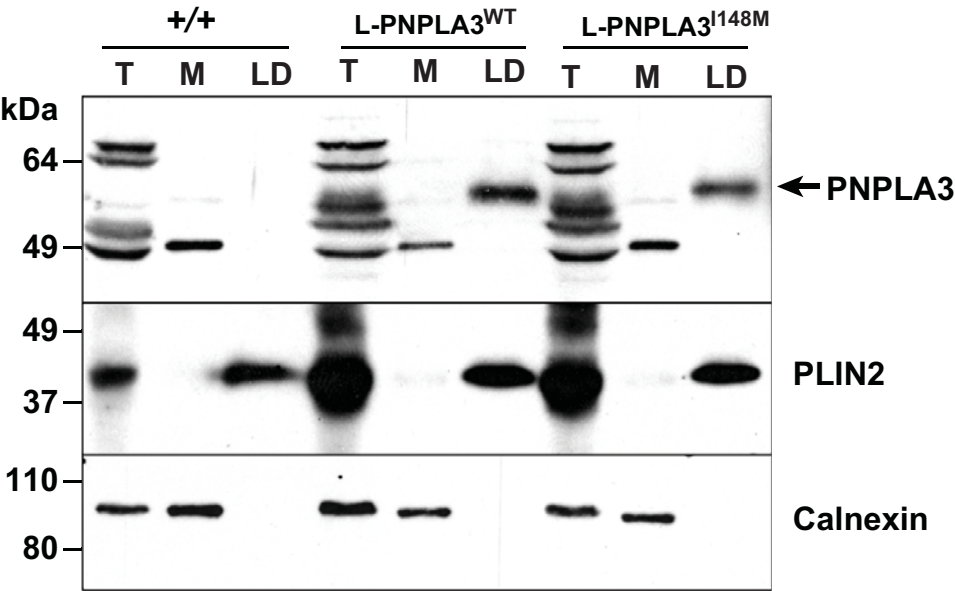
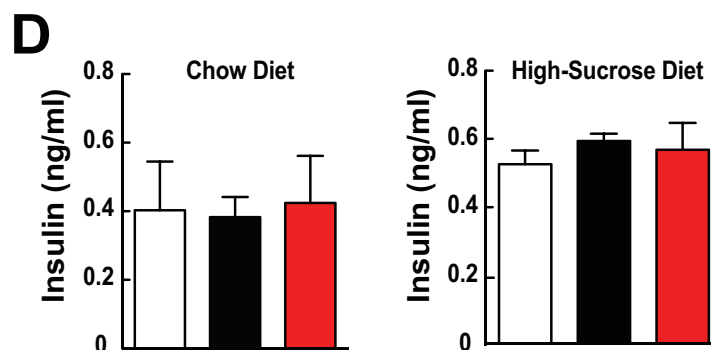
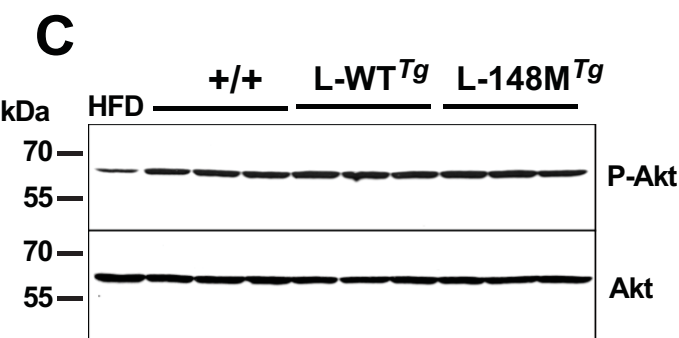
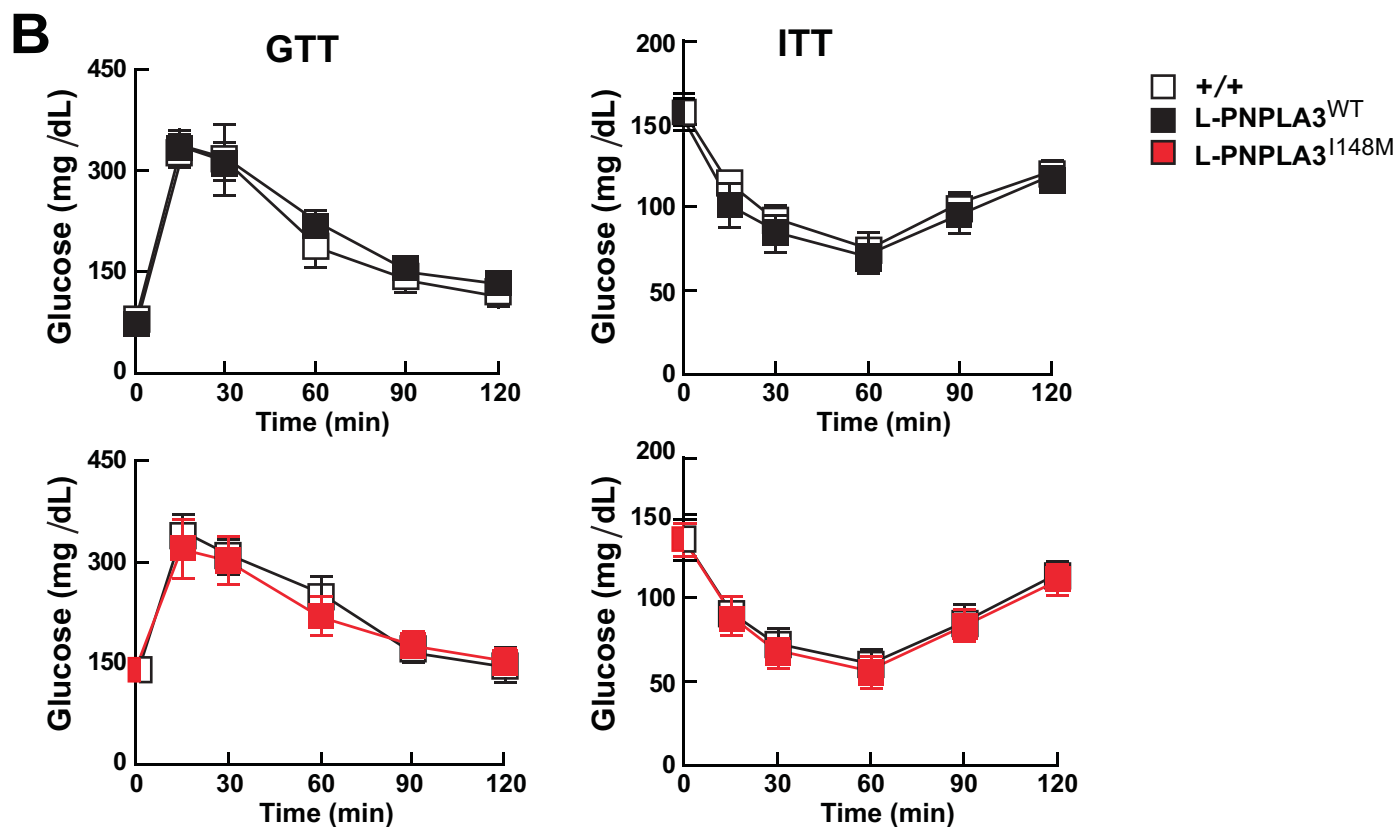
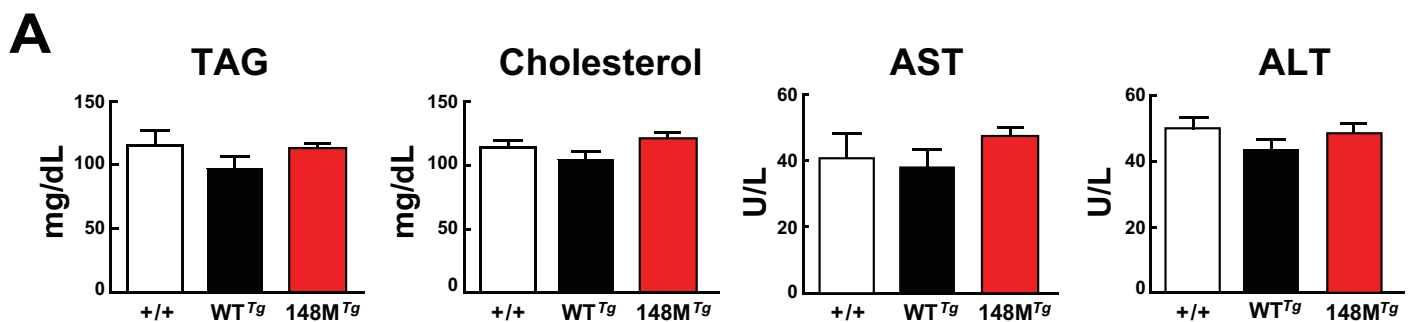


Supplemental Figure 1

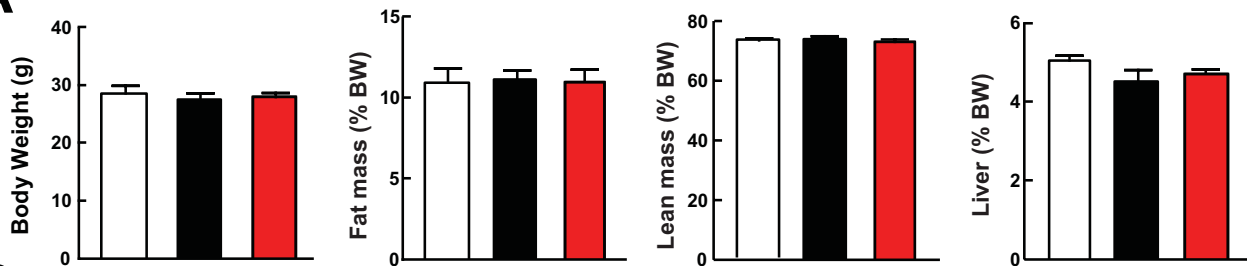
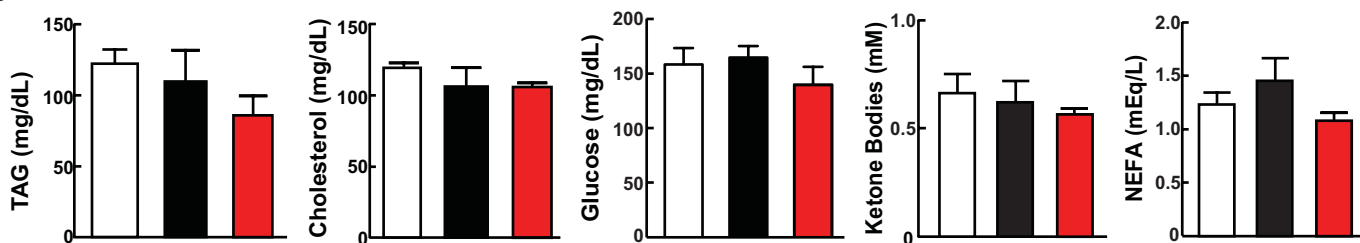
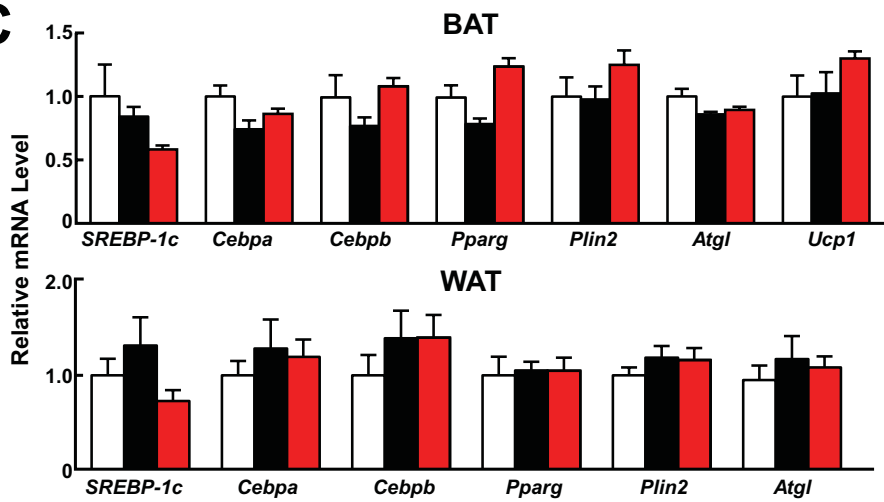
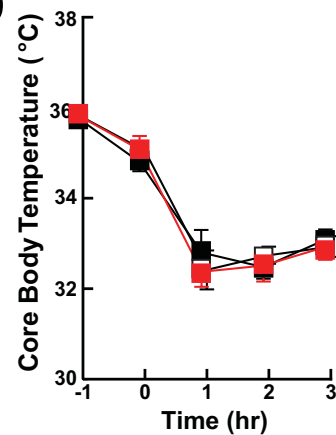


Supplemental Figure 2



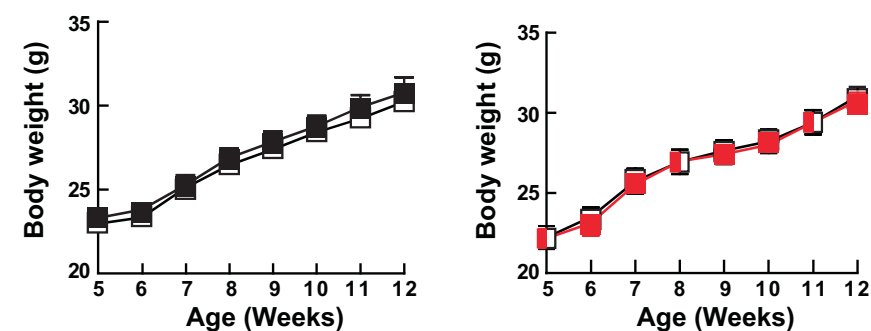
Supplemental Figure 3

□ +/+
■ A-PNPLA3^{WT}
■ A-PNPLA3^{I148M}

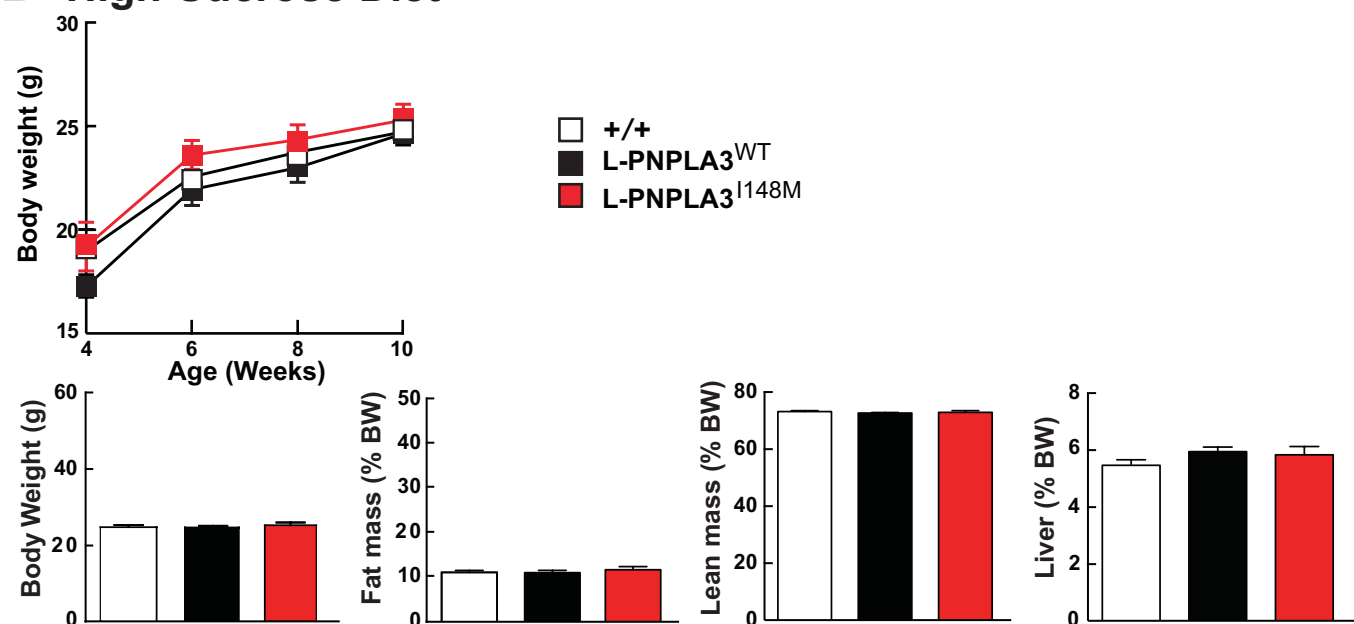
A**B****C****D**

Supplemental Figure 4

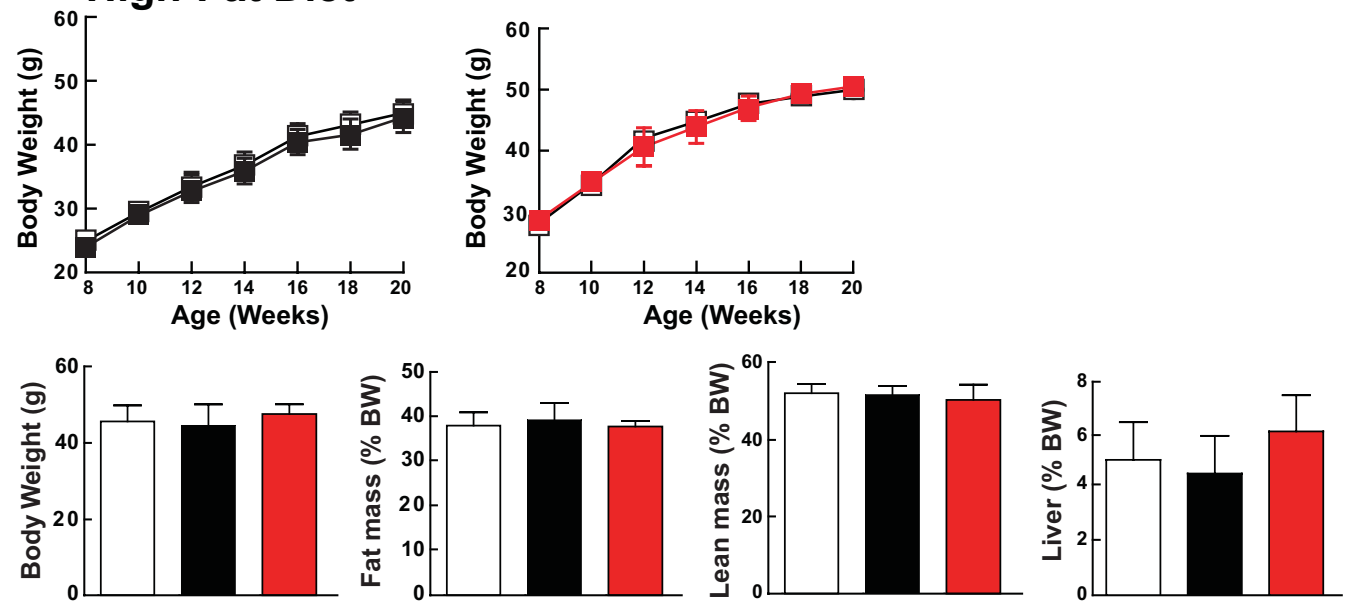
A Chow Diet



B High-Sucrose Diet

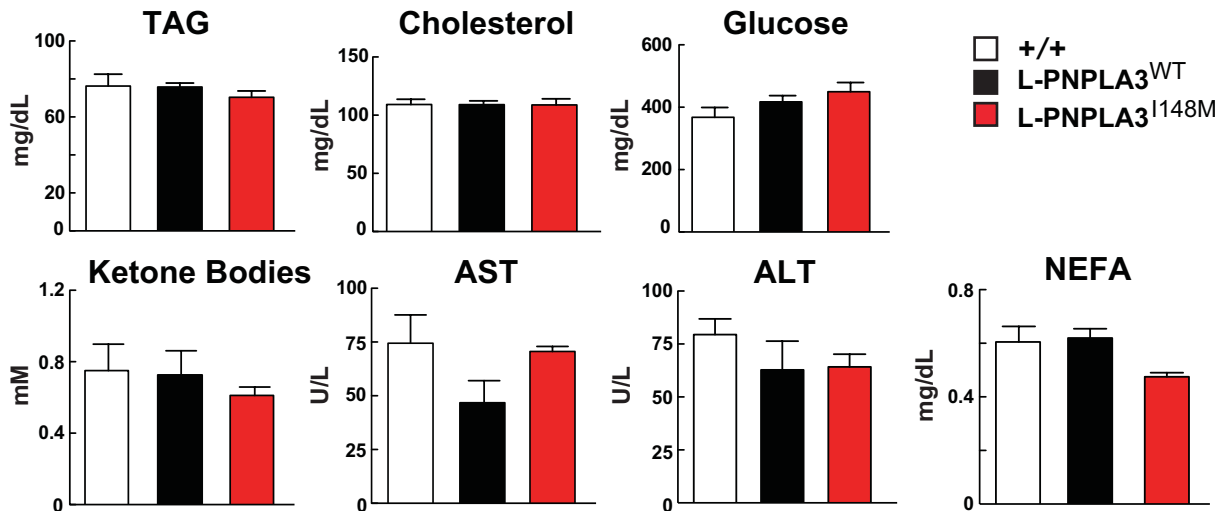


C High-Fat Diet

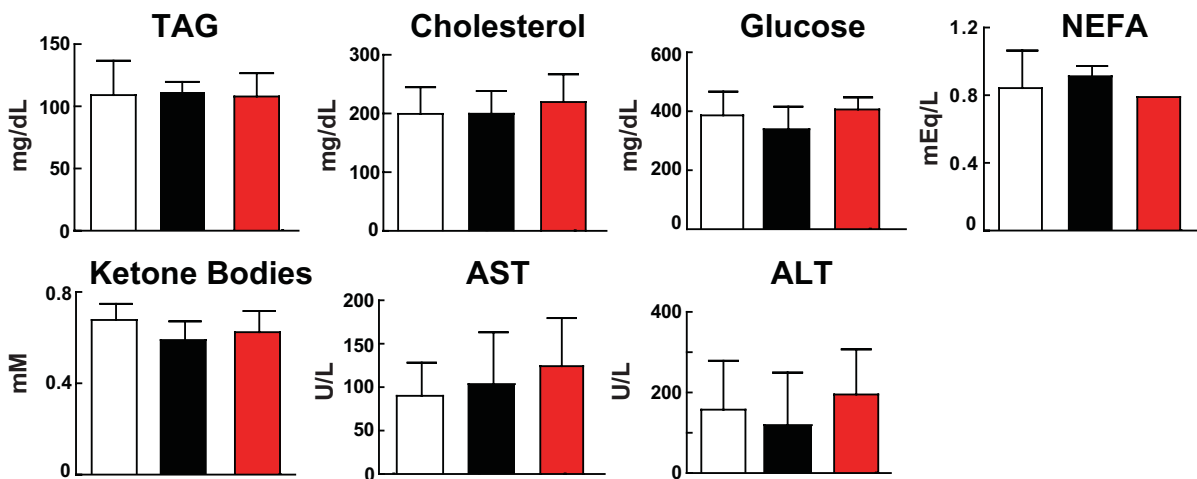


Supplemental Figure 5

A High-Sucrose Diet



B High-Fat Diet



Supplemental Figure 6

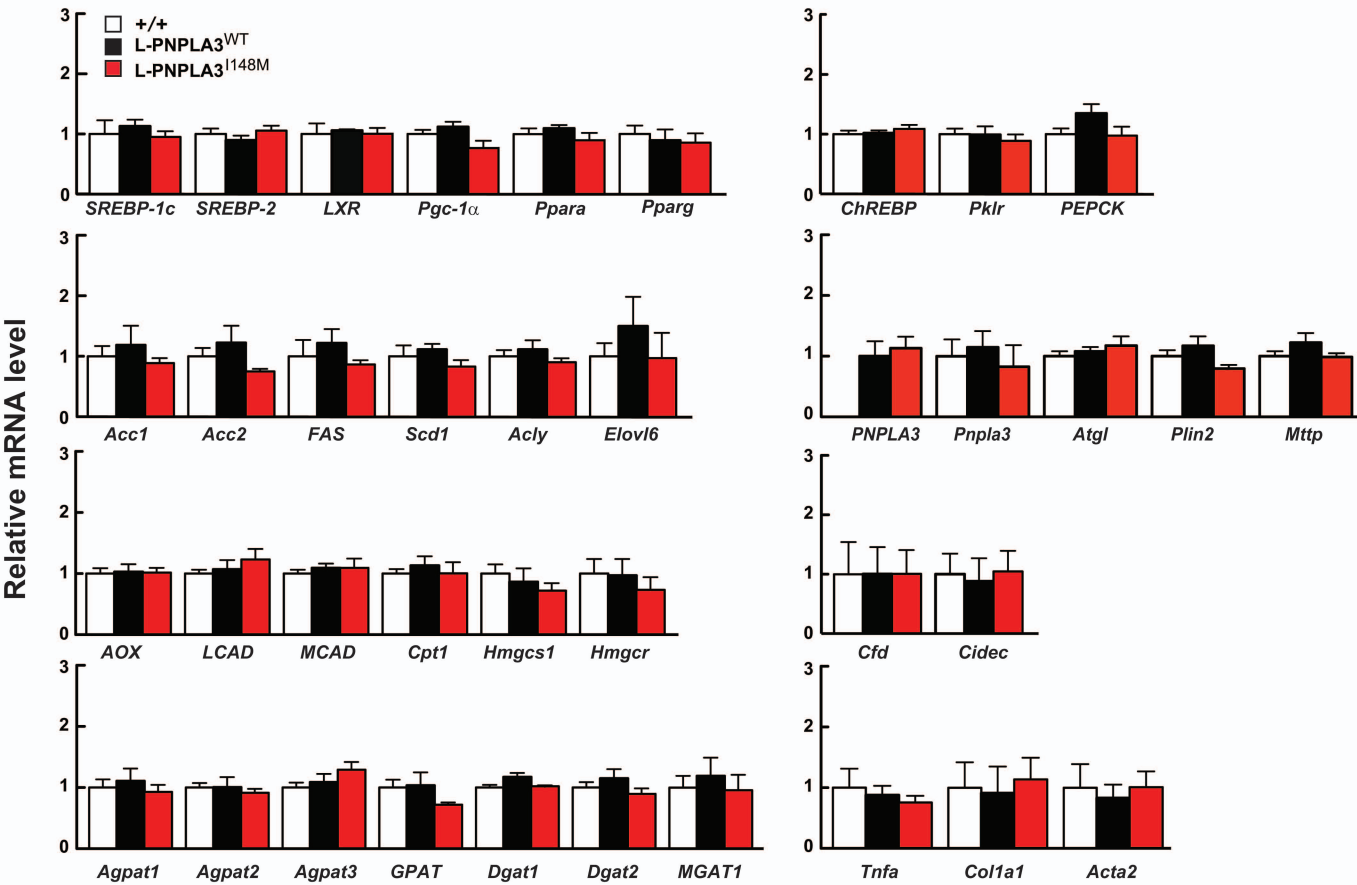


Figure 1B Perilipin 2

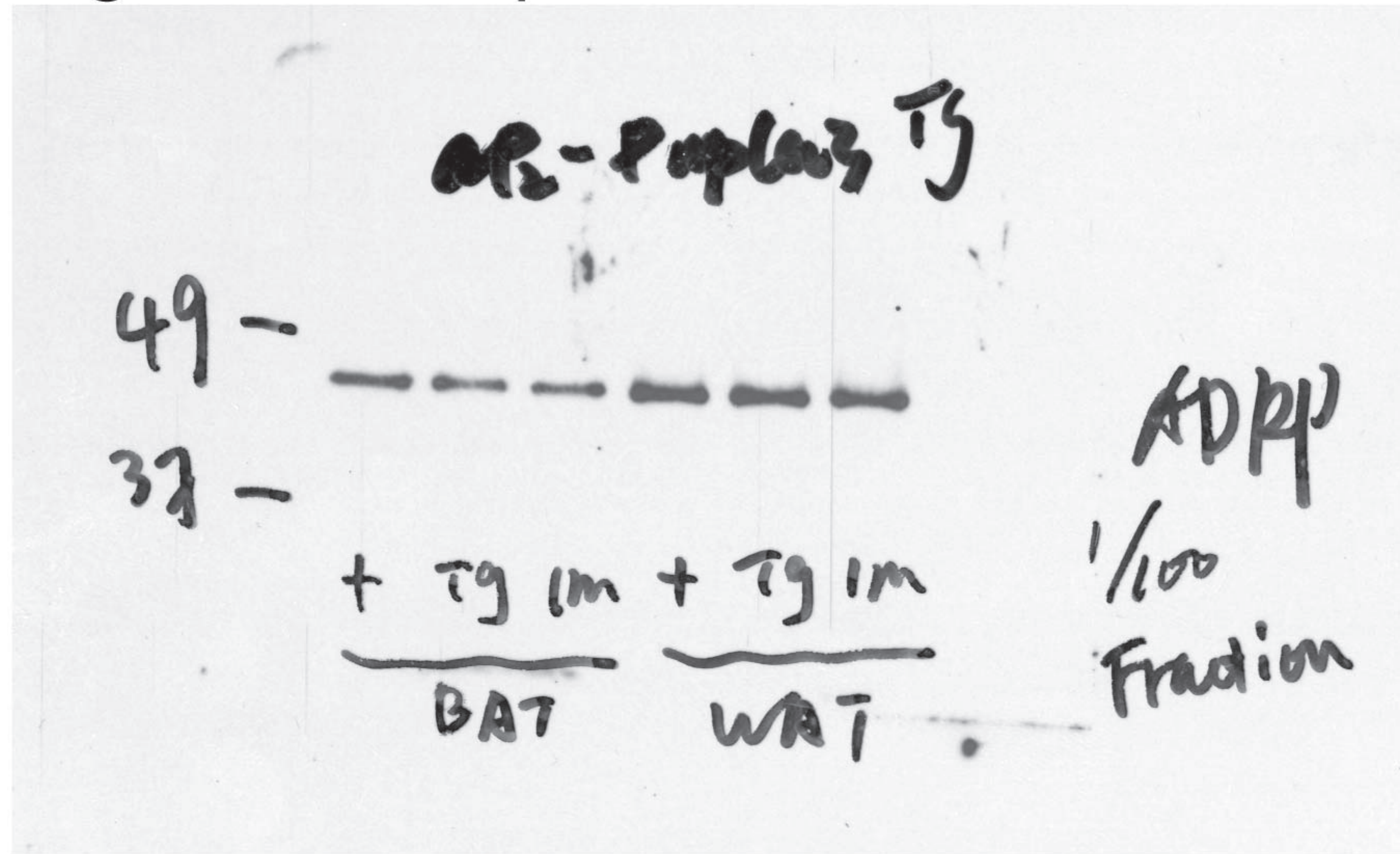


Figure 1A Perilipin 2

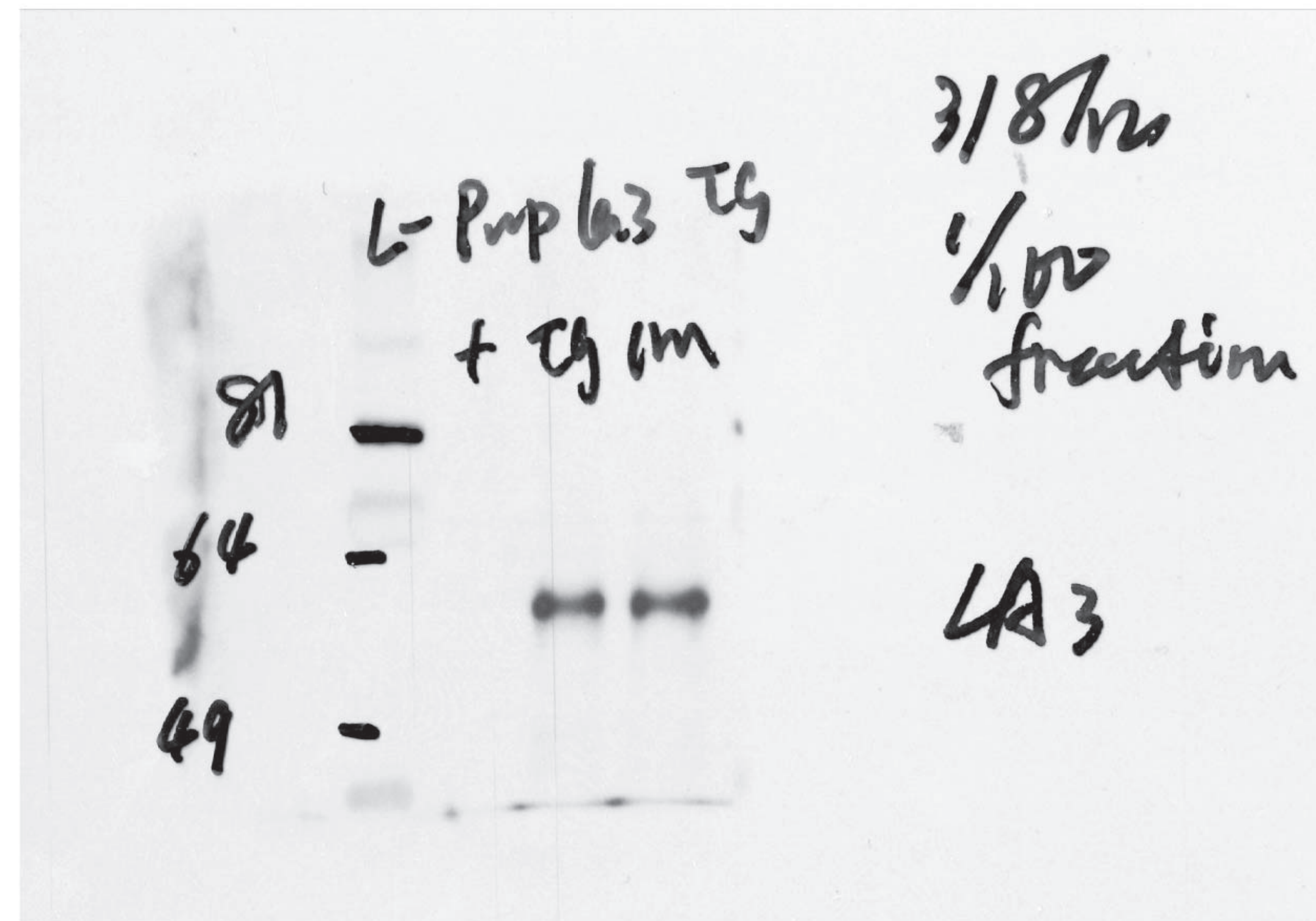
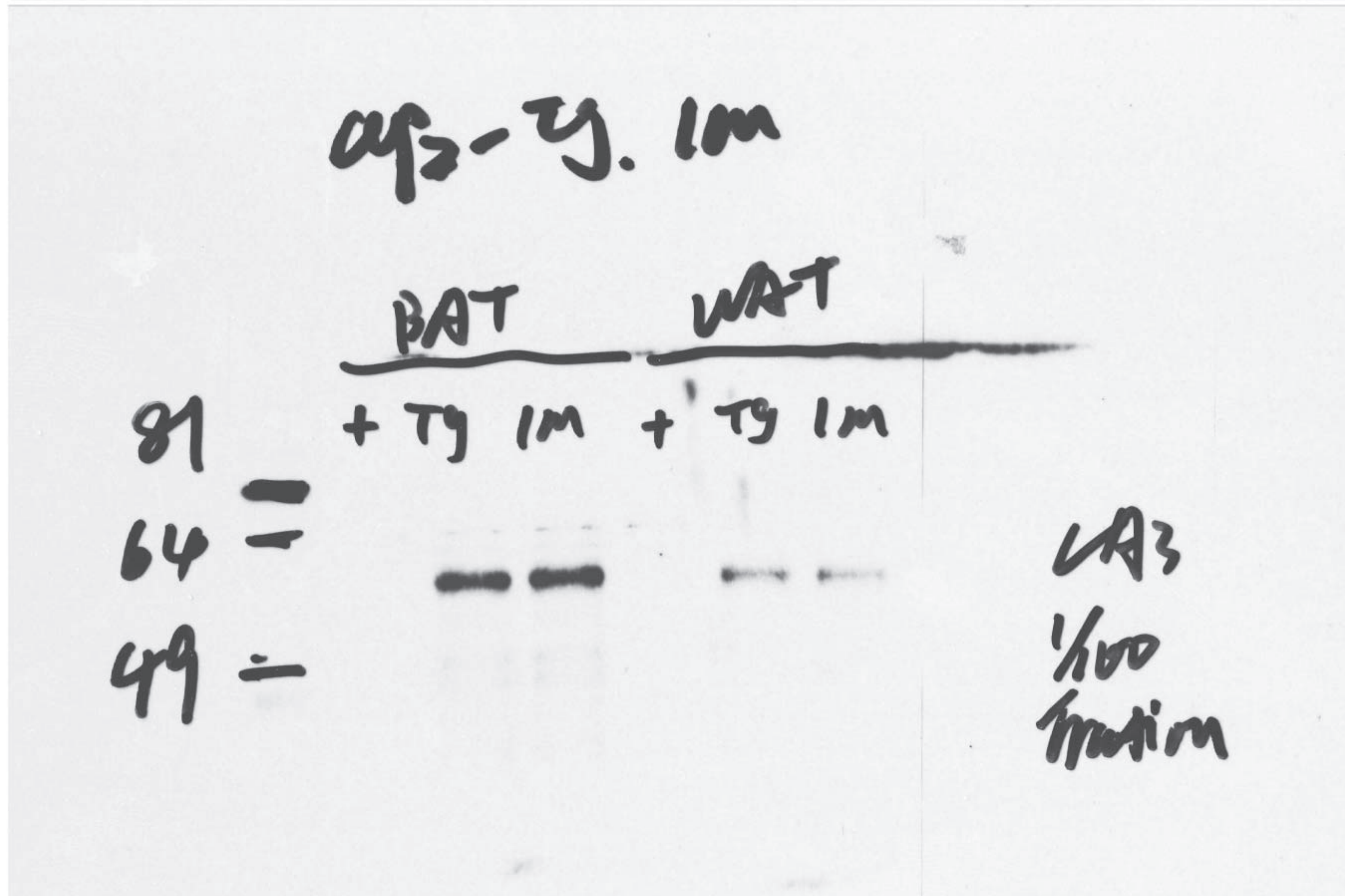
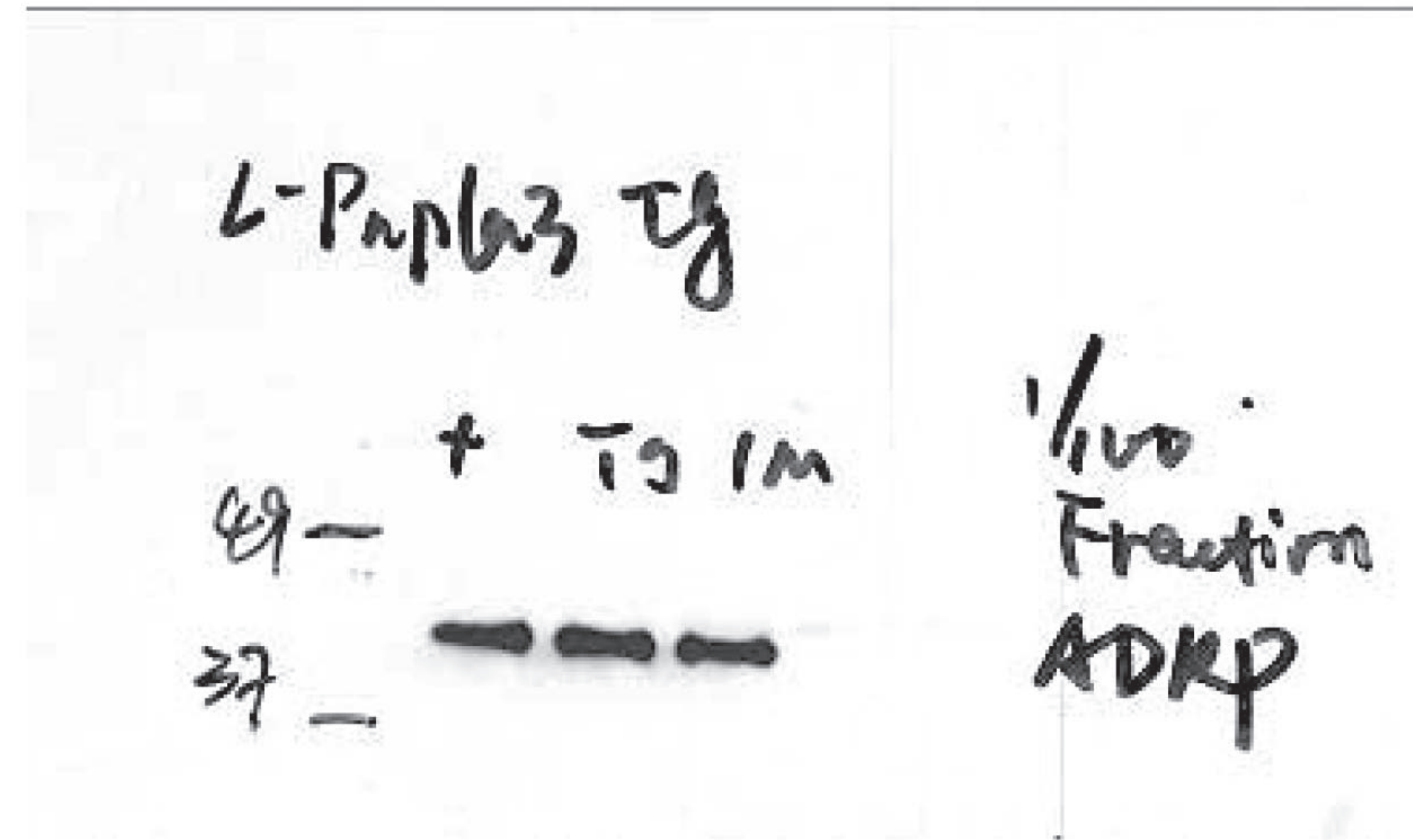
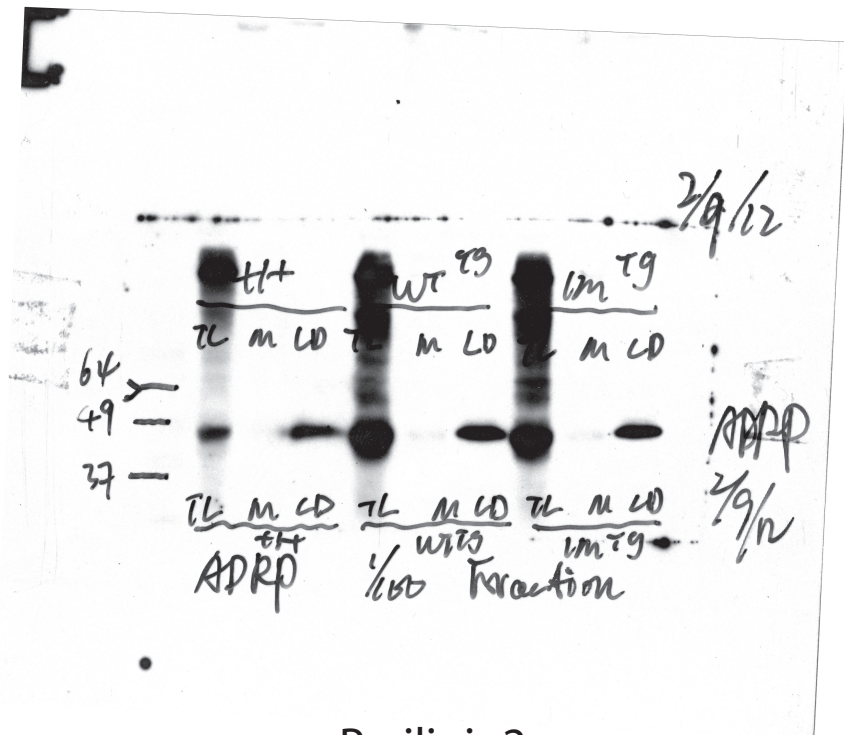
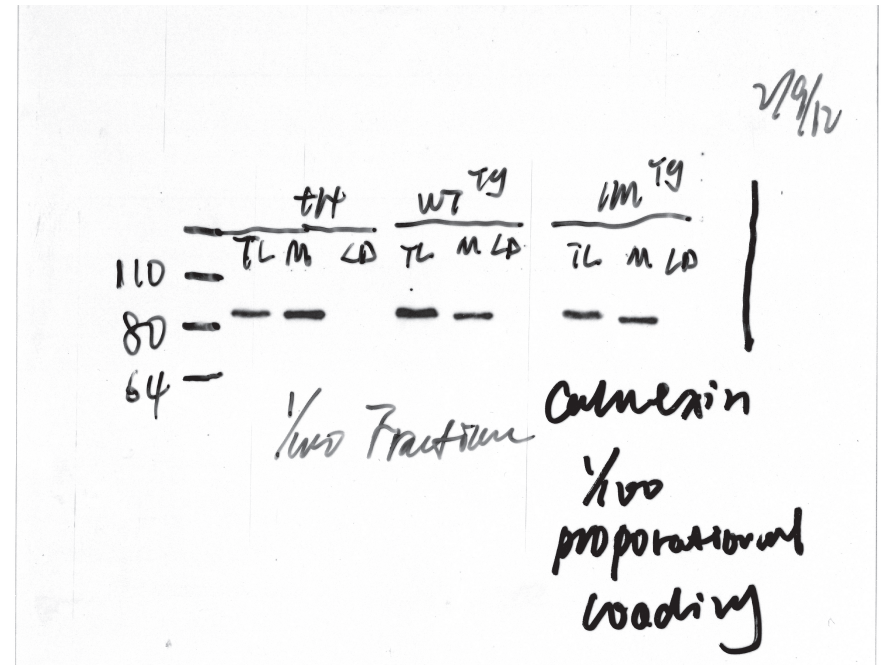


Figure 1B PNPLA3

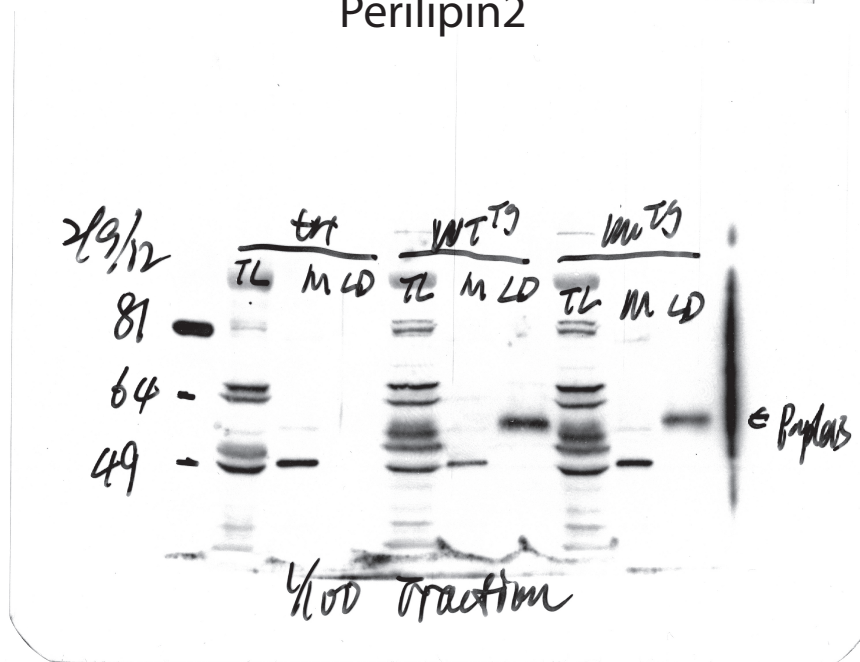
Figure 1A PNPLA3



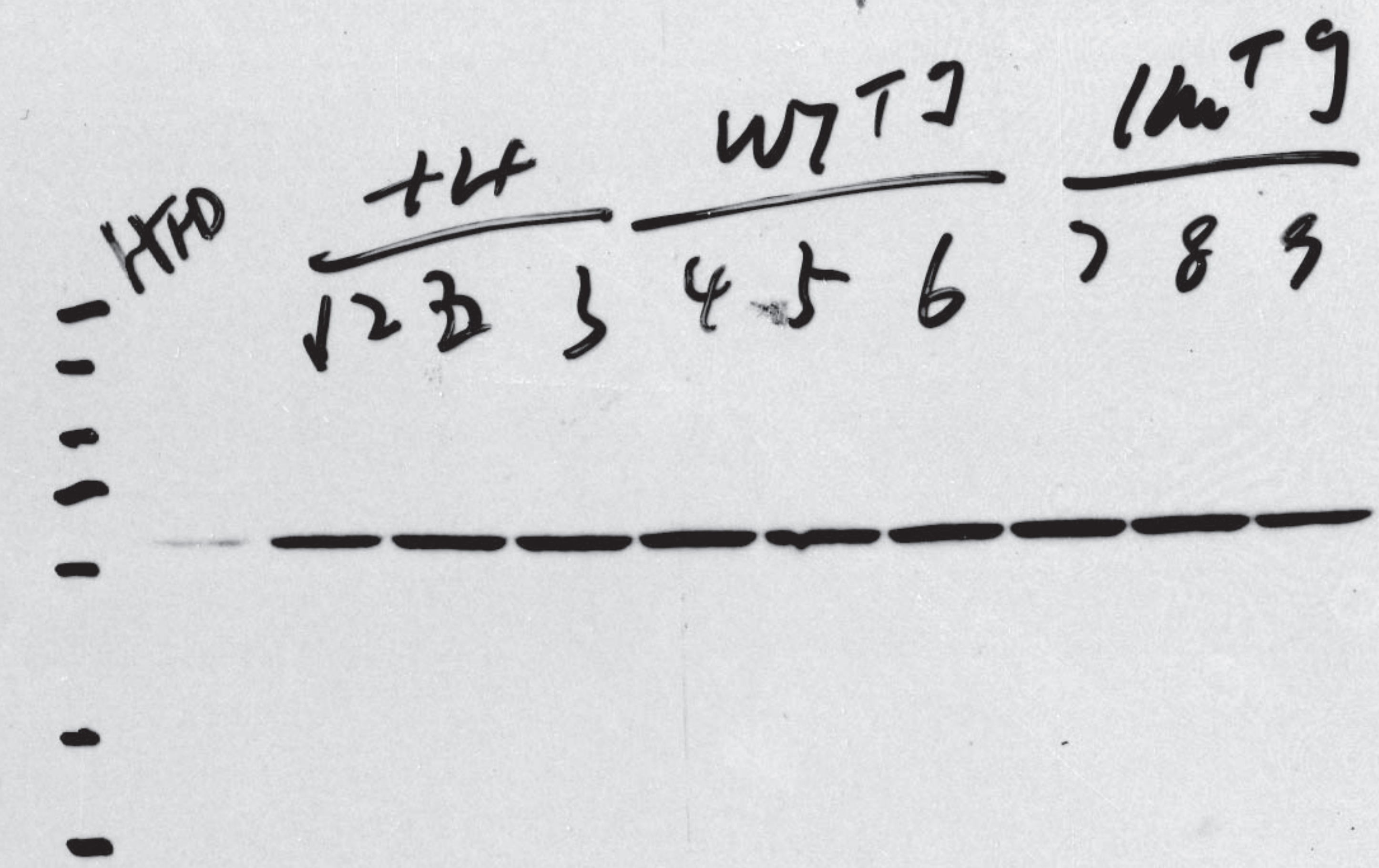
Perilipin2



Calnexin



Human-PNPLA3



P-Akt
 Ser473

Akt

10"
 400µg protein
 5hr 4/12

Supplemental Figures

Supplemental Figure 1

Immunoblot analysis of hPNPLA3 in total cell lysate (T), membranes (M) and lipid droplets (LD) from livers of wild-type (+/+), WT^{Tg} and 148M^{Tg} mice. Total cell lysates, membranes and lipid droplets were isolated from livers of the mice as described (18) and 1% of the total volume of each fraction was size-fractionated on an 8% SDS-PAGE gel. Immunoblotting was performed using a polyclonal anti-human PNPLA3 antibody (18). PLIN2 is a lipid droplet resident protein and calnexin is a membrane resident protein.

Supplemental Figure 2

(A) Plasma levels of lipids and serum levels of aspartate aminotransferase (AST) and alanine aminotransaminase (ALT) in 12-wk-old, chow-fed wild-type (+/+), WT^{Tg}, and 148M^{Tg} male mice (n=4/group) after a 4 h fast. **(B)** Glucose and insulin tolerance tests in 10-wk-old, chow-fed, male mice. After a 16 h fast, 1.5 g/kg of glucose was injected intraperitoneally (IP). Blood samples were collected from the tail vein at indicated times and glucose levels were measured using a glucometer. Insulin tolerance tests (ITT) were performed after a 4 h fast. A total of 0.75 U/kg body weight of human insulin was given IP and blood glucose levels were monitored. Values are means ± SEM (n=6/group). **(C)** Immunoblot analysis of total and phosphorylated hepatic Akt (P-Akt) in livers of 12-13 wk old male wild-type (+/+), WT^{Tg} and 148M^{Tg} mice on an ad lib chow diet (n=3/group). Total cell lysates (40 µg) were subjected to immunoblotting using Akt and phospho-Akt (Ser473) MAb (1:2,000 dilution) (Cell Signaling). Eight-week-old WT mice fed a high-fat diet (HFD) for 12 wks served as a positive control for the experiment. **(D)** Mean serum levels of insulin were measured after a 4 hour fast in plasma from 10-12 week old +/+, WT^{Tg} and 148M^{Tg} male mice (n=5/group) fed either a chow diet (left) or a high-sucrose diet (right) after a 4 hour fast.

Supplemental Figure 3

(A) Body composition of 12-wk-old, chow-fed wild-type (+/+), A-WT^{Tg} and A-148M^{Tg} male mice (n=4-5/group). Total lean mass and fat mass were measured by NMR and normalized to body weight as described in Methods. (B) Plasma levels of lipids, glucose, ketone bodies and nonesterified fatty acids (NEFA) (n=4-5/group). Values are means ± SEM. (C) Relative mRNA levels in BAT and WAT of 12-wk-old, chow-fed WT (+/+), A-WT^{Tg} and A-148M^{Tg} (n=4/group). Total hepatic RNA was subjected to Real-Time PCR for the indicated transcripts. Each value represents the amount of mRNA relative to the level in wildtype mice. Values are means ± SEM. *Cebpa*, CCAAT-enhancer-binding protein α ; *Cebpb*, CAAT-enhancer-binding protein β ; *Plin2*, perilipin 2; *Atgl*, adipose triglyceride lipase; *Ucp1*, uncoupling protein 1. (D) Core body temperature after cold exposure in 11-13 wk-old, chow-fed mice. The core body temperature was monitored hourly, starting 1 hour prior to cold exposure (4°C). Values are means ± SEM from 8 -12 mice in each group.

Supplemental Figure 4

(A) Body weight curves of male wild-type (+/+), WT^{Tg}, and 148M^{Tg} mice (n=5-10 mice/group) on chow (left panel), high-sucrose (middle panel) or high-fat (right panel) diets. Chow and high-sucrose diets were initiated at 4 wks of age and continued for 8 and 6 wks, respectively. The high-fat diet was initiated at 8 wks of age and continued for 12 wks. (B) Body composition of mice maintained on high-sucrose and high fat-diets. Lean and fat masses were measured and normalized as described in Methods.

Supplemental Figure 5

Plasma lipid levels and serum glucose, ketone bodies, NEFA, and liver enzyme (AST and ALT) levels in wild-type (+/+), WT^{Tg} and 148M^{Tg} mice on high-sucrose **(A)** or high-fat diet **(B)**. Mice were fed the indicated diets as described in the legend to supplemental Figure 4. Values are means ± SEM (n=5-10 mice/group).

Supplemental Figure 6

Relative mRNA levels in livers of wild-type (+/+) and PNPLA3 transgenic mice (n=4/group) on a high-fat diet. The mice used in this experiment are described in the legend to Supplemental Figure 4. Total RNA was subjected to Real-Time PCR quantification and mRNA levels were expressed relative to levels in nontransgenic mice. Values are mean ± SEM. *Pgc-1α*, PPAR_γ co-activator 1_α; *ChREBP*, carbohydrate responsive element-binding protein; *Pklr*, liver pyruvate kinase; *PEPCK*, phosphoenolpyruvate carboxykinase; *Acc1*, acetyl-CoA carboxylase 1; *Acc2*, acetyl-CoA carboxylase β; FAS, fatty acid synthase; *Scd1*, stearoyl-CoA desaturase-1; *Acly*, ATP citrate lyase; *Elovl6*, ELOVL family member 6, AOX, acyl-CoA oxidase-1; LCAD, long chain acyl-CoA dehydrogenase; MCAD, medium chain acyl-CoA dehydrogenase; *Cpt1*, carnitine palmitoyltransferase 1; *Hmgcs1*, HMG-CoA synthase; *Hmgcr*, HMG-CoA reductase; *Agpat1*, 1-acylglycerol-3-phosphate-O-acyltransferase-1; *Agpat2*, 1-acylglycerol-3-phosphate-O-acyltransferase-2; *Agpat3*, 1-acylglycerol-3-phosphate-O-acyltransferase-3; *GPAT*, glycerol-3-phosphate acyltransferase; *Dgat1*, diglyceride acyltransferase-1; *Dgat2*, diglyceride acyltransferase-2; *MGAT1*, monoacylglycerol O-acyltransferase 1; *Atgl*, adipose triglyceride lipase; *Plin2*, perilipin 2; *Mttp*, microsomal TAG transfer protein; *Col1a1*, collagen, type 1, α1; *Acta2*, α-smooth muscle actin.