

Supplemental Material for:

**Crystal structures of the Cid1 poly (U) polymerase reveal the
mechanism for UTP selectivity**

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Supplementary figure legends

Supplemental Figure 1: Sequence alignment of *S. pombe* Cid1 with other Cid1 family members. Coloring of the alignment is based on conservation with green being highly conserved to yellow being less conserved. The Cid1 secondary structure is shown above the alignment. The catalytic aspartates are indicated by black boxes. Residues from the Cid1-UTP structure that are important for triphosphate recognition are highlighted with green boxes and residues involved in ribose base recognition are shown by yellow stars. The nucleotide recognition motif is surrounded by a black box and indicated by NRM on the figure. Abbreviations are same as in Figure 4B.

Supplemental Figure 2: UTP recognition by the *Trypanosomal brucei* TUTases. Key residues involved with UTP-specific recognition from each of the TbTUTase structures are shown for (A) *TbRET2*-UTP (PDB ID 2B51), (B) *TbMEAT1*-UTP (PDB ID 3HJ1), and (C) *TbTUT4*-UTP (PDB ID 2IKF). Coloring is the same as in figure 2.

Supplemental Figure 3: Electron density maps for Cid1-NTP structures. 2Fo-Fc electron density maps (blue mesh) contoured at 1σ are shown for each of the bound NTPs in the (A) Cid1-3'dATP, (B) Cid1-GTP, and (C) Cid1-CTP structures.

Supplemental Figure 4: Determination of the kinetic parameters of ATP and UTP addition of Cid1 and the H336N variant. (A) Michaelis-Menten kinetics of ATP incorporation by Cid1 (filled circles) and the H336N mutant (open circles). (B)

Michaelis-Menten kinetics of UTP-incorporation by Cid1 (filled circles) and H336N mutant (open circles).

Supplementary tables

Supplemental Table 1: DNA primers for cloning and mutagenesis used in this work.

cid1_fl_ndeI	5'-CGATCACATATGAGCTACCAAAAGGTCCCTAATTTCG-3'
cid1_dN31_ndeI	5'-CGATCACATATGAGCTACCAAAAGGTCCCTAATTTCG-3'
cid1_xhoI	5'-CGATCTCGAGTCACTCAGAATTGTCACCATCGG-3'
cid1_378stop_f	5'-GATTCATTATTTGAGGAGGCCTAGATTCCGCCTCGTCGCCAGAAA-3'
cid1_378stop_r	5'-TTTCTGGCGACGAGGCGGAATCTAGGCCTCCTCAAATAATGAATC-3'
cid1_h336n_f	5'-GAAGATCCTTTTCGAGATTTCAAATAATGTGGGTAGGACAGTTA-3'
cid1_h336n_r	5'-TAACTGTCCTACCCACATTATTTGAAATCTCGAAAGGATCTTC-3'
cid1_d160a_f	5'-TGGGGCTTCGTTTCAATGTGCTATTGGATTTAACAATCGTC-3'
cid1_d160a_r	5'-GACGATTGTAAATCCAATAGCACATTGAAACGAAGCCCCA-3'

Supplemental Table 2: Data collection and refinement statistics for Cid1 (H336N) crystals

MgATP	
Data collection	
Wavelength (Å)	1.00
Space group	<i>P</i> 2 ₁ 2 ₁ 2 ₁
Cell dimensions	
<i>a, b, c</i> (Å)	53.72, 62.47, 111.83
Resolution (Å) ^a	2.7 (2.77-2.70)
<i>R</i> _{meas} ^{a, b}	10.6 (74.3)
<i>I</i> / σI ^c	11.41 (2.24)
Completeness (%) ^a	98.5 (97.4)
Redundancy ^a	3.5 (3.4)
Refinement	
Resolution (Å)	41.7-2.7
No. reflections	10,674
<i>R</i> _{work} ^c / <i>R</i> _{free} ^d	20.8/28.0
No. atoms	
Protein	2620
NTP	31
Ions (Mg ⁺² , Ca ⁺²)	1
No. water molecules	39
Avg <i>B</i> -factors (Å ²)	
Protein	46.95
NTP	85.58
Ion (Mg ⁺² , Ca ⁺²)	71.9
Water	44.08
R.m.s. deviations ^e	
Bond lengths (Å)	0.0096
Bond angles (°)	1.183
Ramachandran	97.18
Favored (%)	
Ramachandran	0
Disallowed (%)	
Molprobity score	2.04 [97 th]
[percentile]	

^aValues in parentheses are for highest-resolution shell.

^b $R_{\text{meas}} = \sum_h [n_h / (n_h - 1)]^{1/2} \sum_i |I_{h1} - I_{hi}| / \sum_h \sum_i I_{hi}$, where *I*_h is the mean intensity of symmetry-equivalent reflections and *n*_h is the redundancy.

^c $R_{\text{work}} = \sum ||F_o| - |F_c|| / \sum |F_o|$

^d*R*_{free} is the same as *R*_{work}, but calculated based on 5% of the data excluded from refinement

^eR.m.s. deviations from target geometries

Figure S1

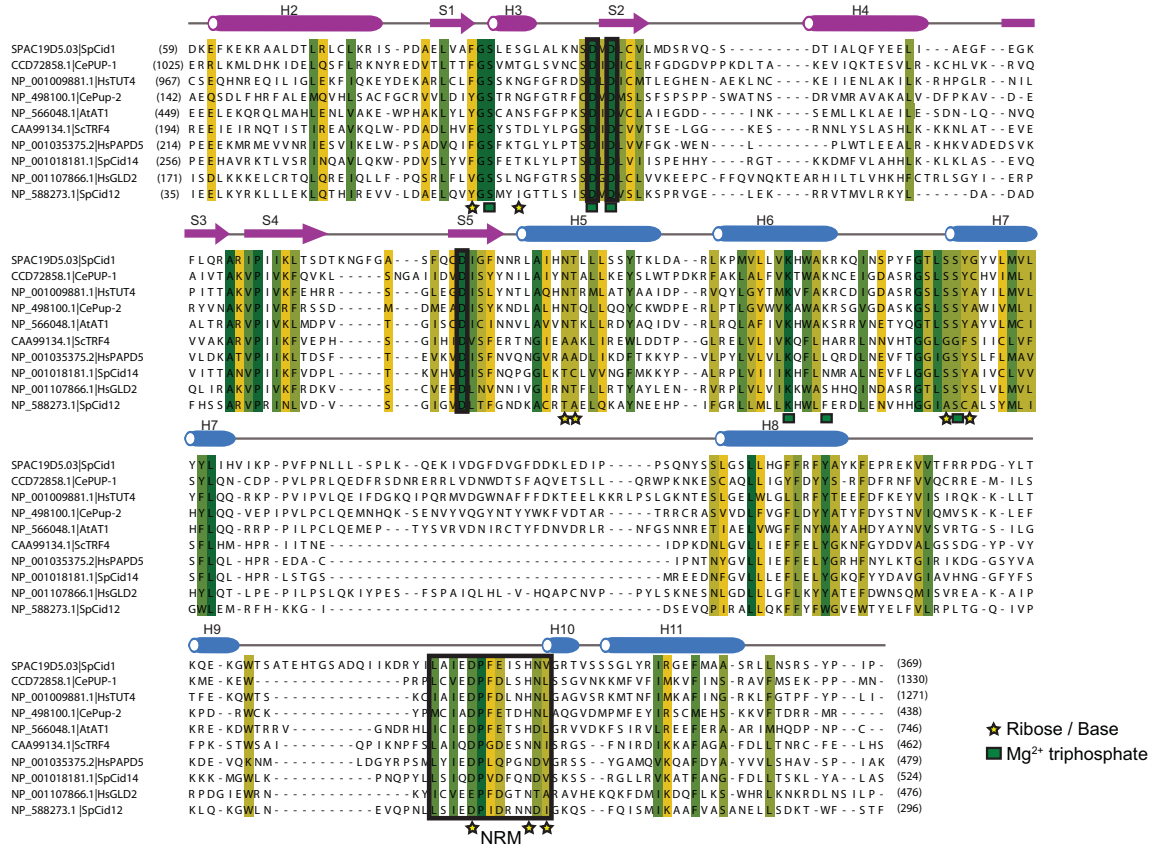
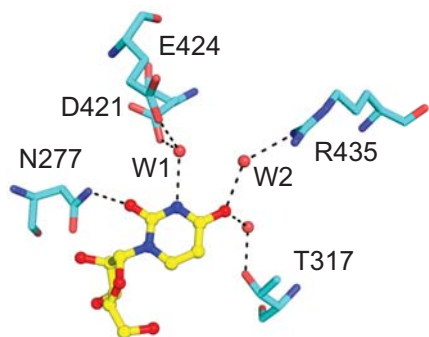
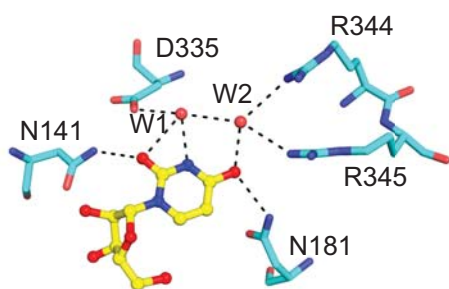


Figure S2

A



B



C

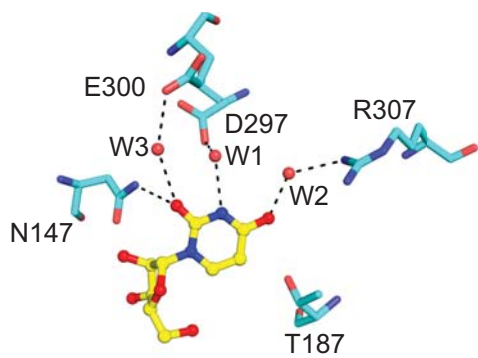
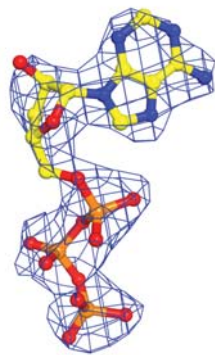
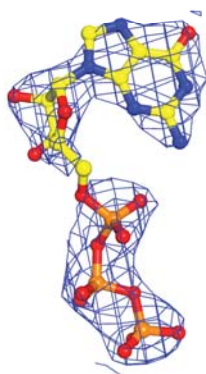


Figure S3

A



B



C

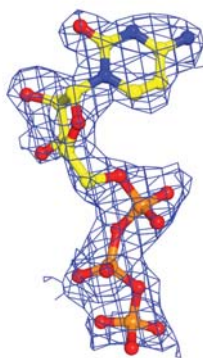


Figure S4

