
27	C	131.20			C	131.49		
28	CH <sub>3</sub>	25.69	1.70	26, 27, 29	CH <sub>3</sub>	25.96	1.69	26, 27, 29
29	CH <sub>3</sub>	17.66	1.62	26, 27, 28	CH <sub>3</sub>	17.94	1.613	26, 27, 28
30	CH <sub>3</sub>	15.98	1.62	22, 23, 24	CH <sub>3</sub>	16.25	1.611	22, 23, 24
31	CH <sub>3</sub>	15.98	1.64	18, 19, 20	CH <sub>3</sub>	16.25	1.63	18, 19, 20
32	CH <sub>3</sub>	23.47	1.73	14, 15, 16	CH <sub>3</sub>	23.722	1.717	14, 15, 16
33	CH <sub>3</sub>	23.47	1.75	10, 11, 12	CH <sub>3</sub>	23.716	1.720	10, 11, 12
34	CH <sub>3</sub>	23.44	1.79	6, 7, 8	CH <sub>3</sub>	16.33	1.75	6, 7, 8
35	CH <sub>3</sub>	23.85	2.04	2, 3, 4	CH <sub>3</sub>	24.28	2.14	2, 3, 4

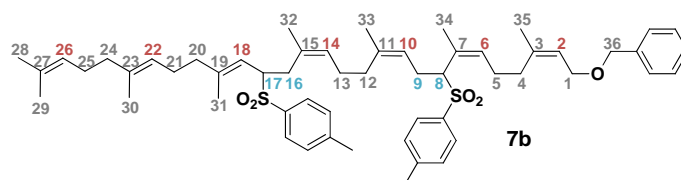
<sup>a</sup> Due to overlaps among signals, the values for the coupling constants or for  $\Sigma J$  could not be always extracted. <sup>b</sup> HMBC correlations, optimized for 8 Hz, are from the stated  $^1\text{H}$  resonance to the indicated carbon resonances.

**Table S3.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for **18-OH** and **20-OH** in DMSO at 298 K (800.13 MHz).



Carbon	Type	18-OH				20-OH			
		$\delta_{\text{C}}$	$\delta_{\text{H}}$	$^1\text{H}$ multiplicity <sup>a</sup> J(H <sub>i</sub> ,H <sub>j</sub> ) [Hz]	HMBC <sup>b</sup>	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$^1\text{H}$ multiplicity <sup>a</sup> J(H <sub>i</sub> ,H <sub>j</sub> ) [Hz]	HMBC <sup>b</sup>
1	CH <sub>2</sub>	59.78	3.83	s	2, 3, 27	59.86	3.91	bs	2, 3, 27
2	C	136.26				136.50			
3	CH	125.39	4.96	t, 6.7	1, 4, 5, 27	125.41	5.06	t, $\Sigma J = 14.6$	1, 4, 5, 27
4	CH <sub>2</sub>	25.51	1.69 1.78	mt mt	2, 3, 5, 6, 12	25.70	2.02	mt	2, 3, 5, 6
5	CH <sub>2</sub>	32.29	1.52 1.77	mt mt	4, 6, 7, 26	32.29	1.90 1.95	mt mt	3, 4, 6, 7, 26
6	C	144.88				138.02			
7	CH	117.71	4.86	d, 10.4	5, 8, 9, 26	119.91	4.84	t, $\Sigma J = 13.5$	5, 9, 26
8	CH	62.58	3.97	ddd, 10.9, 10.9, 3.0	6, 7, 9, 10	23.42 (CH <sub>2</sub> )	2.47 2.61	mt, $\Sigma J = 38.0$ mt, $\Sigma J = 26.0$	6, 7, 9, 10
9	CH <sub>2</sub>	30.31	2.34 2.49	dd, 13.0, 11.3 dd, 13.0, 2.0	7, 8, 10, 11, 25	65.61 (CH)	4.02	dd, 11.4, 3.3	7, 8, 10, 11, 25
10	C	130.79				126.15			
11	CH	128.05	5.13	t, 6.4	9, 12, 13, 25	135.60	5.35	t, $\Sigma J = 13.4$	9, 10, 12, 13, 25
12	CH <sub>2</sub>	26.43	1.93	mt	10, 11, 13, 14	25.88	1.48 1.83	mt mt	10, 11, 13, 14
13	CH <sub>2</sub>	32.07	1.93 1.97	mt mt	12, 14, 15, 24	31.38	1.76 1.82	mt mt	11, 12, 14, 15, 24
14	C	139.23				138.85			
15	CH	122.81	5.33	t, 6.7	13, 14, 16, 24	122.99	5.30	dd, 6.2, 7.1	13, 14, 16, 24
16	CH <sub>2</sub>	66.24	3.93	d, 6.7	14, 15, 17	66.24	3.87 3.89	dd, 6.2, 12.0 dd, 7.1, 12.0	14, 15, 17
17	CH <sub>2</sub>	71.52	4.41	s	16, 18, 22, 23	71.52	4.39	s	16, 18, 22, 23
18, 22	CH	127.82	7.29	d, 7.1	17, 19, 20, 21	127.69	7.27	d, 7.3	17, 20
19, 21	CH	128.59	7.32	t, 7.1	18, 20, 22, 23	128.56	7.31	t, 7.3	23
20	CH	127.75	7.26	t, 7.1	18, 19, 21, 22	127.72	7.25	t, 7.3	18, 22
23	C	139.04				139.04			
24	CH <sub>3</sub>	23.50	1.67	s	13, 14, 15	23.42	1.58	bs	13, 14, 15
25	CH <sub>3</sub>	23.82	1.56	s	9, 10, 11	19.24	1.70	bs	9, 10, 11
26	CH <sub>3</sub>	23.45	1.59	d	5, 6, 7	23.47	1.58	bs	5, 6, 7
27	CH <sub>3</sub>	21.59	1.64	s	1, 2, 3	21.66	1.69	bs	1, 2, 3
28	C	135.09				135.61			
29, 33	CH	129.22	7.70	d, 8.2	30, 31, 32	128.85	7.70	d, 8.2	31
30, 32	CH	129.95	7.41	d, 8.2	28, 29, 33, 34	130.06	7.39	d, 8.2	28, 34
31	C	144.69				144.69			
34	CH <sub>3</sub>	21.51	2.38	s	30, 31, 32	21.44	2.37	s	30, 31, 32

<sup>a</sup> Due to overlaps among signals, the values for the coupling constants or for  $\Sigma J$  could not be always extracted. <sup>b</sup> HMBC correlations, optimized for 8 Hz, are from the stated  $^1\text{H}$  resonance to the indicated carbon.

**Table S4.**  $^1\text{H}$  and  $^{13}\text{C}$  NMR data for **7b** in DMSO at 298 K (800.13 MHz).

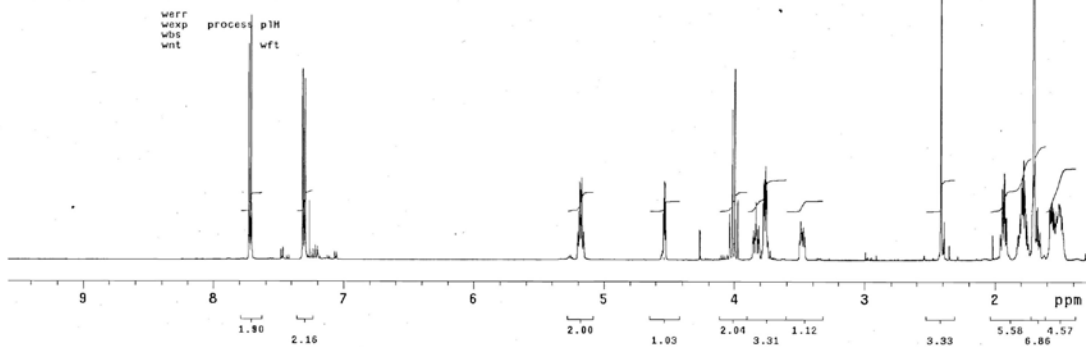
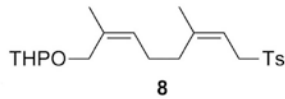
<b>7b</b>						
Carbon	Type	$\delta_{\text{C}}$	$\delta_{\text{H}}$	$^1\text{H}$ multiplicity <sup>a</sup> $J(\text{H}_i, \text{H}_j)$ , [Hz]	HMBC <sup>b</sup>	
1	CH <sub>2</sub>	65.75	3.90 3.87	dd, 11.8, 6.8 dd, 11.8, 6.8	2, 3, 36	
2	CH	122.53	5.30	t, 6.8	1, 3, 4, 35	
3	C	138.48				
4	CH <sub>2</sub>	30.94	1.76 1.82	mt mt	2, 3, 5, 6, 35	
5	CH <sub>2</sub>	25.43	1.48 1.83	mt mt	3, 4, 6, 7	
6	CH	135.14	5.35	t, 6.3	4, 5, 8, 34	
7	C	125.69				
8	CH	65.04	4.02	mt	6, 7, 9, 10, 34	
9	CH <sub>2</sub>	22.97	2.45 2.56	mt mt	7, 8, 10, 11	
10	CH	119.66	4.83	t, 6.9	8, 11, 13, 33	
11	C	137.20				
12	CH <sub>2</sub>	31.49	1.85 1.93	mt mt	10, 11, 13, 14, 33	
13	CH <sub>2</sub>	25.77	1.90 1.93	mt mt	11, 12, 14, 15	
14	CH	127.32	5.08	t, 6.8	12, 13, 16, 32	
15	C	130.49				
16	CH <sub>2</sub>	29.20	2.41 2.52	mt mt	15, 17, 18, 32	
17	CH	62.10	4.03	mt	16, 18, 19	
18	CH	116.73	4.80	ddd, 10.3, 5.1, 0.9	16, 17, 20, 31	
19	C	144.57				
20	CH <sub>2</sub>	39.09	1.88	mt	18, 19, 21, 22, 31	
21	CH <sub>2</sub>	25.60	1.89	mt	19, 20, 22, 23	
22	CH	123.36	4.99	t, 5.5	20, 21, 24, 30	
23	C	134.68				
24	CH <sub>2</sub>	39.17	1.91	mt	22, 23, 25, 26, 30	
25	CH <sub>2</sub>	26.19	2.01	mt	23, 24, 26, 27	
26	CH	124.07	5.05	t, 7.0	24, 25, 28, 29	
27	C	130.68				
28	CH <sub>3</sub>	25.46	1.62	t, 1.0	26, 27, 29	
29	CH <sub>3</sub>	17.47	1.54	bs	26, 27, 28	
30	CH <sub>3</sub>	15.69	1.52	s	22, 23, 24	
31	CH <sub>3</sub>	16.08	1.19	d, 0.9	18, 19, 20	
32	CH <sub>3</sub>	23.12	1.57	bs	15, 16, 17	
33	CH <sub>3</sub>	23.00	1.55	d, 1.0	10, 11, 12	
34	CH <sub>3</sub>	18.79	1.68	d, 5.1	6, 7, 8	
35	CH <sub>3</sub>	22.90	1.59	dd, 3.9, 1.1	2, 3, 4	
36	CH <sub>2</sub>	71.07	4.39	s	1, 37, 38, 42	
37	C	138.59				
38, 42	CH	127.33	7.28	bd, 7.3	36, 40	
39, 41	CH	128.18	7.32	t, 7.3	37	
40	CH	127.30	7.26	tt, 7.3, 1.3	38, 42	
43	C	135.10				
44	CH	128.40	7.696	d, 8.2		
45, 47	CH	129.63	7.398	dq, 8.2, 0.6		
46	C	144.32				
48	CH	128.40	7.698	d, 8.2		
49	CH <sub>3</sub>	21.04	2.37	bs		
50	C	134.68				
51, 55	CH	128.77	7.71	d, 8.2		
52, 54	CH	129.43	7.41	dq, 8.2, 0.6		
53	C	144.21				
56	CH <sub>3</sub>	21.06	2.39	bs		

## References

- (1) Bax, A.; Morris, G. A. *J. Magn. Reson.* **1981**, *42*, 501-505.
- (2) Bax, A.; Summers, M. F. *J. Am. Chem. Soc.* **1986**, *108*, 2093-2094.
- (3) Bodenhausen, G.; Freeman, R.; Niedermeyer, R.; Turner, D. L. *J. Magn. Reson.* **1977**, *26*, 133-164.
- (4) Griesinger, C.; Otting, G.; Wüthrich, K.; Ernst, R. R. *J. Am. Chem. Soc.* **1988**, *110*, 7870-7872.
- (5) Sattler, M.; Schwalbe, H.; Griesinger, C. *J. Am. Chem. Soc.* **1992**, *114*, 1126-1127.
- (6) Wijmenga, S. S.; Hallenga, K.; Hilbers, C. W. *J. Magn. Reson.* **1989**, *84*, 634-642.
- (7) Wishart, D. S.; Bigam, C. G.; Yao, J.; Abildgaard, F.; Dyson, H. J.; Oldfield, E.; Markley, J. L.; Sykes, B. D. *J. Biomol. NMR* **1995**, *6*, 135-140.
- (8) Chang, Y.-F.; Liu, C.-Y.; Guo, C.-W.; Wang, Y.-C.; Fang, J.-M.; Cheng, W.-C. *J. Org. Chem.* **2008**, *73*, 7197-7203.
- (9) Sato, K.; Inoue, S.; Onishi, A.; Uchida, N.; Minowa, N. *J. Chem. Soc., Perkin Trans. 1* **1981**, 761-769.
- (10) Williams, C. M.; Mander, L. N. *Tetrahedron* **2001**, *57*, 425-447.
- (11) Sato, K.; Miyamoto, O.; Inoue, S.; Matsushashi, Y.; Koyama, S.; Kaneko, T. *J. Chem. Soc., Chem. Commun.* **1986**, 1761-1762.

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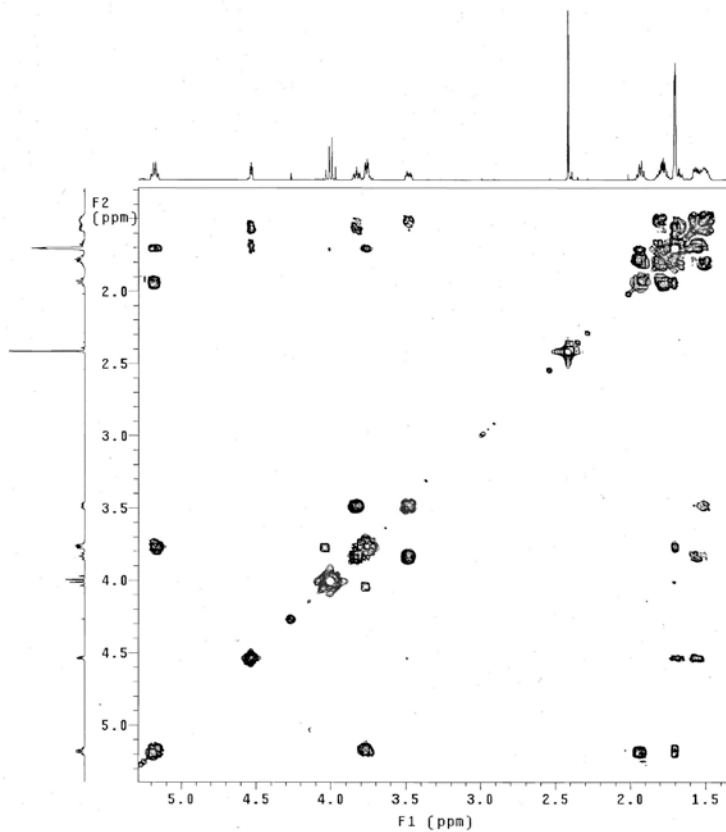
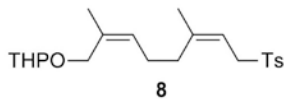
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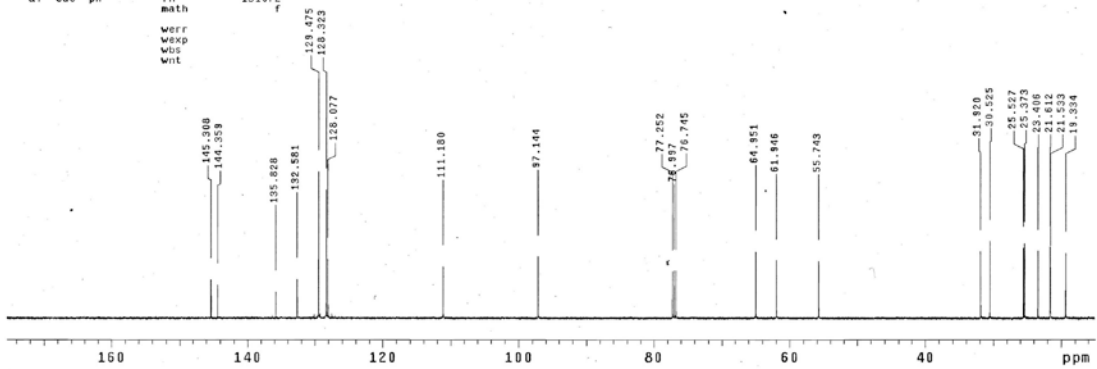
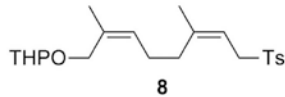
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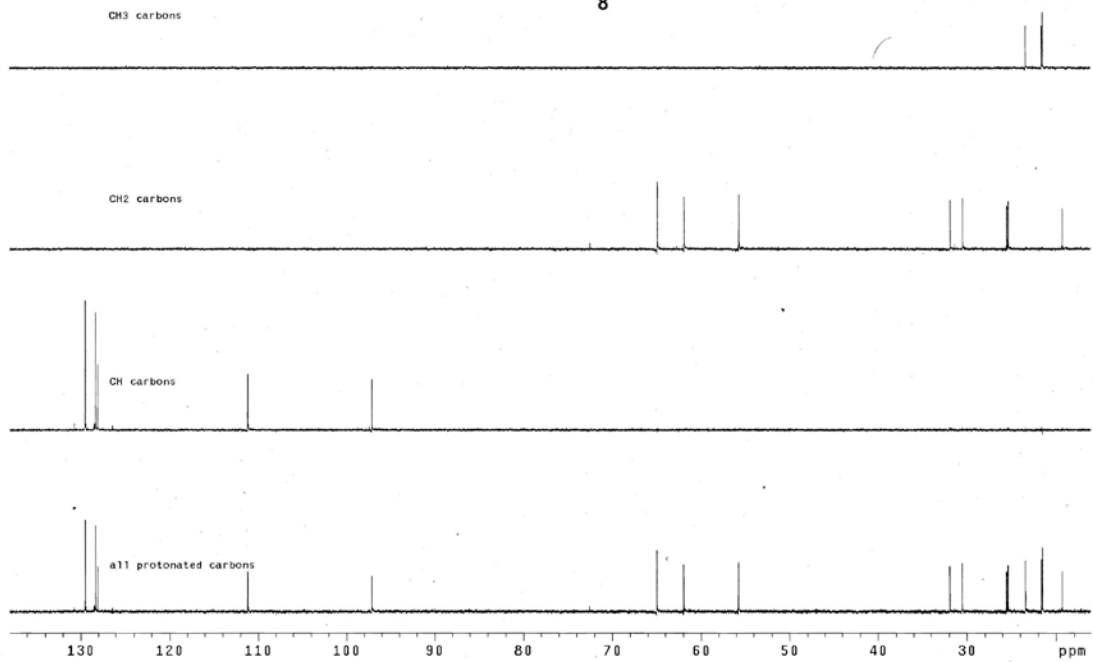
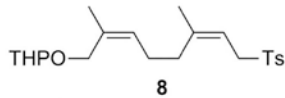
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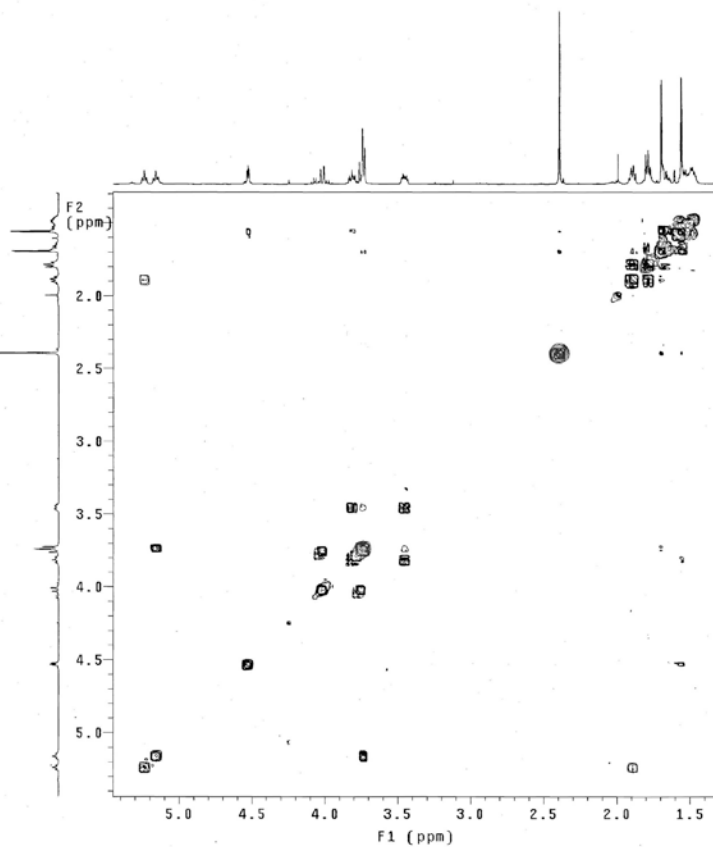
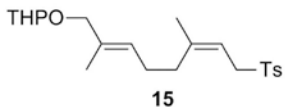
Pulse Sequence: dept





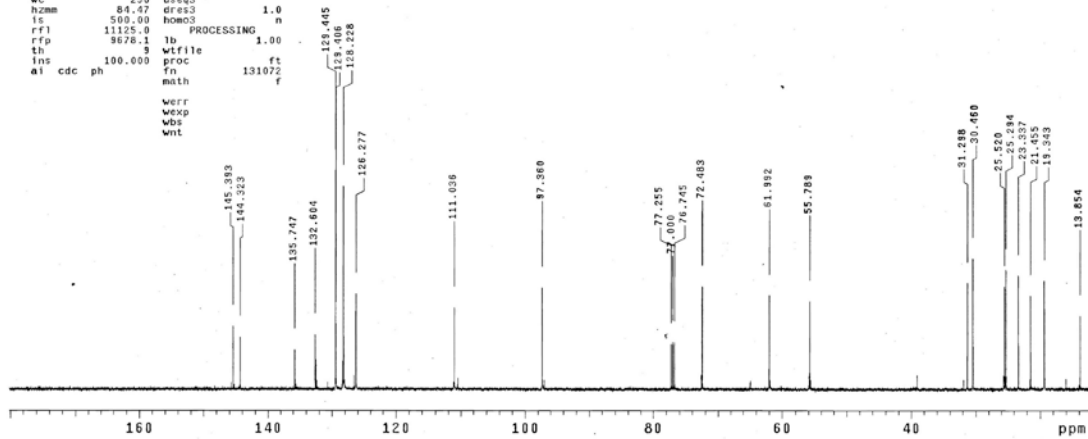
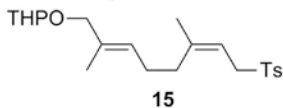


UPN-18  
Pulse Sequence: relayh  
Solvent: CDCl3  
Ambient Temperature  
INOVA-500 "nmr2a.chem.nd.edu"  
Relax. delay 1.300 sec  
COSY 80-90  
Acq. time 0.151 sec  
Width 3599.5 Hz  
2D Width 3599.5 Hz  
4 repetitions  
256 increments  
OBSERVE H1, 499.8611751 MHz  
DATA PROCESSING  
Sine bell 0.075 sec  
F1 DATA PROCESSING  
Sine bell 0.038 sec  
F1 size 1024 x 1024  
Total time 25 min, 46 sec

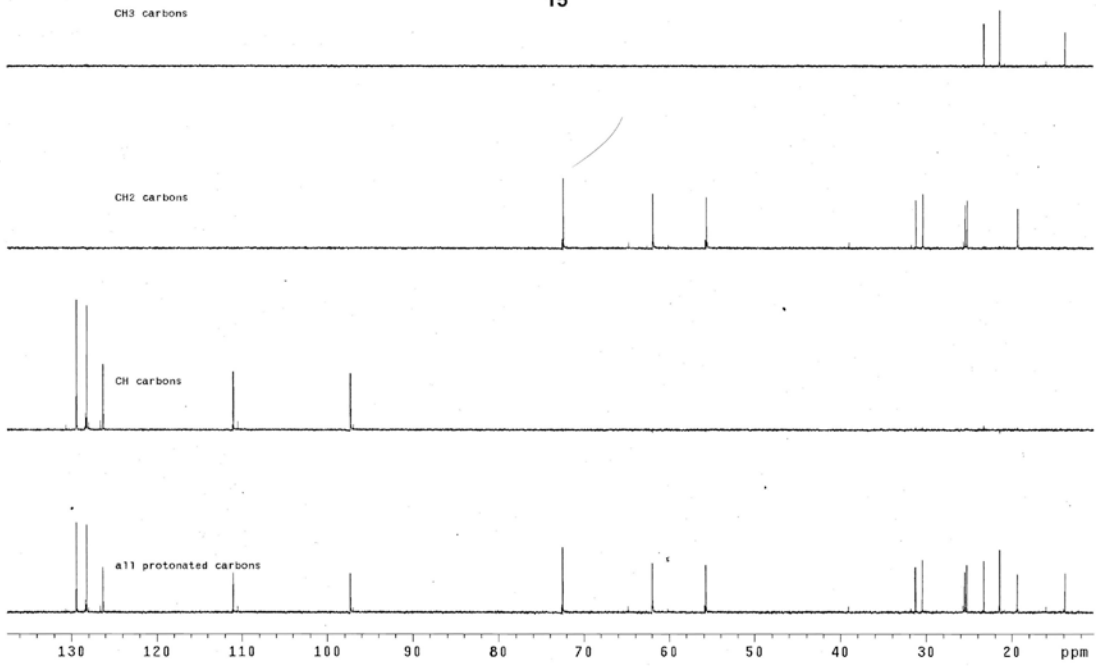
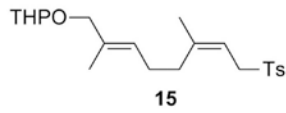


UPN-18  
exp2 s2pu1

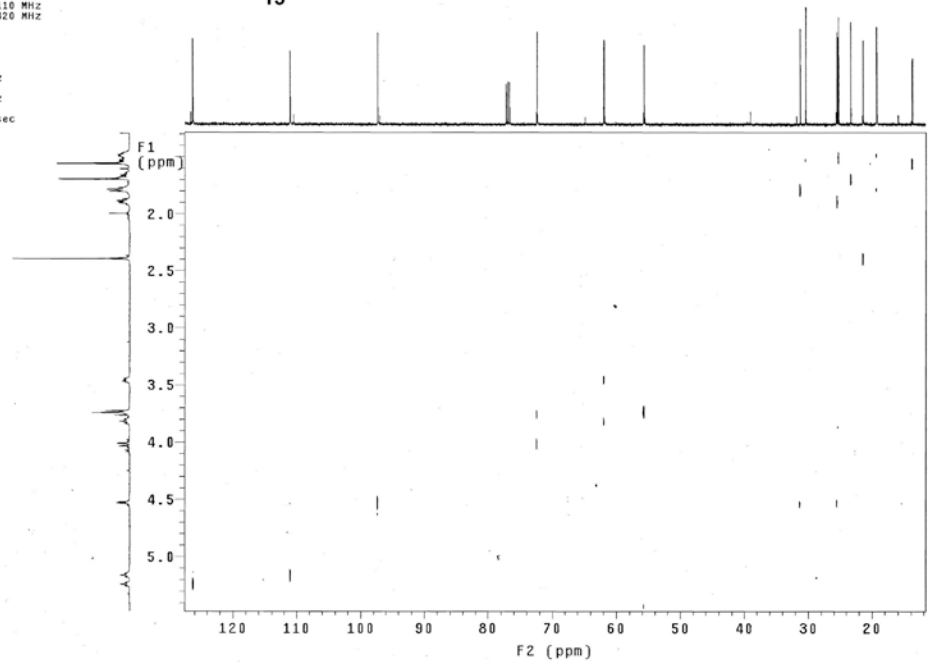
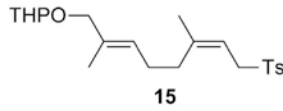
SAMPLE DEC. & VT  
date Dec 11 2008 dfrq 499.864  
solvent CDCl3 dn H1  
file exp dpr 40  
ACQUISITION  
sfrq 125.782 dm dof 0  
tn C13 dm w  
at 1.215 dmf 8787.35  
np 85536 dsq  
sw 28962.3 dres 1.0  
fb 15000 homo n  
bs 4 DEC2  
tpwr 52 dfrq2 0  
pv 18.2 dn2 1  
dl 11.800 dpr2 0  
tof 144.5 dof2 0  
nt 600 dm2 n  
ct 38 dm2 c  
alock n dm2 10000  
gain not used dsq2 1.0  
FLAGS n homo2 DEC3  
il n n  
dp y dfrq3 0  
hs nm dn3 1  
DISPLAY dpr3 0  
sp 1529.4 dof3 0  
wp 21116.5 dm3 n  
vs 81 dm3 c  
sc 0 dm3 10000  
hzam 84.47 dsq3 1.0  
ls 509.00 homo3 n  
rfl 11125.0 PROCESSING  
rfp 9678.1 lb 1.00  
th 9 wfile ft  
ins 100.000 proc fn 131072  
al cdc ph math f



UPN-18  
Pulse Sequence: dept

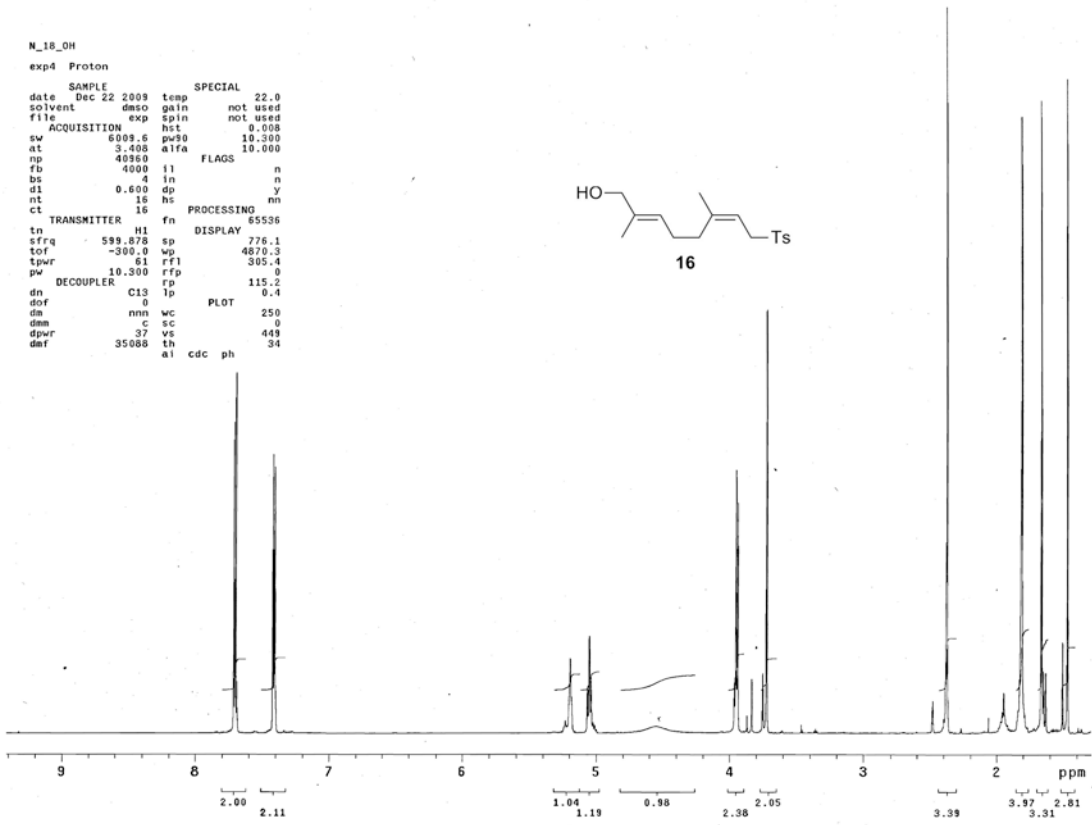
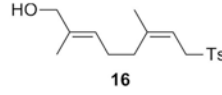


UPN-18  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"  
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18483.5 Hz  
2D Width 3476.7 Hz  
4 repetitions  
256 increments  
OBSERVE C13, 125.6902110 MHz  
DECOUPLE H1, 499.8634420 MHz  
Power 40 dB  
on during acquisition  
off during delay  
VOLTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 Data PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 28 min, 39 sec



N\_18\_OH  
exp4 Proton

date	Dec 22 2003	temp	22.0	SPECIAL
solvent	dms0	gain	not used	
file		spin	not used	
ACQUISITION				
sw	6009.6	pw90	10.300	
at	3.408	alfa	10.000	
np	40969			
fb	4000	fl	n	FLAGS
bs	4	in	n	
dt	0.600	dp	y	
nt	16	hs	nn	
ct	16			
TRANSMITTER				
tn		fn	55536	
DISPLAY				
efrq	599.878	sp	776.1	
torf	-300.0	wp	4670.3	
tpwr	61	rf1	305.4	
pw	10.300	rfp	0	
DECOUPLER				
dn	C13	lp	115.2	
dor	0		0.4	
dm	nnn	wc	250	PLOT
dmm	C	sc	0	
dpar	37	vs	449	
daf	35088	th	449	
	al	cdc	ph	34



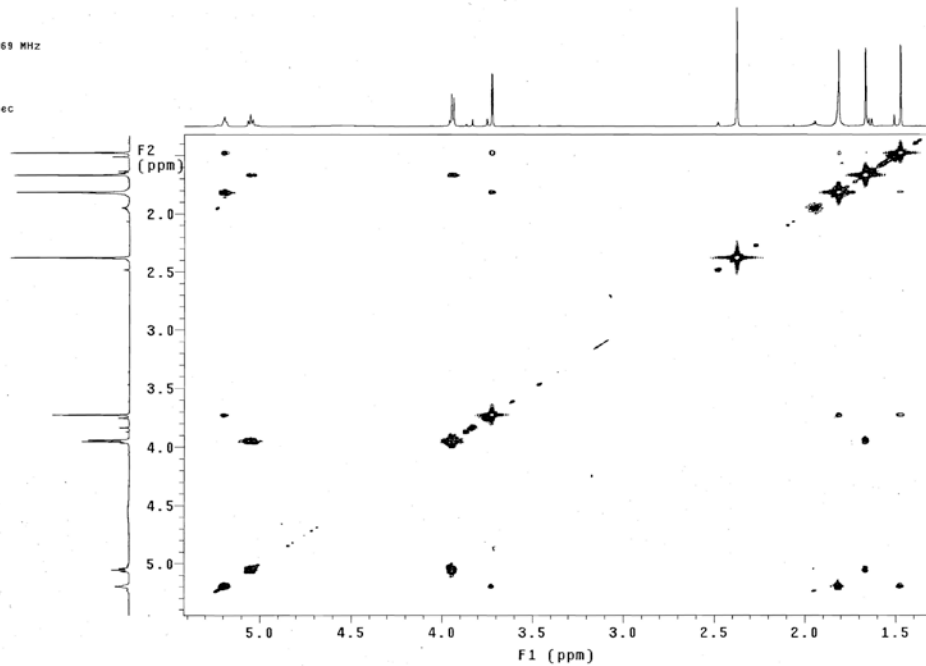
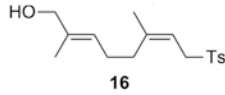
N\_18\_OH

File: afs/nd.edu/user26/dhesek/Private/600UP/DMSO-N-18\_OH-NH.fid

Pulse Sequence: COSY

Solvent: dms0  
Temp: 22.0 C / 295.1 K  
Operator: dhesek  
File: DMSO-N-18\_OH-NH  
VNMR5-600 "nmr600"

Relax. delay 1.000 sec  
Acq. time 0.127 sec  
Width 4032.3 Hz  
2D Width 4032.3 Hz  
8 repetitions  
256 increments  
OBSERVE H1, 599.8757069 MHz  
DATA PROCESSING  
Sine bell 0.083 sec  
F1 Data PROCESSING  
Sine bell 0.127 sec  
FT size 4096 x 4096  
Total time 40 min, 19 sec



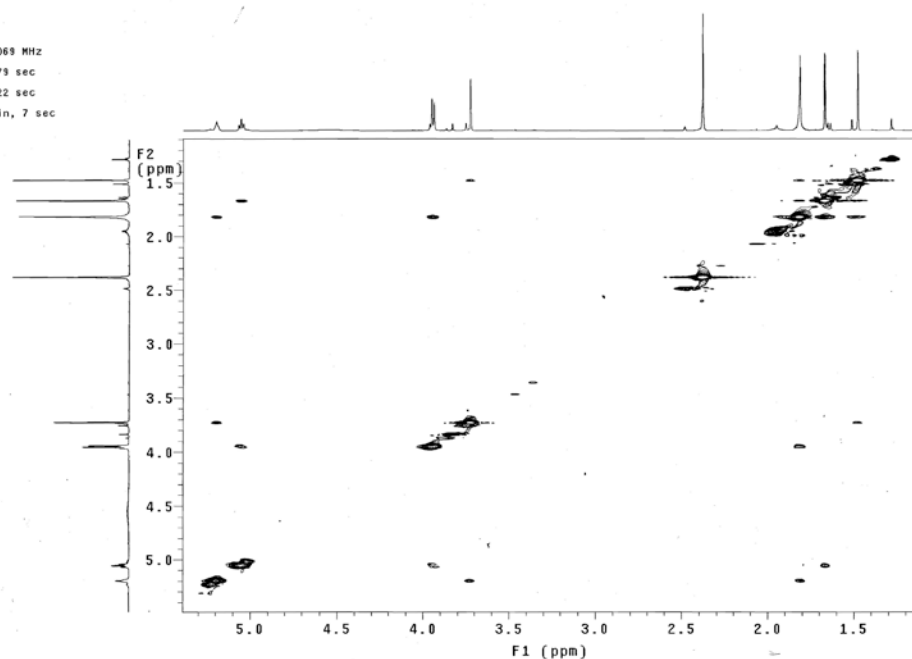
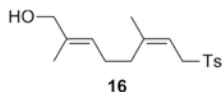
N\_18\_OH

File: xp

Pulse Sequence: RODEY

Solvent: dmsc  
Temp: 22.0 C / 295.1 K  
Operator: dhesek  
VNMR5-600 "nmr600"

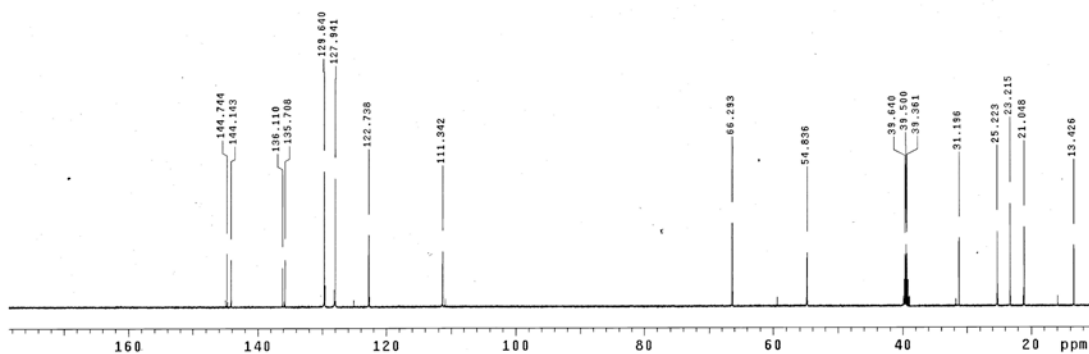
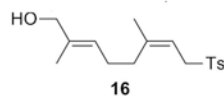
Relax. delay 3.830 sec  
Mixing 0.500 sec  
Acq. time 0.170 sec  
Width 6009.6 Hz  
2D Width 6009.6 Hz  
40 repetitions  
2 x 256 increments  
OBSERVE H1, 599.8757069 MHz  
DATA PROCESSING  
Gauss apodization 0.079 sec  
F1 DATA PROCESSING  
Gauss apodization 0.022 sec  
FT size 2048 x 2048  
Total time 25 hr, 50 min, 7 sec



N\_18\_OH

exp8 Carbon

SAMPLE		SPECIAL	
date	Dec 23 2008	temp	22.0
solvent	dmsc	gain	not used
file		spin	not used
ACQUISITION			
sw	36764.7	pw90	7.400
at	1.783	alfa	10.000
np	131072	FLAGS	
fb	17000	il	n
bs	4	in	n
d1	1.220	op	y
nt	256	hs	nn
ct	60	PROCESSING	
tr	0	lb	0.50
tn	C13	fn	262144
srfq	150.855	DISPLAY	
tof	1542.6	sp	1587.0
tpwr	50	wp	25338.6
pw	7.400	rfl	8571.2
DECOUPLER		rfl	5958.1
dn	H1	rp	-114.0
dof	0	lp	7.8
		PLOT	
dm	yyy	wc	250
dms	w	sc	0
dpwr	44	sc	76
dmsf	15198	th	6
	al	cdc	ph



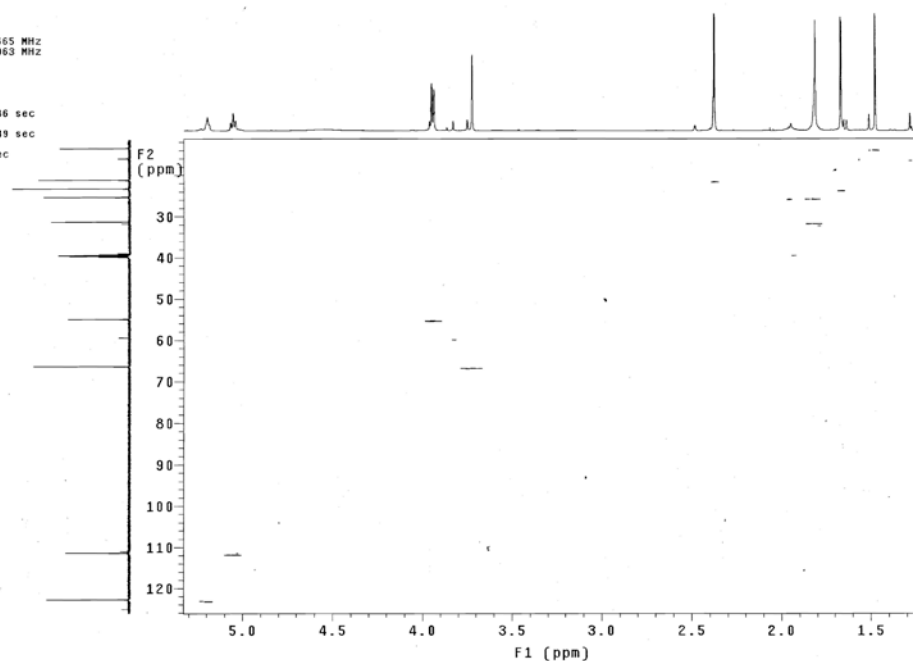
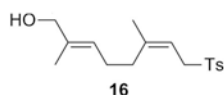
N\_18\_OH

File: xp

Pulse Sequence: HETCOR

Solvent: dmsc  
Temp: 22.0 C / 295.1 K  
Operator: dhsek  
VNMRS-600 "nmr600"

Relax. delay 1.313 sec  
Acq. time 0.187 sec  
Width 25041.7 Hz  
2D Width 4799.0 Hz  
4 repetitions  
2 x 256 increments  
OBSERVE C13, 150.8388665 MHz  
DECOUPLE H1, 599.8784063 MHz  
Power 44 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Gauss apodization 0.086 sec  
F1 DATA PROCESSING  
Gauss apodization 0.049 sec  
FT size 32768 x 2048  
Total time 53 min, 4 sec



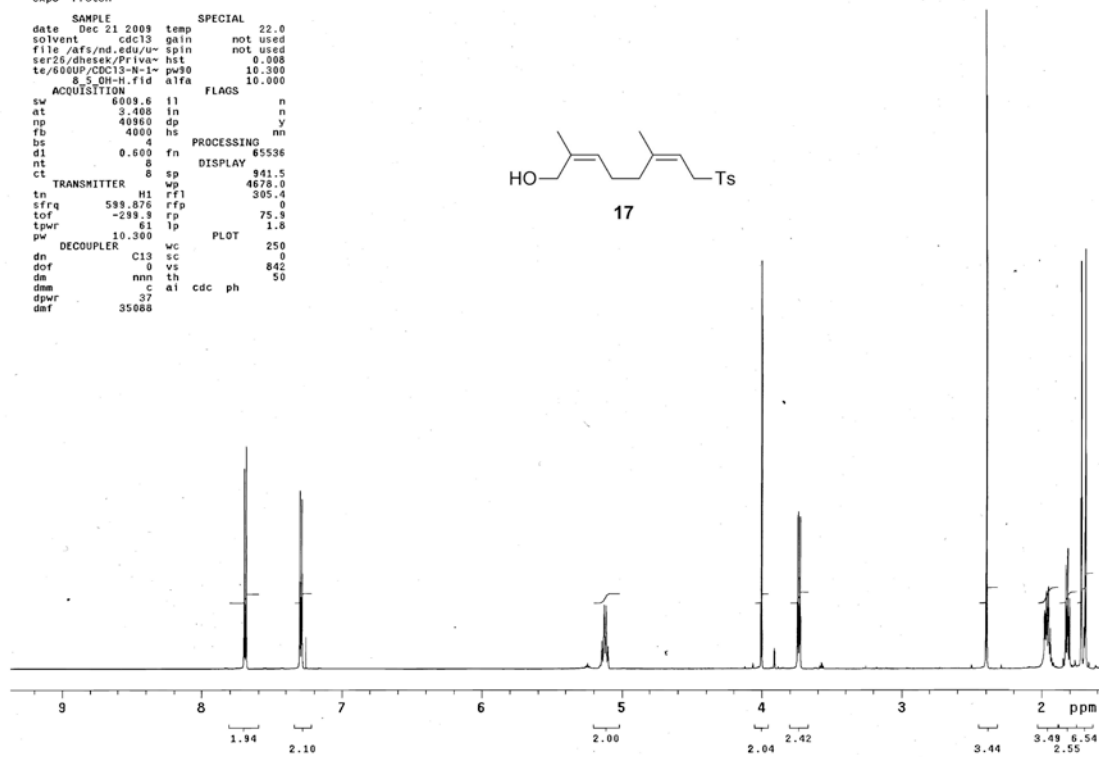
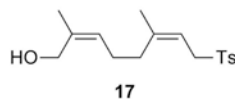
600N-18\_5\_OH

exp8 Proton

SAMPLE		SPECIAL	
date	Dec 21 2009	temp	22.0
solvent	cdcl3	gain	not used
file	/afs/nd.edu/u/~	spin	not used
ser25/dhsek/Priva	hst	0.086	
te/480UP/CDCl3-h-1w	pu90	10.300	
8_5_OH-H.fid	alfa	10.000	

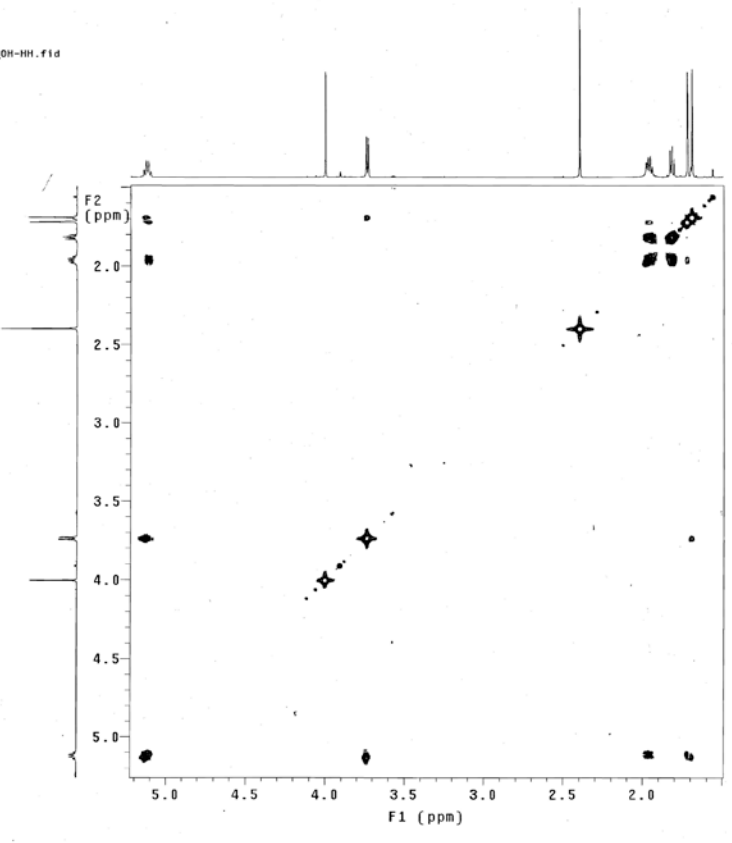
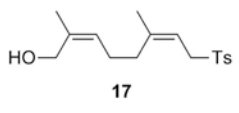
  

ACQUISITION		FLAGS	
sw	6009.6	11	n
at	3.408	in	n
np	40960	dp	y
fb	4000	hs	nm
bs	4	fn	
d1	0.000	fn	
nt	8	DISPLAY	\$5536
ct	8	sp	941.5
TRANSMITTER		wp	4678.0
tn	H1	FF1	305.4
sfrq	599.876	rfrp	0
tof	-289.9	rp	75.9
tpwr	61	lp	1.8
pw	10.300	PLOT	
DECOUPLER		vc	250
dn	C13	sc	0
dof	0	vs	842
dm	nmn	th	50
dmm	c	ai	cdc ph
dpwr	37		
daf	35088		



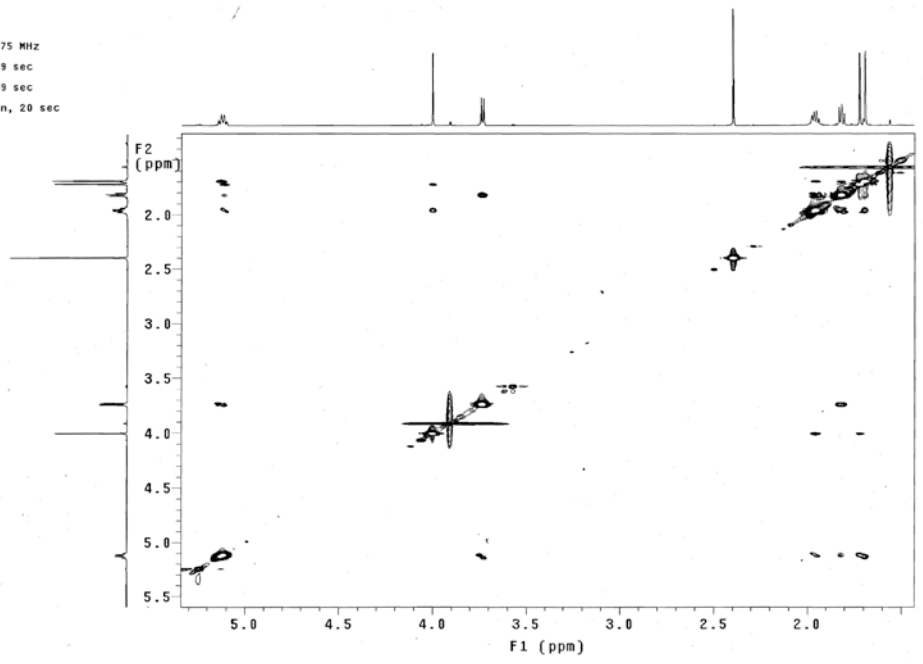
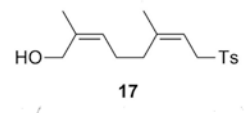
600N-18\_5\_OH  
 File: afs/nd.edu/user26/dhesek/Private/600UP/N-18\_5\_OH-HH.fid  
 Pulse Sequence: CDSY  
 Solvent: cdcl3  
 Temp: 22.0 C / 295.1 K  
 Operator: dhesek  
 File: N-18\_5\_OH-HH  
 VNMR5-600 "nmr600"

Relax. delay 1.000 sec  
 Acq. time 0.242 sec  
 Width 4223.0 Hz  
 2D Width 4223.0 Hz  
 5 repetitions  
 512 increments  
 OBSERVE H1, 599.8728575 MHz  
 DATA PROCESSING  
 Sine bell 0.121 sec  
 F1 DATA PROCESSING  
 Sine bell 0.078 sec  
 FT size 4096 x 4096  
 Total time 1 hr, 30 min, 14 sec



600N-18\_5\_OH  
 File: xp  
 Pulse Sequence: ROESY  
 Solvent: cdcl3  
 Temp: 22.0 C / 295.1 K  
 Operator: dhesek  
 File: N-18\_5\_OH-HH  
 VNMR5-600 "nmr600"

Relax. delay 3.830 sec  
 Mixing 0.500 sec  
 Acq. time 0.170 sec  
 Width 6009.6 Hz  
 2D Width 6009.6 Hz  
 32 repetitions  
 2 x 256 increments  
 OBSERVE H1, 599.8728575 MHz  
 DATA PROCESSING  
 Gauss apodization 0.079 sec  
 F1 DATA PROCESSING  
 Gauss apodization 0.039 sec  
 FT size 2048 x 2048  
 Total time 20 hr, 40 min, 20 sec



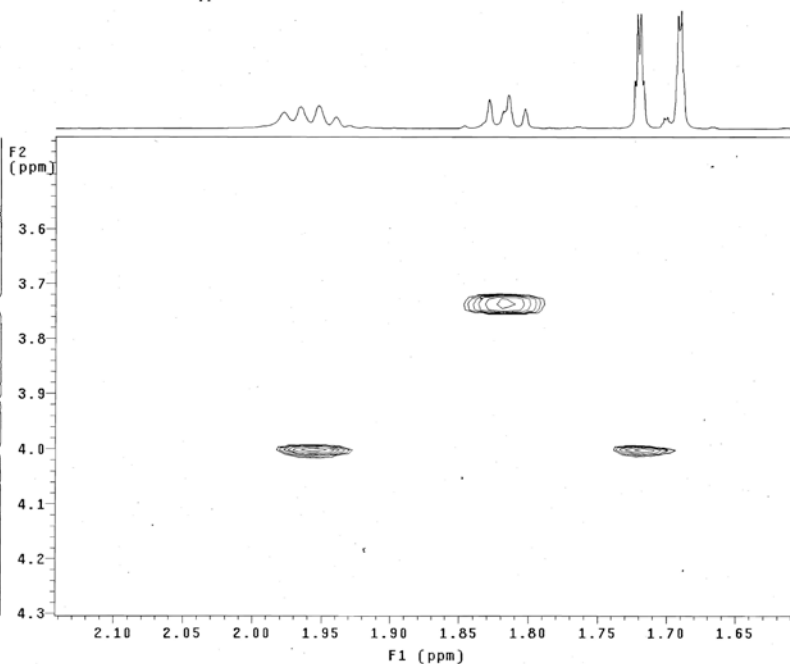
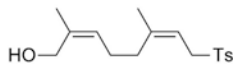
600N-18\_5\_OH

File: xp

Pulse Sequence: ROESY

Solvent: cdc13  
Temp. 22.0 C / 295.1 K  
Operator: dhesek  
VMRS-600 "nmr600"

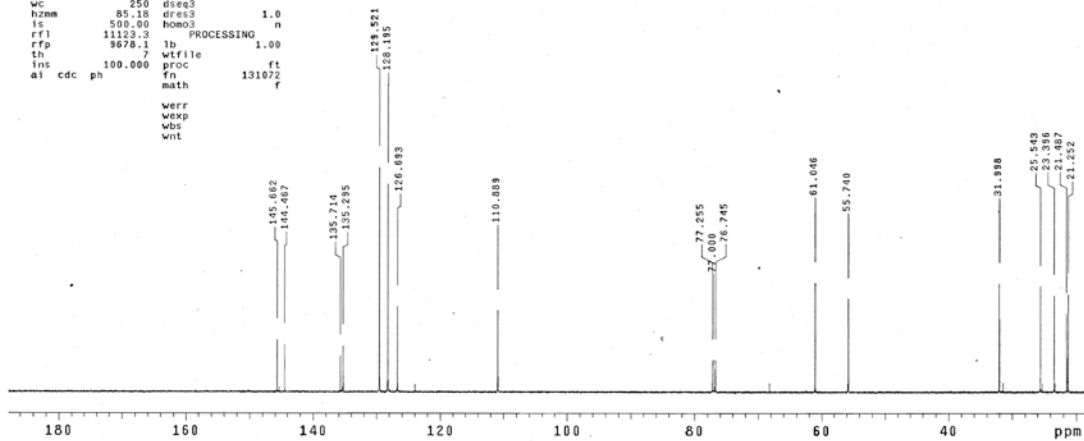
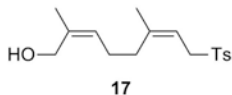
Relax. delay 3.830 sec  
Mixing 0.500 sec  
Acq. time 8.170 sec  
Width 6009.6 Hz  
ZD Width 6009.6 Hz  
32 repetitions  
2 x 256 increments  
OBSERVE H1, 599.8728575 MHz  
DATA PROCESSING  
Gauss apodization 0.079 sec  
F1 DATA PROCESSING  
Gauss apodization 0.039 sec  
F1 size 2048 x 2048  
Total time 20 hr, 40 min, 20 sec



UPN-18\_5\_OH

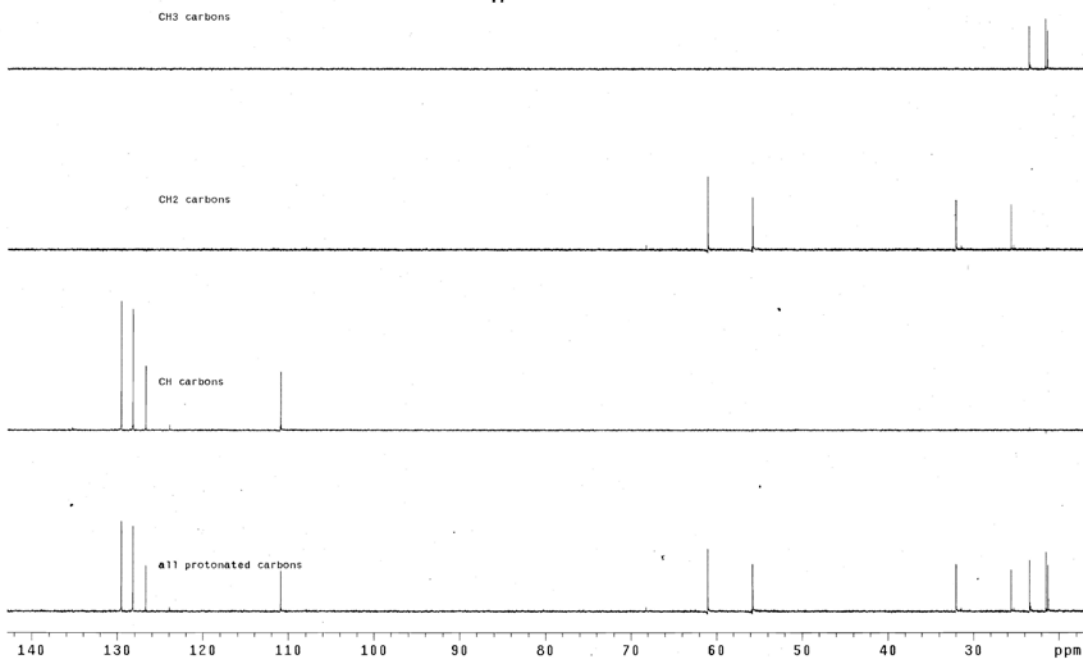
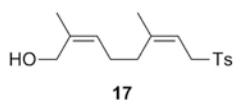
exp2 s2pu1

SAMPLE		DEC. & VT	
date	Dec 13 2009	dfrq	499.864
solvent	CDC13	dn	H1
file		dpur	40
ACQUISITION			
sfrq	125.702	dm	yyy
tn	413	dna	w
at	1.215	daf	8787.35
np	65536	dseq	
sw	26963.3	dres	1.0
fb	15000	homo	n
bs	4		DEC2
tpwr	52	dfrq2	0
pw	10.2	dn2	1
d1	1.800	dpur2	0
tof	144.5	dof2	0
nt	640	dn2	n
ct	75	dna2	c
alock		dn2	10000
gain	not used	dseq2	1.0
flags		dres2	n
il	n	homo2	DEC3
in	n		0
dp	y	dfrq3	0
hs	nn	dn3	1
DISPLAY			
sp	2310.7	dpur3	0
wp	21293.8	dm3	n
vs	45	dna3	c
sc	0	dn3	10000
wc	250	dseq3	
hzm	85.18	dres3	1.0
is	500.00	homo3	n
rf1	11123.3		PROCESSING
rfrp	3678.1	lb	1.00
th		wrf1le	ft
ins	100.000	proc	fn
al	cdc ph	fn	131072
		math	f



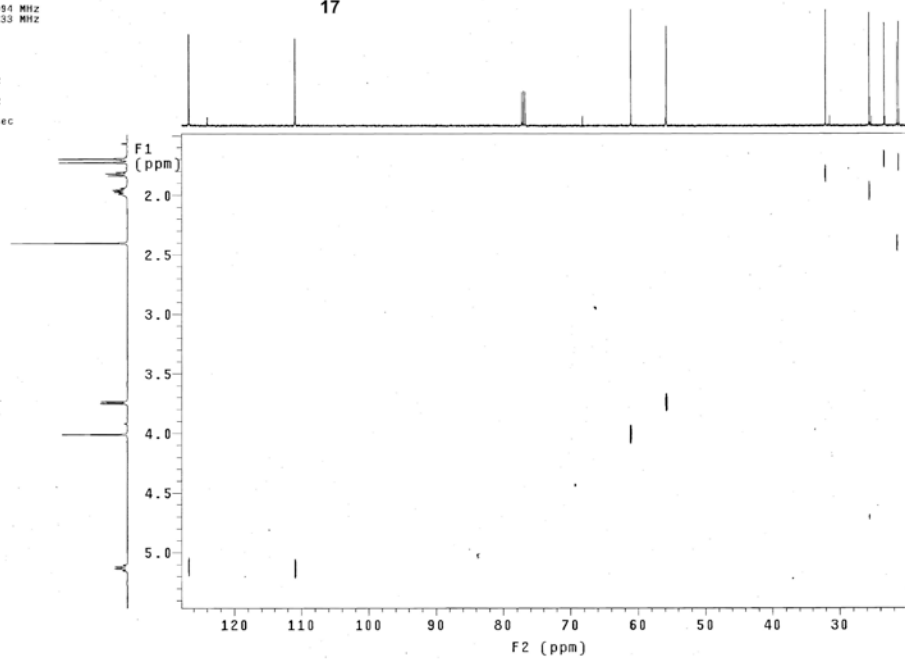
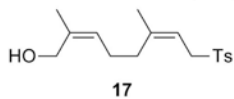


UPN-18\_5\_OH  
Pulse Sequence: dept



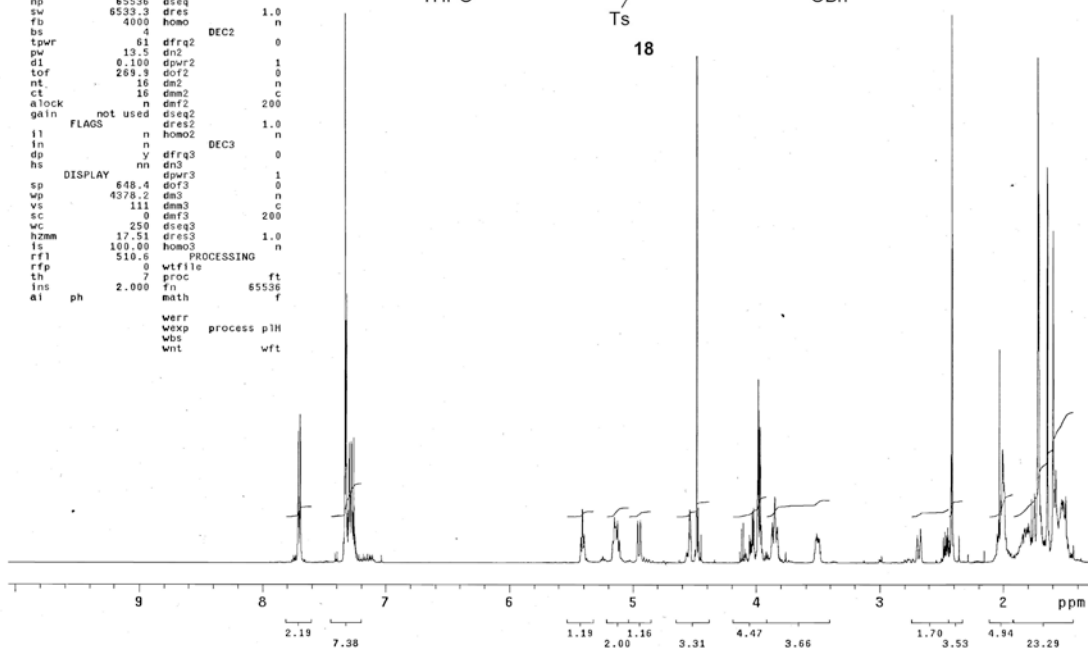
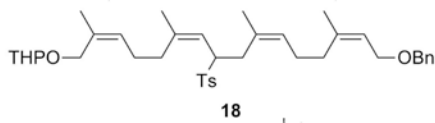
UPN-18\_5\_OH  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: j-18-07  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 3262.2 Hz  
4 repetitions  
256 increments  
OBSERVE ch1: 125.6902094 MHz  
DECUPLE H1: 499.8635233 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 512  
Total time 28 min, 41 sec



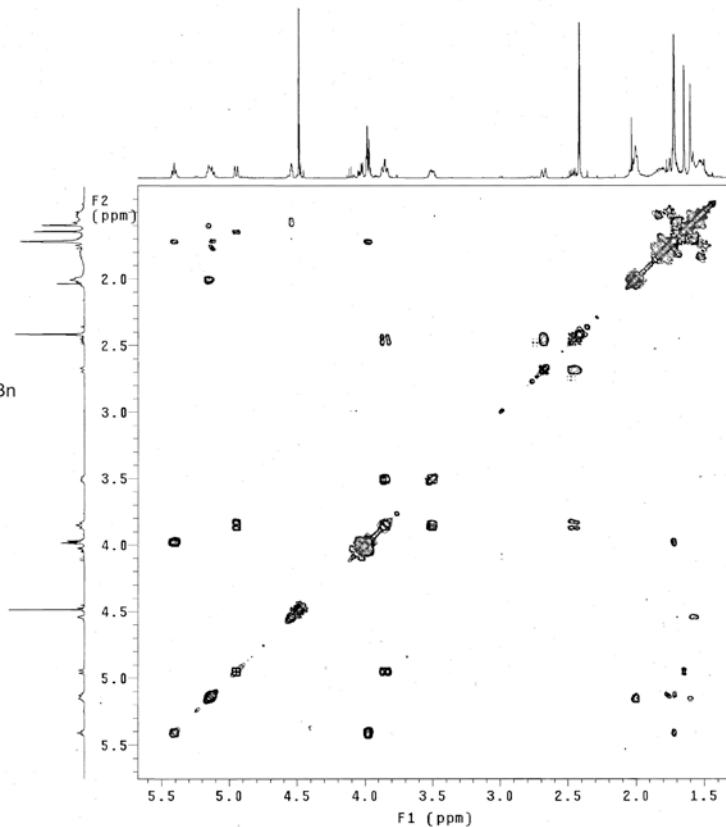
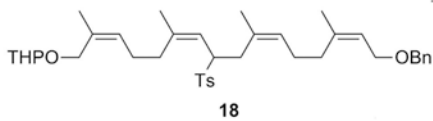
UPN-19  
 exp1 s2pu1

SAMPLE DEC. & VT 499.864  
 date Dec 20 2009 dfrq dn M1  
 solvent CDCl3 dn M1  
 file exp dpr 30  
 ACQUISITION exp dof 0  
 sfrq 499.864 dm nnn c  
 to H1 dm 200  
 at 5.016 dmf 200  
 np 65536 dseq 1.0  
 sw 6535.3 dres c  
 fb 4000 homo n  
 bs 4 DEC2 0  
 tpwr 61 dfrq2 0  
 pv 13.5 dn2 1  
 dl 0.100 dpr2 1  
 tof 269.9 dof2 0  
 nt 16 dm2 n  
 ct 16 dm2 c  
 alock n dmf2 200  
 gain not used dseq2 1.0  
 FLAGS n homo2 DEC3 0  
 il n n 0  
 in n y dfrq3 0  
 dp y dn3 1  
 hs DISPLAY dpr3 0  
 sp 648.4 dof3 0  
 wp 4378.2 dm3 n  
 vs 111 dm3 c  
 sc 0 dmf3 200  
 wc 250 dseq3 1.0  
 hzmm 17.51 dres3 n  
 fs 100.00 homo3 n  
 rf1 510.6 PROCESSING  
 rfp 0 wfile ft  
 th 7 proc 65536  
 ins 2.000 tn math f  
 al ph



UPN-19  
 Pulse Sequence: relayh  
 Solvent: CDCl3  
 Ambient temperature  
 INOVA-500 "nmr2a.chem.nd.edu"

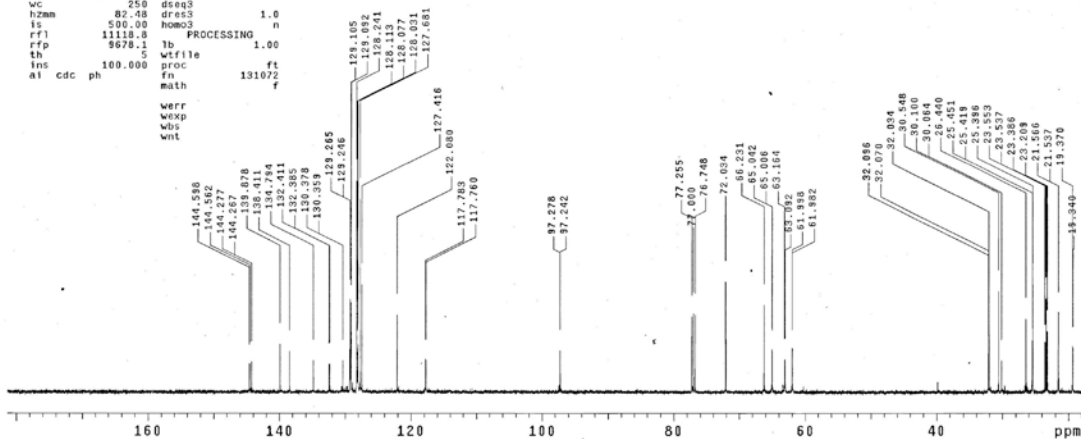
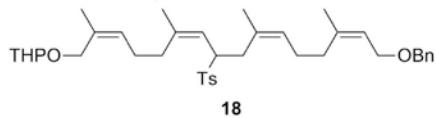
Relax. delay 1.300 sec  
 CDSY 90-90  
 Acq. time 0.148 sec  
 Width 3451.1 Hz  
 2D Width 3451.1 Hz  
 8 repetitions  
 512 increments  
 OBSERVE H1: 499.8611751 MHz  
 DATA PROCESSING  
 Sine bell 0.074 sec  
 F1 DATA PROCESSING  
 Sine bell 0.037 sec  
 FT size 1024 x 1024  
 Total time 1 hr, 04 min, 49 sec



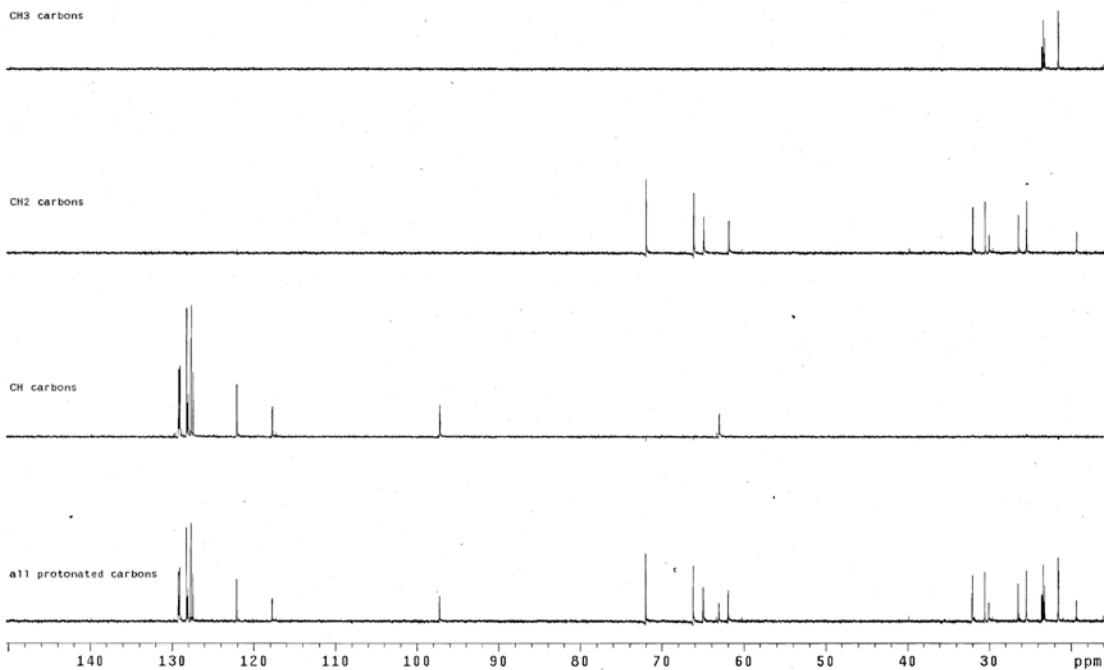
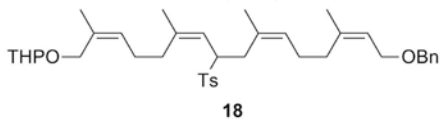
```

UPN-19
exp2 s2pu1
SAMPLE DEC. & VT
date Dec 20 2009 dfrq 459.864
solvent CDC13 dn H1
file exp dpr 48
ACQUISITION dof 0
sfrq 125.702 dm yy
tn C13 dm w
at 1.215 dmf 8787.35
np 65536 dmf 1.0
sv 26953.3 dmf 1.0
fb 15000 homo n
bs 4
tpwr 52 dfrq2 DEC2 0
pv 10.2 dm2 1
d1 1.800 dpr2 1
tof 144.5 dof2 0
nt 840 dm2 n
ct 124 dm2 c
alock not used dmf2 10000
gain n dmf2 1.0
FLAGS n homo2 n
l1 n DEC3 0
ln n
dp y dfrq3 0
hs nn dn3
DISPLAY dpr3 1
sp 2179.3 dof3 0
vp 20620.7 dm3 n
vs 122 dm3 c
sc 0 dmf3 10000
vc 250 dmf3
hzmm 82.48 dmf3 1.0
is 500.00 homo3 n
rf1 111.8 s PROCESSING 1.00
rfp 9678.1 lb
th 5 wfille ft
ins 100.000 proc fn 131072
al cdc ph math f

```

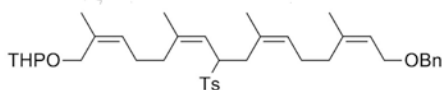


UPN-19  
Pulse Sequence: dept

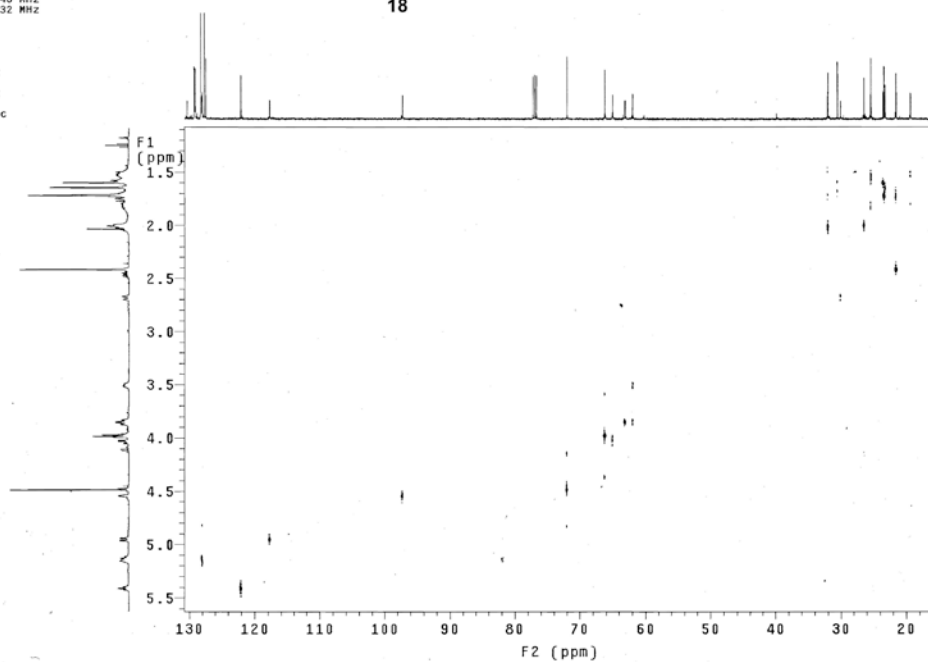


UPN-19  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

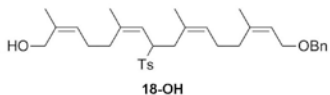
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18400.5 Hz  
2D Width 3682.9 Hz  
8 repetitions  
256 increments  
OBSERVE C13, 125.6902048 MHz  
DECOUPLE H1, 499.8634332 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4095 x 1024  
Total time 57 min, 7 sec



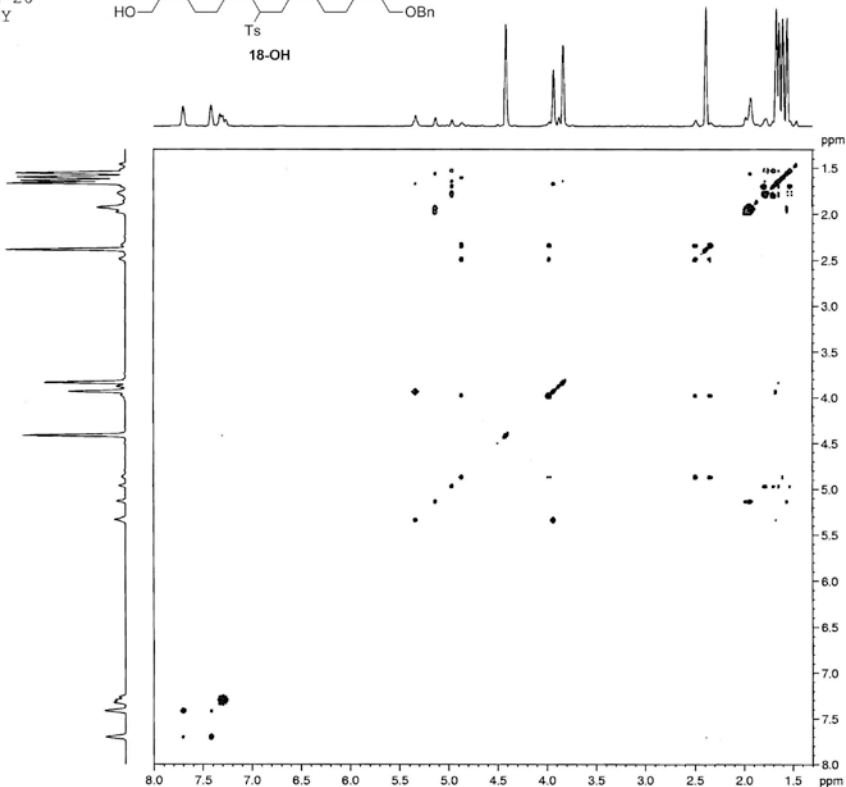
18



800N-20  
TOCSY



18-OH



Current Data Parameters  
NAME 800N-20  
EXPNO 3  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 2001121  
Time 22.14  
INSTRUM spect  
PROBHD 5 mm CPYCI 1H-  
PULPROG dipol2detgpc1  
ID 2048  
SOLVENT DMSO  
NS 16  
DS 16  
SWH 8223.688 Hz  
FIDRES 4.015471 Hz  
AQ 0.1245884 sec  
RG 60.800 usec  
DE 16.00 usec  
TE 301.9 K  
G0 0.0000000 sec  
D1 1.399999999 sec  
d11 0.030000000 sec  
D16 0.000200000 sec  
D20 0.000100000 sec  
D21 0.000100000 sec  
D9 0.050000000 sec  
DELTA 0.001200000 sec  
DELTA1 0.001208000 sec  
FACTOR1 11  
RG1 0.000121600 sec  
I1 22  
ETCINT 0

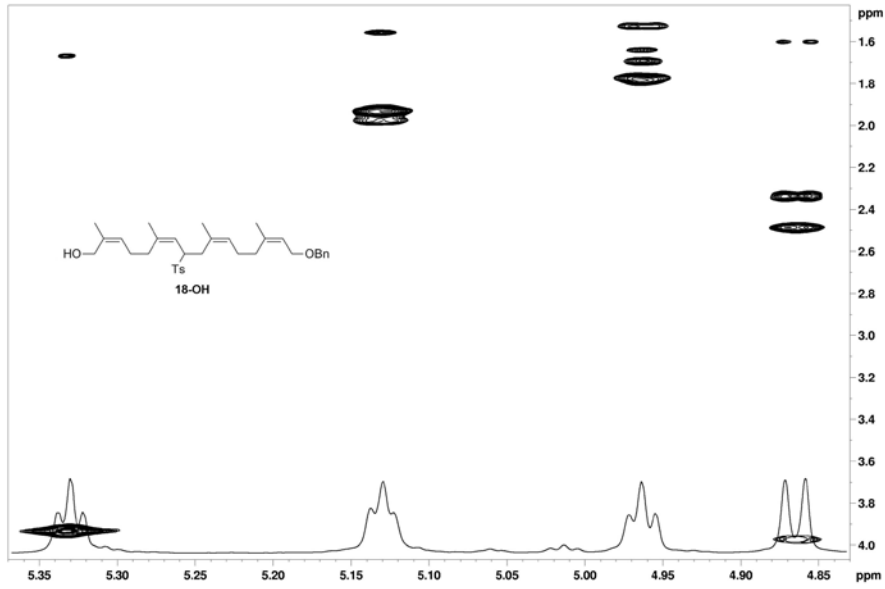
----- CHANNEL f1 -----  
NUC1 13  
P1 9.15 usec  
P2 18.30 usec  
PE 24.00 usec  
PC1 0.20 dB  
PL10 8.58 dB  
GFC1 860.1831640 MHz

----- GRADIENT CHANNEL -----  
GNAM1 SING.100  
GNAM2 SING.100  
GPE1 30.00 %  
GPE2 30.00 %  
P15 1000.00 usec

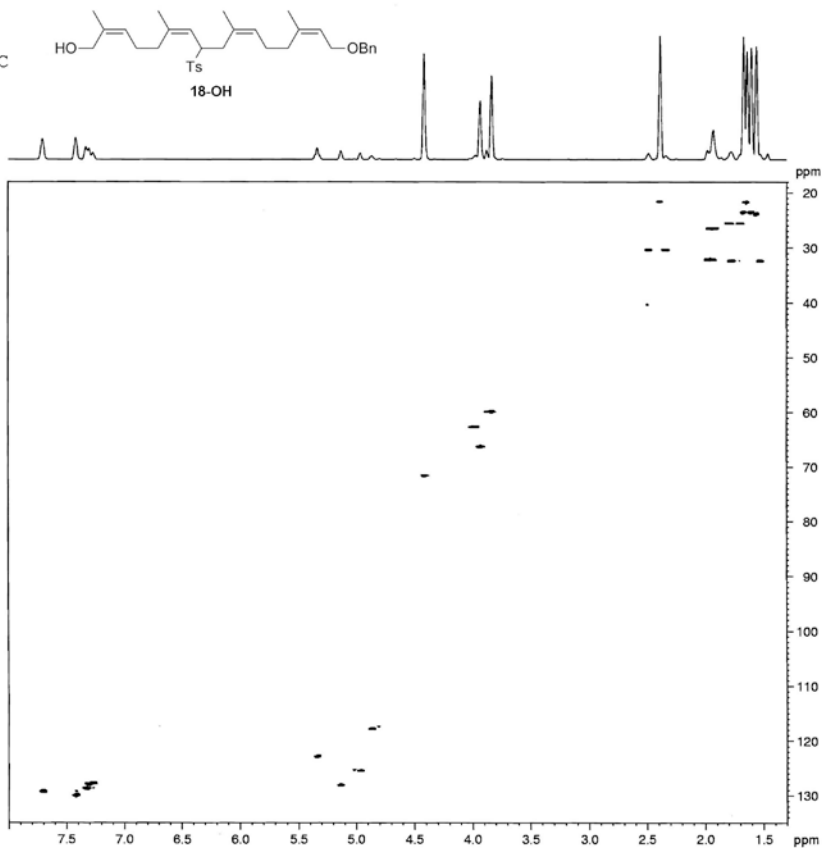
F1 - Acquisition parameters  
ID 1024  
SFO1 800.1838 MHz  
FIDRES 8.030942 Hz  
SW 10.277 ppm  
FOMODE Echo-Antiecho

F2 - Processing parameters  
SI 2048  
SF 800.1800112 MHz  
WDW QP  
SSB 2  
LB 0 Hz  
GB 0  
PC 1.40

F1 - Processing parameters  
SI 2048  
MC2 echo-antiecho  
SF 800.1800101 MHz  
WDW States-TPP1  
SSB 2  
LB 0 Hz  
GB 0



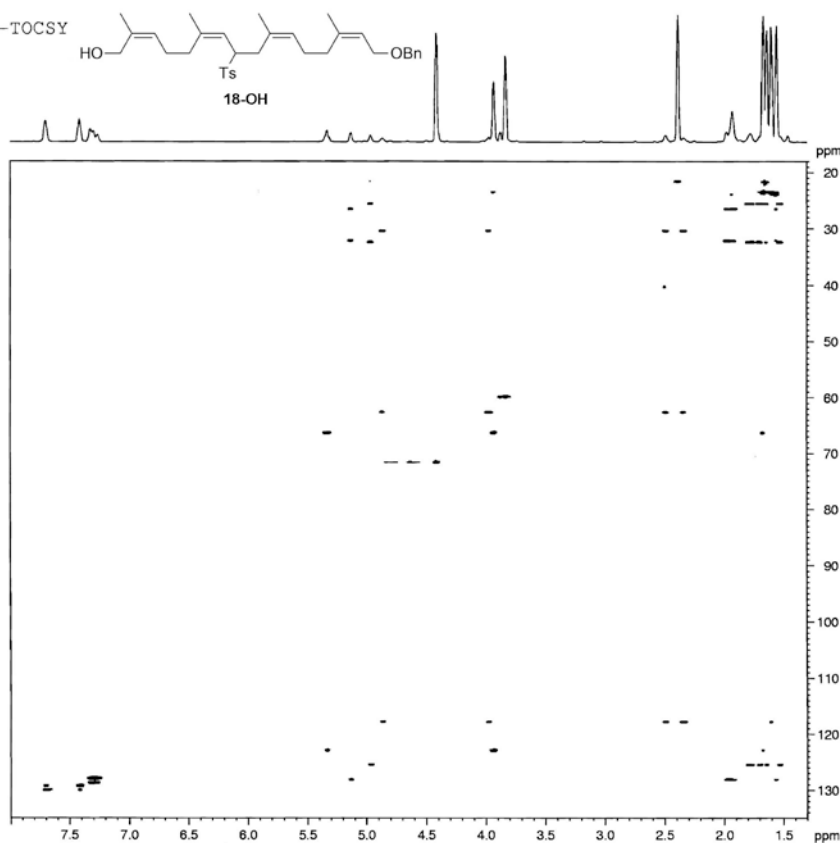
800N-20  
13C-HSQC



```

Current Data Parameters
NAME      800N-20
EXPNO    4
PROCNO   1
F2 - Acquisition Parameters
Date_    20121222
Time     08:47
INSTRUM  spect
PROBHD   5 mm CPIC 1H-
PULPROG  zgpg30
SOLVENT  benzene-d6
NUC1      13C
NUC2      1H
SFO      125
SF      125.760 MHz
AQ      0.462092 sec
RG      655.000
DE      10.000 uM
TE      303.2 K
CQ      0.000000
CZ      119.000000
SI      0.000000 sec
SFO2     500.136 MHz
P1      12.000000 sec
PC      100.000000 uM
P2      12.000000 sec
PC2     100.000000 uM
P3      12.000000 sec
PC3     100.000000 uM
P4      12.000000 sec
PC4     100.000000 uM
P5      12.000000 sec
PC5     100.000000 uM
P6      12.000000 sec
PC6     100.000000 uM
P7      12.000000 sec
PC7     100.000000 uM
P8      12.000000 sec
PC8     100.000000 uM
P9      12.000000 sec
PC9     100.000000 uM
P10     12.000000 sec
PC10    100.000000 uM
P11     12.000000 sec
PC11    100.000000 uM
P12     12.000000 sec
PC12    100.000000 uM
P13     12.000000 sec
PC13    100.000000 uM
P14     12.000000 sec
PC14    100.000000 uM
P15     12.000000 sec
PC15    100.000000 uM
P16     12.000000 sec
PC16    100.000000 uM
P17     12.000000 sec
PC17    100.000000 uM
P18     12.000000 sec
PC18    100.000000 uM
P19     12.000000 sec
PC19    100.000000 uM
F2 - Processing parameters
SI      1
SF      500.136 MHz
WDW     EM
SSB     0
GB      0
PC      1.00
F1 - Processing parameters
SI      1
SF      125.760 MHz
WDW     EM
SSB     0
GB      0
PC      1.00
  
```

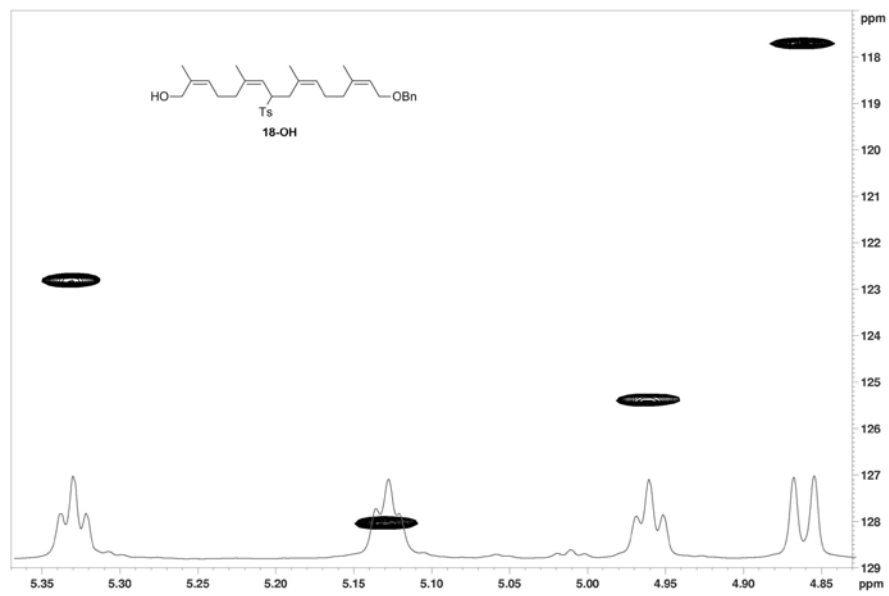
800N-20  
13C-HSQC-TOCSY

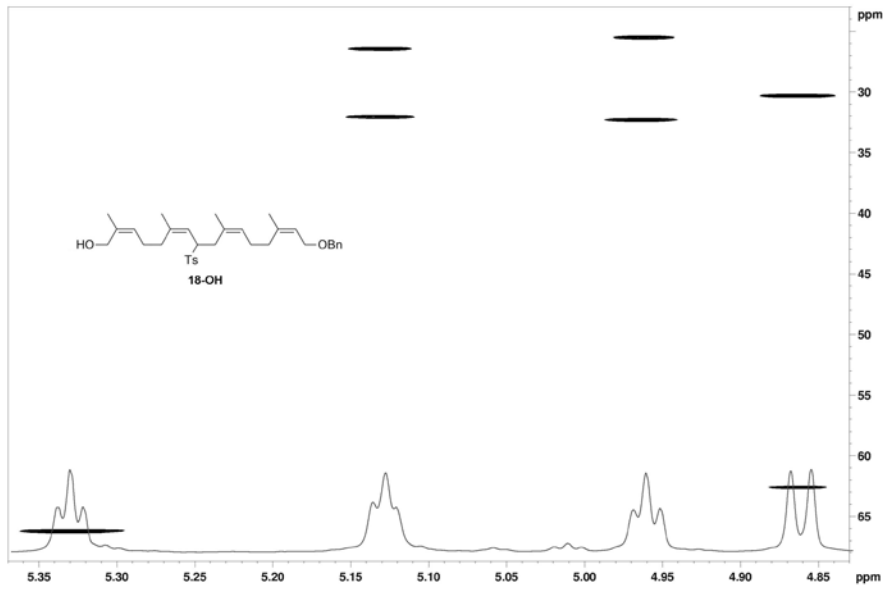


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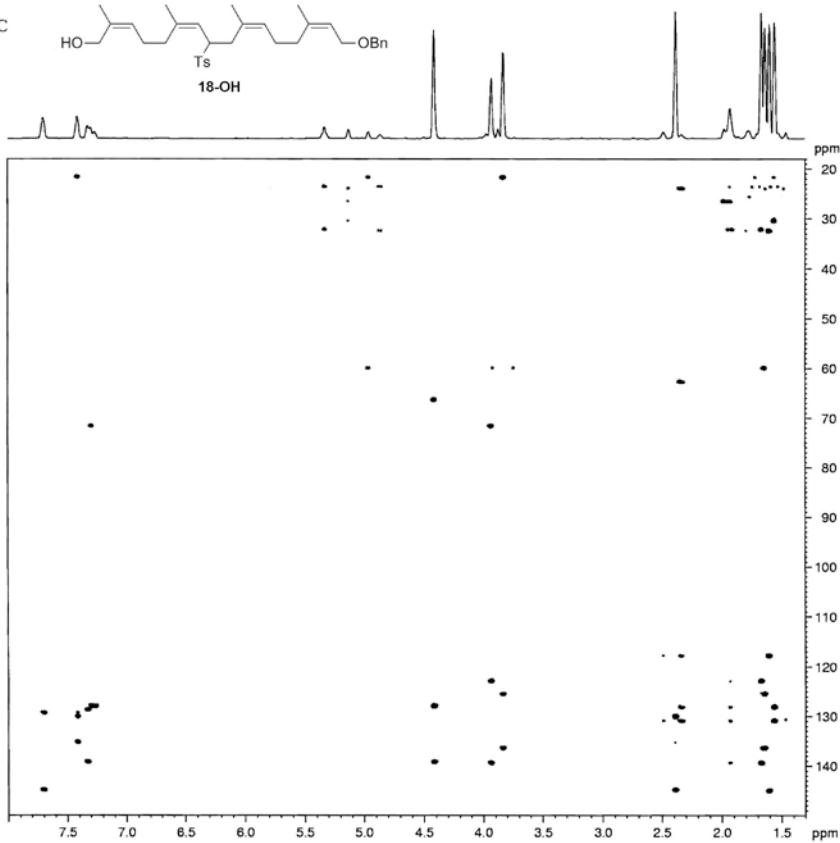
Current Data Parameters
NAME      800N-20
PROCNO    1
PR - Acquisition Parameters
Date_     2001222
Time      11.41
INSTRUM   spect
PROBHD    5 mm CPY131
PULPROG   zgpg30
TD         13104
SOLVENT   DMSO
NS         32
DS         4
SWH        8221.601 Hz
FIDRES     0.18042 Hz
AQ         0.0421012 sec
RG         69.800 umm
DE         20.00 umm
TE         301.2 K
CH3111    -0.000000
CH3112    145.000000
G1         0.0000000 sec
G11        0.0000000 sec
G12        0.0000000 sec
G13        0.0000000 sec
G14        0.0000000 sec
G15        0.0000000 sec
G16        0.0000000 sec
G17        0.0000000 sec
G18        0.0000000 sec
G19        0.0000000 sec
DELTA1     0.0012230 sec
DELTA2     0.0012000 sec
DELTA3     0.0014714 sec
DELTA4     0.0013100 sec
FACTORS    1 1
IN         0.0000111
SI         32
SI1        0
SI2        0
----- CHANNEL f1 -----
NUC1       13C
P1         9.15 umm
PC1        18.30 umm
PR1        1800.00 umm
PL1        24.00 umm
PL12       0.00 dB
PL13       0.00 dB
SFO1       800.1837460 MHz
----- CHANNEL f2 -----
CPDPRG2    p0w0t0g0
NUC2       1H
P2         500.00 umm
PC2        200.00 umm
PR2        15.00 umm
PL2        120.00 dB
PL22       10.00 dB
PL23       10.00 dB
SFO2       201.260316 MHz
SFO12      1.00 MHz
SFO13      1.00 MHz
SFO14      1.00 MHz
SFO15      1.00 MHz
SFO16      1.00 MHz
SFO17      1.00 MHz
SFO18      1.00 MHz
SFO19      1.00 MHz
SFO20      1.00 MHz
SFO21      1.00 MHz
SFO22      1.00 MHz
SFO23      1.00 MHz
SFO24      1.00 MHz
SFO25      1.00 MHz
SFO26      1.00 MHz
SFO27      1.00 MHz
SFO28      1.00 MHz
SFO29      1.00 MHz
SFO30      1.00 MHz
SFO31      1.00 MHz
SFO32      1.00 MHz
SFO33      1.00 MHz
SFO34      1.00 MHz
SFO35      1.00 MHz
SFO36      1.00 MHz
SFO37      1.00 MHz
SFO38      1.00 MHz
SFO39      1.00 MHz
SFO40      1.00 MHz
SFO41      1.00 MHz
SFO42      1.00 MHz
SFO43      1.00 MHz
SFO44      1.00 MHz
SFO45      1.00 MHz
SFO46      1.00 MHz
SFO47      1.00 MHz
SFO48      1.00 MHz
SFO49      1.00 MHz
SFO50      1.00 MHz
SFO51      1.00 MHz
SFO52      1.00 MHz
SFO53      1.00 MHz
SFO54      1.00 MHz
SFO55      1.00 MHz
SFO56      1.00 MHz
SFO57      1.00 MHz
SFO58      1.00 MHz
SFO59      1.00 MHz
SFO60      1.00 MHz
SFO61      1.00 MHz
SFO62      1.00 MHz
SFO63      1.00 MHz
SFO64      1.00 MHz
SFO65      1.00 MHz
SFO66      1.00 MHz
SFO67      1.00 MHz
SFO68      1.00 MHz
SFO69      1.00 MHz
SFO70      1.00 MHz
SFO71      1.00 MHz
SFO72      1.00 MHz
SFO73      1.00 MHz
SFO74      1.00 MHz
SFO75      1.00 MHz
SFO76      1.00 MHz
SFO77      1.00 MHz
SFO78      1.00 MHz
SFO79      1.00 MHz
SFO80      1.00 MHz
SFO81      1.00 MHz
SFO82      1.00 MHz
SFO83      1.00 MHz
SFO84      1.00 MHz
SFO85      1.00 MHz
SFO86      1.00 MHz
SFO87      1.00 MHz
SFO88      1.00 MHz
SFO89      1.00 MHz
SFO90      1.00 MHz
SFO91      1.00 MHz
SFO92      1.00 MHz
SFO93      1.00 MHz
SFO94      1.00 MHz
SFO95      1.00 MHz
SFO96      1.00 MHz
SFO97      1.00 MHz
SFO98      1.00 MHz
SFO99      1.00 MHz
SFO100     1.00 MHz
----- GRADIENT CHANNEL -----
GR1NAME    GR1
GR1P1      100.00
GR1P2      100.00
GR1P3      100.00
GR1P4      100.00
GR1P5      100.00
GR1P6      100.00
GR1P7      100.00
GR1P8      100.00
GR1P9      100.00
GR1P10     100.00
GR1P11     100.00
GR1P12     100.00
GR1P13     100.00
GR1P14     100.00
GR1P15     100.00
GR1P16     100.00
GR1P17     100.00
GR1P18     100.00
GR1P19     100.00
GR1P20     100.00
GR1P21     100.00
GR1P22     100.00
GR1P23     100.00
GR1P24     100.00
GR1P25     100.00
GR1P26     100.00
GR1P27     100.00
GR1P28     100.00
GR1P29     100.00
GR1P30     100.00
GR1P31     100.00
GR1P32     100.00
GR1P33     100.00
GR1P34     100.00
GR1P35     100.00
GR1P36     100.00
GR1P37     100.00
GR1P38     100.00
GR1P39     100.00
GR1P40     100.00
GR1P41     100.00
GR1P42     100.00
GR1P43     100.00
GR1P44     100.00
GR1P45     100.00
GR1P46     100.00
GR1P47     100.00
GR1P48     100.00
GR1P49     100.00
GR1P50     100.00
GR1P51     100.00
GR1P52     100.00
GR1P53     100.00
GR1P54     100.00
GR1P55     100.00
GR1P56     100.00
GR1P57     100.00
GR1P58     100.00
GR1P59     100.00
GR1P60     100.00
GR1P61     100.00
GR1P62     100.00
GR1P63     100.00
GR1P64     100.00
GR1P65     100.00
GR1P66     100.00
GR1P67     100.00
GR1P68     100.00
GR1P69     100.00
GR1P70     100.00
GR1P71     100.00
GR1P72     100.00
GR1P73     100.00
GR1P74     100.00
GR1P75     100.00
GR1P76     100.00
GR1P77     100.00
GR1P78     100.00
GR1P79     100.00
GR1P80     100.00
GR1P81     100.00
GR1P82     100.00
GR1P83     100.00
GR1P84     100.00
GR1P85     100.00
GR1P86     100.00
GR1P87     100.00
GR1P88     100.00
GR1P89     100.00
GR1P90     100.00
GR1P91     100.00
GR1P92     100.00
GR1P93     100.00
GR1P94     100.00
GR1P95     100.00
GR1P96     100.00
GR1P97     100.00
GR1P98     100.00
GR1P99     100.00
GR1P100    100.00
PR - Acquisition parameters
TD         13104
SFO1       201.260316 MHz
FIDRES     0.18042 Hz
AQ         0.0421012 sec
PRGPROG    f0w0t0g0
PR - Processing parameters
SI         32
SFO12      1.00 MHz
SFO13      1.00 MHz
SFO14      1.00 MHz
SFO15      1.00 MHz
SFO16      1.00 MHz
SFO17      1.00 MHz
SFO18      1.00 MHz
SFO19      1.00 MHz
SFO20      1.00 MHz
SFO21      1.00 MHz
SFO22      1.00 MHz
SFO23      1.00 MHz
SFO24      1.00 MHz
SFO25      1.00 MHz
SFO26      1.00 MHz
SFO27      1.00 MHz
SFO28      1.00 MHz
SFO29      1.00 MHz
SFO30      1.00 MHz
SFO31      1.00 MHz
SFO32      1.00 MHz
SFO33      1.00 MHz
SFO34      1.00 MHz
SFO35      1.00 MHz
SFO36      1.00 MHz
SFO37      1.00 MHz
SFO38      1.00 MHz
SFO39      1.00 MHz
SFO40      1.00 MHz
SFO41      1.00 MHz
SFO42      1.00 MHz
SFO43      1.00 MHz
SFO44      1.00 MHz
SFO45      1.00 MHz
SFO46      1.00 MHz
SFO47      1.00 MHz
SFO48      1.00 MHz
SFO49      1.00 MHz
SFO50      1.00 MHz
SFO51      1.00 MHz
SFO52      1.00 MHz
SFO53      1.00 MHz
SFO54      1.00 MHz
SFO55      1.00 MHz
SFO56      1.00 MHz
SFO57      1.00 MHz
SFO58      1.00 MHz
SFO59      1.00 MHz
SFO60      1.00 MHz
SFO61      1.00 MHz
SFO62      1.00 MHz
SFO63      1.00 MHz
SFO64      1.00 MHz
SFO65      1.00 MHz
SFO66      1.00 MHz
SFO67      1.00 MHz
SFO68      1.00 MHz
SFO69      1.00 MHz
SFO70      1.00 MHz
SFO71      1.00 MHz
SFO72      1.00 MHz
SFO73      1.00 MHz
SFO74      1.00 MHz
SFO75      1.00 MHz
SFO76      1.00 MHz
SFO77      1.00 MHz
SFO78      1.00 MHz
SFO79      1.00 MHz
SFO80      1.00 MHz
SFO81      1.00 MHz
SFO82      1.00 MHz
SFO83      1.00 MHz
SFO84      1.00 MHz
SFO85      1.00 MHz
SFO86      1.00 MHz
SFO87      1.00 MHz
SFO88      1.00 MHz
SFO89      1.00 MHz
SFO90      1.00 MHz
SFO91      1.00 MHz
SFO92      1.00 MHz
SFO93      1.00 MHz
SFO94      1.00 MHz
SFO95      1.00 MHz
SFO96      1.00 MHz
SFO97      1.00 MHz
SFO98      1.00 MHz
SFO99      1.00 MHz
SFO100     1.00 MHz

```





800N-20  
13C-HMBC



```

Current Data Parameters
NAME      800N-20
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20091223
Time     11:53
SYSTEM   spect
PROBHD   5 mm CPTCI 1H-
PULPROG  hmcgpp12zmgf
TD        2048
SOLVENT  DMSO
NS        64
DS        4
SFR      8223.850 Hz
F2FRES   4.015471 Hz
AQ        0.124584 sec
RG        2500
DM        60.800 usec
DE        30.00 usec
TE        301.9 K
CST13    8.0000000
CST16    123.0000000
CST17    173.0000000
d0        0.0000000 sec
d1        1.3999998 sec
d16       0.0002000 sec
d8        0.0625000 sec
DELTA1   0.0238000 sec
DELTA2   0.00165714 sec
DELTA3   0.0612800 sec
ISO       0.00001310 sec

===== CHANNEL F1 =====
NUC1      13C
P1        9.15 usec
P2        28.20 usec
PL1       0.20 dB
SFO1     800.1837640 MHz

===== CHANNEL F2 =====
NUC2      13C
P3        14.80 usec
PL2       -2.00 dB
SFO2     201.2245355 MHz

===== GRADIENT CHANNEL =====
GRAN1    SENSE:100
GRAN2    SENSE:100
GRAN3    SENSE:100
GRAN4    SENSE:100
GRAN5    SENSE:100
GRAN6    SENSE:100
GFE1     50.00 %
GFE2     30.00 %
GFE3     40.10 %
GFE4     15.00 %
GFE5     -10.00 %
GFE6     -5.00 %
P16      1000.00 usec

F1 - Acquisition parameters
TD        1024
SFO1     201.2245 MHz
F1FRES    37.29376 Hz
TM        189.478 ppm
F1NDCOE   QF

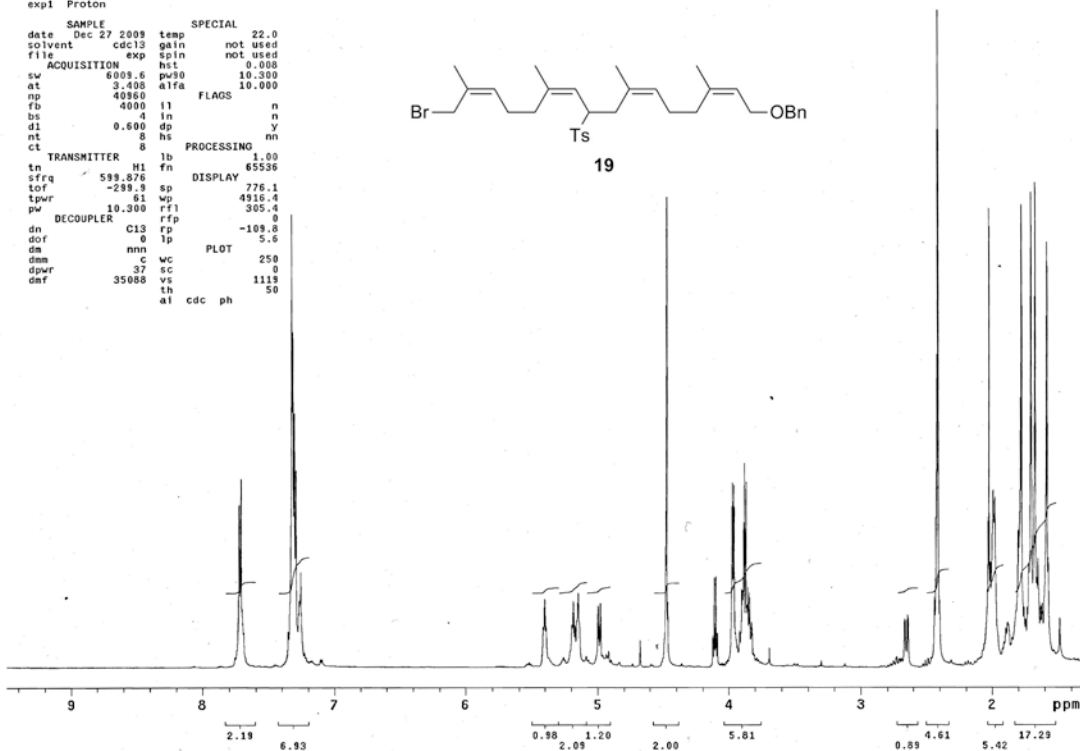
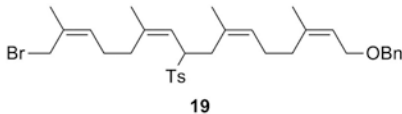
F2 - Processing parameters
SI        2048
SF        800.1800135 MHz
WDM       QFIND
SFB       0
IS        0 Hz
GB        0
PC        1.40

F1 - Processing parameters
SI        QF
SF        201.20541 MHz
WDM       States-TFPI
SFB       0
IS        0 Hz
GB        0
  
```

UPN-22\_600

exp1 Proton

```
SAMPLE SPECIAL 22.0
date Dec 27 2009 temp gain not used
solvent cdc13 exp spin not used
file 6009.6 hst 0.000
ACQUISITION pw30 10.300
sw 3.400 alfa 10.000
np 40960 FLAGS
fb 4000 l1 n
bs 4 in n
d1 0.600 dp y
nt 8 hs nn
ct 8 PROCESSING
TRANSMITTER lb 1.00
tn H1 fn 65536
sfreq 599.876 DISPLAY
tof -289.9 sp 776.1
tpwr 81 wp 4916.4
pk 10.300 rfl 305.4
DECOUPLER C13 rfp 0
dn 0 lp -109.8
dor 0 lp 5.6
dm nnn WC PLOT 250
dsm c sc 0
dprf 37 vs 1113
dmf 35088 th 50
al cdc ph
```



600N-18\_5\_OH

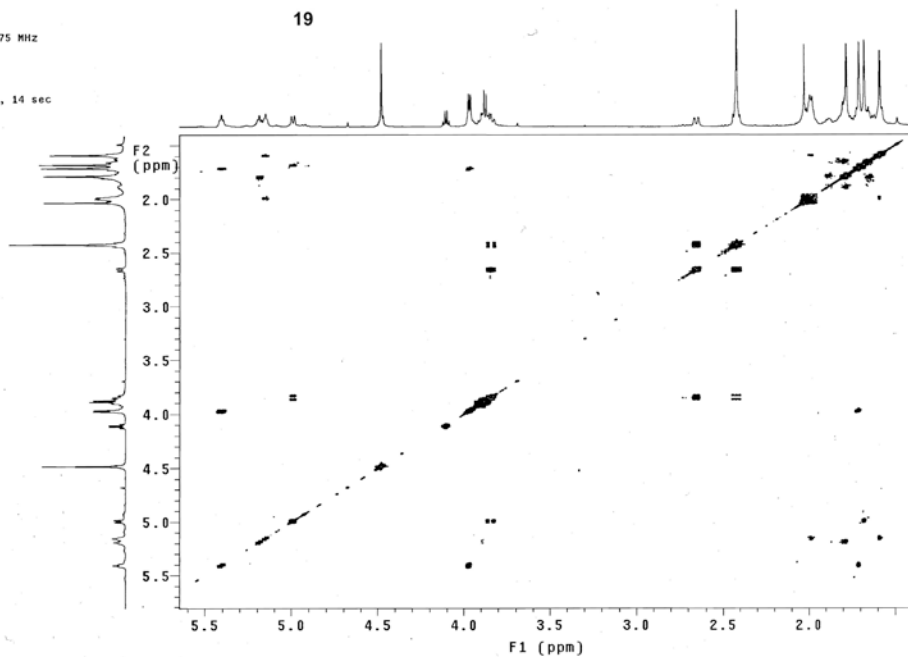
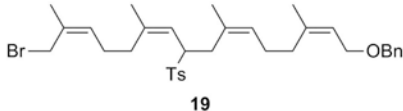
File: xp

Pulse Sequence: COSY

Solvent: cdc13  
Temp: 22.0 C / 295.1 K  
Operator: dhesek  
VNMRS-600 "nmr600"

Relax. delay 1.000 sec  
Acq. time 0.242 sec  
Width 4223.0 Hz  
2D Width 4223.0 Hz  
8 repetitions  
512 increments

OBSERVE H1, 599.8728575 MHz  
DATA PROCESSING  
Sine bell 0.121 sec  
F1 DATA PROCESSING  
Sine bell 0.242 sec  
FT size 8192 x 8192  
Total time 1 hr, 30 min, 14 sec





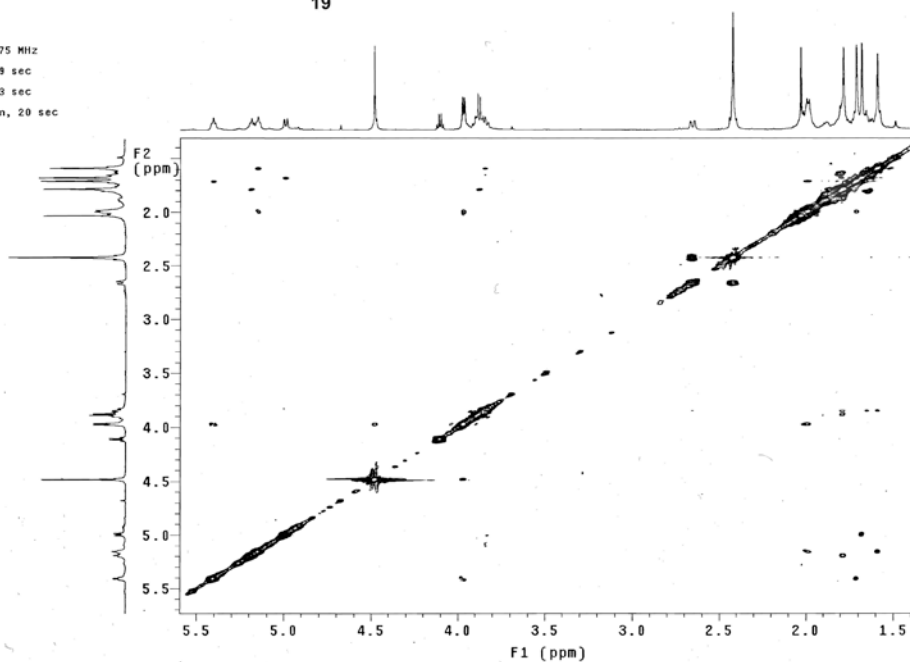
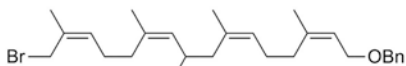
600N-18\_5\_OH

File: xp

Pulse Sequence: ROESY

Solvent: cdc13  
Temp: 22.0 C / 295.1 K  
Operator: dhsek  
VNMR5-600 "mar600"

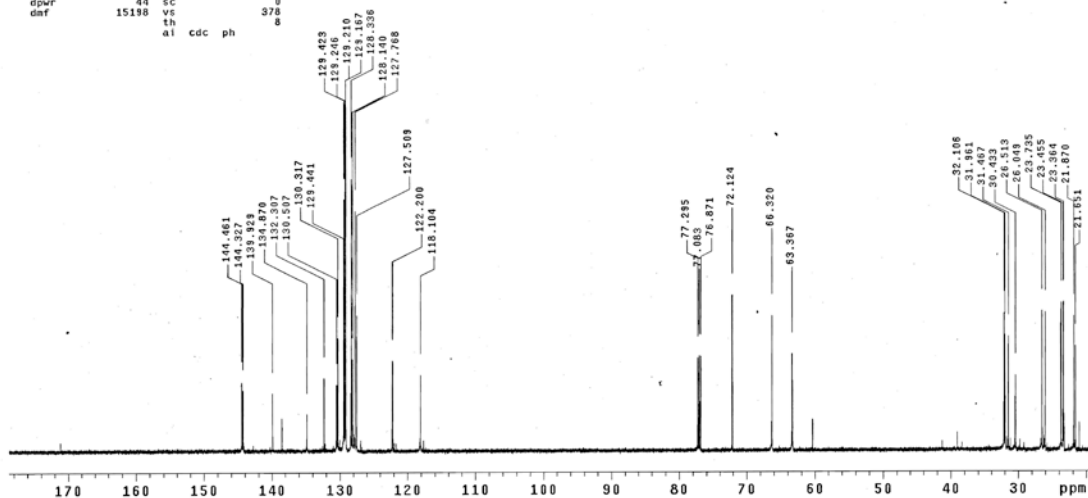
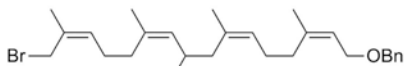
Relax. delay 3.830 sec  
Mixing 0.500 sec  
Acq. time 0.170 sec  
Width 6009.6 Hz  
2D Width 6009.6 Hz  
32 Repetitions  
2 x 256 Increments  
OBSERVE H1, 599.8728575 MHz  
DATA PROCESSING  
Gauss apodization 0.079 sec  
F1 DATA PROCESSING  
Gauss apodization 0.063 sec  
FT size 4096 x 4096  
Total time 20 hr, 40 min, 20 sec



UPN-22\_600

exp2 Carbon

SAMPLE		SPECIAL	
date	Dec 27 2009	temp	22.0
solvent	cdc13	gain	not used
file	exp	spin	not used
ACQUISITION		SPECIAL	
sw	30487.8	hst	0.008
at	1.783	pw90	7.400
np	108634	alpha	10.000
fb	17000	FLAGS	
bs	4	n	n
d1	1.220	in	n
nt	256	dp	y
Ct	256	hs	m
TRANSMITTER		PROCESSING	
tn	C13	fb	0.50
sfrq	150.852	fn	262144
tof	-720.0	sp	2309.8
tpwr	50	wp	24047.5
pw	7.400	rfl	1668.5
DECOUPLER		PLOT	
dn	H1	fp	63.9
dof	0	lp	-2.2
dm	yvy	vc	250
dpr	44	sc	0
dpr	15190	vc	378
dpr		th	8
dpr		al	cdc ph



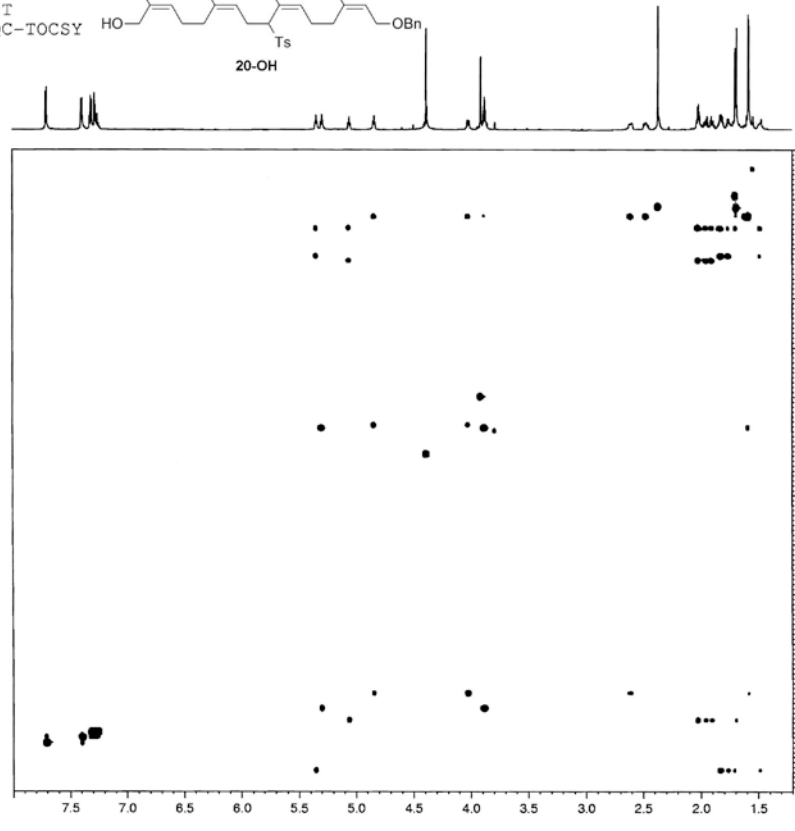
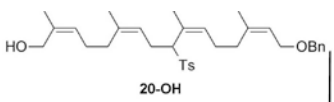








800N-20T  
13C-HSQC-TOCSY

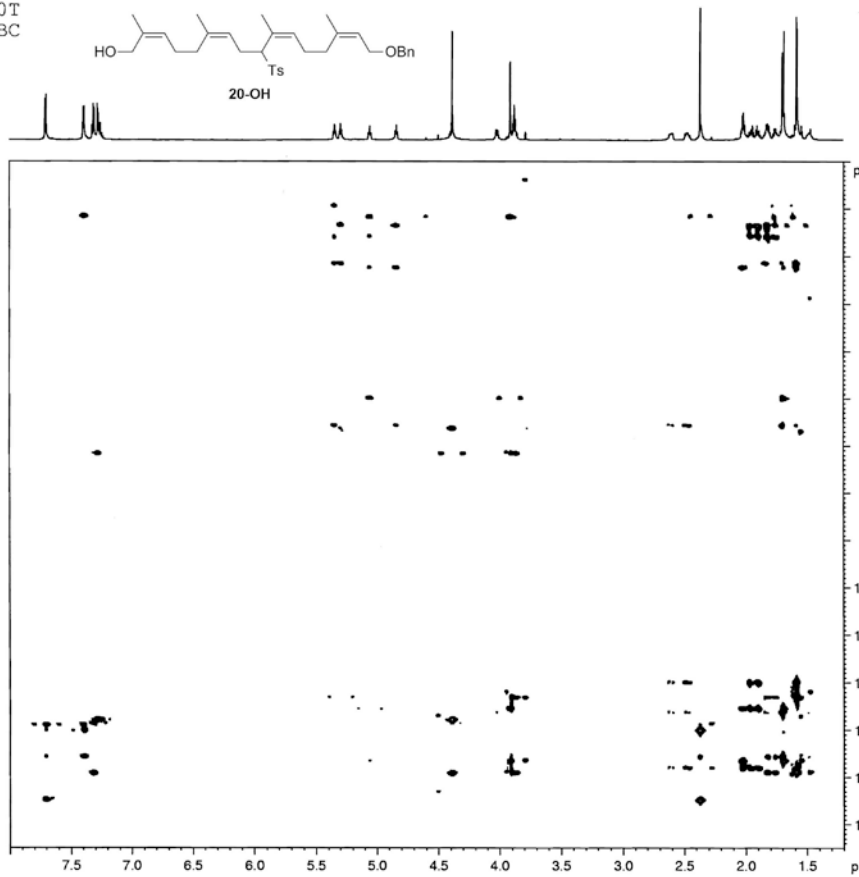
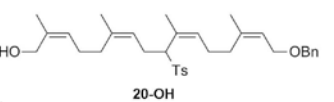


```

Current Data Parameters
NAME      DMSO-d6-20T
EXPNO     1
PROCNO    1

F2 - Acquisition Parameters
Date_     20121214
Time      11:49
INSTRUM   spect
PROBHD    5 mm CPIC1 1H-
PULPROG   zgpg30
ID        1224
SOLVENT   DMSO
NS        64
DS        4
FIDRES    0.023162 Hz
AQ        0.0423012 sec
RG        2050
DE        60.800 usec
TE        300.2 K
CDELT1    0.0000100 sec
CDELT2    1.0000000 sec
CDELT3    0.0000000 sec
CDELT4    0.0000000 sec
CDELT5    0.0000000 sec
DELTA1    0.0020000 sec
DELTA2    0.0012000 sec
DELTA3    0.0012000 sec
DELTA4    0.0012000 sec
DELTA5    0.0012000 sec
IMD       0.0001210 sec
SFO1      800.137640 MHz
===== CHANNEL f1 =====
NUC1       13C
P1         9.30 usec
PL1        0.00 dB
PC1        1.00
===== CHANNEL f2 =====
NUC2       1H
P2         18.60 usec
PL2        0.00 dB
PC2        1.00
===== GRADIENT CHANNEL =====
GPRAM1     SINE.100
GPRAM2     SINE.100
GPRAM3     SINE.100
GPRAM4     SINE.100
GPRAM5     SINE.100
GPRAM6     SINE.100
OP1        50.00 %
OP2        40.10 %
OP3        15.00 %
OP4        -10.00 %
OP5        -3.00 %
OP6        1000.00 usec
F1 - Acquisition parameters
ID         1224
NAME      20121214
EXPNO     1
PROCNO    1
PROBHD    5 mm CPIC1 1H-
PULPROG   zgpg30
ID        1224
SOLVENT   DMSO
NS        64
DS        4
FIDRES    0.023162 Hz
AQ        0.0423012 sec
RG        2050
DE        60.800 usec
TE        300.2 K
CDELT1    0.0000100 sec
CDELT2    1.0000000 sec
CDELT3    0.0000000 sec
CDELT4    0.0000000 sec
CDELT5    0.0000000 sec
DELTA1    0.0020000 sec
DELTA2    0.0012000 sec
DELTA3    0.0012000 sec
DELTA4    0.0012000 sec
DELTA5    0.0012000 sec
IMD       0.0001210 sec
SFO1      800.137640 MHz
===== CHANNEL f1 =====
NUC1       13C
P1         9.30 usec
PL1        0.00 dB
PC1        1.00
===== CHANNEL f2 =====
NUC2       1H
P2         18.60 usec
PL2        0.00 dB
PC2        1.00
===== GRADIENT CHANNEL =====
GPRAM1     SINE.100
GPRAM2     SINE.100
GPRAM3     SINE.100
GPRAM4     SINE.100
GPRAM5     SINE.100
GPRAM6     SINE.100
OP1        50.00 %
OP2        40.10 %
OP3        15.00 %
OP4        -10.00 %
OP5        -3.00 %
OP6        1000.00 usec
F1 - Processing parameters
SI         2048
SF         800.1301156 MHz
WDW        EM
SSB        0
LB         0 Hz
GB         0
PC         1.40
F1 - Processing parameters
SI         2048
SF         201.2245 MHz
WDW        EM
SSB        0
LB         0 Hz
GB         0
  
```

800N-20T  
13C-HMBC



```

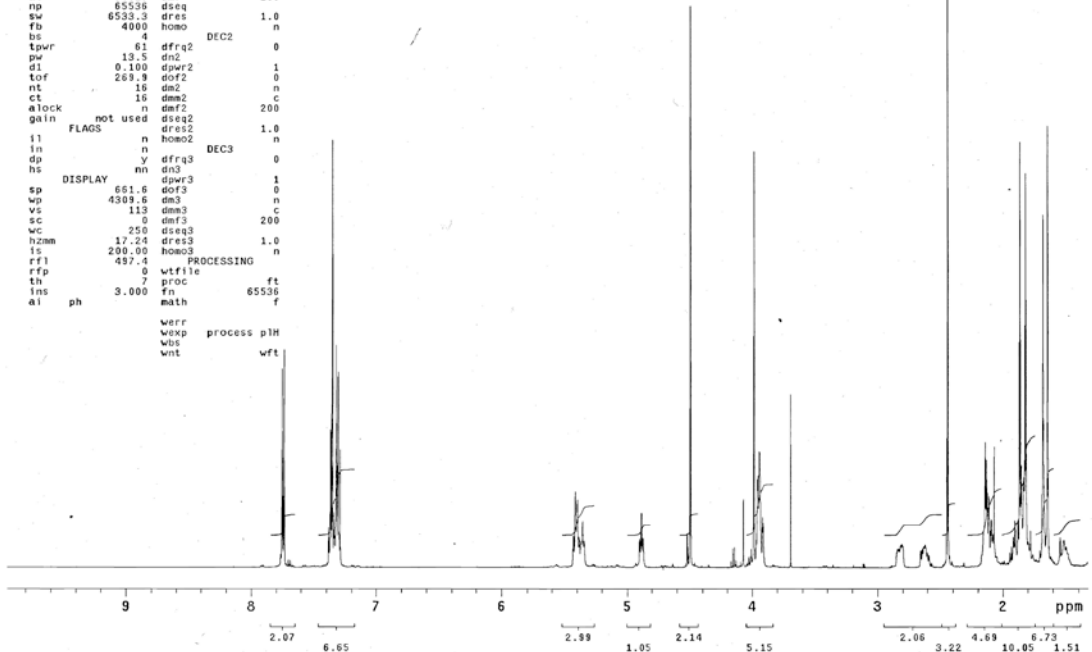
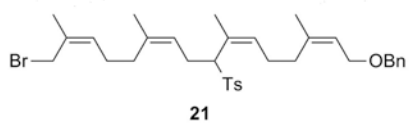
Current Data Parameters
NAME      DMSO-d6-20T
EXPNO     7
PROCNO    1

F2 - Acquisition Parameters
Date_     20121214
Time      11:49
INSTRUM   spect
PROBHD    5 mm CPIC1 1H-
PULPROG   zgpg30
ID        1224
SOLVENT   DMSO
NS        64
DS        4
FIDRES    0.023162 Hz
AQ        0.0423012 sec
RG        2050
DE        60.800 usec
TE        300.2 K
CDELT1    0.0000100 sec
CDELT2    1.0000000 sec
CDELT3    0.0000000 sec
CDELT4    0.0000000 sec
CDELT5    0.0000000 sec
DELTA1    0.0020000 sec
DELTA2    0.0012000 sec
DELTA3    0.0012000 sec
DELTA4    0.0012000 sec
DELTA5    0.0012000 sec
IMD       0.0001210 sec
SFO1      800.137640 MHz
===== CHANNEL f1 =====
NUC1       13C
P1         9.30 usec
PL1        0.00 dB
PC1        1.00
===== CHANNEL f2 =====
NUC2       1H
P2         18.60 usec
PL2        0.00 dB
PC2        1.00
===== GRADIENT CHANNEL =====
GPRAM1     SINE.100
GPRAM2     SINE.100
GPRAM3     SINE.100
GPRAM4     SINE.100
GPRAM5     SINE.100
GPRAM6     SINE.100
OP1        50.00 %
OP2        40.10 %
OP3        15.00 %
OP4        -10.00 %
OP5        -3.00 %
OP6        1000.00 usec
F1 - Acquisition parameters
ID         1224
NAME      20121214
EXPNO     7
PROCNO    1
PROBHD    5 mm CPIC1 1H-
PULPROG   zgpg30
ID        1224
SOLVENT   DMSO
NS        64
DS        4
FIDRES    0.023162 Hz
AQ        0.0423012 sec
RG        2050
DE        60.800 usec
TE        300.2 K
CDELT1    0.0000100 sec
CDELT2    1.0000000 sec
CDELT3    0.0000000 sec
CDELT4    0.0000000 sec
CDELT5    0.0000000 sec
DELTA1    0.0020000 sec
DELTA2    0.0012000 sec
DELTA3    0.0012000 sec
DELTA4    0.0012000 sec
DELTA5    0.0012000 sec
IMD       0.0001210 sec
SFO1      800.137640 MHz
===== CHANNEL f1 =====
NUC1       13C
P1         9.30 usec
PL1        0.00 dB
PC1        1.00
===== CHANNEL f2 =====
NUC2       1H
P2         18.60 usec
PL2        0.00 dB
PC2        1.00
===== GRADIENT CHANNEL =====
GPRAM1     SINE.100
GPRAM2     SINE.100
GPRAM3     SINE.100
GPRAM4     SINE.100
GPRAM5     SINE.100
GPRAM6     SINE.100
OP1        50.00 %
OP2        40.10 %
OP3        15.00 %
OP4        -10.00 %
OP5        -3.00 %
OP6        1000.00 usec
F1 - Processing parameters
SI         2048
SF         800.1301156 MHz
WDW        EM
SSB        0
LB         0 Hz
GB         0
PC         1.40
F1 - Processing parameters
SI         2048
SF         201.2245 MHz
WDW        EM
SSB        0
LB         0 Hz
GB         0
  
```

UPN-22T

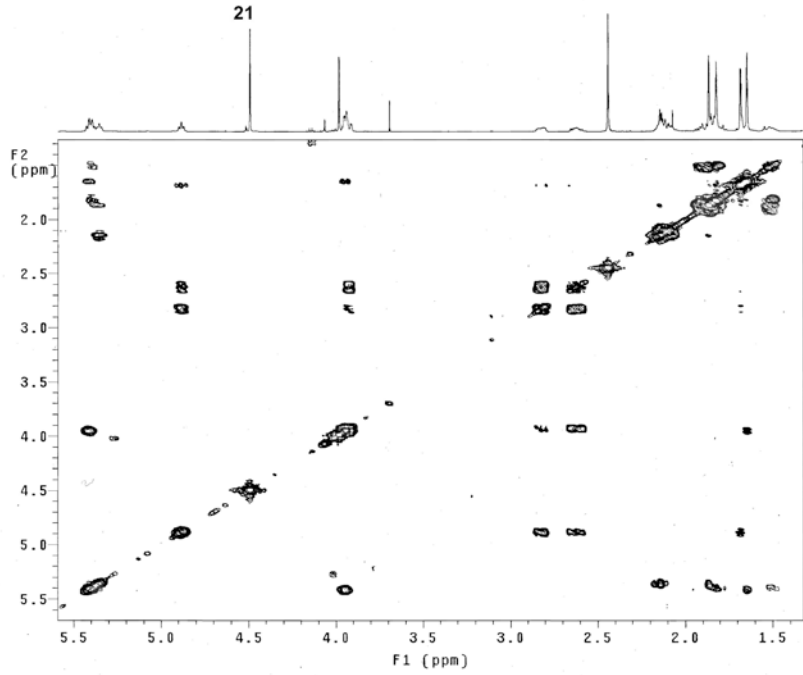
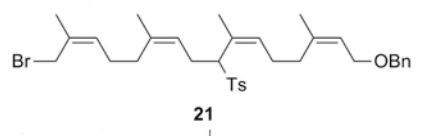
exp3 s2pu1

SAMPLE	DEC. & VT	499.864
date	Dec 26 2009	dfrq
solvent	CDCl3	dn
file	exp	dpr
ACQUISITION	exp	dof
sfrq	499.864	dm
tn	H1	dms
at	5.016	dmf
np	65536	dseq
sw	6533.3	dres
fb	4000	homo
bs	4	DEC2
tpwr	61	dfrq2
pw	13.5	dn2
d1	0.100	dpr2
tof	269.9	dof2
nt	16	dm2
ct	16	dms2
alock	not used	dmf2
gain	not used	dseq2
FLAGS	n	dres2
h1	n	homo2
dp	y	DEC3
hs	mn	dfrq3
DISPLAY	mn	dn3
sp	651.6	dpr3
wp	4309.6	dof3
vs	115	dm3
sc	0	dms3
wc	250	dmf3
hzmm	17.24	dseq3
is	200.00	dres3
rfl	497.4	homo3
rpf	0	PROCESSING
th	7	wfile
ins	3.000	proc
al	ph	fn
		math
		werr
		wexp
		wbs
		wnt
		process pH
		wft



UPN-22T

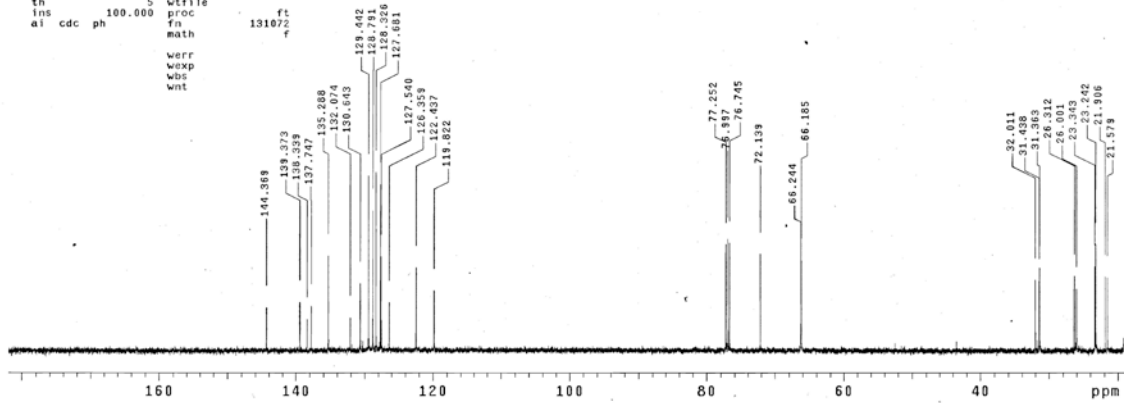
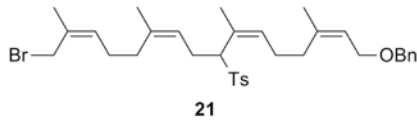
Pulse Sequence: relayh  
 Solvent: CDCl3  
 Ambient temperature  
 INOVA-500 "nmr2a.chem.nd.edu"  
 Relax. delay 1.300 sec  
 COSY 90-90  
 Acq. time 0.139 sec  
 Width 3691.6 Hz  
 2D Width 3691.6 Hz  
 8 repetitions  
 256 increments  
 OBSERVE H1, 499.8611619 MHz  
 DATA PROCESSING  
 Sine bell 0.989 sec  
 F1 DATA PROCESSING  
 Sine bell 0.035 sec  
 FT size 1024 x 1024  
 Total time 50 min, 50 sec



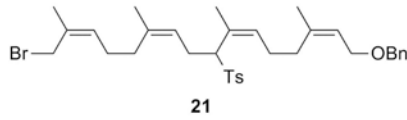
```

UPN-22T
exp2 s2pu1
SAMPLE DEC. & VT
date Dec 26 2009 dfrq 499.864
solvent CDC13 dn H1
file exp dpwr 40
ACQUISITION dof 0
sfrq 125.702 dm yyy
tn C13 dm w
at 1.215 dmf 8787.35
np 65536 dseq
sw 26963.3 dres 1.0
fb 15000 homo n
bs 4 DEC2 0
tpwr 52 dfrq2
pw 10.2 dn2
d1 1.800 dpwr2 1
tof 144.5 dof2 0
nt 640 dm2 n
ct 98 dmm2 c
clock n dmf2 10000
gain not used dseq2
FLAGS n dres2 1.0
in n homo2 n
dp y dfrq3 DEC3 0
hs nm dn3
DISPLAY dpwr3 1
sp 2391.0 dof3 0
wp 20476.3 dm3 n
vs 160 dmm3 c
sc 0 dmf3 10000
wc 250 dseq3
hzmm 81.92 dres3 1.0
is 500.00 homo3 n
rf1 11113.9 PROCESSING
rfp 9678.1 lb wtfille 1.00
th 5 proc
lms 100.000 ft
ai cdc ph fn 131072 f
werr
wexp
wbs
wnt

```



UPN-22T  
Pulse Sequence: dept



CH3 carbons



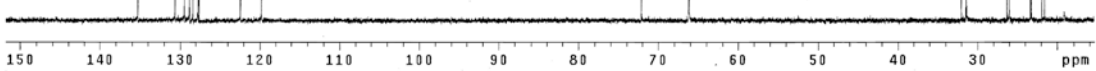
CH2 carbons



CH carbons



all protonated carbons

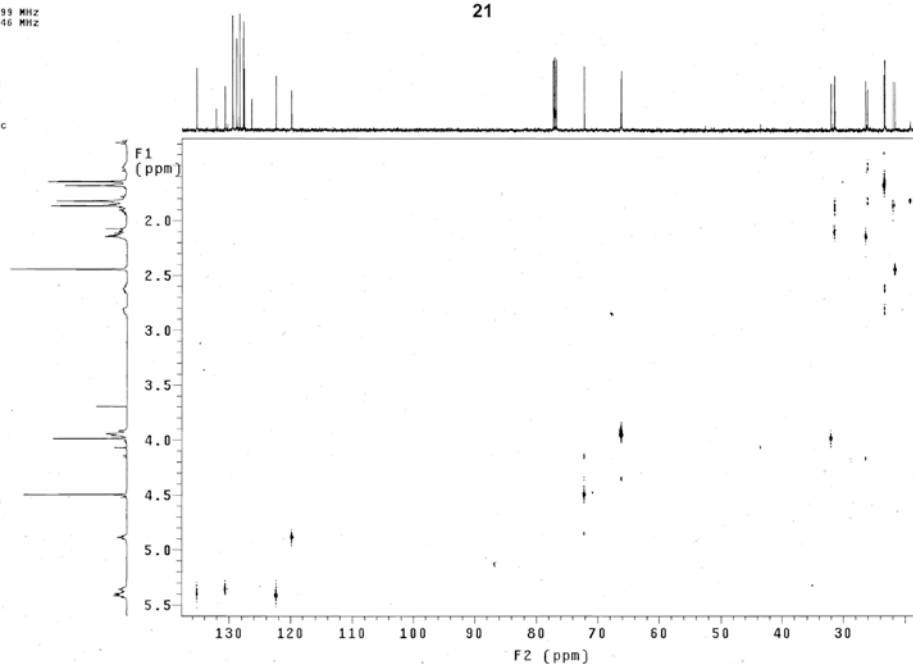
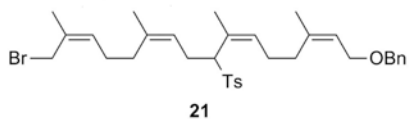




UPN-22T

Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient Temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

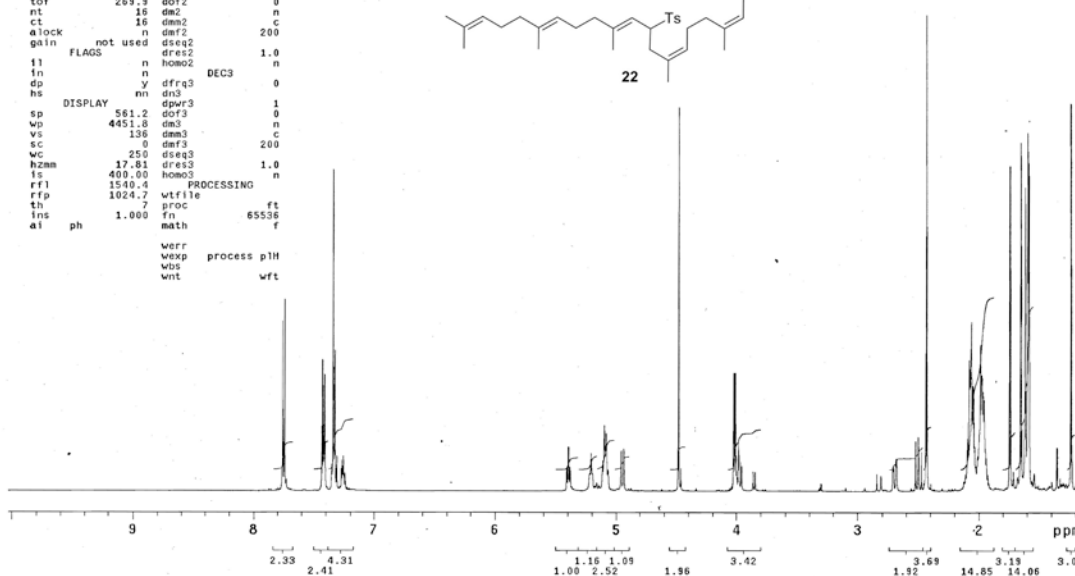
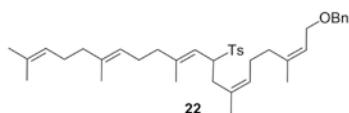
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D width 3717.3 Hz  
8 repetitions  
258 increments  
OBSERVE C13, 125.6901999 MHz  
DECOUPLE H1, 499.863646 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 57 min, 6 sec



UPH-7

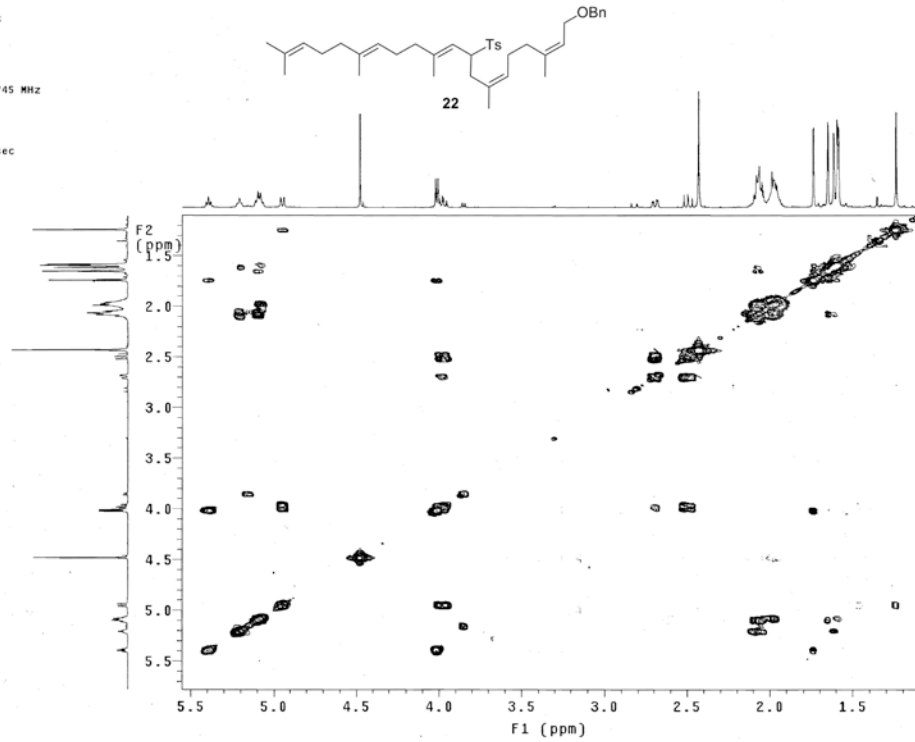
expl s2pu1

SAMPLE		DEC. & VT	
date	Jan 13 2010	dfrq	499.856
solvent	acetone	dn	H1
file	exp	spwr	30
ACQUISITION		dot	0
sfrq	499.867	dm	nm
tn	H1	dms	c
at	5.016	dmf	200
mp	65536	dsq	0
sw	6553.3	dres	1.0
fb	4000	homo	n
bs	4	DECE2	0
tpwr	81	dfrq2	0
pw	13.5	dn2	0
d1	0.100	spwr2	1
tof	269.3	dot2	0
nt	16	dm2	n
ct	16	dms2	c
alock	n	dmf2	200
gain	not used	dsq2	0
fl	FLAGS	dres2	1.0
tn	n	homo2	n
dp	n	dfrq3	0
hs	nn	dn3	0
DISPLAY		spwr3	1
sp	561.2	dot3	0
wp	4451.8	dm3	n
vs	136	dms3	c
sc	0	dmf3	200
wc	250	dsq3	0
hzsm	17.81	dres3	1.0
fs	400.00	homo3	n
rf1	1544.9	PROCESSION	ft
rfp	1024.7	wtfile	ft
th	7	proc	65536
ins	1.000	fn	f
al	ph	math	
		werr	process pH
		wexp	wnt
		wnt	wft

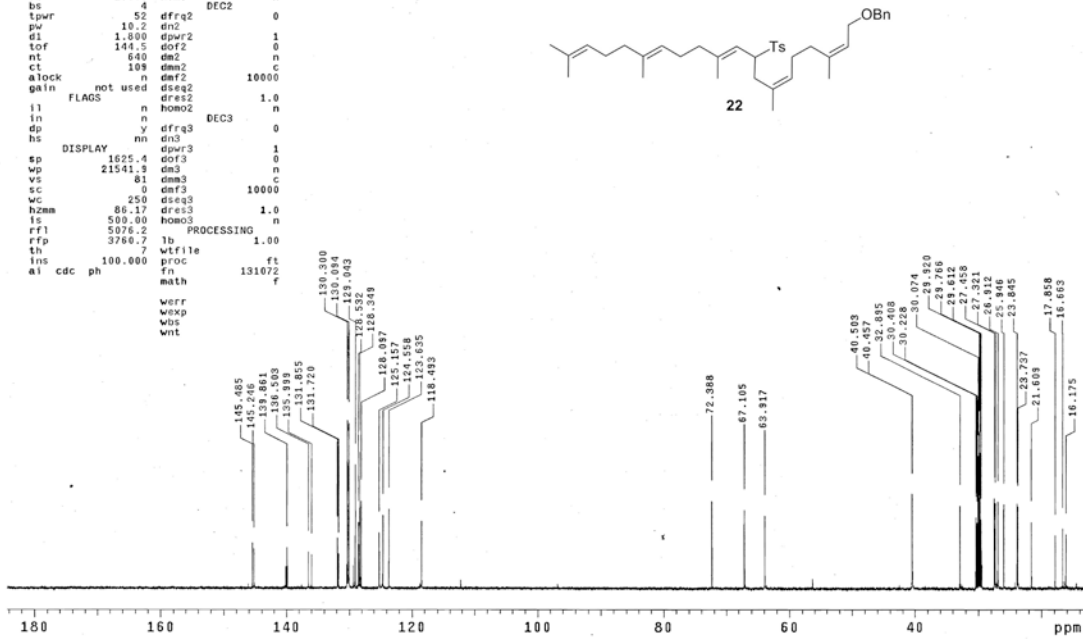


UPH-7  
Pulse Sequence: relayh  
Solvent: acetone  
Ambient Temperature  
INOVA-500 "mr2a.chem.nd.edu"

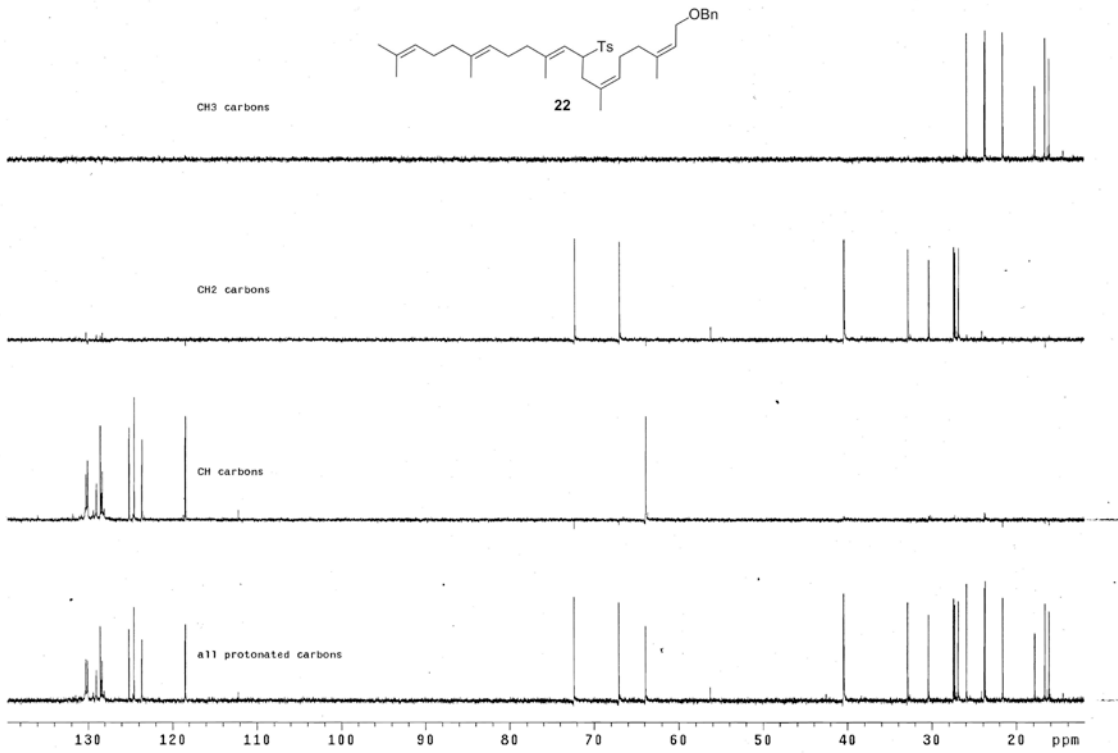
Relax, delay 1.300 sec  
CDSV 36-90  
Acq. time 0.134 sec  
Width 3820.1 Hz  
2D Width 3820.1 Hz  
8 repetitions  
256 increments  
OBSERVE H1 499.8637745 MHz  
DATA PROCESSING  
Sine bell 0.087 sec  
F1 DATA PROCESSING  
Sine bell 0.034 sec  
FT size 1024 x 1024  
Total time 50 min, 38 sec



UPH-7  
exp2 s2pu1  
SAMPLE date Jan 13 2010 DEC. & VT 499.866  
solvent Acetone dn HI  
file exp epwr 40  
ACQUISITION srrq 125.703 dm yyv  
tn 213 dms 8787.35  
at 1.115 dmf  
np 65536 dsq  
sw 26963.3 dres 1.0  
fb 15000 homo n  
bs 4 DEC2  
tpwr 52 dfra2 0  
pw 18.2 dn2  
d1 1.800 epwr2 1  
tof 144.5 dof2 0  
nt 640 dm2 n  
ct 109 dms2 c  
alock n dmf2 10000  
gain not used dsq2 1.0  
ll FLAGS n dsq2 n  
ll n homo2  
dp y dfra3 0  
hs mn dn3  
DISPLAY epwr3 1  
sp 1625.4 dof3 0  
wp 21541.3 dm3 n  
vs 81 dms3 c  
sc 0 dmf3 10000  
wc 250 dsq3  
hzma 86.17 dres3 1.0  
ls 500.00 homo3 n  
rf1 5076.2 PROCESSING  
rfp 3760.7 lb  
th 7 wfile 1.00  
ins 100,000 proc fn  
ai cdc ph math 131072 r

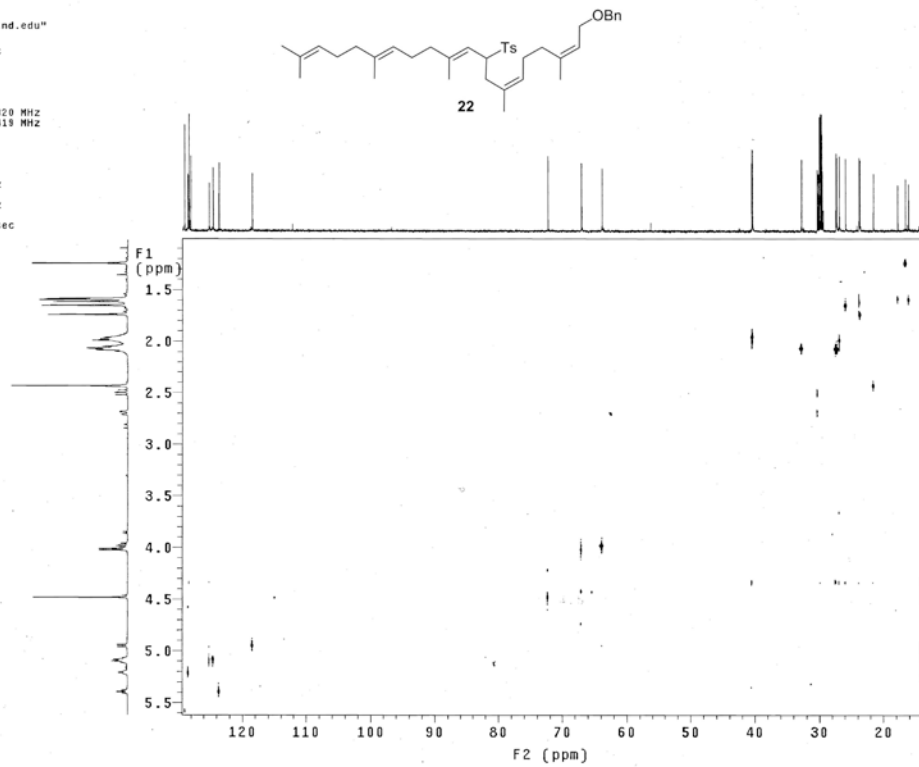


UPH-7  
Pulse Sequence: dept



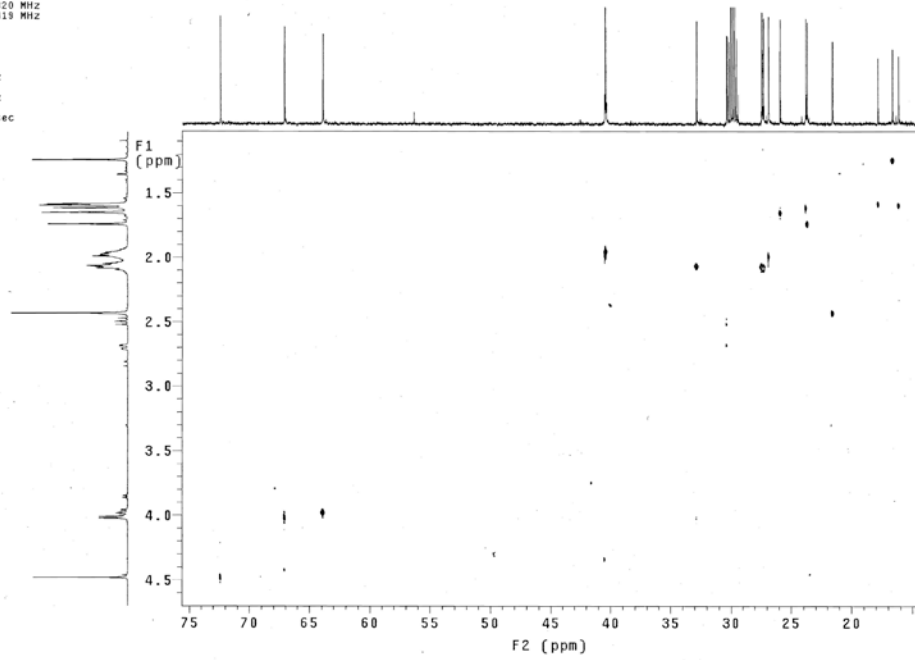
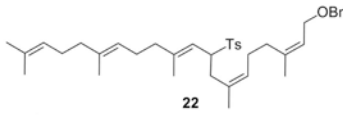
UPH-7  
Pulse Sequence: hetcor  
Solvent: Acetone  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 38403.5 Hz  
2D Width 3562.7 Hz  
4 repetitions  
256 increments  
OBSERVE C13, 125.6907320 MHz  
DECUPLE H1, 499.8659419 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 28 min, 38 sec

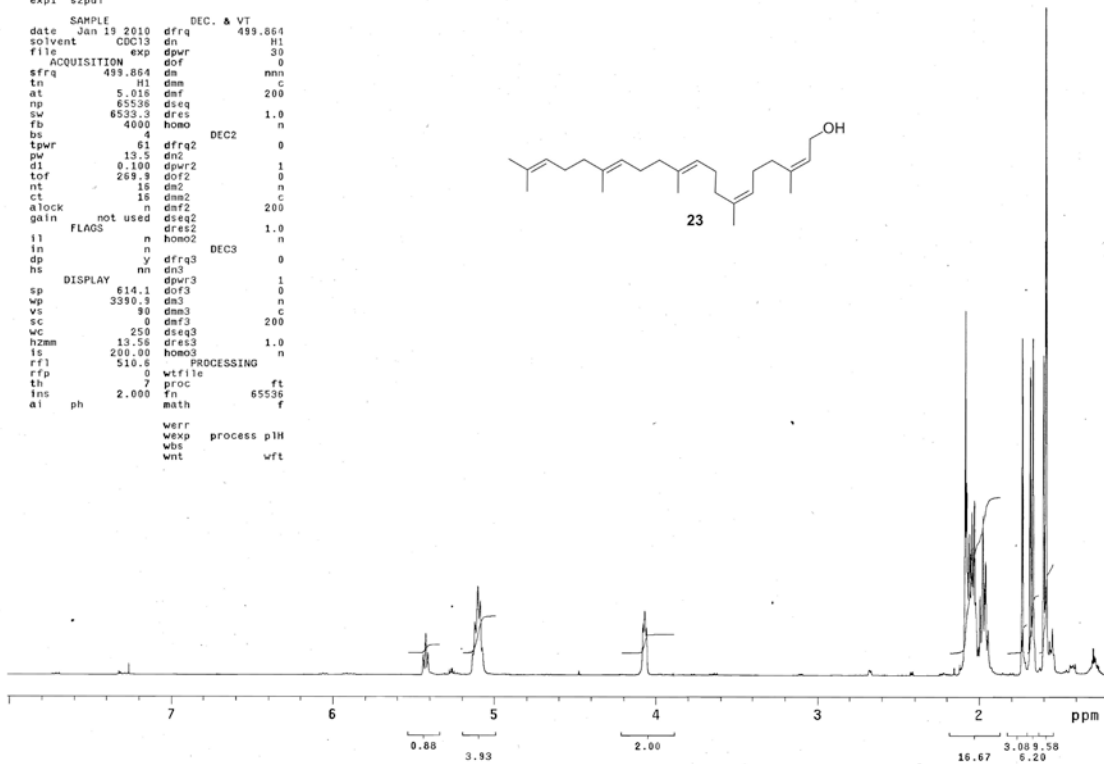
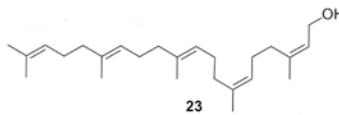


UPH-7  
Pulse Sequence: hetcor  
Solvent: Acetone  
Ambient temperature  
User: 1-14-87  
INGVA-500 "nmr2a.chem.nd.edu"

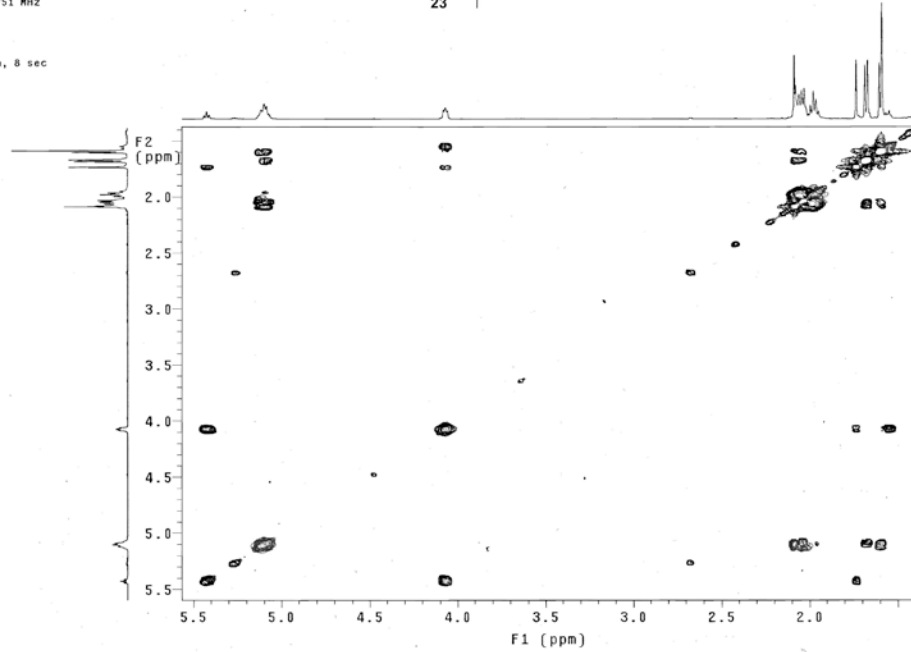
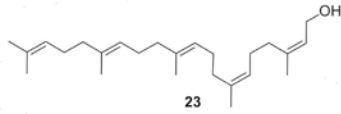
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 3562.7 Hz  
4 repetitions  
256 increments  
OBSERVE C13, 125.6907320 MHz  
DECOUPLE H1, 499.8659419 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 28 min, 38 sec



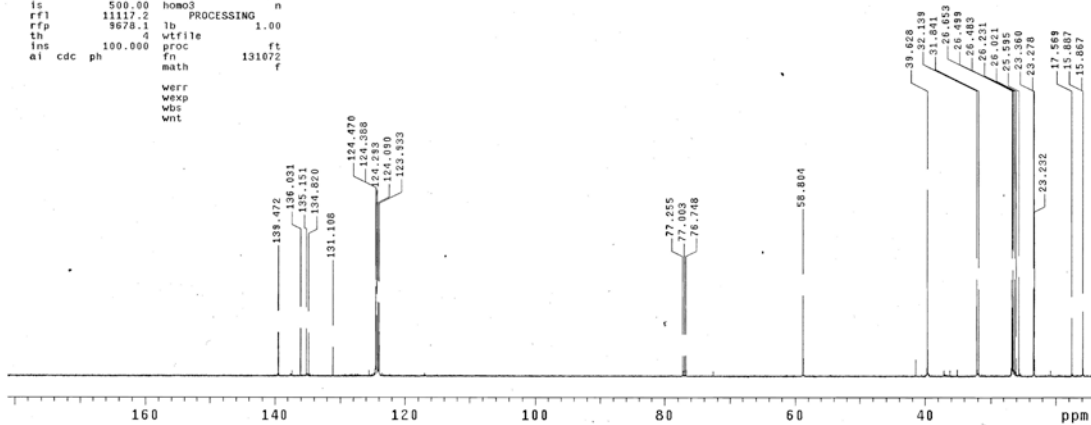
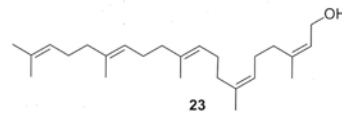
UPH-8  
exp1 s2pu1  
SAMPLE DEC. & VT  
date Jan 19 2010 dfrq 499.864  
solvent CDCl3 dn H1  
file exp dpwr 30  
ACQUISITION dof 0  
sfrq 499.864 da nnn  
tn H1 dnm C  
at 5.016 dmf 200  
np 85536 dseq  
sw 6533.3 dres 1.0  
fb 4000 homo n  
bs 4 DEC2  
tpwr 61 dfrq2 0  
pw 13.5 dnc 1  
d1 0.100 dpwr2 1  
tof 269.9 dof2 0  
nt 16 dnc n  
ct 16 dnm2 C  
alock n dmf2 200  
gain not used dseq2 1.0  
FLAGS n homo2 n  
in n DEC3  
dp y dfrq3 0  
hs nn dnc3 1  
DISPLAY dpwr3 1  
sp 614.1 dof3 0  
wp 3590.3 dnc3 n  
vs 90 dnm3 C  
sc 0 dmf3 200  
wc 250 dseq3  
hzmm 13.56 dres3 1.0  
is 200.00 homo3 n  
rfl 510.6 PROCESSING  
rfp 0 wfile ft  
th 7 proc ft  
ins 2.000 tn 65536  
al ph math f  
werr  
wexp process pH  
wbt  
wnt wft



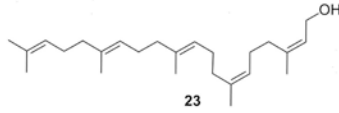
UPH-8  
Pulse Sequence: relayh  
Solvent: CDCl3  
Ambient temperature  
INOVA-500 "mr2a.chem.nd.edu"  
Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.151 sec  
Width 3382.2 Hz  
ZD Width 3382.2 Hz  
8 repetitions  
512 increments  
OBSERVE H1, 499.8611751 MHz  
DATA PROCESSING  
Sine bell 0.078 sec  
F1 DATA PROCESSING  
Sine bell 0.038 sec  
FT size 1024 x 1024  
Total time 1 hr, 45 min, 8 sec



UPH-8  
exp2 s2pul  
SAMPLE DEC. & VT  
date Jan 19 2010 dfrq 499.864  
solvent CDCl3 dn H1  
file exp dpr 40  
ACQUISITION  
sfrq 125.762 dm 0  
tn C13 dma yyy  
at 1.215 dmf 8787.35  
np 65536 dseq  
sv 26963.3 dres 1.0  
fb 15000 homo n  
bs 4 DEC2  
tpwr 52 dfrq2 0  
pw 18.2 dn2 1  
d1 1.890 dpr2 0  
tof 144.5 dof2 0  
nt 640 dm2 n  
ct 93 dma2 c  
atock n dmf2 10000  
gain not used dseq2  
FLAGS n homo2 1.0  
l1 n DEC3  
ln n  
dp y dfrq3 0  
hs nn dn3  
DISPLAY dpr3 1  
sp 1785.3 dof3 0  
wp 20974.9 dm3 n  
vs 32 dma3 c  
sc 0 dmf3 10000  
wc 250 dseq3  
hzmm 83.90 dres3 1.0  
is 500.00 homo3 n  
rfl 11117.2 PROCESSING  
rfp 9678.1 lb 1.00  
th 4 wtf1le ft  
lms 100.000 proc fn 131072  
ai cdc ph math f  
werr  
wexp  
wbs  
wnt



UPH-8  
Pulse Sequence: dept

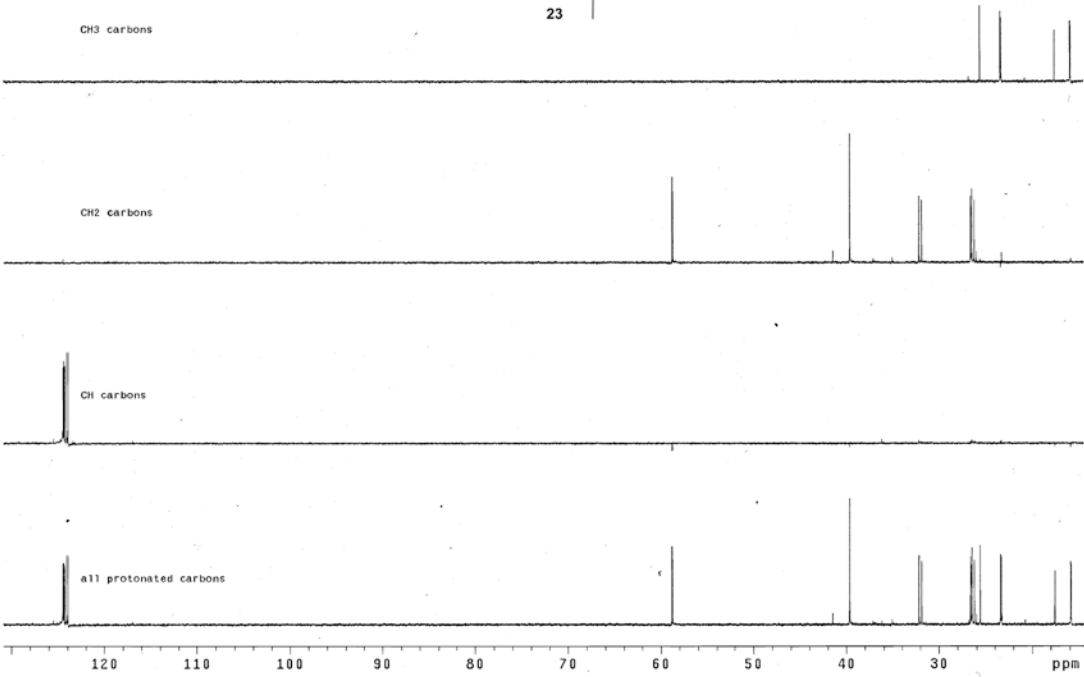


CH3 carbons

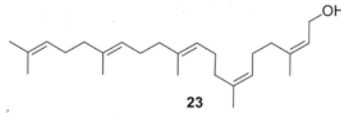
CH2 carbons

CH carbons

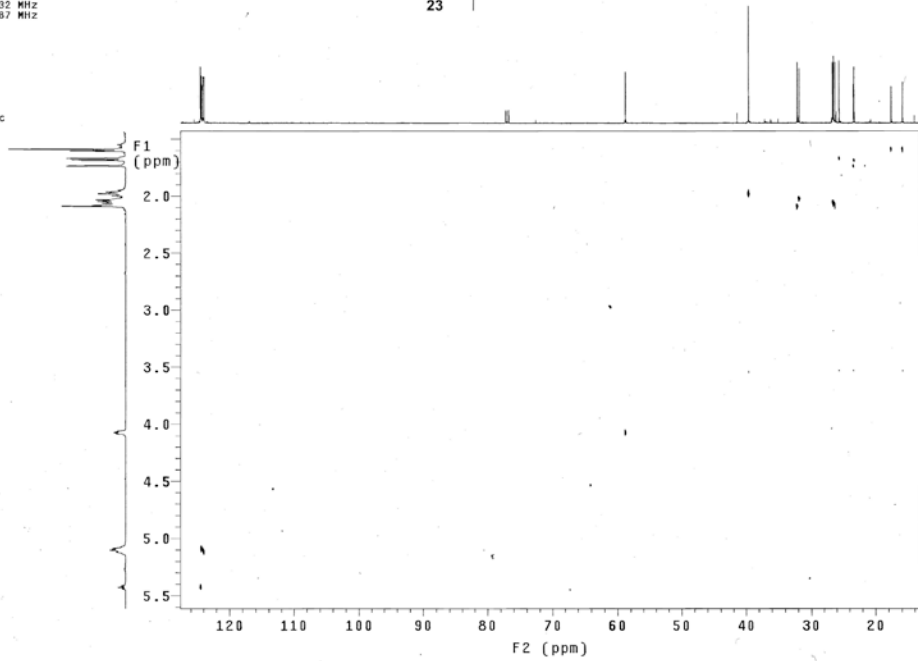
all protonated carbons



UPH-8  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INDVA-500 "nmr2a.chem.nd.edu"

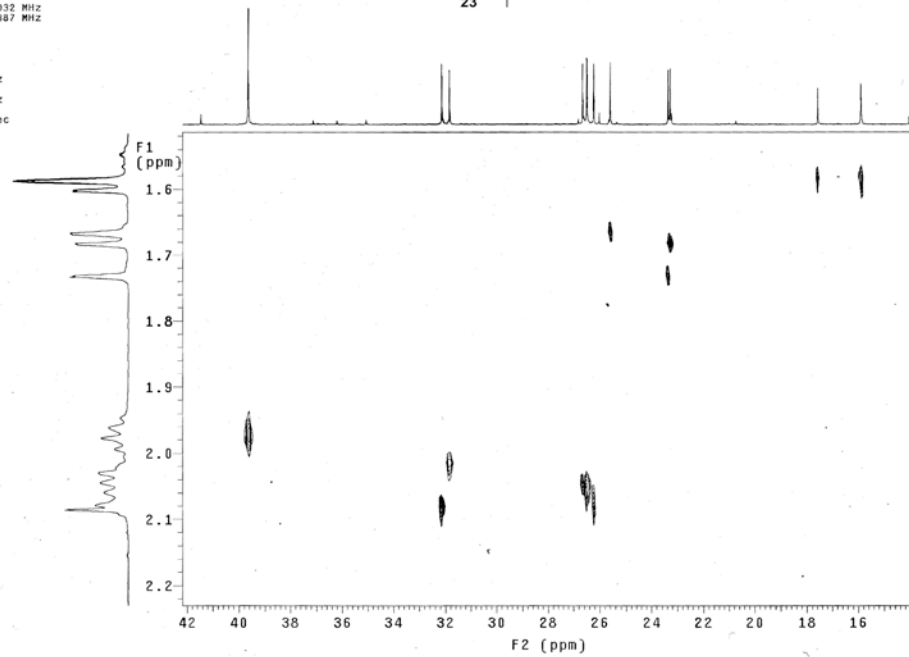
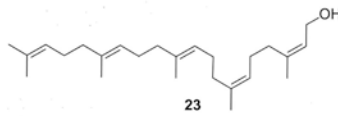


Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 2129.8 Hz  
4 repetitions  
258 increments  
OBSERVE C13, 125.6902032 MHz  
DECOUPLE H1, 499.8629387 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 4096 x 512  
Total time 29 min, 2 sec



UPH-8  
Pulse Sequence: **hstcor**  
Solvent: CDCl3  
Ambient temperature  
User: 1-10-87  
INOVA-500 "mm2a.chem.nd.edu"

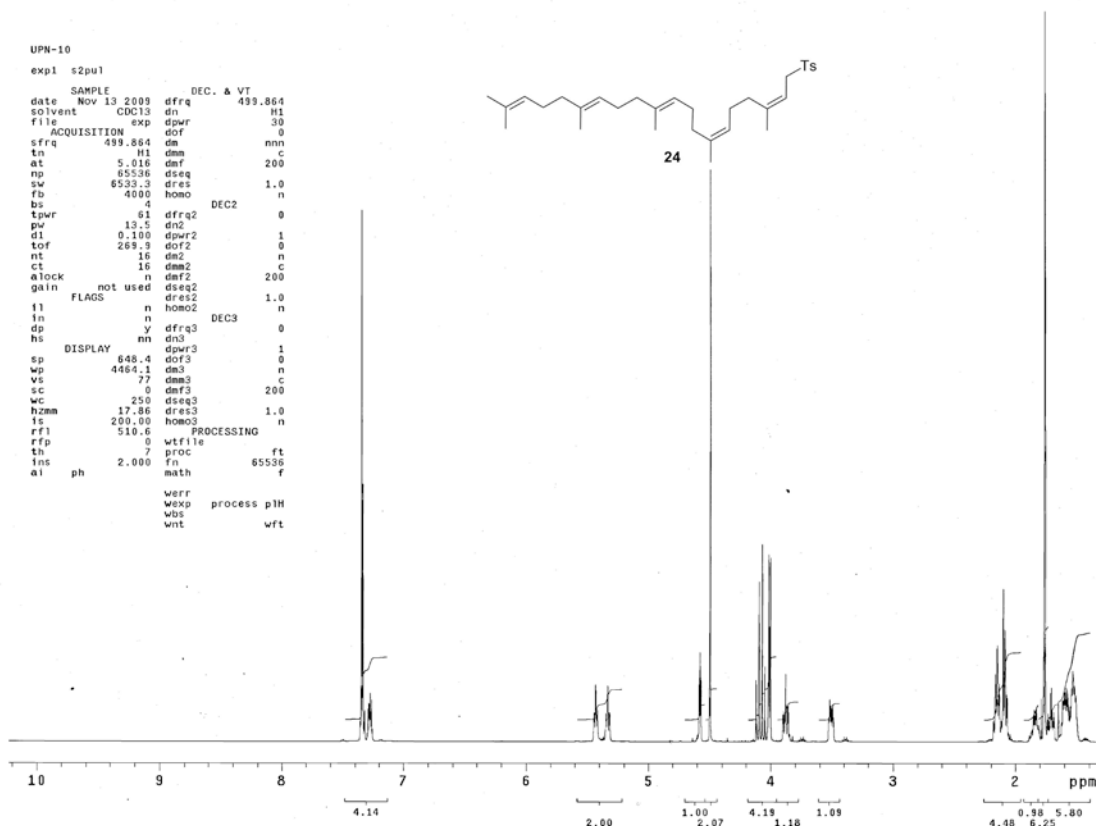
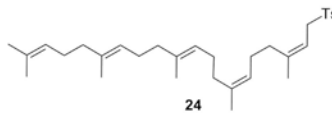
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 19401.5 Hz  
2D Width 2179.0 Hz  
4 repetitions  
256 increments  
OBSERVE C13, 125.6902032 MHz  
DECOUPLE H1, 499.8623387 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 4096 x 512  
Total time 29 min, 2 sec



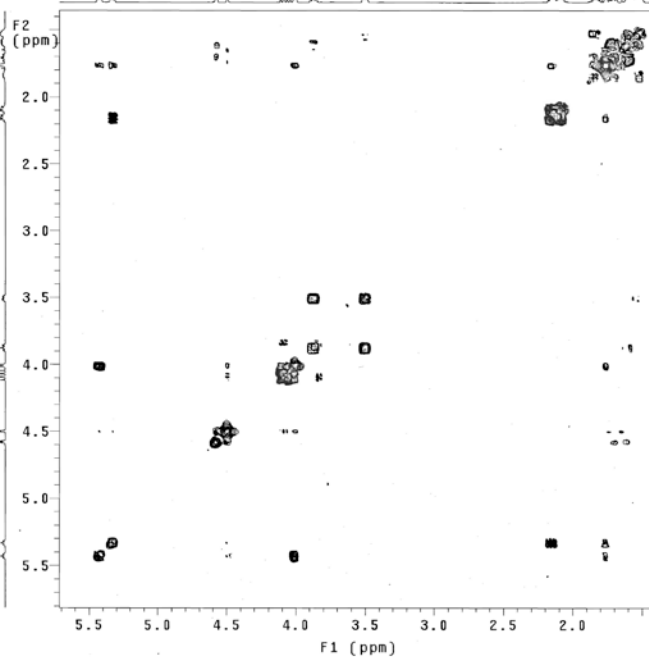
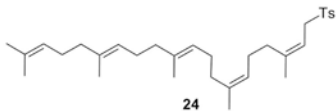
UPN-10  
expl s2pul

SAMPLE DEC. & VT  
date Nov 13 2009 dfrq 499.864  
solvent CDCl3 dn H1  
file exp dpwr 30  
ACQUISITION dof 0  
sfrq 499.864 da mn  
tn H1 dm c  
at 5.016 dmf 200  
np 85536 dseq  
sw 6533.3 dres 1.0  
fb 4000 homo n  
bs 4  
tpwr 61 dfrq2 DEC2 0  
pw 13.5 dn2  
d1 0.100 dpwr2 1  
tof 269.3 dof2 0  
nt 16 dm2 n  
ct 16 dm2 c  
alock n dmf2 200  
gain not used dseq2  
FLAGS dres2 1.0  
ll n homo2 n  
dp y dfrq3 DEC3 0  
hs nn dn3  
DISPLAY dpwr3 1  
sp 648.4 dof3 0  
wp 4464.1 dm3 n  
vs 77 dm3 c  
sc 0 dmf3 200  
wc 250 dseq3  
hzmm 17.86 dres3 1.0  
ls 200.00 homo3 n  
rfl 510.6  
rfp 0 wtfile PROCESSING  
th 7 proc ft  
ins 2.000 fa 65536 f  
al ph math

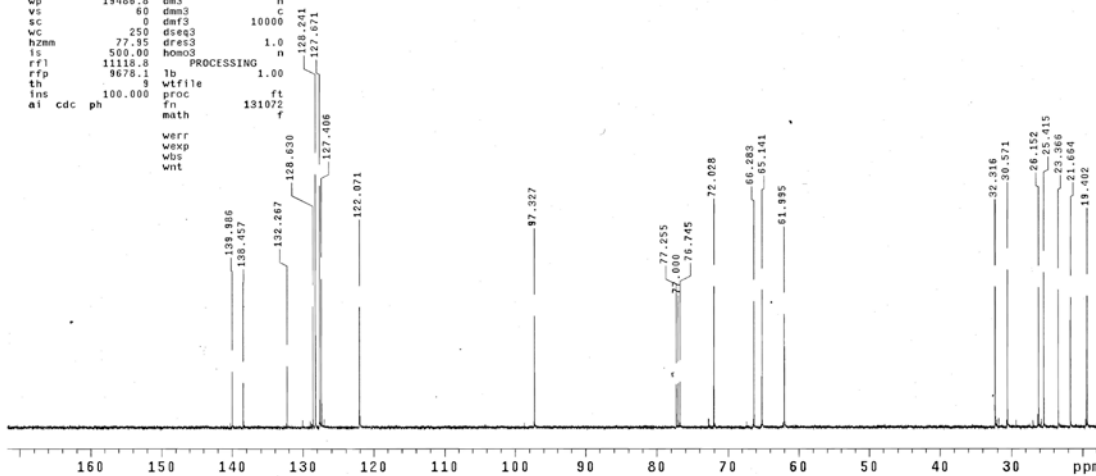
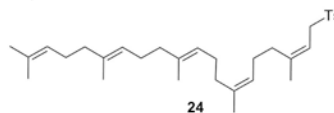
werr  
wexp process pH  
wbs  
wrt



UPN-10  
Pulse Sequence: relayh  
Solvent: CDC13  
Ambient temperature  
INOVA-500 "mrca.chem.nd.edu"  
Relax. delay 1.300 sec  
COSY 90-90  
Acq. time 0.162 sec  
Width 3167.7 Hz  
2D Width 3167.7 Hz  
4 repetitions  
256 increments  
OBSERVE HI 499.8611751 MHz  
DATA PROCESSING  
Sine bell 0.091 sec  
F1 DATA PROCESSING  
Sine bell 0.040 sec  
FT size 1024 x 1024  
Total time 26 min, 0 sec

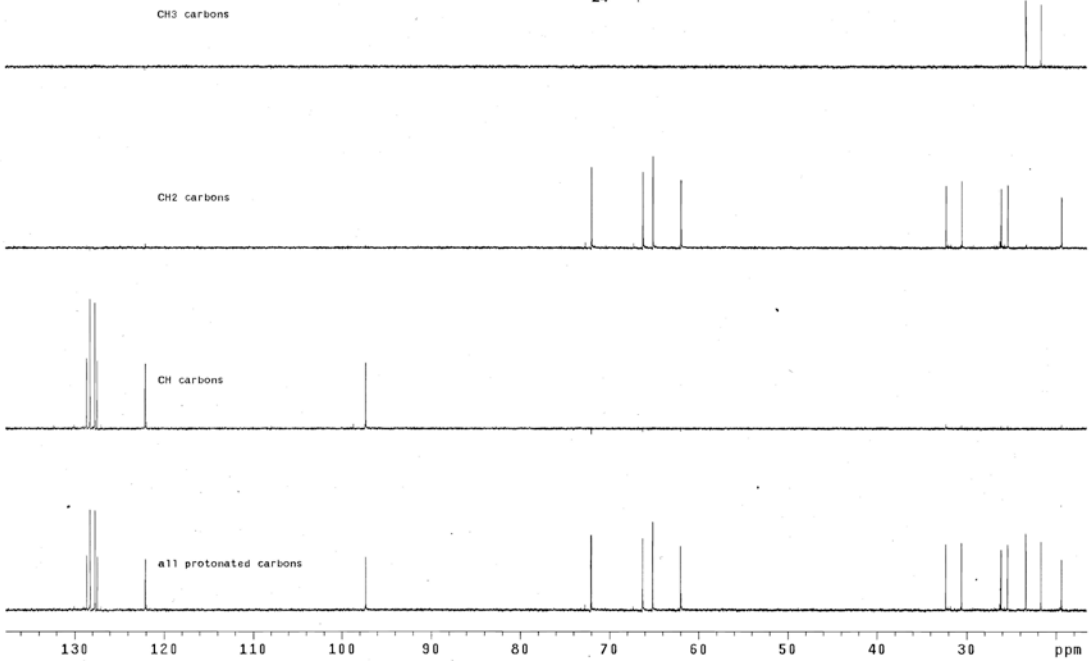
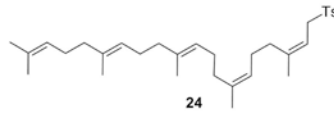


UPN-10  
exp2 s2pu1  
SAMPLE DEC. & VT  
date Nov 13 2008 dfrq 499.864  
solvent CDC13 dn HI  
file exp dpwr 40  
ACQUISITION dof 0  
sfrq 125.702 dm VVY  
tn C13 dm 8787.35  
at 1.215 dmf  
np 65536 dseq  
sw 26963.3 dres 1.0  
fb 15000 homo n  
bs 4  
tpwr 52 dfrq2 0  
pv 10.2 dn2  
d1 1.800 dpwr2 1  
tof 144.5 dof2 0  
nt 640 dn2 0  
ct 46 dm2 c  
atock n dm2 10000  
gain not used dres2 1.0  
il FLAGS n homo2 n  
in n  
dp y dfrq3 0  
hs nn dn3  
DISPLAY dpwr3 1  
sp 2102.5 dof3 0  
wp 19466.8 dm3  
vs 60 dm3  
sc 0 dm3 10000  
wc 250 dres3  
hzm 77.95 dm3 1.0  
ls 500.00 homo3 n  
rf1 11118.6 PROCESSING  
rfp 9678.1 lb wtfile 1.00  
th 9  
ins 100.000 proc ft  
ai cdc ph fn 131072  
math f  
werr  
wexp  
wbs  
wnt



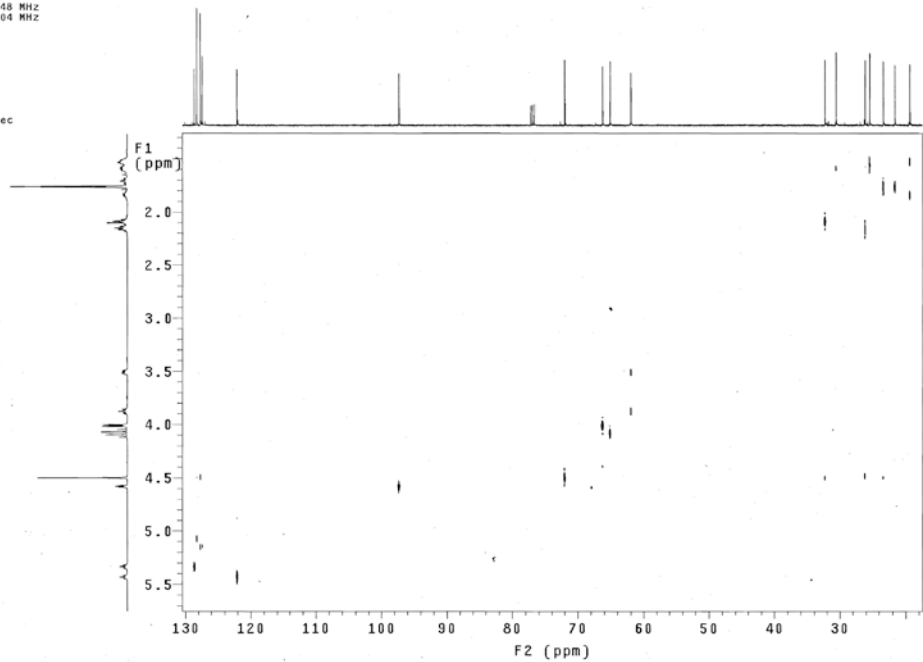
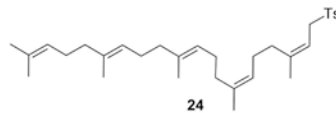


UPN-10  
Pulse Sequence: dept



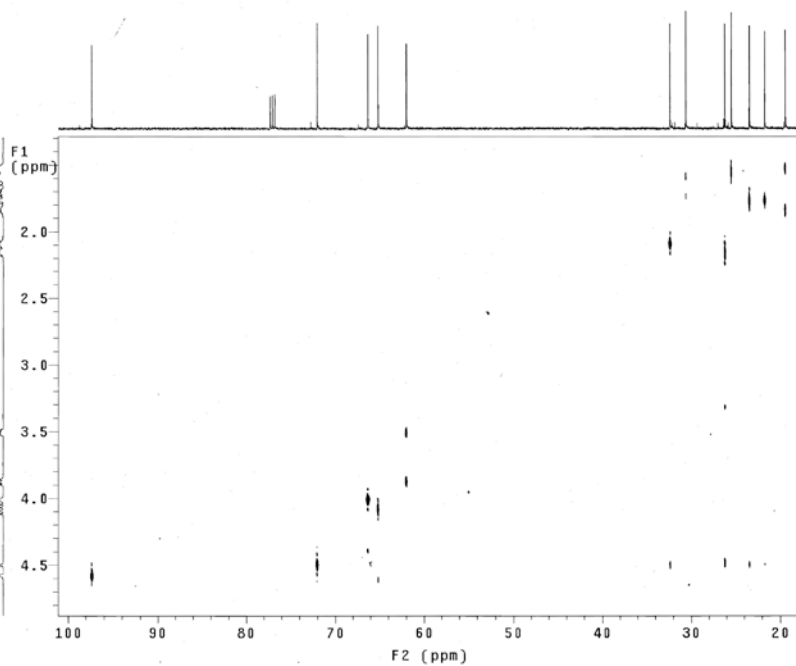
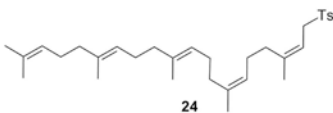
UPN-10  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 3279.4 Hz  
4 repetitions  
128 increments  
OBSERVE C13, 125.6902048 MHz  
DECOUPLE H1, 499.8634204 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 4096 x 512  
Total time 14 min, 13 sec

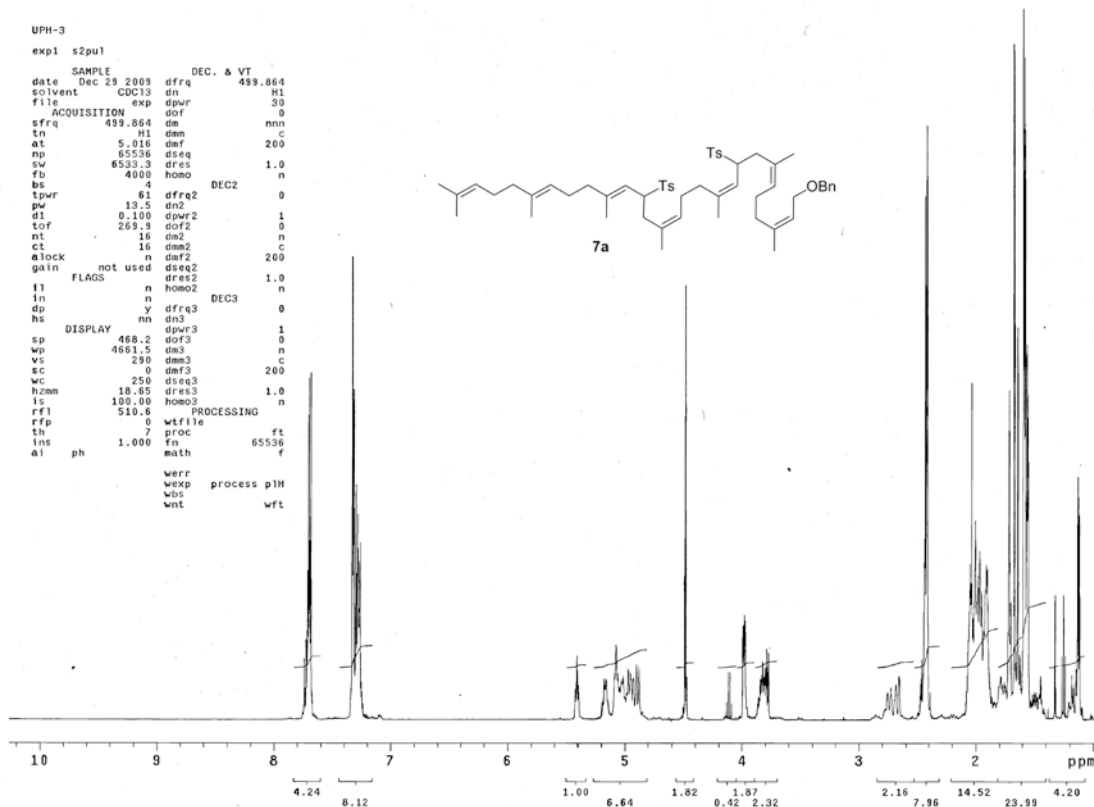
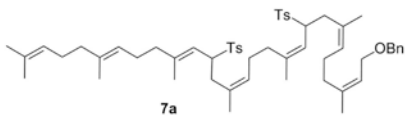


UPH-10  
 Pulse Sequence: hetcor  
 Solvent: CDCl3  
 Ambient temperature  
 User: 1-14-97  
 INOVA-500 "nmr2a.chem.nd.edu"

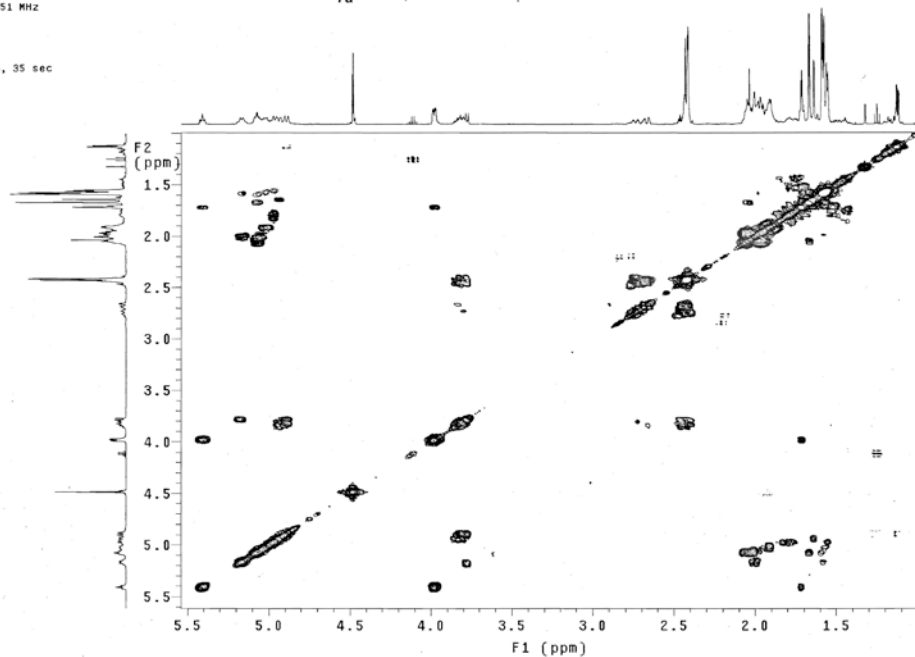
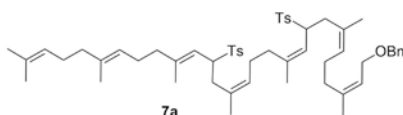
Relax. delay 1.500 sec  
 Acq. time 0.111 sec  
 Width 18403.5 Hz  
 2D Width 3273.4 Hz  
 4 repetitions  
 128 increments  
 OBSERVE C15, 125.6902048 MHz  
 DECOUPLE H1, 499.8634204 MHz  
 Power 40 db  
 on during acquisition  
 off during delay  
 WALTZ-16 modulated  
 DATA PROCESSING  
 Line broadening 1.0 Hz  
 F1 DATA PROCESSING  
 Line broadening 0.3 Hz  
 FT size 4096 x 512  
 Total time 14 min, 13 sec



UPH-3  
 exp1 s2pu1  
 SAMPLE DEC. & VT  
 date Dec 29 2003 dfrq 499.864  
 solvent CDC13 dn H1  
 file exp dpr 30  
 ACQUISITION  
 sfrq 499.864 dm mnn  
 tn H1 dmn C  
 at 5.016 def 200  
 np 65536 dseq C  
 sw 6533.3 dres 1.0  
 fb 4000 homo n  
 bs 4 DEC2  
 tpwr 61 dfrq2 0  
 pw 13.5 dn2 1  
 dt 0.100 dpr2 1  
 tof 269.3 dof2 0  
 nt 16 dm2 n  
 ct 16 dmq2 C  
 alock not used daf2 200  
 gain FLAGS dseq2 dres2 1.0  
 in n homo2 n  
 dp y dfrq3 DEC3 0  
 hs nn dn3  
 DISPLAY dpr3 1  
 sp 468.2 dof3 0  
 wp 4601.5 dm3 n  
 vs 290 dmq3 C  
 sc 0 dmf3 200  
 wc 250 dseq3 1.0  
 hzm 18.65 dres3 n  
 is 100.00 homo3 n  
 rfl 510.6 PROCESSING  
 rfp 0 wfile ft  
 th 7 proc fa 65536  
 lns ph 1.000 math f  
 al  
 werr  
 wexp process pH  
 wbs  
 wnt wft

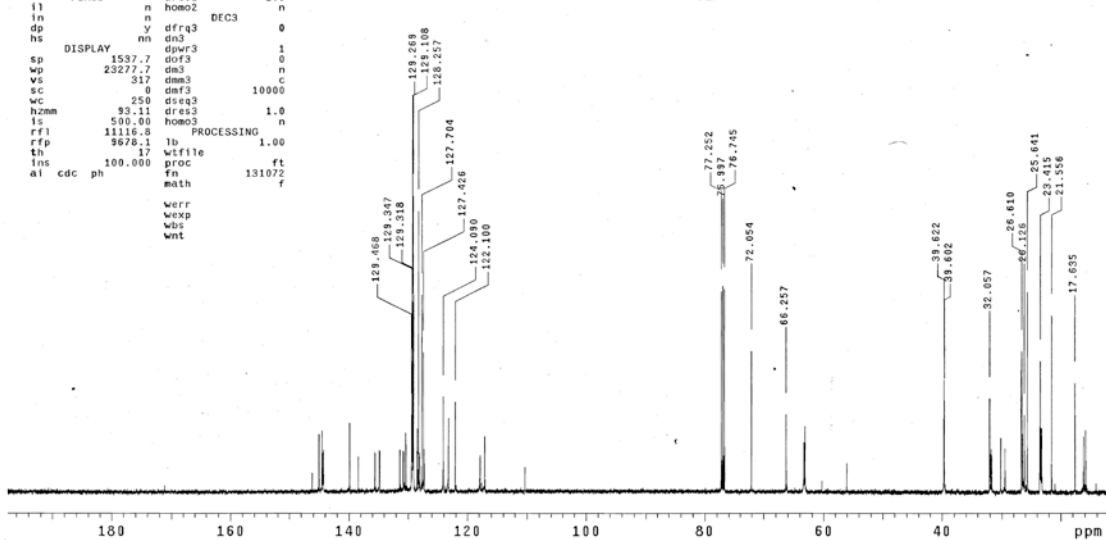
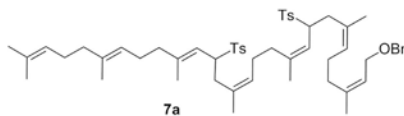


UPH-3  
Pulse Sequence: relayh  
Solvent: CDCl3  
Ambient temperature  
INOVA-500 nmr@chem.nd.edu  
Relax: delay 1.300 sec  
COSY 90-90  
Acq. time 0.140 sec  
Width 3657.1 Hz  
2D Width 3657.1 Hz  
16 repetitions  
256 increments  
OBSERVE: H1, 499.8611751 MHz  
DATA PROCESSING  
Sine bell 0.070 sec  
F1 DATA PROCESSING  
Sine bell 0.035 sec  
FT size 1024 x 1024  
Total time 1 hr, 41 min, 35 sec

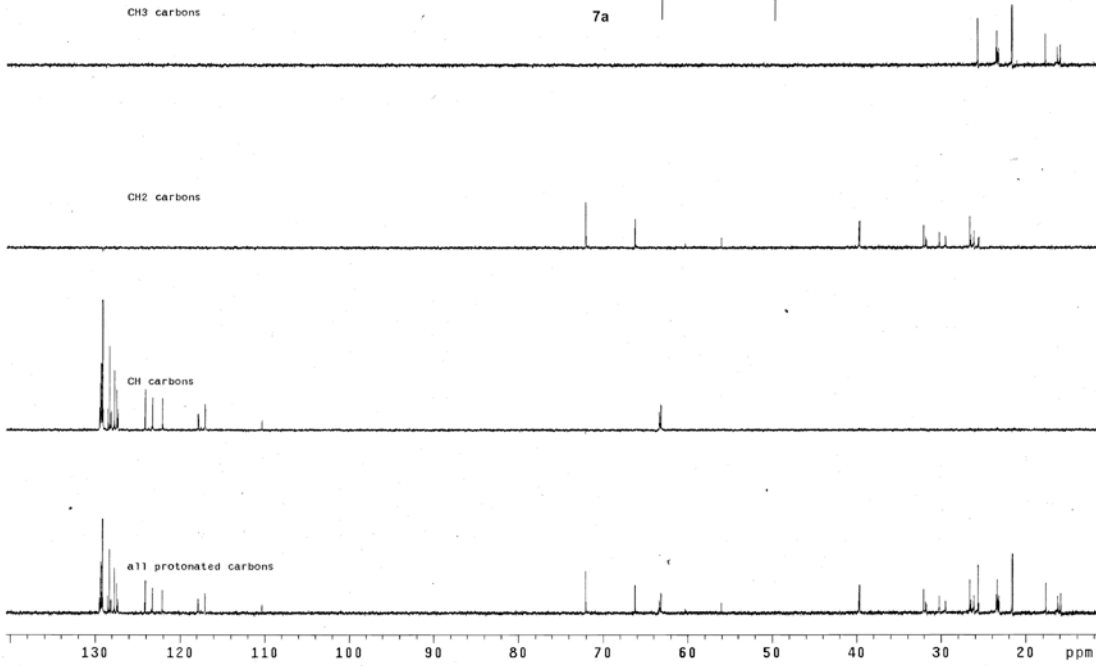
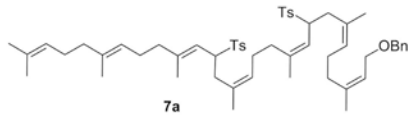


UPH-3  
exp2 s2pul

SAMPLE		DEC. & VT	
date	Dec 29 2009	dfrq	499.864
solvent	CDCl3	dn	H1
file	exp	dpvr	40
ACQUISITION			
sfrq	125.702	dm	yyv
tn	C13	dmm	y
at	1.215	dmf	8787.35
np	65536	dseq	
sv	26983.3	dres	1.0
fb	15000	homo	n
bs	4	DEC2	n
tpvr	52	dfrq2	0
pw	10.2	dm2	1
d1	1.800	dvr2	1
tof	144.5	dof2	0
nt	640	dm2	n
ct	640	dmm2	c
alock	n	dmf2	10000
gain	not used	dseq2	1.0
ll	FLAGS	dres2	1.0
ln	n	homo2	n
dp	y	dfrq3	0
hs	nn	dn3	0
DISPLAY			
sp	1537.7	dof3	1
wp	23277.7	dm3	n
vs	317	dmm3	c
sc	0	dmf3	10000
wc	250	dseq3	1.0
hzm	93.11	dres3	n
is	500.00	homo3	n
rfl	11116.0	PROCESSION	ft
rfp	9678.1	lb	1.00
th	100.000	wfille	
lms	100.000	PROC	ft
al	cdc	fn	131072
	ph	math	f

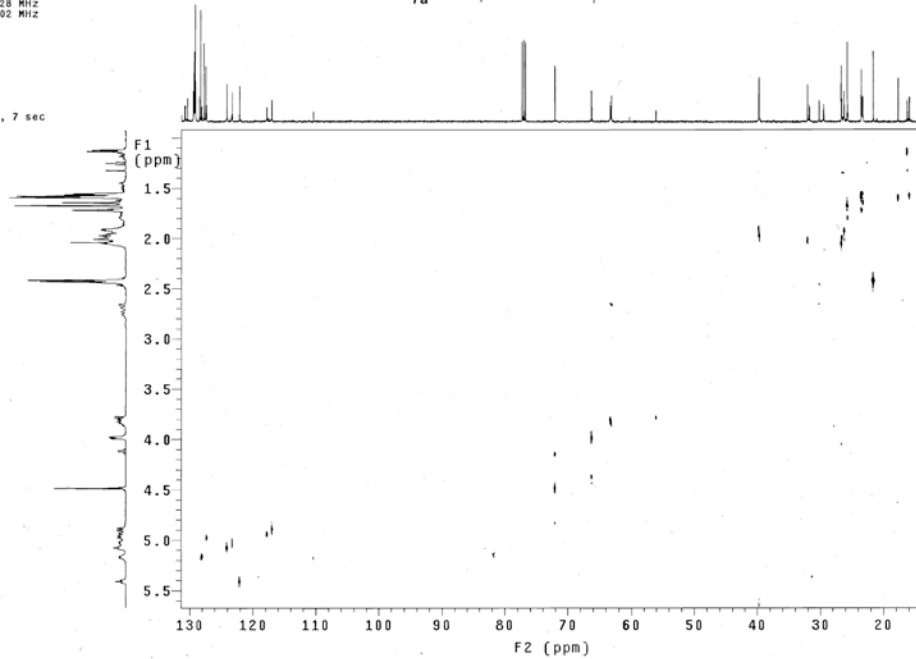
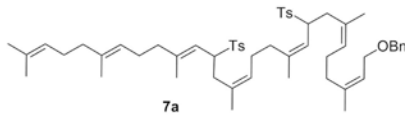


UPH-3  
Pulse Sequence: dept

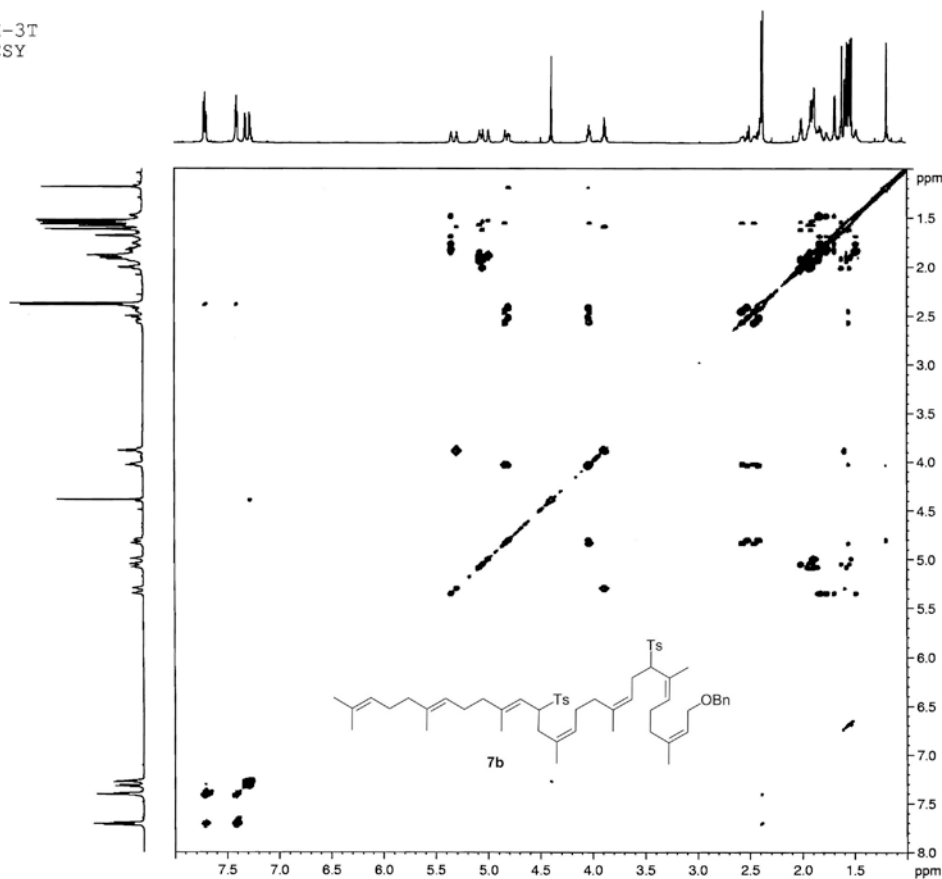


UPH-3  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 15403.5 Hz  
2D Width 3682.7 Hz  
16 repetitions  
256 increments  
OBSERVE C13, 125.6902028 MHz  
DECOUPLE H1, 499.8633902 MHz  
Power 40 db  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 1024  
Total time 1 hr, 54 min, 7 sec



UPH-3T  
TOCSY



```

Current Data Parameters
NAME      UPH-3T
EXPNO    3
PROCNO   1

F2 - Acquisition Parameters
Date_    2010110
Time     1.49
INSTRUM  spect
PROBHD   5 mm CPTCI 1H-
PULPROG  dipz1zetepzi
TD       1024
SOLVENT  DMSO
NS       16
DS       16
SWH      8223.685 Hz
FIDRES   8.030942 Hz
AQ       0.0021092 sec
RG       14.2
DM       50.800 usec
DE       30.00 usec
TE       301.9 K
dC       0.0000000 sec
D1       1.39999998 sec
d11      0.01000000 sec
D16      0.00000000 sec
D20      0.00001000 sec
D21      0.00001000 sec
D9       0.34000000 sec
DELTA1   0.00120300 sec
DELTA13  0.00120800 sec
FACTOR1  11
IN0      0.00012160 sec
I1       22
ST1CNT   0

----- CHANNEL f1 -----
NUC1     1H
P1       9.30 usec
P2       16.60 usec
P6       24.00 usec
PL1      0.20 dB
PL10     9.43 dB
SFO1     800.1837640 MHz

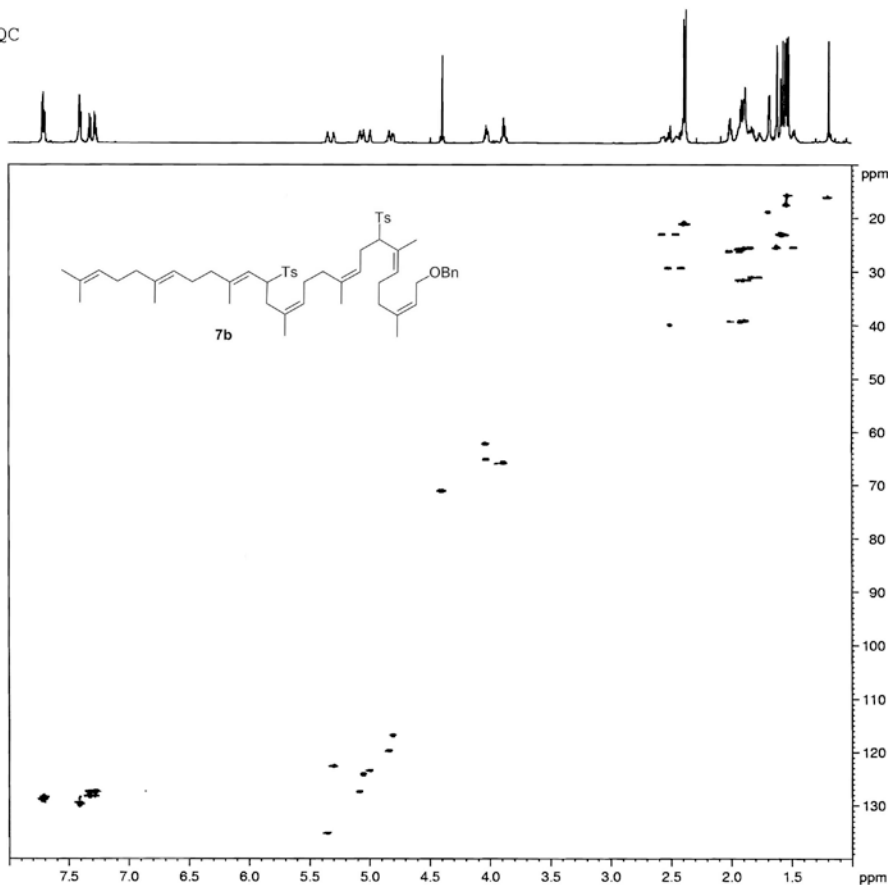
----- GRADIENT CHANNEL -----
GPNAM1   SINE-100
GPNAM2   SINE-100
GP21     30.00 A
GP22     30.00 A
PL6      1000.00 usec

F1 - Acquisition parameters
TD       1024
SFO1     800.1838 MHz
FIDRES   8.030942 Hz
SW       16.277 ppm
PRMODE   Echo-Antiecho

F2 - Processing parameters
SI       2048
SF       800.1800073 MHz
WDW      QSI
SSB      2
LB       0 Hz
GB       0
PC       1.40

F1 - Processing parameters
SI       2048
MC2      echo-antiecho
SF       800.1800072 MHz
WDW      States-TFPI
SSB      2
LB       0 Hz
GB       0
  
```

UPH-3T  
13C-HSQC



```

Current Data Parameters
NAME      UPH-3T
EXPNO    1
PROCNO   1

F2 - Acquisition Parameters
Date_    20100110
Time     9.44
INSTRUM  spect
PROBHD   5 mm CPTCI 1H-
PULPROG  hsqz1epzi
TD       1024
SOLVENT  DMSO
NS       16
DS       16
SWH      8223.685 Hz
FIDRES   8.030942 Hz
AQ       0.0021092 sec
RG       14.2
DM       50.800 usec
DE       30.00 usec
TE       301.9 K
dC       0.0000000 sec
D1       1.39999998 sec
d11      0.01000000 sec
D16      0.00000000 sec
D20      0.00001000 sec
D21      0.00001000 sec
D9       0.34000000 sec
DELTA1   0.00120300 sec
DELTA13  0.00120800 sec
FACTOR1  11
IN0      0.00012160 sec
I1       22
ST1CNT   0

----- CHANNEL f1 -----
NUC1     1H
P1       9.30 usec
P2       16.60 usec
P6       24.00 usec
PL1      0.20 dB
PL10     9.43 dB
SFO1     800.1837640 MHz

----- CHANNEL f2 -----
CPDPRG2  p04ep180
NUC2     13C
P11      500.00 usec
P21      2000.00 usec
P3       14.80 usec
PCPD2    1500.00 usec
PC12     10.00 dB
PC13     10.00 dB
PC14     10.00 dB
SFO2     201.2603114 MHz
SFO3     1.00 MHz
SFO4     1.00 MHz
SFO5     Exp.80, 1.10, 1.00
SFO6     Exp.80, 0.5, 20, 1
SFO7     0.500
SFO8     0.500
SFO9     0.500
SFO10    0 Hz
SFO11    0 Hz
SFO12    0 Hz
SFO13    0 Hz
SFO14    0 Hz
SFO15    0 Hz
SFO16    0 Hz
SFO17    0 Hz

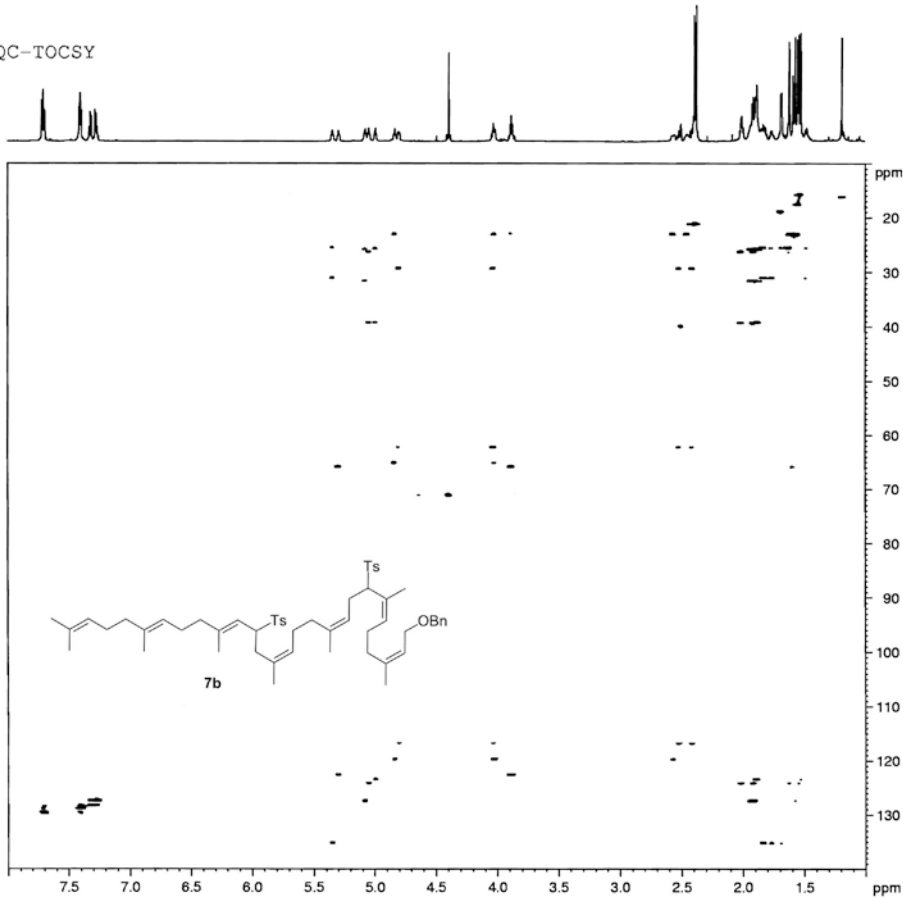
----- GRADIENT CHANNEL -----
GPNAM1   SINE-100
GPNAM2   SINE-100
GPNAM3   SINE-100
GPNAM4   SINE-100
GP11     30.00 A
GP12     30.00 A
GP13     11.00 A
GP14     -5.00 A
GP15     -5.00 A
PL6      1000.00 usec
PL9      500.00 usec

F1 - Acquisition parameters
TD       1024
SFO1     201.2603114 MHz
FIDRES   25.533399 Hz
SW       16.277 ppm
PRMODE   Echo-Antiecho

F2 - Processing parameters
SI       2048
SF       800.1800074 MHz
WDW      QSI
SSB      3
LB       0 Hz
GB       0
PC       1.40

F1 - Processing parameters
SI       2048
MC2      echo-antiecho
SF       201.2603112 MHz
WDW      States-TFPI
SSB      3
LB       0 Hz
GB       0
  
```

UPH-3T  
13C-HSQC-TOCSY

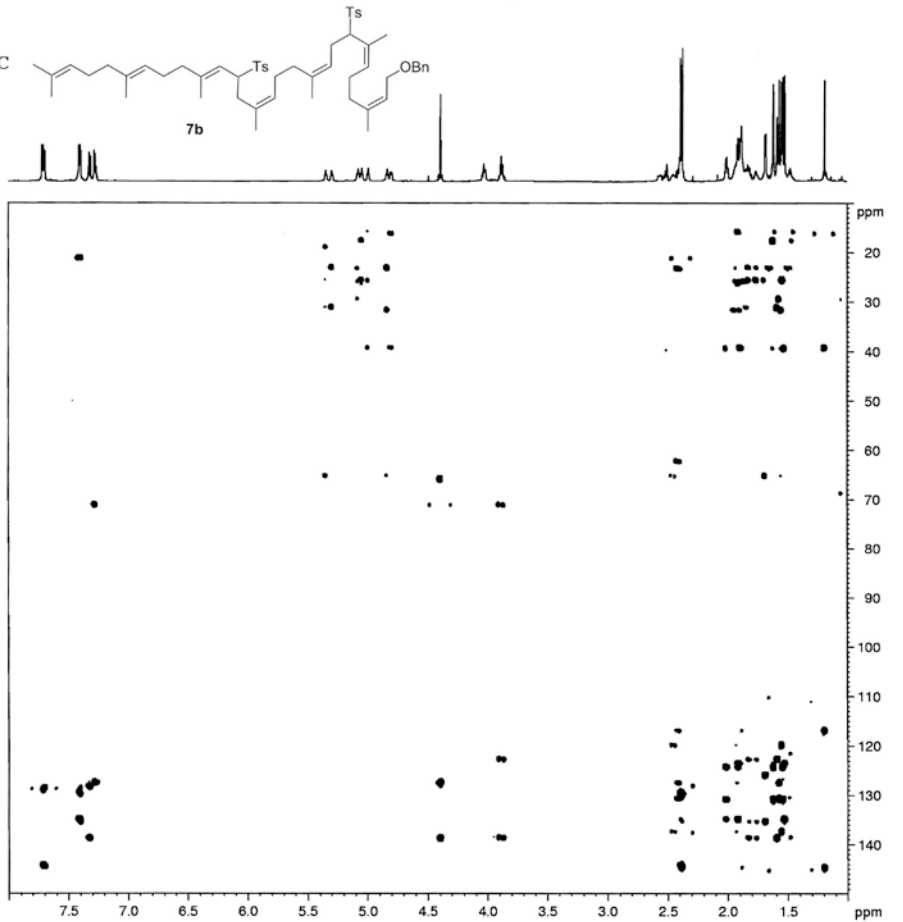


```

Current Data Parameters
NAME      UPH-3T
EXPNO    1
PROCNO   1
F2 - Acquisition Parameters
Date_    20100210
Time     22.44
INSTRUM  spect
PROBHD   5 mm CPXI 1H-
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        2048
DS        4
SWH       8223.683 Hz
FIDRES   0.23542 Hz
AQ        0.042392 sec
RG        3200
DM        60.800 usec
DE        30.00 usec
TE        301.2 K
CNS17    -0.1000000
CNS18    14.0000000
CNS19    1.0000000 sec
SI        0.0000000 sec
SI1       0.0300000 sec
SI2       0.0020000 sec
SI3       0.0012814 sec
SI4       0.00086200 sec
SI5       0.0400000 sec
DELTA    0.0012240 sec
DELTA1   0.0012880 sec
DELTA2   0.0014704 sec
DELTA3   0.0013900 sec
FACTORS  0.0001615 sec
IN0      0.0001615 sec
SI        0
STICHT   0
----- CHANNEL f1 -----
NUC1      13C
P1        9.30 usec
P2        18.60 usec
P3        1000.00 usec
P4        24.00 usec
PL1       0.20 dB
PL2       0.20 dB
PL3       0.43 dB
SFO1      800.1837440 MHz
----- CHANNEL f2 -----
CPDPRG2  f4mzg30
NUC2      13C
P1        9.30 usec
P2        18.60 usec
P3        1000.00 usec
P4        24.00 usec
PL1       0.20 dB
PL2       0.20 dB
PL3       0.43 dB
SFO2      201.2551102 MHz
SFO1S    Cyp,80.1,5,10,ns
SFO1M    Cyp80.1,5,10,1
SFO1MT   Cyp80comp,4
SFO1MS   0.500
SFO1SL   0.500
SFO1S15  0 Hz
SFO1S16  0 Hz
SFO1S17  0 Hz
----- GRADIENT CHANNEL -----
GPRAM1   SINE.100
GPRAM2   SINE.100
GPRAM3   SINE.100
GPRAM4   SINE.100
GPRAM5   SINE.100
GPRAM6   SINE.100
GP21     30.00 %
GP22     30.00 %
GP23     40.10 %
GP24     15.00 %
GP25     -10.00 %
GP26     -5.00 %
P16      1000.00 usec
F1 - Acquisition parameters
TD        201.2551102 MHz
SFO1      201.2551102 MHz
SFO2      150.141 ppm
F2 - Processing parameters
SI        2048
SF        800.1837440 MHz
WDW       QFDM
SSB       0
LB        0 Hz
GB        0
PC        1.00
F1 - Processing parameters
SI        2048
SF        echo-echo
SF2       201.2551102 MHz
WDW       States-TPPI
SSB       0
LB        0 Hz
GB        0
PC        0

```

UPH-3T  
13C-HMBC



```

Current Data Parameters
NAME      UPH-3T
EXPNO    6
PROCNO   1
F2 - Acquisition Parameters
Date_    20100211
Time     13.54
INSTRUM  spect
PROBHD   5 mm CPXI 1H-
PULPROG  zgpg30
TD        65536
SOLVENT  DMSO
NS        2048
DS        4
SWH       8223.683 Hz
FIDRES   0.215471 Hz
AQ        0.1245684 sec
RG        3200
DM        60.800 usec
DE        30.00 usec
TE        301.2 K
CNS17    8.0000000
CNS18    125.0000000
CNS19    175.0000000
SI        0.0000000 sec
SI1       1.1949998 sec
SI2       0.0000000 sec
SI3       0.0000000 sec
SI4       0.0400000 sec
DELTA    0.0028000 sec
DELTA1   0.0018514 sec
DELTA2   0.0612960 sec
IN0      0.0001310 sec
----- CHANNEL f1 -----
NUC1      13C
P1        9.30 usec
P2        18.60 usec
P3        1000.00 usec
P4        24.00 usec
PL1       0.20 dB
PL2       0.20 dB
SFO1      800.1837440 MHz
----- CHANNEL f2 -----
NUC2      13C
P1        9.30 usec
P2        18.60 usec
P3        1000.00 usec
P4        24.00 usec
PL1       0.20 dB
PL2       0.20 dB
SFO2      201.2245355 MHz
----- GRADIENT CHANNEL -----
GPRAM1   SINE.100
GPRAM2   SINE.100
GPRAM3   SINE.100
GPRAM4   SINE.100
GPRAM5   SINE.100
GPRAM6   SINE.100
GP21     30.00 %
GP22     40.10 %
GP23     15.00 %
GP24     -10.00 %
GP25     -5.00 %
P16      1000.00 usec
F1 - Acquisition parameters
TD        512
SFO1      201.2245 MHz
SFO2      74.546753 Hz
SF        189.678 ppm
F2 - Processing parameters
SI        2048
SF        800.183066 MHz
WDW       QFDM
SSB       0
LB        0 Hz
GB        0
PC        1.40
F1 - Processing parameters
SI        512
NUC2      13C
SF        201.2055102 MHz
WDW       States-TPPI
SSB       0
LB        0 Hz
GB        0
PC        0

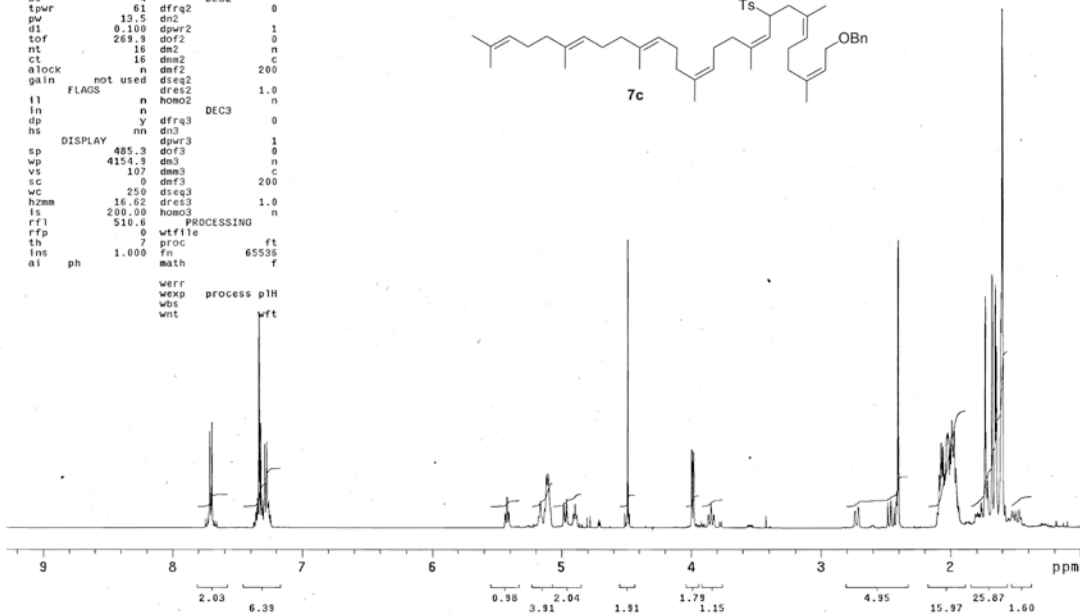
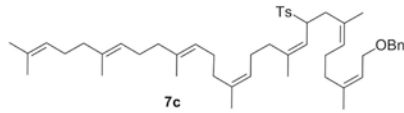
```

UPH-11  
exp1 s2pul

```

SAMPLE          DEC. & VT
date Jan 22 2010 dfrq 499.864
solvent CDC13  dn      H1
file exp dpr      30
ACQUISITION
effq 499.864  dn      nmh
in H1  dm      c
at 5.016  dmf      200
np 65536  dseq     c
bv 6533.3  dres     1.0
fb 4000    hoba    n
bs 4
lpwr 61  dfrq2    DEC2  0
pw 13.5  dn2      1
d1 0.100  dpr2    1
tof 269.9  dof2    0
nt 16    dm2     n
ct 18    dms2    c
alock n    dmf2    200
gain not used  dseq2  1.0
FLAGS
il n hoba2    DEC3  0
ln n
dp y dfrq3    0
hs nn dn3
DISPLAY dpr3    1
sp 485.3  dof3    0
wp 4154.8  dm3     n
vs 107    dms3    c
sc 0      dmf3    200
wc 250    dseq3   1.0
hzmm 16.62  dres3  1.0
ls 200.00  hoba3   n
rfl 510.6  PROCESSING
rfp 0      wtfile   ft
th 7      proc      65536
ins 1.000  fr        f
al ph     math

```



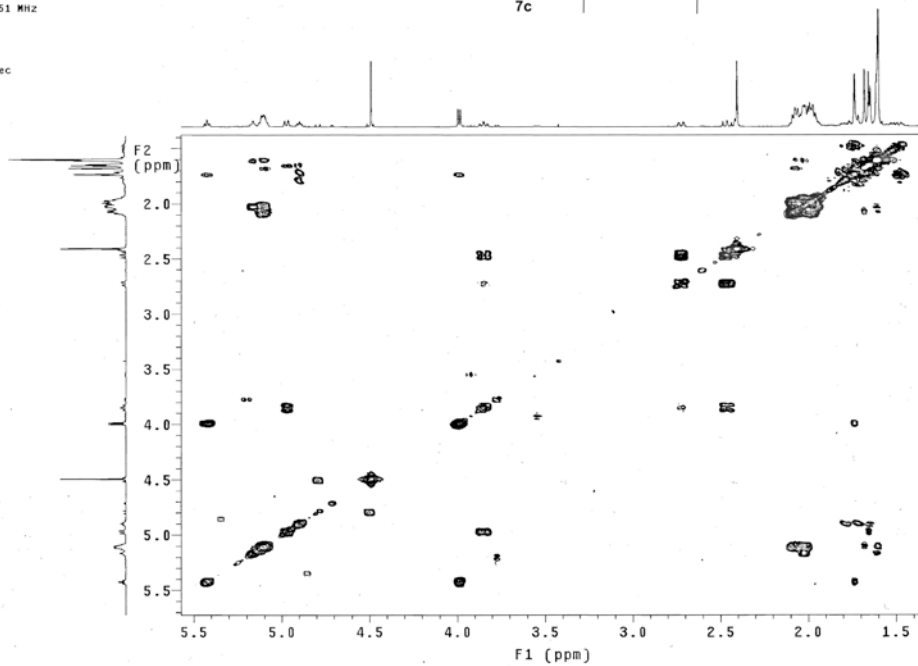
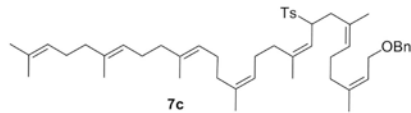
UPH-11

Pulse Sequence: relayh  
Solvent: CDC13  
Ambient Temperature  
INOVA-500 "nmr2a.chem.nd.edu"

```

Relax. delay 1.300 sec
COSY 30-30
Acq. time 0.150 sec
Width 3406.9 Hz
2D Width 3406.9 Hz
8 repetitions
256 increments
OBSERVE H1, 499.8611751 MHz
DATA PROCESSING
Sine bell 0.075 sec
F1 DATA PROCESSING
Sine bell 0.038 sec
FT size 1024 x 1024
Total time 51 min, 19 sec

```

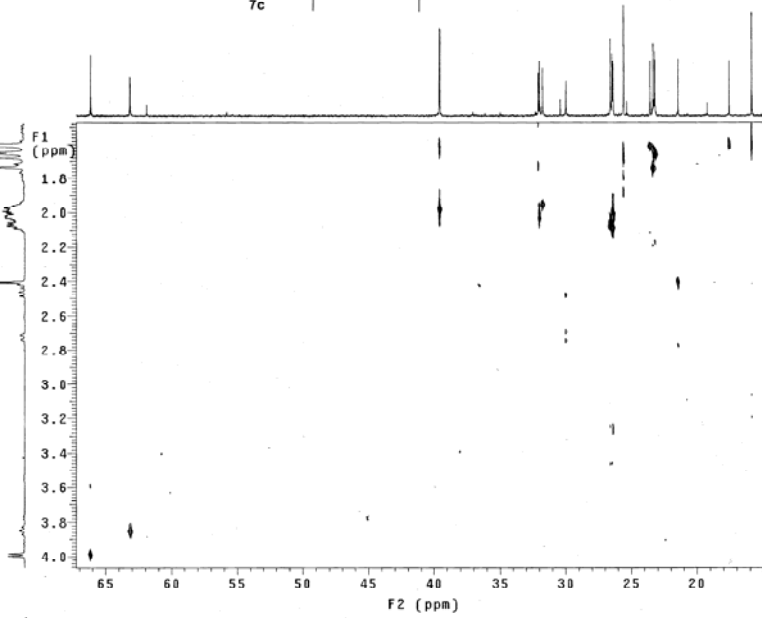
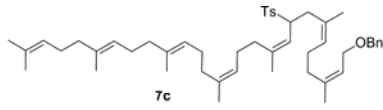






UPH-11  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INOVA-500 "nmr2a.chem.nd.edu"

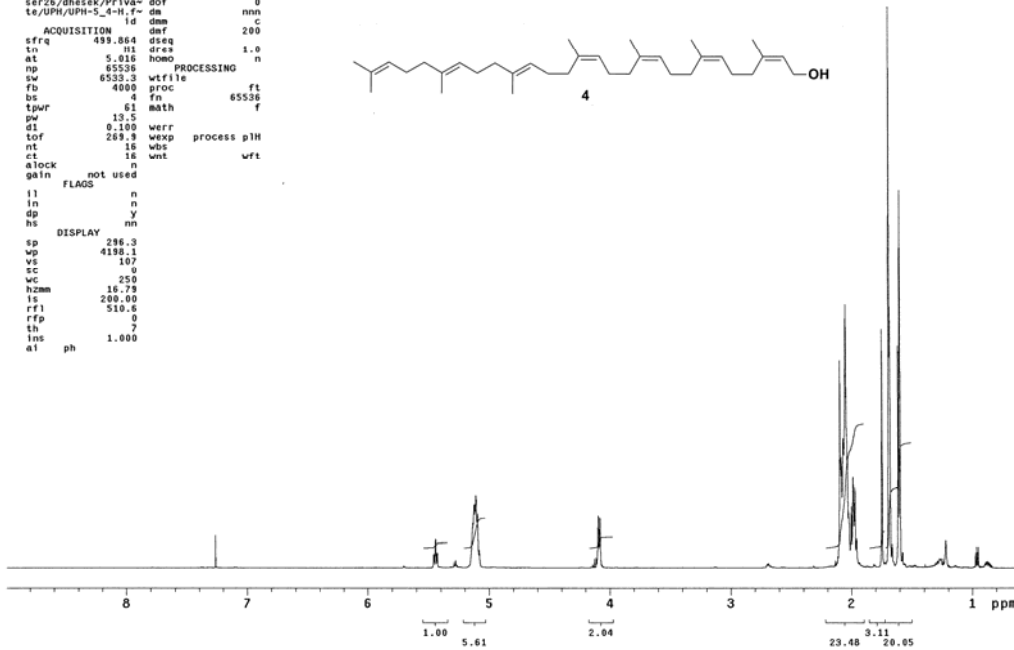
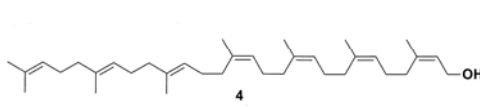
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18409.5 Hz  
ZD Width 3382.2 Hz  
4 repetitions  
256 increments  
OBSERVE C13, 125.6302147 MHz  
DECOUPLE H1, 499.8634976 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 9.3 Hz  
FT size 4096 x 512  
Total time 28 min, 40 sec



UPH-5\_4

exp1 s2pu1

SAMPLE DEC. & VT 499.864  
date Jan 22 2010 dfrq dn H1  
solvent CDCl3 dn 30  
file /afs/nd.edu/u/ dpwr 0  
ser26/dhasek/Privat dof 0  
te/UPH/UPH-5\_4-H.f- da nnn  
c  
ACQUISITION id dm 200  
sfrq 499.864 dseq 1.0  
sn 01 dres n  
at 5.016 homo n  
np 65536 PROCESSING  
sw 6533.3 wtrfile ft  
fb 4000 proc 65536  
bs 4 fn  
tpwr 61 math f  
pw 13.5  
d1 0.100 werr  
tof 269.9 wexp process pH  
nt 16 wbs  
ct 16 wnt wft  
alock n  
gain not used  
FLAGS n  
ll n  
in n  
dp y  
hs m  
DISPLAY 286.3  
wp 4198.1  
vc 107  
sc 0  
wc 250  
hzmm 18.78  
ls 200.00  
rfl 510.6  
rfg 0  
th 7  
ins 1.000  
al ph

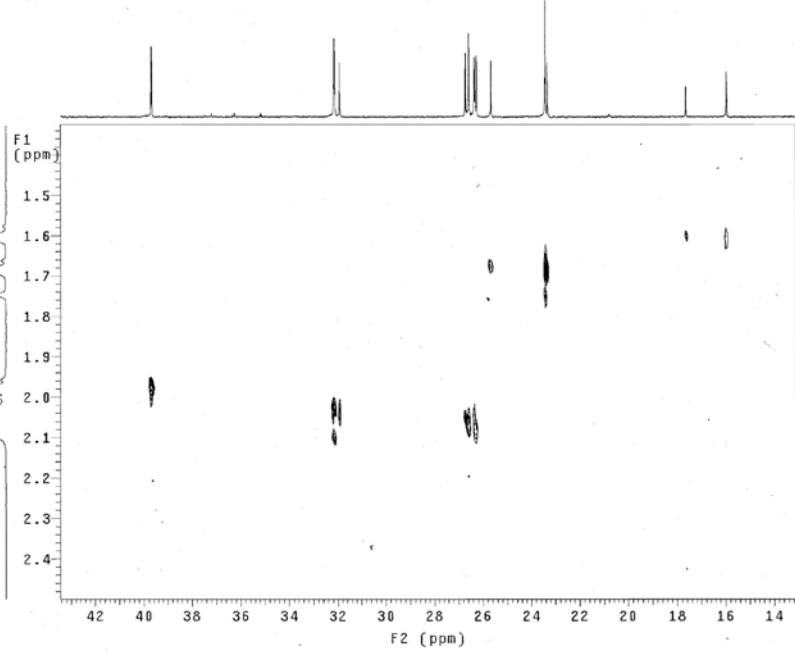
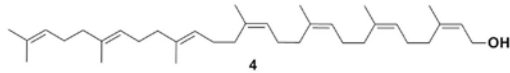




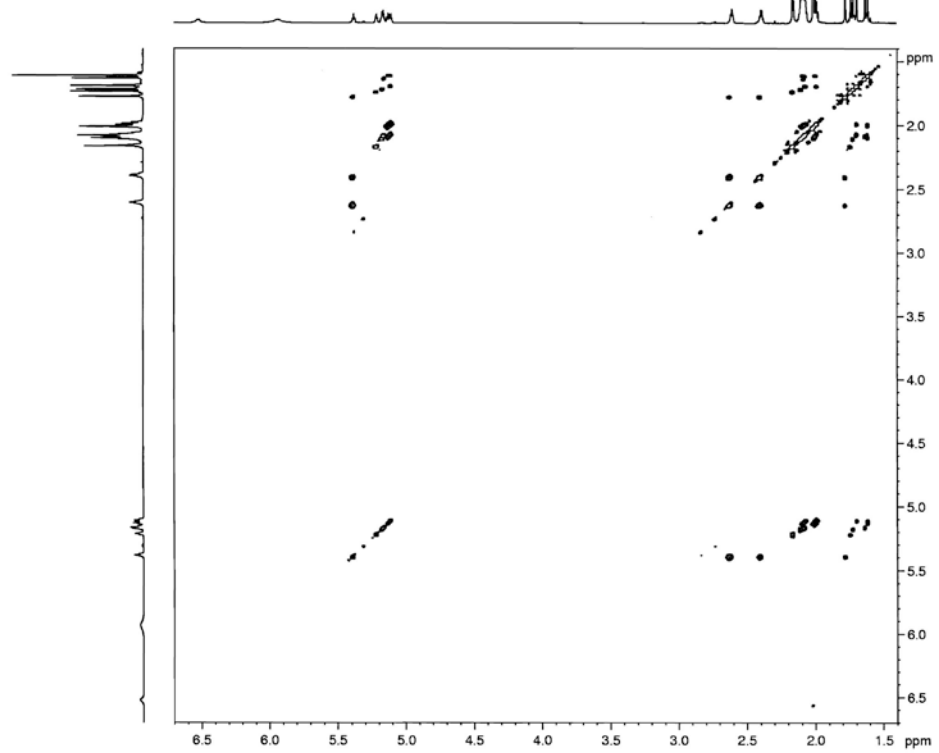
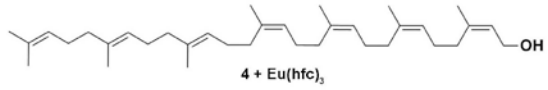


UPH-5\_3  
Pulse Sequence: hetcor  
Solvent: CDCl3  
Ambient temperature  
User: 1-14-87  
INNOVA-500 "nmr2a.chem.nu.edu"

Relax delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
ZD Width 5193.5 Hz  
32 repetitions  
512 increments  
OBSERVE C13, 125.6901970 MHz  
DECOUPLE H1, 499.8633346 MHz  
Power 40 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
FT size 4096 x 512  
Total time 7 hr, 48 min, 35 sec



UPH-5\_3  
TOCSY



Current Data Parameters  
NAME UPH-5-3-1  
EXPNO 3  
PROCNO 1

F2 - Acquisition Parameters  
Date\_ 20100517  
Time 21.19  
INSTRUM spect  
PROBHD 5 mm CPTCI IH-  
PULPROG dipolwetp1  
TD 1024  
SOLVENT CDCl3  
NS 16  
DS 16  
SWH 8012.820 Hz  
FIDRES 7.825020 Hz  
AQ 0.0639476 sec  
RG 22.8  
DW 62.400 usec  
DE 19.00 usec  
TE 301.2 K  
d0 0.0000300 sec  
D1 1.2999995 sec  
d11 0.0300000 sec  
D16 0.0002000 sec  
D30 0.0001000 sec  
D21 0.0001000 sec  
D9 0.0600000 sec  
DELTA 0.0012000 sec  
DELTA1 0.0012000 sec  
FACTOR1 11  
INO 0.00012480 sec  
I1 22  
SICNT 0

----- CHANNEL f1 -----  
NUC1 1H  
P1 8.30 usec  
p2 16.00 usec  
P6 24.00 usec  
PL1 0.20 dB  
PL10 9.42 dB  
SFO1 800.1828373 MHz

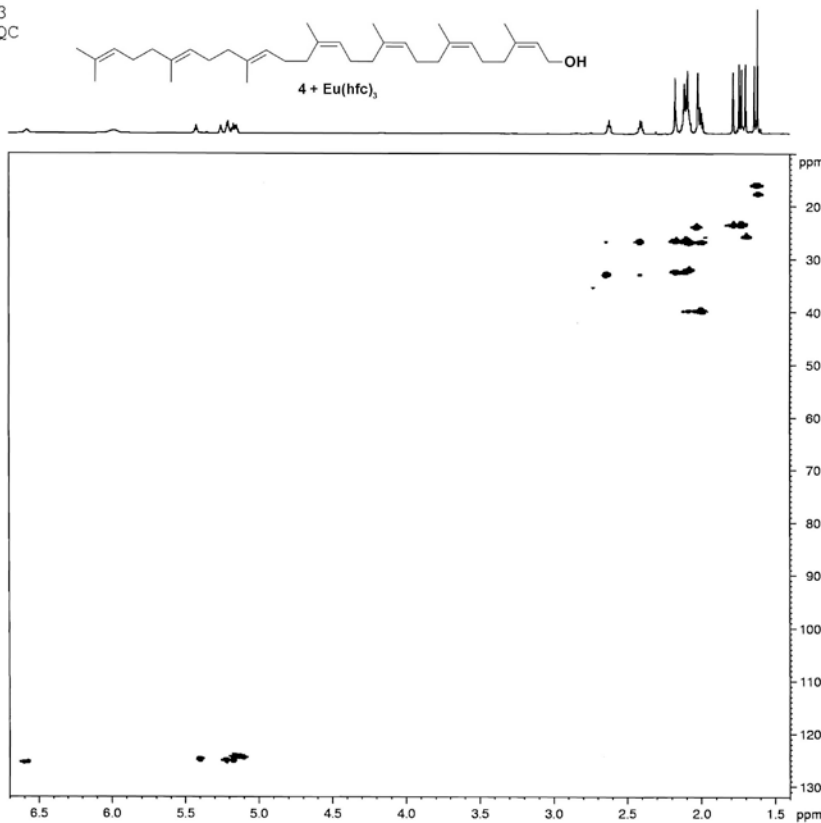
----- GRADIENT CHANNEL -----  
GPNAM1 SINE.100  
GPNAM2 SINE.100  
GP1 10.00  
GP2 10.00  
F15 1000.00 usec

F1 - Acquisition parameters  
TD 1024  
SFO1 800.1828 MHz  
FIDRES 7.825020 Hz  
SW 10.014 ppm  
FMODE Echo-Antiecho

F2 - Processing parameters  
SI 3048  
SF 800.180111 MHz  
WUW QSINE  
SSB 4  
LB 0 Hz  
GB 0

F1 - Processing parameters  
SI 3048  
MC2 echo-antiecho  
SF 800.180111 MHz  
WUW States-TFPI  
SSB 4  
LB 0 Hz  
GB 0

UPH-5\_3  
13C-HSQC

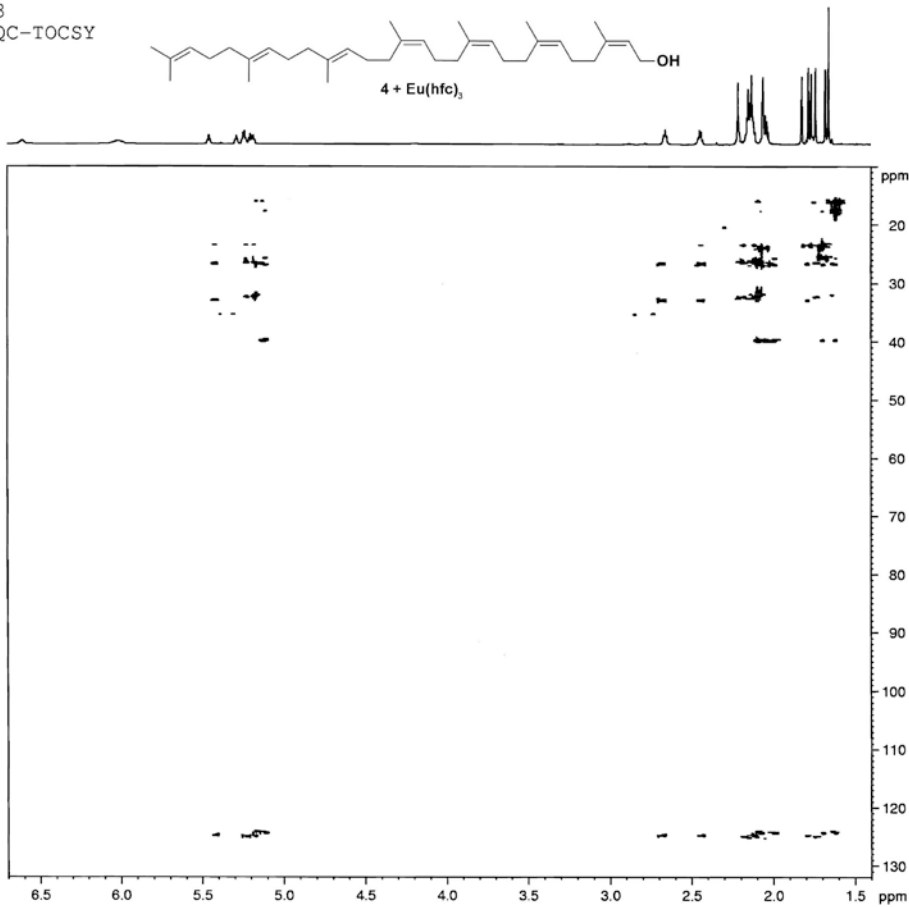


```

Current Data Parameters
NAME           UPH-5_3
EXPNO          1
PROCNO         1
F2 - Acquisition Parameters
Date_          20100118
Time           4.10
INSTRUM        spect
PROBHD         3 mm QNP1H
PULPROG        zgpg30
SOLVENT        CDCl3
NS              32
DS              4
SWH            8012.820 Hz
FIDRES        7.82520 Hz
AQ            0.043976 sec
RG            64.00
AQW           42.400 usec
DE            30.00 usec
TE            301.2 K
CROSSP1        -0.50000000
CROSSP2        145.00000000
CROSSP3        0
CROSSP4        1.3999999999
CROSSP5        0.00000000
CROSSP6        0.00000000
CROSSP7        0.0017414
CROSSP8        0.0017414
CROSSP9        0.0017414
CROSSP10       0.0017414
CROSSP11       0.0017414
CROSSP12       0.0017414
CROSSP13       0.0017414
CROSSP14       0.0017414
CROSSP15       0.00000000
CROSSP16       0.00000000
CROSSP17       0
CROSSP18       0
CROSSP19       0
CROSSP20       0
CHANNEL f1 -----
NUC1            13C
P1             18.00 usec
PL1            0.00 dB
PC1            1.00
SFO1           101.625375 MHz
CHANNEL f2 -----
CPDPRG2        p54ep180
NUC2            13C
P2             18.00 usec
PL2            0.00 dB
PC2            1.00
SFO2           101.625375 MHz
----- GRADIENT CHANNEL -----
GPMAX1         200.00 usec
GPMAX2         200.00 usec
GPMAX3         200.00 usec
GPMAX4         200.00 usec
GPMAX5         200.00 usec
GPMAX6         200.00 usec
GPMAX7         200.00 usec
GPMAX8         200.00 usec
GPMAX9         200.00 usec
GPMAX10        200.00 usec
GPMAX11        200.00 usec
GPMAX12        200.00 usec
GPMAX13        200.00 usec
GPMAX14        200.00 usec
GPMAX15        200.00 usec
GPMAX16        200.00 usec
GPMAX17        200.00 usec
GPMAX18        200.00 usec
GPMAX19        200.00 usec
GPMAX20        200.00 usec
F1 - Acquisition parameters
TD              65536
SFO1           201.25239 MHz
SFO2           101.625375 MHz
SFO3           101.625375 MHz
P1             18.00 usec
PL1            0.00 dB
PC1            1.00
F2 - Processing parameters
SI              65536
SF              800.1802630 MHz
WDW              EM
SSB              0
LB               0 Hz
GB               0 Hz
PC              1.00
F1 - Processing parameters
SI              65536
SF              201.2554289 MHz
WDW              EM
SSB              0
LB               0 Hz
GB               0 Hz
PC              1.00

```

UPH-5\_3  
13C-HSQC-TOCSY

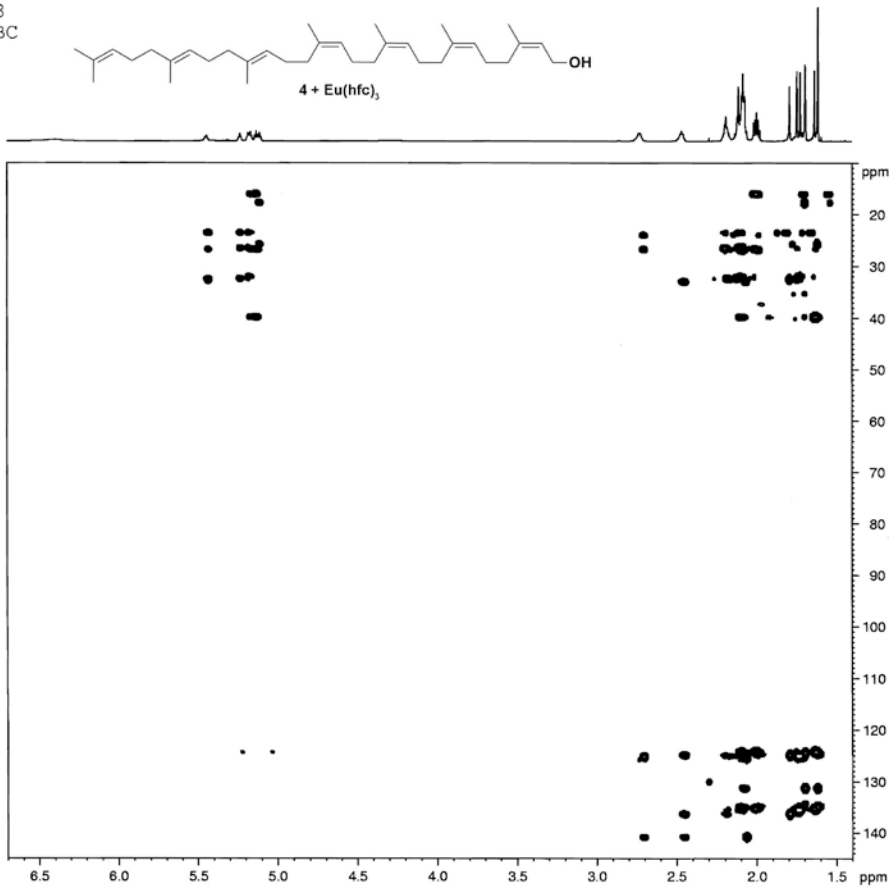
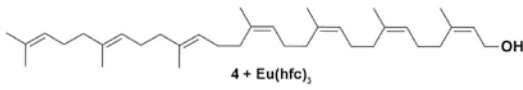


```

Current Data Parameters
NAME           UPH-5_3
EXPNO          1
PROCNO         1
F2 - Acquisition Parameters
Date_          20100118
Time           4.10
INSTRUM        spect
PROBHD         3 mm QNP1H
PULPROG        zgpg30
SOLVENT        CDCl3
NS              32
DS              4
SWH            8012.820 Hz
FIDRES        7.82520 Hz
AQ            0.043976 sec
RG            64.00
AQW           42.400 usec
DE            30.00 usec
TE            301.2 K
CROSSP1        -0.50000000
CROSSP2        145.00000000
CROSSP3        0
CROSSP4        1.3999999999
CROSSP5        0.00000000
CROSSP6        0.00000000
CROSSP7        0.0017414
CROSSP8        0.0017414
CROSSP9        0.0017414
CROSSP10       0.0017414
CROSSP11       0.0017414
CROSSP12       0.0017414
CROSSP13       0.0017414
CROSSP14       0.0017414
CROSSP15       0.00000000
CROSSP16       0.00000000
CROSSP17       0
CROSSP18       0
CROSSP19       0
CROSSP20       0
CHANNEL f1 -----
NUC1            13C
P1             18.00 usec
PL1            0.00 dB
PC1            1.00
SFO1           101.625375 MHz
CHANNEL f2 -----
CPDPRG2        p54ep180
NUC2            13C
P2             18.00 usec
PL2            0.00 dB
PC2            1.00
SFO2           101.625375 MHz
----- GRADIENT CHANNEL -----
GPMAX1         200.00 usec
GPMAX2         200.00 usec
GPMAX3         200.00 usec
GPMAX4         200.00 usec
GPMAX5         200.00 usec
GPMAX6         200.00 usec
GPMAX7         200.00 usec
GPMAX8         200.00 usec
GPMAX9         200.00 usec
GPMAX10        200.00 usec
GPMAX11        200.00 usec
GPMAX12        200.00 usec
GPMAX13        200.00 usec
GPMAX14        200.00 usec
GPMAX15        200.00 usec
GPMAX16        200.00 usec
GPMAX17        200.00 usec
GPMAX18        200.00 usec
GPMAX19        200.00 usec
GPMAX20        200.00 usec
F1 - Acquisition parameters
TD              65536
SFO1           201.25239 MHz
SFO2           101.625375 MHz
SFO3           101.625375 MHz
P1             18.00 usec
PL1            0.00 dB
PC1            1.00
F2 - Processing parameters
SI              65536
SF              800.1802630 MHz
WDW              EM
SSB              0
LB               0 Hz
GB               0 Hz
PC              1.00
F1 - Processing parameters
SI              65536
SF              201.2554289 MHz
WDW              EM
SSB              0
LB               0 Hz
GB               0 Hz
PC              1.00

```

UPH-5\_3  
13C-HMBC



```

Current Data Parameters
NAME      UPH-5-3-1
EXPNO    4
PROCNO   1

F2 - Acquisition Parameters
Date_    20100519
Time     13:23
INSTRUM  spect
PROBHD   5 mm CPIC1 1H-
PULPROG  zgpg30
ID       2048
SOLVENT  CDCl3
NS       64
DS       2
SWH      8223.685 Hz
FIDRES   4.015471 Hz
AQ       0.1245681 sec
RG       4600
DM       60.800 usec
DE       30.00 usec
TE       301.9 K
CRS13    8.0000000
CRS16    125.0000000
CRS17    175.0000000
d0       0.0000300 sec
d1       1.3999998 sec
d16      0.0020000 sec
d6       0.0620000 sec
DELTA1   0.0028000 sec
DELTA2   0.00169714 sec
DELTA3   0.0612900 sec
IND      0.00001310 sec

===== CHANNEL f1 =====
NUC1     13C
P1       8.10 usec
P2       16.20 usec
P11      0.00 dB
SF01     800.182873 MHz

===== CHANNEL f2 =====
NUC2     1H
P3       14.80 usec
P12      -2.00 dB
SF02     201.224533 MHz

===== GRADIENT CHANNEL =====
GPRAM1   SINE.100
GPRAM2   SINE.100
GPRAM3   SINE.100
GPRAM4   SINE.100
GPRAM5   SINE.100
GPRAM6   SINE.100
GP1      50.00 %
GP2      30.00 %
GP3      40.10 %
GP4      15.00 %
GP5      -10.00 %
GP6      -3.00 %
P16      1000.00 usec

F1 - Acquisition parameters
TD       312
SF01     201.2245 MHz
FIDRES   74.546753 Hz
SM       189.478 ppm
FMODE    QF

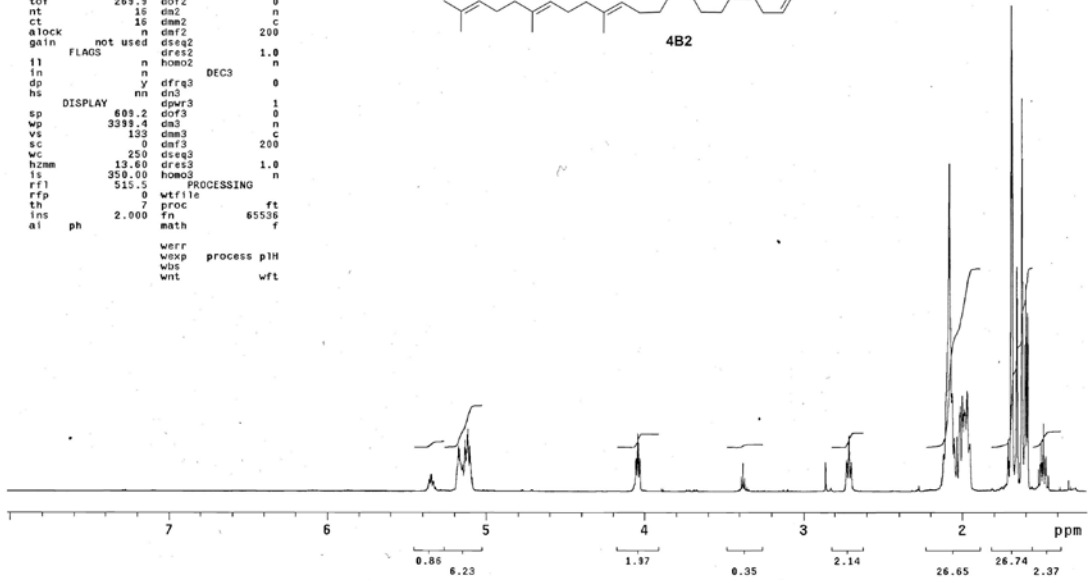
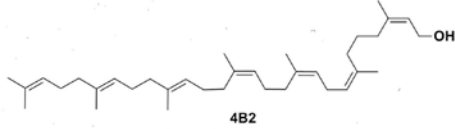
F2 - Processing parameters
SI       2048
SF       800.180130 MHz
WDW      0
SSB      0
LB       0 Hz
GB       0
PC       1.40

F1 - Processing parameters
SI       4096
SF       201.2054289 MHz
WDW      States-TPPI
SSB      0
LB       0 Hz
GB       0
  
```

UPH\_5\_2

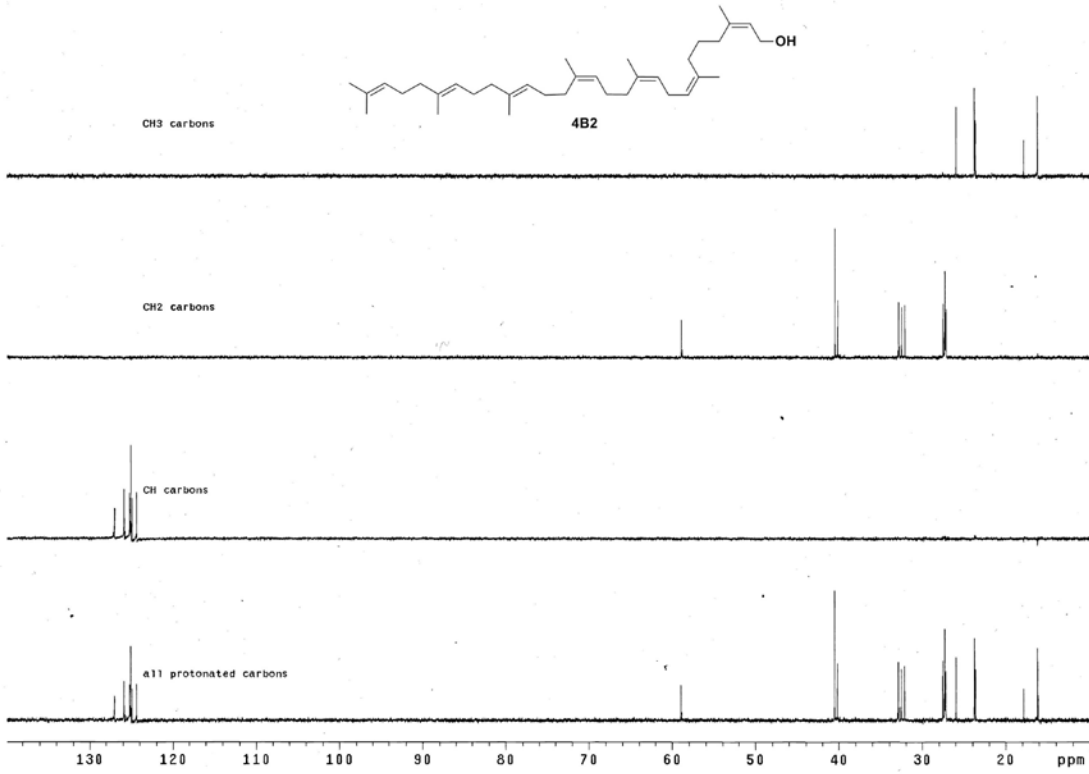
```

exp4 s2pu1
SAMPLE
date Jan 4 2010 dfrq DEC. & VT 499.866
solvent acetone dn H1
file exp spwr 30
ACQUISITION
sfrq 499.867 dof 0
tn H1 dm nnn
at 5.016 dmf 200
np 85536 dfrq c
sw 6533.3 dss 1.0
fb 4000 homo n
bs 4 DEC2 0
tpwr 61 dfrq2 0
pw 13.5 dn2 1
dt 0.100 spwr2 0
tof 269.3 dof2 0
nt 16 dm2 n
ct 16 dmw2 c
alock n dmf2 200
gain not used dssq2 1.0
flags n homo2 n
i1 n DEC3 0
dp y dfrq3 0
hs nn dn3 1
DISPLAY
sp 609.2 dof3 0
wp 3393.4 dm3 n
vs 133 dmw3 c
sc 0 dmf3 200
wc 250 dssq3 1.0
hzmm 13.60 homo3 n
is 350.00 wtrfile ft
rfl 515.5 PROCESSING
rfp 0 proc ft
th 2.000 fn 65536
als ph math f
werr process pH
wds wft
  
```



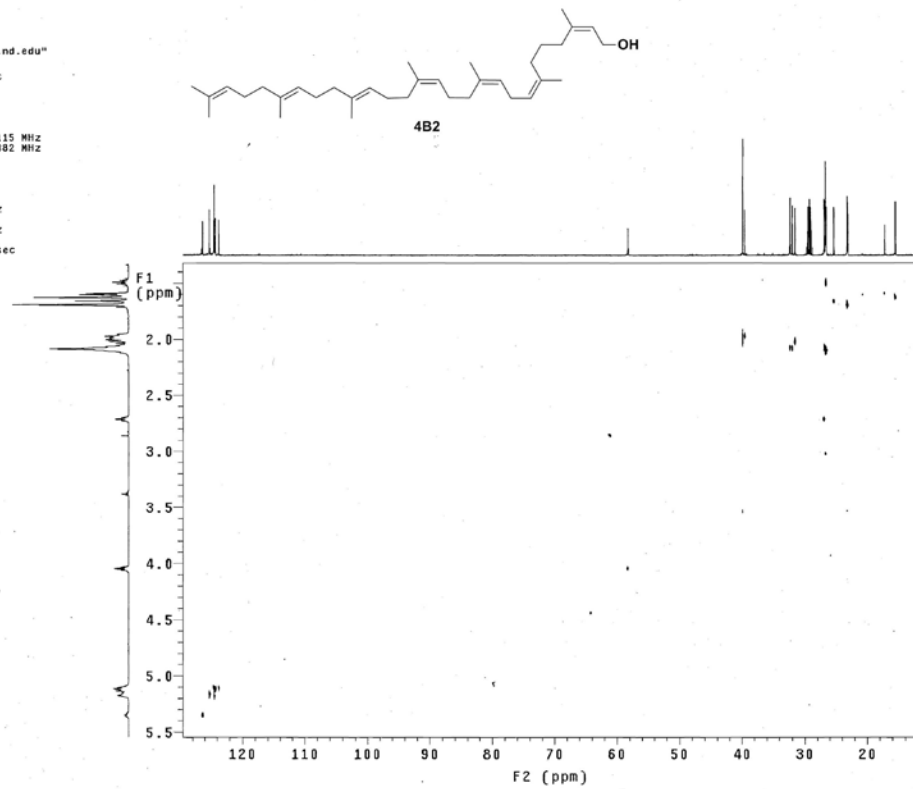


UPH\_5\_2  
Pulse Sequence: dept



UPH\_5\_2  
Pulse Sequence: hetcor  
Solvent: Acetone  
Ambient temperature  
User: j-14-97  
INOVA-500 "nmr2a.chem.nd.edu"

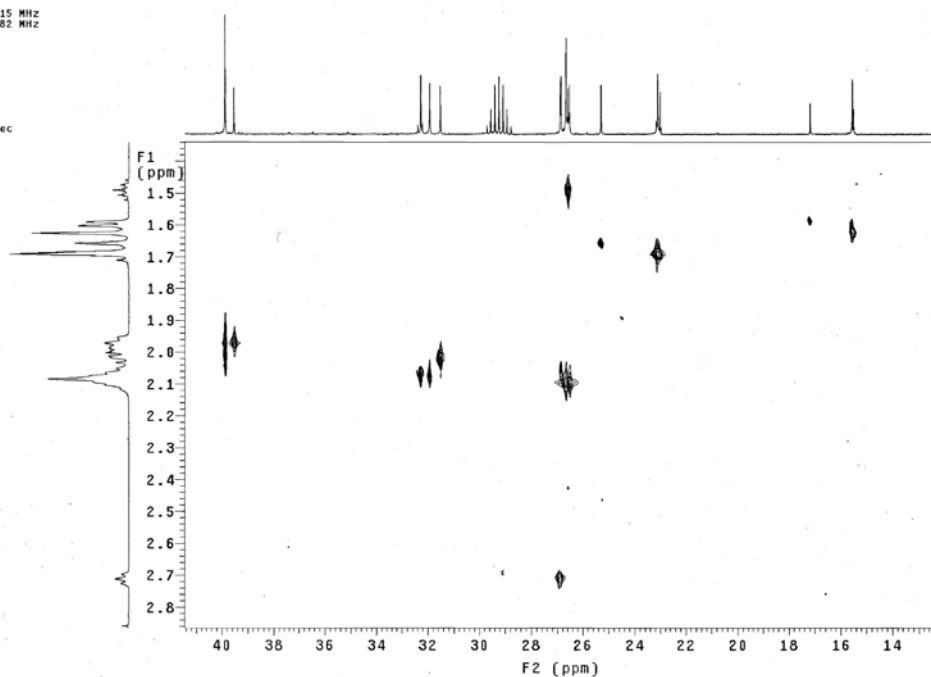
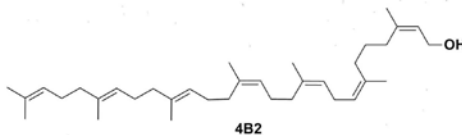
Relax. delay 1.500 sec  
Acq. time 0.111 sec  
Width 18403.5 Hz  
2D Width 2240.6 Hz  
4 repetitions  
256 increments  
OBSERVE c13, 125.6908115 MHz  
DECOUPLE H1, 499.8655382 MHz  
Power 00 dB  
on during acquisition  
off during delay  
WALTZ-16 modulated  
DATA PROCESSING  
Line broadening 1.0 Hz  
F1 DATA PROCESSING  
Line broadening 0.3 Hz  
F1 size 4096 x 512  
Total time 28 min, 59 sec



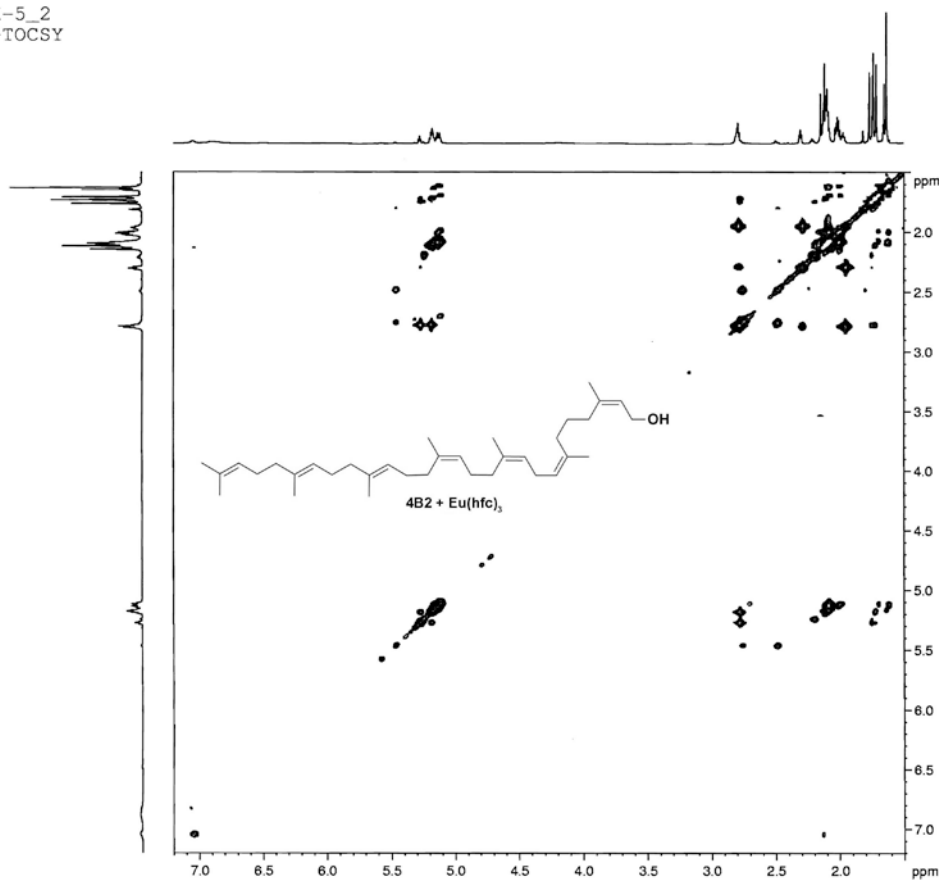


UPH\_5\_2  
 Pulse Sequence: hetcor  
 Solvent: Acetone  
 Ambient Temperature  
 User: 1-14-07  
 INOVA-500 "nmr2a.chem.nd.edu"

Relax. delay 1.500 sec  
 Acq. time 0.111 sec  
 Width 18403.5 Hz  
 2D Width 2240.6 Hz  
 4 repetitions  
 256 increments  
 OBSERVE C13, 125.6908115 MHz  
 DECOUPLE H1, 499.8655382 MHz  
 Power 40 db  
 on during acquisition  
 off during delay  
 WALTZ-16 modulated  
 DATA PROCESSING  
 Line broadening 1.0 Hz  
 F1 DATA PROCESSING  
 Line broadening 0.3 Hz  
 FT size 4096 x 512  
 Total time 28 min, 59 sec



UPH-5\_2  
 1H-TOCSY



Current Data Parameters  
 NAME UPH-5-2a-1  
 EXPNO 4  
 PROCNO 1

F2 - Acquisition Parameters  
 Date\_ 20100710  
 Time 16.56  
 INSTRUM spect  
 PROBHD 5 mm CPTCI 1H-  
 PULPROG dipzltztpsi  
 TD 1024  
 SOLVENT ccd13  
 NS 16  
 DS 16  
 SWH 8012.820 Hz  
 FIDRES 7.825020 Hz  
 AQ 0.0539476 sec  
 RG 22.6  
 DW 62.400 usec  
 DE 30.00 usec  
 TE 301.9 K  
 D0 0.00900300 sec  
 D1 1.29499991 sec  
 d11 0.03000000 sec  
 D16 0.00020000 sec  
 D20 0.00001000 sec  
 D21 0.00001000 sec  
 D3 0.03000000 sec  
 DELTA 0.00120300 sec  
 DELTA1 0.00120800 sec  
 FACTOR1  
 IN0 0.00012480 sec  
 I1 10  
 STCNT 0

----- CHANNEL f1 -----  
 NUCL1 1H  
 P1 8.20 usec  
 p2 16.40 usec  
 P6 24.00 usec  
 PF1 6.20 dB  
 PS10 9.53 dB  
 SFO1 800.1828373 MHz

----- GRADIENT CHANNEL -----  
 GPNAM1 SINE.100  
 GPNAM2 SINE.100  
 GP21 30.00 \*  
 GP22 30.00 \*  
 F16 1000.00 usec

F1 - Acquisition parameters  
 TD 1024  
 SFO1 800.1828 MHz  
 FIDRES 7.825020 Hz  
 SW 10.014 ppm  
 FMODE Echo-Antiecho

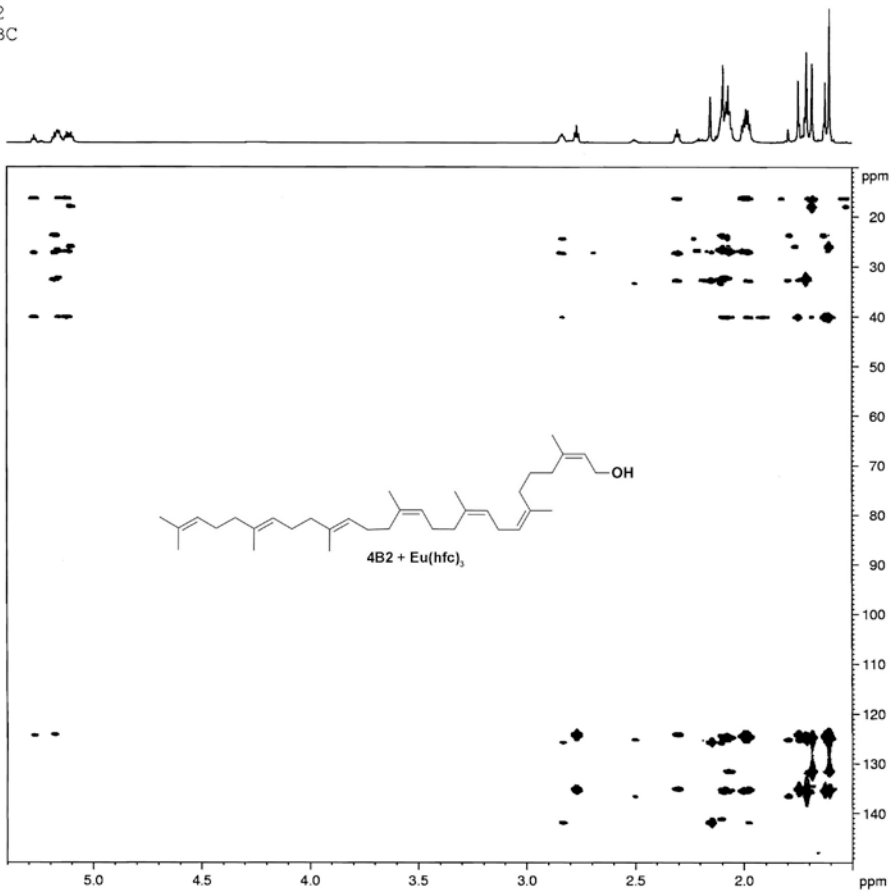
F2 - Processing parameters  
 SI 2048  
 SF 800.1800106 MHz  
 WSW QSIKE  
 SSB 2  
 LB 0 Hz  
 GB 0  
 PC 1.40

F1 - Processing parameters  
 SI 2048  
 MC2 echo-antiecho  
 SF 800.1800106 MHz  
 WSW States-TF1  
 SSB 2  
 LB 0 Hz  
 GB 0





UPH-5\_2  
13C-HMBC



```

Current Data Parameters
NAME      UPH-5_2a-1
EXPNO    12
PROCNO   1

F2 - Acquisition Parameters
Date_    20100712
Time     14.52
INSTRUM  spect
PROBHD   5 mm CPIC 1H-
PULPROG  zgpg30
TD        2048
SOLVENT  CDCl3
NS        64
DS        128
SWH       8223.685 Hz
FIDRES   0.015471 Hz
AQ        0.1245684 sec
RG        5790
DM        65.800 usec
DE        30.00 usec
TE        301.9 K
CNS113   8.0000000
CRS16    125.0000000
CNS17    175.0000000
d0        0.0000000 sec
d1        1.3999998 sec
d16       0.0020000 sec
de        0.0625000 sec
delta1    0.0210000 sec
delta2    0.00145714 sec
delta3    0.06129600 sec
IND       0.0001310 sec

===== CHANNEL f1 =====
NUC1      13C
P1        8.00 usec
P2        16.40 usec
PL1       0.00 dB
PL2       0.00 dB
SFO1     800.1828373 MHz

===== CHANNEL f2 =====
NUC2      1H
P2        14.80 usec
PL2       -2.00 dB
SFO2    201.2245255 MHz

===== GRADIENT CHANNEL =====
GPNAM1   SINE.100
GPNAM2   SINE.100
GPNAM3   SINE.100
GPNAM4   SINE.100
GPNAM5   SINE.100
GPNAM6   SINE.100
GP21     50.00 %
GP22     30.00 %
GP23     40.10 %
GP24     15.00 %
GP25     -10.00 %
GP26     -5.00 %
P16      1000.00 usec

F1 - Acquisition parameters
TD        512
SFO1     201.2245 MHz
FIDRES   74.546753 Hz
SW        189.678 ppm
FREQ0DE  QF

F2 - Processing parameters
SI        2048
SF        800.1800169 MHz
WDW       QSI
SSB       4
LB        0 Hz
GB        0
PC        1.40

F1 - Processing parameters
SI        4096
NUC1      13C
SF        201.2053682 MHz
WDW       States-TPPI
SSB       4
LB        0 Hz
GB        0
  
```