

Supplemental Materials and methods

Cloning and expression of the diffocin gene clusters in *Bacillus subtilis*

pETcoco1 (Novagen) was the starting backbone vector that was first modified to remove the two XhoI sites with primers AV1419-AV1420 which have BbsI ends (primer sequences are shown in Supplemental table 2). Then a region was amplified from pETcoco1 DNA with these primers, and the PCR product was cut with BbsI and was ligated back into the larger pETcoco1 vector fragment that was previously cut with XhoI. The result of this ligation destroyed the two XhoI sites. This plasmid was further modified by a similar strategy to destroy the EcoRI sites using primers AV1416 and AV1245. The resulting vector is termed pSW251.

Vector pUC19 was used to accept fragments of the diffocin cluster to assemble into pETcoco. The polylinker of pUC19 was modified by cutting with EcoRI and HindIII and ligating in oligos AV1372, AV1373, AV1374, AV1375. This changed the polylinker to NotI-NheI-KpnI-XhoI-EcoRV-BstBI-BbsI-EcoRI-NsiI-SphI-BamHI-AscI. The construct was termed pSW232. Three fragments of the diffocin cluster were individually amplified by PCR from *C. difficile* 16 (CD16) DNA and cloned into pSW232. The 5' fragment was amplified with primers AV1368 and AV1289 which have NotI and XhoI ends respectively. The middle fragment was amplified with primers AV1288 and AV1366 which have XhoI and EcoRI ends respectively. The 3' fragment was amplified with primers AV1367 and AV1300 which have EcoRI and BamHI ends respectively. These three PCR fragments were separately cloned into pSW232, and termed pSW243, pSW241, and pSW242 for the 5', middle, and 3' portions respectively.

The three diffocin cluster fragments were excised from the pSW241-3 vectors. The pSW243 was cut with NotI-XhoI, pSW241 was cut with XhoI-EcoRI, and pSW242 was cut with EcoRI-AscI (note that this AscI sites is part of the modified SW232 polylinker). These three fragments were

assembled into pSW251 that had been cut with NotI and AscI. The resulting plasmid was termed pDG461 and contains the entire CD16 diffocin cluster.

To make a *B. subtilis* integration vector we used sequences from pDR111 (gift from David Rudner, Harvard Medical School) that includes portions of the *amyE* gene flanking a cloning/promoter region and a spectinomycin marker. This plasmid was modified by cutting with NcoI and SpeI and religating to create pDG481. The pDG481 polylinker was then modified by cutting the vector with HindIII and SphI and ligating in oligos DG1 and DG2. This added NotI and AscI sites to create pDG482. The region containing the entire *amyE* front and back region with the modified polylinker was then amplified using primers DG9 and DG10 both of which have BsaI ends. This fragment was cloned into the NotI and AscI sites of pSW251 (resulting in destruction of the two sites) creating pDG487. Note that there are new NotI and AscI sites that were introduced by the modified poly linker insertion. The NotI/AscI fragment containing the diffocin cluster from pDG461 was then excised and cloned into the NotI/AscI site of pDG487. This new construct was termed pDG488 and is the vector used to introduce the entire diffocin gene cluster into *B. subtilis* BDR123 (see below) to generate BDR123-488. To delete ORFs 1377-1379, a backbone was made by cutting pDG488 with SphI which has sites outside the integration sequence and one within 1374. Two fragments were amplified from pDG488. The first used primers DG13/14 which generated a fragment from the SphI site to the end of ORF 1376 in which an AscI site was incorporated. The second fragment, also with SphI/AscI sites used primers DG15/16 which span from the SphI site outside the integration site to sequence downstream of 1379 (AscI incorporated in the primer). The three way ligation resulted in pDG491. This was transformed into BDR123 to generate BDR123-491.

Diffocin 4 was cloned similarly to diffocin 16 with some modifications due to the absence of the EcoRI site within the CD4 gene cluster. The pSW251 was modified to have an XhoI site in the polylinker using oligos DG211 and DG212 (NotI and AscI). This created vector pDG577. The diffocin cluster was amplified in three fragments. The first was using primers DG210 and AV1288 (XhoI/NcoI). The second used primers DG209 and DG15 (NcoI and AscI). These two were cloned into pDG577 (cut with XhoI/AscI) to create pDG578. The third fragment was amplified using AV1368 and AV1289 (XhoI/NotI) and cloned into pDG578 (cut with XhoI/NotI) to create pDG579. The integration vector was made by taking the NotI/AscI fragment from pDG579 and cloning it into pDG487. This construct was termed pDG580 and was transformed into BDR123 to generate BDG12.

Deletion of the PBSX locus.

The PBSX prophage knockout was constructed by following the procedure outlined in Liu S, et al. 2008. Microbiology. 154:2562-2570. Briefly, the promoter of the *araA* gene, PCR amplified from *B. subtilis* strain BDR11 (David Rudner, Harvard Medical School), was fused to a neomycin resistance gene (*neo*), PCR amplified from plasmid pUB110, by overlapped extension PCR. Overlapped extension PCR was then used to fuse this arabinose regulated neomycin gene to about 1 kb of sequence flanking each end of the *araR* gene, PCR amplified from BDR11.

This PCR product was transformed into BDR11 to make strain BDG2, deleting the *araR* gene and replacing it with a constitutively expressed *neo*. The deletion of *araR* and replacement with P_{araA}-neo was confirmed by PCR and by the conferral of resistance to kanamycin. This strain is functionally equivalent to AR1 from Lui et al., 2008 (17).

Next, a construct was made to knockout the PBSX locus itself. To make this construct, the following five PCR products were spliced by overlapped extension PCR in order, into one large

product: 1 kb of sequence 5' of the *xlyB* gene, amplified from BDR11; 1 kb of sequence 3' of the *xlyA* gene, amplified from BDR11; a chloramphenicol resistance gene, *cat*, amplified from plasmid pJW034; *araR*, amplified from BDR11; and finally, 1 kb of sequence containing the *xlyB* gene, amplified from BDR11. The overlapped extension PCR product was cloned into the *Xma*I and *Spe*I sites of pBluescript SK+. This construct was then linearized with *Sca*I, transformed into strain BDG2, and plated onto LB agar plates supplemented with 5 µg/ml chloramphenicol. Colonies were picked from this plate and patched onto LB agar plates supplemented with either 5 µg/ml chloramphenicol, or 20 µg/ml kanamycin. Successful recombinants were those that are resistant to chloramphenicol, but sensitive to kanamycin due to repression of the *neo* gene by AraR. These strains will have replaced the PBSX target with the *cat* and *araR* genes. Such isolates were selected after growth for 4 hours in LB broth with no antibiotic selection, and then plated onto LB agar plates supplemented with 20 µg/ml kanamycin. This was to select for recombinants that loop out the *cat* and *araR* genes resulting in a clean deletion of the target PBSX genes. The colonies that grew on these plates were tested by colony PCR for the presence of a PBSX knockout. The deletion was confirmed by PCR in strain BDG9. To further optimize expression, additional constructs were made by first switching the selective marker from spectinomycin to chloramphenicol and by designing the integration construct as a single crossover into *amyE*. The pDG487 was modified by digestion with *Asc*I and *Nhe*I to remove the *amyE* 5' portion and *lacI*, and cloning in oligos DG310 and DG 311 which destroys the *Nhe*I site and adds an *Rsr*II site, generating pDG588. This was cut with *Asc*I and *Not*I, and the *Asc*I/*Not*I fragment from pDG579 containing the diffocin 4 cluster was ligated in to generate pDG589. This construct was transformed into BDG9 to generate BDG21. BDG 21 contained the

entire diffocin 4 locus as well as the spectinomycin marker flanked by identical back ends of the *amyE* gene.

To switch the selective marker, the following construct was made. The pDG481 was cut with BbsI and PstI to remove the front portion of the *amyE* gene. Two PCR fragments were then prepared for a three way ligation into this vector; the first was from pJW034 (David Rudner, Harvard Medical School) using primers DG488 and DG489 that contains the *cat* gene. The second fragment was amplified from diffocin 4 using DNA primers DG367 and DG368. The three way ligation resulted in pDG621. This was transformed into BDG21 and recombinants resistant to chloramphenicol and sensitive to spectinomycin were selected resulting in strain BDG45.

Switching the bactericidal spectrum of diffocins.

To replace the diffocin 4 *1374* with that of diffocin 16, a backbone was made by cutting pDG580 with AscI and PmlI and isolating the larger fragment. Two other fragments were amplified from other constructs. The first was from plasmid pDG461 using primers DG281 (PmlI) and DG316 (BspQI). The second was amplified from pDG579 using primers DG284 (AscI) and DG317 (BspQI). The three way ligation created pDG587. This plasmid contained the diffocin 16 *1374* and part of ORF 1373. To reconstruct the diffocin 4 ORF 1373, such that the only replacement in the cluster is *1374* gene, the following PCR products were produced and cloned into the pDG461 PmlI/BspQI backbone: pDG587 was amplified using primers DG284 and DG 339 and cut with AscI and BsmBI and pDG579 was amplified with primers DG281 and DG340 and was cut with BsmBI and PmlI. This three way ligation resulted in pDG603. pDG603 was transformed into *B. subtilis* strain BDG9 to make BDG26 which was transformed again with pDG621 to produce BDG55.

Supplemental Table 1. Bacterial strains used in this study. All *C. difficile* strains are human clinical isolates.

Species	Strain designation	Ribotype	Source	Comments
<i>Clostridium difficile</i>	CD4	024	L-C Fournier	Diffocin producer
<i>Clostridium difficile</i>	CD16	027	L-C Fournier	Diffocin producer
<i>Clostridium difficile</i>	630	012	ATCC	
<i>Clostridium difficile</i>	43593	060	ATCC	Diffocin producer
<i>Clostridium difficile</i>	19135	001	Diane Citron	
<i>Clostridium difficile</i>	19137	015	Diane Citron	
<i>Clostridium difficile</i>	19099	002	Diane Citron	
<i>Clostridium difficile</i>	19145	153	Diane Citron	
<i>Clostridium difficile</i>	19103	001	Diane Citron	
<i>Clostridium difficile</i>	19155	NA	Diane Citron	
<i>Clostridium difficile</i>	19104	027	Diane Citron	
<i>Clostridium difficile</i>	19142	NA	Diane Citron	
<i>Escherichia coli</i>	Top10		Invitrogen	Electrocompetent cells for subcloning.
<i>Bacillus subtilis</i>	BDR11		David Rudner	Expression strain for diffocins
<i>Bacillus subtilis</i>	BDR123		David Rudner	Expression strain for diffocins
<i>Clostridium acetobutylicum</i>	824		ATCC	
<i>Clostridium sordellii</i>	9714		ATCC	
<i>Clostridium sporogenes</i>	3584		ATCC	
<i>Clostridium biofermentans</i>	638		ATCC	
<i>Listeria Ivanovvii</i>	19119		ATCC	
<i>Listeria monocytogenes</i>	23074		ATCC	
<i>Listeria innocua</i>	33090		ATCC	
<i>Bifidobacterium breve</i>	15700		ATCC	
<i>Bacteroides fragilis</i>	23745		ATCC	
<i>Bacteroides fragilis</i>	25285		ATCC	

Supplemental Table 2. Oligonucleotides/primers used in this study.

Primer name	Sequence 5'-3'
AV1245	cccttgaagaccaatttcgtatggcaatgaaagacgg
AV1288	gaagaaagagatgggactcgagatg
AV1289	tatacatctcgagtcccatctctt
AV1300	tccccggatccacactacaatctactctaaactcagg
AV1367	ggaaaagggattgctaatagtg
AV1368	tttttgcggccgcaatacccactacaccttcgtc
AV1372	aattgcggccgcagctcgctagcggctacccgaggatatttcgaagaagacacatccg
AV1373	aattccgggatgcatgcctctaggatccggcgcgcc
AV1374	agctggcgcgccggatcctagaggcatgcatcccgaattcggatgtgtctt
AV1375	cgaagatatactcaggtaccgctagcgagctgcggccgc
AV1416	ttccttgaagacctaatttggggcaatcccgaaggag
AV1419	ttcttgaagaccatcgaagcaccaccaccaccactg
AV1420	tttttgaagacaatcgaagggttcgccctgtcgtcgcac
DG1	ggcgcgccactagtaccggtgccatggcggccgc
DG2	agctgcggccgccatggcaccggtactagtggcgcgccatg
DG9	ttccttggctcagcgaacaaaattctccagtcttc
DG10	ttccttggctcaggccgtcgcgactaagaaaatgcc
DG13	gtgagcggataacaattccc
DG14	agattgtagtgtggatccgg
DG15	tccttcggcgcgcctcaaatttaagcttaactcc
DG16	tttagggactactcactcgc
DG209	actggataccatggtacttc
DG210	tgaagtaccatggtatccag
DG211	ggccgcctcgaggg
DG212	cgcgccctcgaggc
DG281	gtataagccacgtgctgaag
DG284	aggcgcgcctcaaatttaag
DG316	tcttctgctcttcaagggtctccaagatttaaattctt
DG317	ctccttgccttccactatataatcaattaataataaaac
DG339	tcctctcgtctcaatgaagcaaaaataaacttttac
DG340	tcctctcgtctcttattataaaaacctcctaattat

Supplemental Figure 1S. A) Clustal analysis of ORF 1374 from srtains CD4, CD16, CD43593, and 630. B) Dendrogram of the clustal analysis.

A.

```

4          MKRRTKLLQRGNFFGDKNMVVDEFDEGYDNYDFINFFTGCCNYTFGLKNNNIIYCGDNSN
16         MKQNKLLQRGAYFNDKNI LIDDDFKRYNDYDFVEFFTGISNSTFGLKSDGNLYACGDNTG
43593     MKQNKLLQRGAYFNDKNI LIDDDFKRYNDYDFVEFFTGISNSTFGLKSDGNLYACGNNTG
630       MKQNKLLQRGAYFNDKNI LIDDDFKRYNDYDFVEFFTGISNSTFGLKSDGNLYACGDNTG
          **:..***** :*.***:::***: *::****:***** .* *****:.. **.**:*:..

4          FQLGLGEDNTRKLFTKIPNISTNIKKVACGESHAVILTSDGELLVAGINTDGOGLGLE
16         FQLGLGKDSERRMFSKVK--IDNVKYVSCGSKHSVAVTKDGFAYGAGTSNVGQLGVIES
43593     FPLGLGKDSERRMFSKVK--IDNVKYVSCGSKHSVAVTKDGFAYGAGTSNVGQLGVIES
630       FQLGLGKDSERRMFSKVK--IDNVKYVSCGSKHSVAVTKDGFAYGAGTSNVGQLGVIES
          * *****:*. :*:***: *:* *.*.*..*:* .*.* ** .. ***:*. .

4          KVGKTVSTFEKVPKIGVKDIACGLQSTYLLYNDGTLYVAGNNLYGQLGLGTNGASANVN
16         TVYYEFTKLP----IDDVKTVACGYDFTFVLKNDGTLYSAGLNSSGQLGLG---DTNNRA
43593     TVYYEFTKLP----IDDVKTVACGYDFTFVLKNDGTLYSAGLNSSGQLGLG---DTNNRA
630       TVYYEFTKLP----IDDVKTVACGYDFTFVLKNDGTLYSAGLNSSGQLGLG---DTNNRV
          .* ..: : *..** :*** : *::* ***** ** * ***** : *

4          TFTKVDVDNVKAVFSYNKSAFI IKNDNKCYSTGFNNQQLGLGDKNNRDLFSLVS-INDV
16         TFTKVNIDSVKDVVTYNQSVFI IKMDGTAHACGLNSNGQLGINSTLNKSVFNKIEGMDNV
43593     TFTKVNIDSVKDVVTYNQSVFI IKMDGTAHACGLNSNGQLGINSTLNKSVFNKIEGMDNV
630       TFTKVNIDSVKDVVTYNQSVFI IKMDGTAHACGLNSNGQLGINSTLNKSVFNKIEGMDNV
          *****:*. ** *.:***:*.***** *...: :*:.:*****:.... *:.:*. .. :*:

4          KTIACGSEHTVLMTYNNDIYCGCK---EKCFGNALQSSLFTKIEEVNIKTIACGHGNTM
16         KQIACGSSHTILIKNDGTMYTTGYNGVGQLGTGNNNNSIVFTLSSINNVKYASCNNHTM
43593     KQIACGSSHTILIKNDGTMYTTGYNGVGQLGTGNNNNSIVFTLSSINNVKYASCNNHTM
630       KQIACGSSHTILIKNDGTMYTTGSNGYGQLGTGNNNNSIVFTLSSINNVKYASCNNHTM
          * *****.**:*:.. :. :* * : ** :* :** . *.* :***:*.**

4          LIDNKGTCLKVAGNNDIYQLGIANYSENIDNSFIDLKNIIVAKNIFIGLSHSILIDSNNDSY
16         ILKYDNTLSTGQNNYGQLANANKDVASRNTFAKVNVENIKDIKCGSQFNFLINGSKEIF
43593     ILKYDNTLSTGQNTYQQLANANKDVASRNTFAKVNVENIKDIKCGSQFNFLINGSKEIF
630       ILKYDNTLSTGQNNYGQLANANKDVASRNTFVKVNVENIKDIKCGSQFNFLINGSKEIF
          :.. ..** :*: * * . ** . *:* :. : *:* * ...:***:..: :

4          CTGDNTYQQLGSFFDDMHIVEFKMDSEKYSYSNYINLIKSEDKLTLLEEMEIKDIELP
16         VSGCNLAGQLGSFFHTTFLYEFQSS--NLDNYSGLLVNDDYLYVTKDNSEFLNVKLS
43593     VSGCNLAGQLGSFFHTTFLYEFQSS--NLDNYSGLLVNDDYLYVTKDNSEFLNVKLS
630       VSGCNLAGQLGSFFHTTFLYEFQSS--NLDNYSGLLVNDDYLYVTKDNSEFLNVKLS
          :* * *****. :. :***:*. . ** .* :.* * : *:. :*: :*:

4          LDHSVRDVVFSFYCTLVILGNGDVYGLGNNRYKMGSDLPQLNELTKLSISNVKSIVA
16         DNFQDYKKIELTDSNMFIVMNDGTLYACGLNNGQLGLGDTVNRVMTKVDIDNVLDIKG
43593     DNFQDYKKIELTDNNMFIVMNDGTLYACGLNNGQLGLGDTVNRVMTKVDIDNVLDIKG
630       DNFQDYKKIELTDNNMFIVMNDGTLYACGLNNGQLGLGDTVNRVMTKVDIDNVLDIKG
          :... :. : : : : : : * :. * *.* :* . . : . :***:*.** .* .

4          SKNISGGIFYIKNDDTCYSGPNSNSIAGVLP-SNSDVFKKISIDNVKKVVINTDLSNWF
16         NGNST---FVLKNNGTLYSCGLNSNGQLGLRDEVNRNIFTKIEIENVKDFCVGSNY----
43593     NGNST---FVLKNNGTLYSCGYNSSGILGLKDNTNRNIFTKIEIENVKDFCVESNY----
630       NGNST---FVLKNNGTLYSCGYNSSGILGLKDNTNRNIFTKIEIENIKDFCVESNY----
          . * : * :***:*. * * **.. * : * :*.**.**:*. . : :

4          SLIVTNNKQIYTSKSSSYVNGLSNALISQYTEISLSNVTDAYSSYNATFIVVDEKKVYA
16         --VIALNHSKEVYGWGNPNYNNIEKTSNYPYKQG- ISNIEKIAAYDYSVYMINSEGKLYV

```


43593 --IVVLNHSKELYGWGNESYIVYGNRNYPKDTRVSNVEKIATWSDTLYILDSTGATKT
630 --IVALNHSKELYGWGNQSYIVYGDNRNYPKDTRVSNVEKIATWSDTLYILDSTGATKT
:. * : . * * : : * : . : : : . . .

4 TGINTNYLLGFSTSDGSNVNGLLSDWYIINISGSSYSRVSCTNNITKINNI I IYEYVTV
16 SGYNYNYQLGKGNNSNQSK---ALVSQCRNSTSSTSNGLR--TLPKITNVFPFYDGCAI
43593 IGYSYNGSGGY PAPSST---YRDGGYINKNTSYRTLEFYNTSKTKLVNLFAYNGCVF
630 IGYSYNGSGGY PAPSST---YREGGYINKNTSYRTLEFYNTSKTKLVNLFAYNGCVF
* . * * : : . . . * : * : . .

4 FCTNIGSFLTGYHGTSWTKPTDSSYRVQYQGISYAGYLDYIYNYYPTRCTQSSSSTTFA
16 IDEGGYVYLTGYHGYLRTLNSPISDYSR---YGTFFIATNSNHN-----TYFI
43593 VDENGLAYCIGENNINFRGNSTTNNNSLR---FINNSGVYTTNTDGTDTYCYQWYTKLI
630 VDENGLAYCIGENNINFRGGSTTNNNSLR---FINNSGVYTTNTDGTDTYCYQWYTKLI
. . : * : . : . . : : * : :

4 YLYNGESSNLKVNPNLNLISGSSYIHQYGRNYLNNQSSNNAASNINSGPITSDKAI
16 QETDFSGIEKVI GMSNNILFFKKGSSYITGYP-KTFGSTITGHRYSYTSINSESSNLGSN-
43593 RCSI F DSPQNI IGNSKNILYLSKNNSTFKCTG-NCITYGINSQNWYSYFSDSSNGAIALG
630 RCSI F DSPQNI IGNSKNILYLSKNNSTFKCTG-NCITYGINSQNWYSYFSDSSNGAIALG
. . : : . . : * : . . * : : : : . . : :

4 FLYKALLYLSSNTLYGFGNISESAKELDVSDTQDGYNATNYKKVMKNIKNIFIPPYDLR
16 ----FIIYHSNSKLYGK----IANSGQFGNSTNIDGTSNYDTGLKDIKDIIVK-----
43593 NEFILKNYSGECLLKGYG----KATNGEFGNSTNISSISNYDTGLKDIKDIIVK-----
630 NEFILKNYSGECLLKGYG----KATNGEFGNSTNISSISNYDTGLKDIKDIIVK-----
* . . * * * * . . : : : : . : * * . . : * : * : * : . .

4 DKTRFAILTDKSLFICGYNSKGTGHISVNSSLNLNKNINYNKKNSSSEISSNIQEIYSHS
16 GNTVVVVVDKNNNIYVTGMNQNNKLGIGEYNNPVKKFTNITEQSNSFIFMDDIKEITTSR
43593 NNTVVVVVDKNNNIYVTGANQFNKLGIGEYNNQPIKFTNITEQSNSFIFMDDIKEITTSR
630 NNTVVVVVDKNNNIYVTGANQFNKLGIGEYNNQPIRFTNITEQSNSFIFMDDIKEITTSR
. : * . . : : : : : * * . . * * . . : : * . : : . . * : : * : * : . .

4 KSTYLLTNNMMLYSVGLNDVQGLGVGDEINRKVFTKINIDNIKSINVNRFTDNSKHAFAI
16 NTMFIVKNDGTAYATGNSSGQLGLGDTINRNKFTQINLDNIKKIST---SIDGNTTFAI
43593 NTMFIVKNDGTAYATGNSSGQLGLGDTINRNKFTQINLDNIKKIST---SIDGNTTFAI
630 NTMFIVKNDGTAYATGNSSGQLGLGDTINRNKFTQINLDNIKKIST---SIDGNTTFAI
: : : : . * : . * : * * . * * * * : * * : * * : * * * * . . : : : * * *

4 KNDNTCYAVGLNNSGQLGIGDENVNRNIFTKINVENVKYVAVYGNTSLLLTNDGLLYGAGN
16 RNDGTLYSTGLNTKGQLGLGDIVNRNFTTKVNIQNVRDVVLGTTHSHAIKDDNTLYSCGE
43593 RNDGTLYSTGLNTKGQLGLGDIVNRNFTTKVNIQNVRDVVLGTTHSHAIKDDNTLYSCGE
630 RNDGTLYSTGLNTKGQLGLGDIVNRNFTTKVNIQNVRDVVLGTTHSHAIKDDNTLYSCGE
: * * * * : * : * * * * * * * * * * * * * * * * * : * : . * : : * . * * * * :

4 NGKGQLGLGDTTSR---NIFTRIPINGVRDVYLCNDVSIIVKNDNTCYCVGLVNGYFGFT
16 NTHGQLGLGSESNHPDVLTFVNNITNVRDVYCSDTTTFIVKDTNIAAYCCGYNNNSQLGM
43593 NTHGQLGLGSESNHPDVLTFVNNITNVRDVYCSDTTTFIVKDTNIAAYCCGYNNNSQLGM
630 NTHGQLGLGSESNHPDVLTFVNNITNVRDVYCSDTTTFIVKDTNIAAYCCGYNNNSQLGM
* : * * * * . : : * * * . * * * * . : : : * * : * * * * .

4 EGSISTFTKINIENVKSVVTAGSEATFFITNDNMIYTTGKKERVFFSTETNDIKGIRVIN
16 GNTTDQYSFIKCMENVKEVIPNEINTYIITIYNTAYSTGLNTDYCLGLNSNSNQSSFSEI
43593 GNTTDQYSFIKCMENVKEVIPNEINTYIITIYNTAYSTGLNTDYCLGLNSNSNQSSFSEI
630 GNTTDQYSFIKCMENVKEVIPNEINTYIITIYNTAYSTGLNTDYCLGLNSNSNQSSFSEI
. : . : : * : . * . . * : * * * * * * * * : . . : * . . .

4 NIINAKKIVVN-GYTSAILTNDNKLFGV---GLSGYGSIANNNNTNSVEDVKDFVTAN
16 PISNVVKVAPNRNNAVLLLTSEGDVYTAGKCSNGSGTGSETPEKIKKIASKAKDIGMNYR
43593 PISNVVKVAPNRNNAVLLLTSEGDVYTAGKCSNGSGTGSETPEKIKKIASKAKDIGMNYR
630 PISNVVKVAPNRNNAVLLLTSEGDVYTAGKCSNGSGTGSETPEKIKKIASKAKDIGMNYR
* * . * : . * . : : * * * * * * : : : . . . * * : . .

4 NTLYIDNNNNLISSGRDITYGISDESRYRDMSPVYKVSIKKDVDTVFSSYNTIFIKDIYGK
16 CGHYVSDNGDLYGTGFNNGQLGVGDVTKRDTFIKTNTR-VKKILPLEYANIAIKDTNDI
43593 CGHYVSDNGDLYGTGFNDCGQLGVGDVTKRDTFIKTNTR-VKKILPLEYANIAIKDTNDI
630 CGHYVSDNGDLYGTGFNDCGQLGVGNVTKRDTFIKTNTR-VKKILPLEYANIAIKDTNDI

.:.:* .:* : * . . . : *.. : . : .* .* *** .

4 FYSSTRDNRYNHLGIHHRDNDKNEALEGSLHSYFKTDNTSDKIVFNKKNKLVFNDKY
16 YICGLNN--YGQLGVGNRYDSRNN---DNRI FNYKHMNFVMDLTSIKNRHNFILLNNKI
43593 YICGLNN--YGQLGVGNRYDSRNN---DNRI FNYKHMNFVMDLTSIKNRHNFILLNNKI
630 YICGLNN--YGQLGVGNRYDSRNN---DNRI FNYKHMNFVMDLTSIKNRHNFILLNNKI

: . . . : *.:***: :***. :* :. :..* : : *:.....**:*

4 IKTNNKYINYKNIFKDNFK--YTSIILPFEVSDIDISKTHSLAVAKDGKLYGIGSNSYKE
16 VIPTTKDIDYGLVLGNLYKGDLYTELPYEDIKEVSISKTHIIILLNDGTMYGCGTNYHGE
43593 VIPTTKDIDYGLVLGNLYKGDLYTELPYEDIKEVSISKTHIIILLNDGTMYGCGTNYHGE
630 VIPTTKDIDYGLVLGNLYKGDLYTELPYEDIKEVSISKTHIIILLNDGTMYGCGTNYHGE

: ...* *:* : : : * : : : : : : : : : : : : : : : : : : : *

4 INQTLEDIELLTLTEVNIISDVKKVACGDNYSYIIKTDNTLWSYGKNTYQLGVGHNNVDV
16 LLQDLSINQVDEFVQINVSVDVKKHVSCGDNFTYFIKSDSLWSIGKNSEYQLGIGHNNPVT
43593 LLQDLSINQVDEFVQINVSVDVKKHVSCGDNFTYFIKSDSLWSIGKNSEYQLGIGHNNPVT
630 LLQDLSINQVDEFVQINVSVDVKKHVSCGDNFTYFIKSDSLWSIGKNSEYQLGIGHNNPVT

: * * . : * * * * * * * * * * *

4 ELQKVTGLPSVKDISIYNSMTLVLTNEGELYAQGYNTNGLFGLGESEKDKIIRTFTKVL
16 ELQRITTISSCKEVHCGKNYTLVVTGNELFVQGYNDKALGLGSDSENTIIKFFTKALT
43593 ELQRITTISSCKEVHCGKNYTLVVTGNELFVQGYNDKALGLGSDSENTIIKFFTKALT
630 ELQRITTISSCKEVHCGKNYTLVVTGNELFVQGYNDKALGLGSDSENTIIKFFTKALT

:* :.* *:* : . : ***:* . **:* *** : * :.....**:* ***.**

4 NVKEIKSHNDHILVIKNDNSLWITGKNKSMYKISISITDLYEFTKIPIPEHLNDILDIE
16 DIREIKSYGSDHILVLKNDNSVWVTGKNRDVYKIEQPVEFLKEFTIIPISEDVNTVKDVL
43593 DIREIKSYGSDHILVLKNDNSVWVTGKNRDVYKIEQPVEFLKEFTIIPISEDVNTVKDVL
630 DIREIKSYGSDHILVLKNDNSVWVTGKNRDVYKIEQPVEFLKEFTIIPISEDVNTVKDVL

: : : : : : : : : : : : : : : : : : * * * * : * * . : * : *

4 LSDDTIYMITKVDTSKASIEIVEKSIISQVRVVVQDPNNVIEKLEMFINDLISTKTNLEI
16 ATDNTLYIISEVGTNAAEITEKSISSIKIKIQDPNKDISRIEMLINGESVKSVDLIT
43593 ATDNTLYIISEVGTNAAEITEKSISSIKIKIQDPNKDISRIEMLINGESVKSVDLIT
630 ATDNTLYIISEVGTNAAEITEKSISSIKIKIQDPNKDISRIEMLINGESVKSVDLIT

: * : * : * : * : * : * : * : * : * : * : * : * : * : * : * : * : * : *

4 NSIIFEIPQNKIVLGENKILIKASSPTGDLYSSMFIFKSETGLKVKKDSILMINNKVYSI
16 EKISFEVPPDKIKIGENKILFRAYCKGDDLYASLFIFKESTGNSIIKDSYVMIGNRMKYV
43593 EKISFEVPPDKIKIGENKILFRAYCKGDDLYASLFIFKESTGNSIIKDSYVMIGNRMKYV
630 EKISFEVPPDKIKIGENKILFRAYCKGDDLYASLFIFKESTGNSIIKDSYVMIGNRMKYV

: . * * * : * : * : * : * : * : * : * : * : * : * : * : * : * : * : *

4 INITENNTDLIVTLNEGLKDDMMENPIYQLINKTKVQVKINKSDLFKDMKLVEIKKSDS
16 VNTTSNEQDITITLDRGLEEDLNLGDPIYQLINKTKVQVKINKSDLFKDMKLVEIKKSDS
43593 VNTTSNEQDITITLDRGLEEDLNLGDPIYQLINKTKVQVKINKSDLFKDMKLVEIKKSDS
630 VNTTSNEQDITITLDRGLEEDLNLGDPIYQLINKTKVQVKINKSDLFKDMKLVEIKKSDS

: * * . : * : * : * : * : * : * : * : * : * : * : * : * : * : * : *

4 SYQEIYELEEANIKSAQPKIIVEKGDKWTAIKRPSMIFRYDAENNEPQA
16 SYQEIYELEEANIKSAQPKIIVEKGDKWTAIKRPSMIFRYDAENNEPQA
43593 SYQEIYELEEANIKSAQPKIIVEKGDKWTAIKRPSMIFRYDAENNEPQA
630 SYQEIYELEEANIKSAQPKIIVEKGDKWTAIKRPSMIFRYDAENNEPQA

B.

