Fig. S1 pre-miR-LRRK2-5 and pre-mi R-LRRK2-5NB direct silencing of a complementary dual-luciferase target sequence A) Dual-luciferase reporter ass ay showing knockdown of LRRK2 target seque nce following co-transfection with indicated pre-m iRNA variants. Valu es rep resent m ean ratios of *Renilla*:Firefly luciferase +/- S.D. from n=6. Pre-miRNA variants are normalised to cells transfected with a non-specific U6 pre-miRNA hairpin. *= P<0.05 relative to normalising control.

Fig. S2 mirt-LRRK2-5NB directs silenc ing in a sequence-specific manner A) Codon-modified scrambled target sequence to mirt-LRRK2-5NB. B) Dual-luciferase reporter assays showing knockdown of a c odon-modified scrambled LRRK2 target following co-transfection with indic ated mirtron (left panel) and pre-m iRNA control (right panel) variants. V alues represent mean ratios of *Renilla*:Firefly luciferase +/-S.D. from n=6. Mirtron variants are norm alized to cells transf ected with the NAD variant. Pre-miRNA variants a re normalised to c ells transfected with a non-specific U6 pre-miRNA hairpin. *= P<0.05 relative to respective normalising control.

Fig. S3 Direct comparison of syntheti c mirtrons to U6-transcribed pre-miRNAs (shRNAs) A) Dual-luciferase reporter assays showing knockdown of a LRRK2 target following co-transfection with indic ated mirtron (left panel) and pre-m iRNA control (right panel) variants at equivalent hairpin concentrations of 2.5 *f*m. Values represent mean ratios of *Renilla*:Firefly lu ciferase +/- S. D. f rom n=3. Mirtron variants are normalized to cells transf ected with the NAD variant. Pre-m iRNA variants are normalised to cells transfected with a non-specific U6 pre-miRNA hairpin. *= P<0.05 relative to respective normalising control.

Fig. S4 Algorithm design of a synthetic mirtron ta rgeting α -synuclein. A) Predicted hairpin alignments of α -synuclein targeting synthetic mirtron, α -syn-mirt-1, designed using an in-house algorithm . B) Representative fluorescent m icroscopy images of different intron variants 48hr s after transfection in HEK-293 cells. C) Quantification of eGFP fluorescence following expression indicated intron variants in HEK-293 cells. Values represent m ean fluorescence +/- S.D. from n=3. *= P<0.05 relative to non-transf ected cells. D) Du al-luciferase reporter assays showing knockdown of an α -synuclein target following co-trans fection with indicated m irtron (left panel) and pre-m iRNA control (right panel) variants. Values represent m ean ratios of *Renilla*:Firefly luciferase +/- S.D. from n=3. Mirtron variants are normalized to cells transfected with the NAD variant. Pre-miRNA variants are normalised to cells transfected with a non -specific U6 pre- miRNA hairpin. *= P<0.05 relative to respective n ormalising contro l. E) Quantif ication of mCherry tagged α -synuclein fluorescence following co-expression with the NAD intron or mirt- α -Syn-1 in HEK-293 cells. Values represent m ean fluor escence +/- S.D. from n=3. Mirt- α -Syn-1 fluorescence is norm alized to cells transfected with the NAD intron . *= P<0.05 relative to respective normalising control.

Fig. S5 A) Quantification of eGFP fluorescence fo llowing expression of indicated intron variants in SH-SY5Y cells and Hela cells. Values represent mean fluorescence \pm - S.D. from n=3. *= P<0.05 relative to non-transfected cells.



A.

B.

| LRRK2 sequence: | ac <mark>c</mark> | ttt | att | cct | gac | tct | tct | <u>at</u> g |
|-----------------|-------------------|-----|-------------|-------------|-------------|-------------|-------------|-------------|
| Amino-acid seq: | Thr | Phe | Ile | Pro | Asp | Ser | Ser | Met |
| Codon replaced: | ac <mark>G</mark> | ttC | at C | cc G | ga T | tc G | tc G | atg |







AUCAU G С UGGAGC AGG AGC ACAUAGAGAA UAGAGC A mirt-a-syn-1 |||ACCUCG UGUAUCUCUU UCC UCG GUCUCG G G CU CCCCUCU ACUCU A _



NAD

B.



mirt-a-syn-1



12

D.





E.

C.





SH-SY5Y cells

