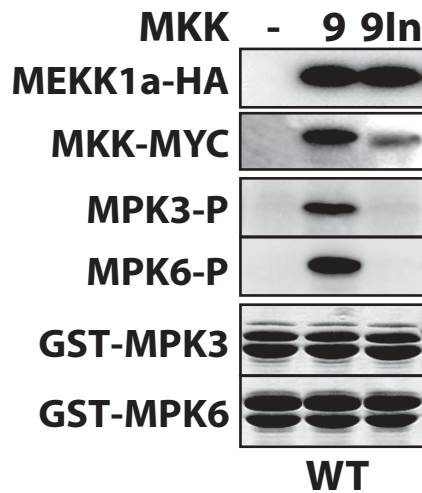
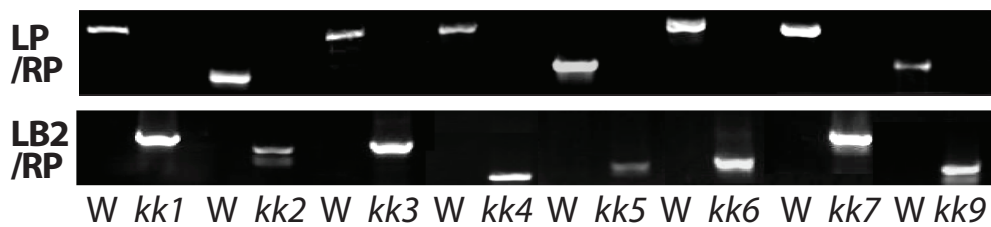


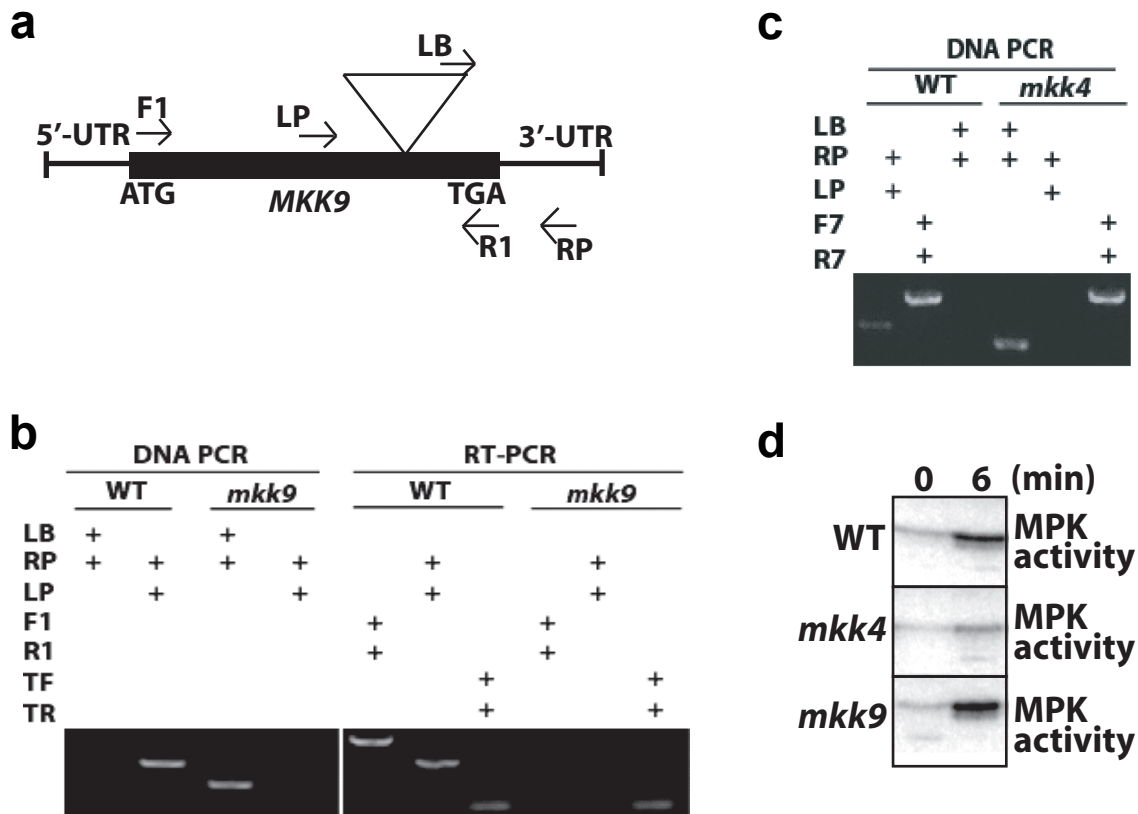
**Supplementary Figure S1 | Specific *MPK*, *MKK* and *CTR1* expression and function in ethylene signalling.** **a**, *MPK3* and *MPK6* are expressed in protoplasts and leaves. Quantitative RT-PCR (black, protoplasts) and ATH1 GeneChip data (white, mature leaves) retrieved using Genevestigator<sup>50</sup> are shown. Error bars, s.d. (n=3). **b**, Transient expression of WT *MKK7* or *MKK9* preferentially activates endogenous MAPKs in *ctr1* protoplasts. Empty vector (-) or an inactive form of *MKK7*<sup>K64M</sup> (*7In*) was used as a control. *In-gel* MAPK assay<sup>34-36</sup> was performed to reveal endogenous protein kinase activities. Endogenous *MPK6* protein level (anti-*MPK6*) was shown as a loading control. Experiments were repeated at least three times with similar results. **c**, WT *MKK4* and *MKK5* did not activate MAPKs in WT protoplasts. **d**, *EBS-LUC* activity is suppressed by expression of the *CTR1a* in *ctr1* protoplasts. Inactive *CTR1* (*InCTR1*) was used as a control. **e**, Transcript level of *MKK9* is relatively more abundant than those of *MKK6* and *MKK7* in protoplasts and adult leaves. Error bars, s.d. (n=3).



**Supplementary Figure S2 | Active MKK9 directly phosphorylates MPK3 and MPK6.** MKK9-MYC was activated by constitutively active MEKK1a-HA<sup>36</sup> in WT protoplasts and immunoprecipitated for an *in vitro* phosphorylation assay with GST-MPK3<sup>K67R, K68R</sup> or GST-MPK6<sup>K92M, K93M</sup>, generated from *E. coli* with no autophosphorylation activity as previously described<sup>36</sup>. Empty vector (-) or an inactive mutant MKK9<sup>S195A, S201A</sup> (9In) served as a control. Expression of proteins was detected by immunoblot analysis using anti-HA (MEKK1a-HA) and anti-MYC (MKK-MYC) antibodies. Levels of GST-MPK3<sup>K67R, K68R</sup> and GST-MPK6<sup>K92M, K93M</sup> were determined by Coomassie Blue staining.

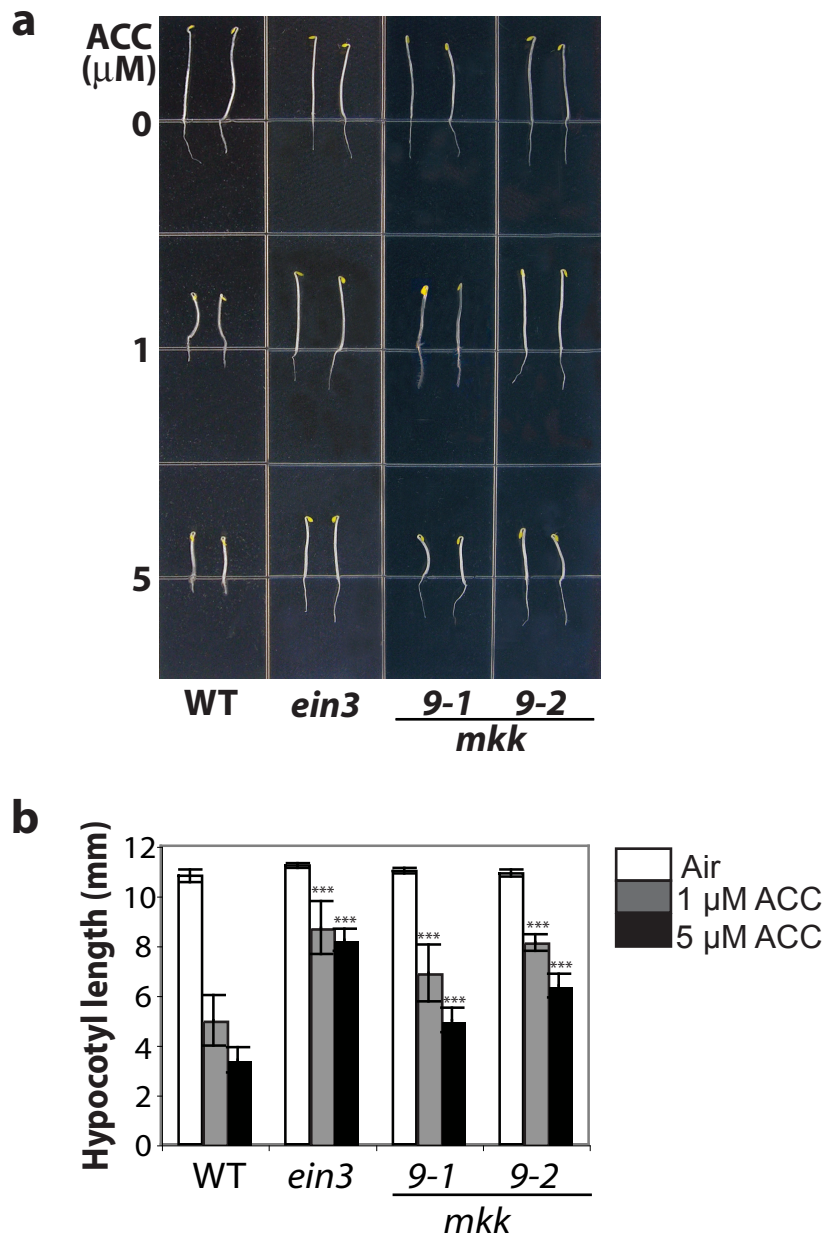


**Supplementary Figure S3 | T-DNA insertion of the *mkk* mutants.** Genomic DNA analyses of *mkk* (*kk1-9*) mutants and WT (W) by PCR. LB: T-DNA left border primer, LP/RP: primer sets for genomic DNA analyses of *MKKs*. The detailed primer sequence information is provided in Supplementary Table S1. The *mkk4*, *mkk5* and *mkk6* mutants are not null alleles.



**Supplementary Figure S4 | Analysis of *mkk9-1* and *mkk4*.** **a**, Schematic presentation of the *mkk9-1* gene structure with the T-DNA insertion. **b**, Genomic DNA PCR and RT-PCR analyses of WT and *mkk9-1*. RP/LP: Right and left primer set for *MKK9* DNA and RNA analyses. F1/R1: forward and reverse primer set for RT-PCR analysis of *MKK9*. TF/TR: forward and reverse primer set for RT-PCR analysis of *Tubulin4* as a positive control. The primer sequence information is provided in Supplementary Table S1. **c**, Genomic DNA PCR analysis of *mkk4*. A PCR control (At1g18350) was included using F7: AAGCACACGACGCACATTAAC and R7: CAGGCTCCTCACTTAAATCCC. **d**, Specific MAPK activation defect in *mkk4*. Transient mechanical stress activates strong endogenous MAPKs in WT and *mkk9*, but not in *mkk4*. *In-gel* MAPK assay was performed with minimal manipulation<sup>35,36</sup>.

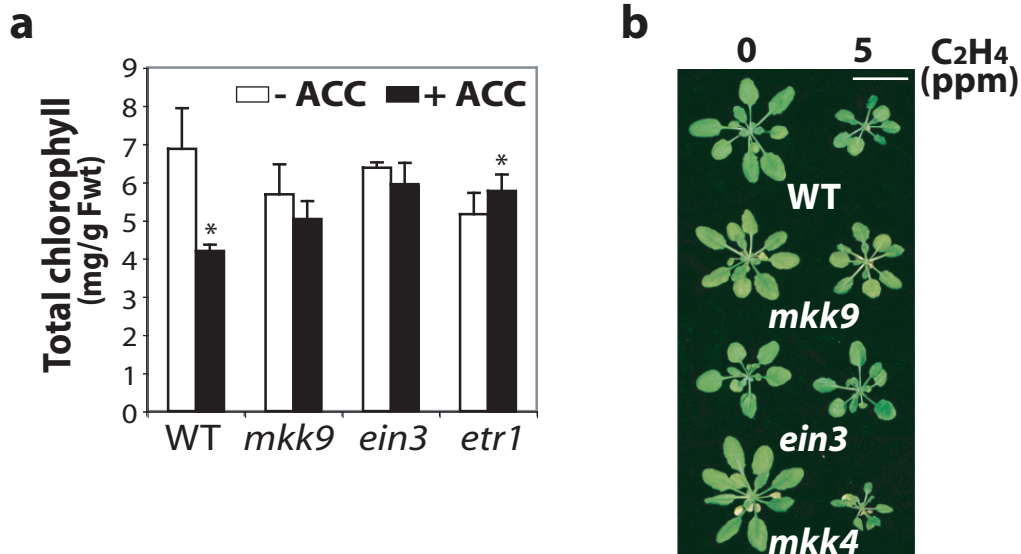




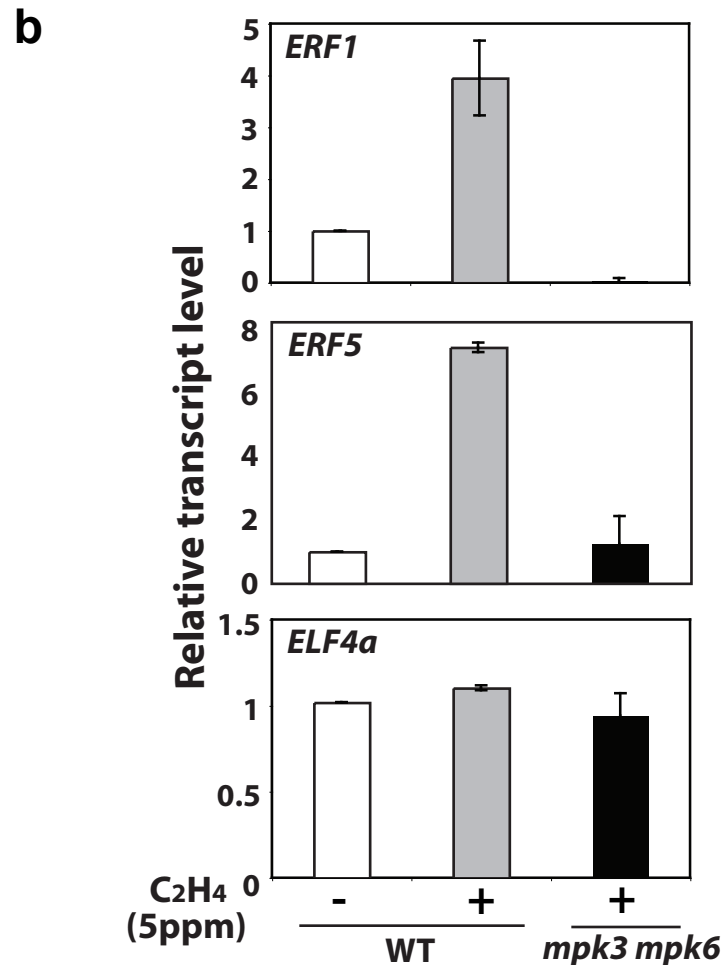
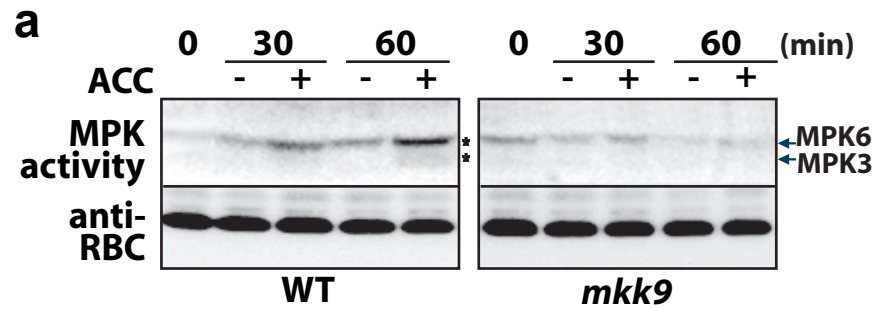
**Supplementary Figure S5 | Analysis of ACC inhibition of hypocotyl elongation**

**in *mkk9-1* and *mkk9-2*.** **a**, Two independent *mkk9* alleles (3-d-old) show ACC insensitivity. Representative seedlings from each genotype population (n=20) are shown. Experiments were repeated five times with reproducible results.

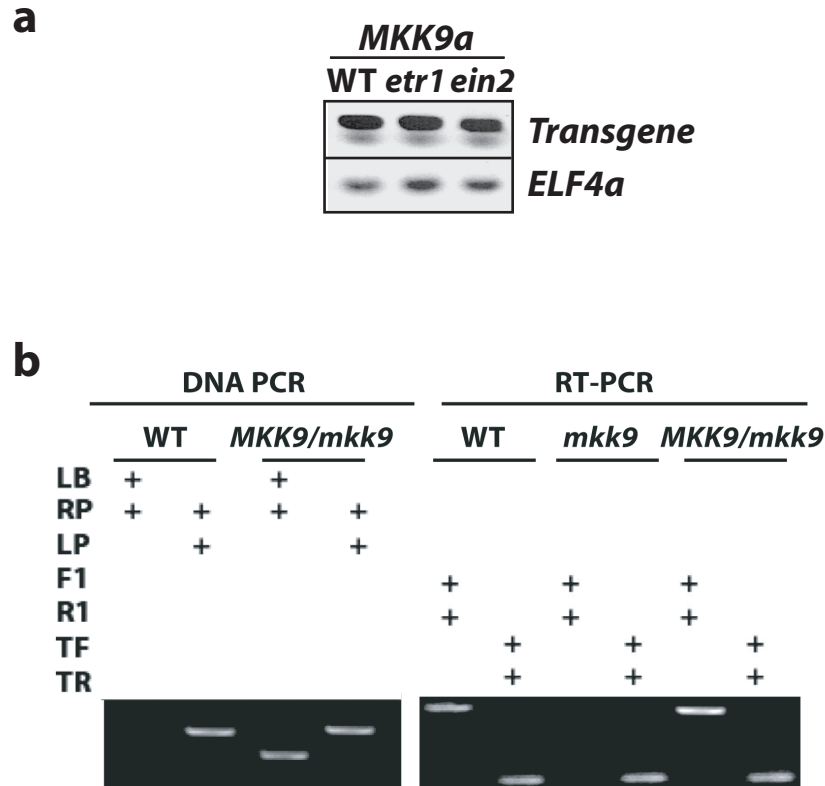
**b**, Quantitative analysis of hypocotyl elongation in the presence of ACC. Error bar, s.d. (n=20). Experiments were repeated three times with similar results. Asterisks indicate differences between WT and mutant with statistical significance at \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001 (t-test).



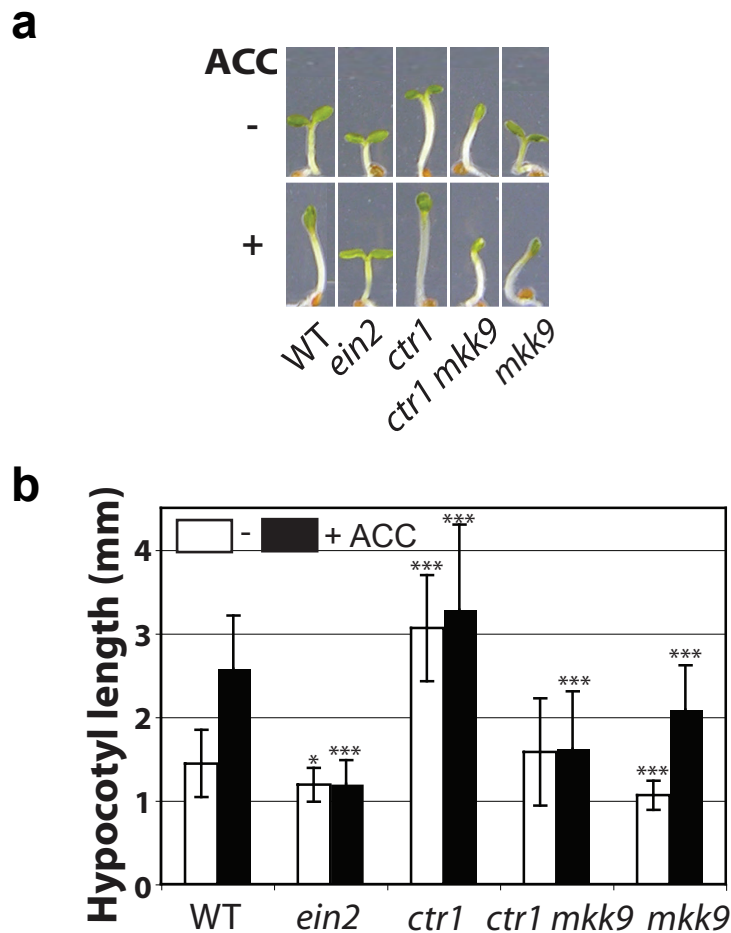
**Supplementary Figure S6 | Diverse ethylene insensitive phenotypes of the *mkk9* mutant.** **a**, Reduced chlorophyll degradation by 1  $\mu$ M ACC in the detached *mkk9* leaves. Error bars, s.d. (n=3). The experiments were repeated twice with similar results. Asterisks over bars indicate differences between WT and mutant with statistical significance at \*P<0.05 (t-test). **b**, Rosette development in *mkk9* and *ein3* is relatively resistant to C<sub>2</sub>H<sub>4</sub> inhibition. Scale bar, 10 mm. Representative seedlings from each genotype population (n=10) are shown. Experiments were repeated three times with reproducible results.



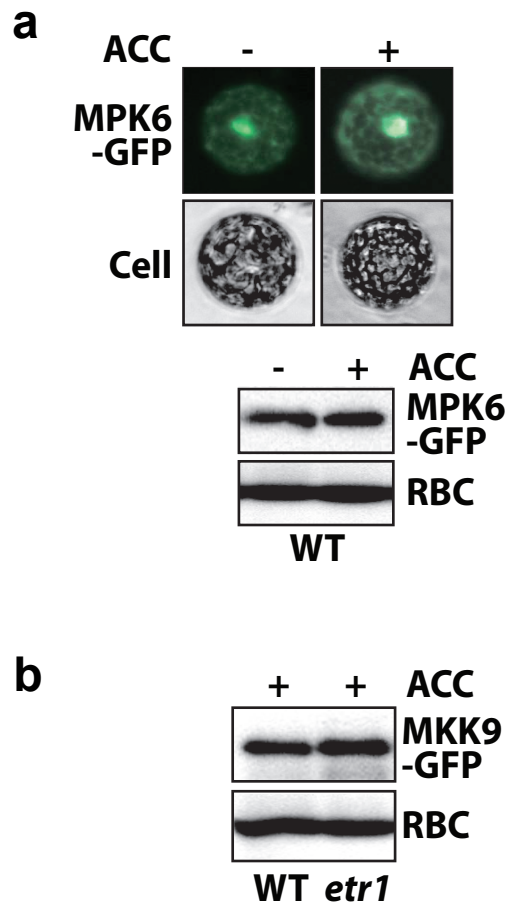
**Supplementary Figure S7 | MAPKs and immediate early gene induction by ethylene in leaves.** **a**, ACC (200  $\mu$ M) was fed through the leaf petiole for 30 or 60 min. *In-gel* MAPK assay was performed with minimal manipulation. Endogenous RBC (anti-RBC) was used as a loading control. **b**, The transcript levels of *ERF1*, *ERF5* and *ELF4a* (Elongation Initiation Factor as control) were measured by qRT-PCR after 1h C<sub>2</sub>H<sub>4</sub> treatment. The conditional *mpk3 mpk6* double mutant plants were generated by using virus-induced gene silencing<sup>44</sup> of *MPK3* in the *mpk6* T-DNA insertion mutant<sup>29</sup>. Error bars, s.d. (n=3).



**Supplementary Figure S8 | Transgenic plant analysis and complementation of *mkk9-1*.** **a**, Similar levels of *MKK9a* transgene expression by RT-PCR. The transcript of *ELF4a* is an endogenous gene control. **b**, Genomic DNA PCR and RT-PCR analyses of WT and transgenic *mkk9* lines are shown. The *mkk9-1* mutant was complemented (*MKK9/mkk9*) with a 4.2kb genomic fragment containing 1.5kb upstream of the translation start site and 1.4kb downstream of the translation stop site using a binary vector *pBin19*. The detailed primer sequence information is provided in Supplementary Table S1.



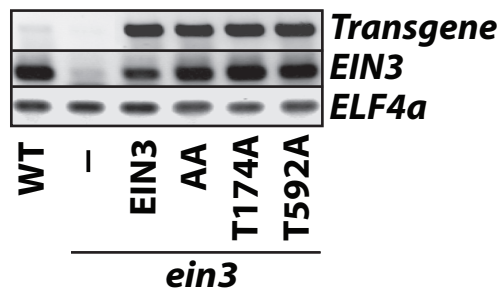
**Supplementary Figure S9 | Ethylene responses in the *ctr1 mkk9* double mutant. a**, Seedlings (5-d-old) germinated in constant light on water-agar plates (no salts or sugars) containing no (upper panels) and 50  $\mu$ M ACC (lower panels). Ethylene insensitivity is shown in *ein2*, *mkk9* and *ctr1 mkk9*. **b**, Quantitative analysis of hypocotyl elongation promoted by ACC. Error bar, s.d. (n=20). The experiments were repeated twice with similar results. Asterisks over bars indicate differences between WT and mutants with statistical significance at \*P<0.05, \*\*P<0.01 and \*\*\*P<0.001 (t-test).



**Supplementary Figure S10 | Analysis of MPK6-GFP and MKK9-GFP.**

**a**, MPK6-GFP localizes in both the cytosol and nucleus without (-) or with (+) of 200  $\mu$ M ACC. Immunoblot analysis of MPK6-GFP shows similar protein abundance. Endogenous RBC was used as a loading control.

**b**, MKK9-GFP protein abundance is similar in WT and *etr1* protoplasts in the presence of ACC(200  $\mu$ M) despite its differential nuclear localization in WT.



**Supplementary Figure S11 | Transgenic plant analysis.** Similar RNA expression of EIN3 and EIN3 mutants in transgenic *ein3* lines. The transcript of *ELF4a* is an endogenous gene control.

**Supplemental Table1. Oligonucleotides used in this study.**

Oligo name	Oligonucleotide (5' - 3')
<b>qRT-PCR primers for <i>MAPKs</i></b>	
<i>MPK1_f</i> :	ATATGACCCGAAT GCAAACC
<i>MPK1_r</i> :	TTCCATGGCATCACTATTCG
<i>MPK3_f</i> :	AATGGCCATTGATCTTGTTGA
<i>MPK3_r</i> :	GTTACAAGATTA CATATGTGGAA
<i>MPK6_f</i> :	CCCGACAGTGCATCCTTTAG
<i>MPK6_r</i> :	G TTCCTTCATCTGCTCCTCTG
<i>MKK6_f</i> :	ACA ACTAT ATGTCGCCTGAGAGG
<i>MKK6_r</i> :	CCCAA ACTCCAAATGTC ACTG
<i>MKK7_f</i> :	GTCGTGTGCTTTGGAGAACC
<i>MKK7_r</i> :	AAGGGTGACCGAGAAGCTG
<i>MKK9_f</i> :	AGGAGCTTCGTTGAGTGTTG
<i>MKK9_r</i> :	TCCCCTAACATTCTGGAGTATA
<b>qRT-PCR primers for ethylene response genes</b>	
<i>ERF1_f</i> :	GAGGAAACTCG ATGAGACG
<i>ERF1_r</i> :	GGAGCGGT GATCAAAGTCAC
<i>ERF5_f</i> :	TGGA GAGACGTTTCCGTTTG
<i>ERF5_r</i> :	TGAGGAGATAACGGCGACAG
<i>ACS2_f</i> :	AATGGACGCAGACCAATCTT
<i>ACS2_r</i> :	GCTCGGAGAAGAGGTGAGTG
<b>qRT-PCR primers for controls</b>	
<i>TF (Tubulin4)</i> :	AGGGAAAGGAAGAGAGGAAG
<i>TR (Tubulin4)</i> :	GCTGGCTAATCCTACCTTTGG
<i>ELF4F</i> :	TCATAGATCTGGTCCTTGAAAC
<i>ELF4R</i> :	GGCAGTCTCTTCGTG CTGAC
<b>T-DNA screening primers (Salk)</b>	
<i>T-DNA_LB</i> :	TGGTTCACGTAGTGGGCCATCG
MKK1 (Salk_027645)	
<i>MKK1_LP</i> :	GACAAGTCTCTTAAGTCATAACATCTCG
<i>MKK1_RP</i> :	AACATGCTATCTGCCATCTGC



MKK3 (Salk\_051970)

*MKK3\_LP*: GAACAAACGTTTTCTCATGTGTG

*MKK3\_RP*: AGAAGGATCCAGATGCTCGAC

MKK5 (Salk\_067321)

*MKK5\_LP*: TAACCAGGCAACCATCTCAAG

*MKK5\_RP*: TGGAAAGAGCGTGGAATACAC

MKK6 (Salk\_084332)

*MKK6\_LP*: CGCAGTCCTGTTTTCAAATTC

*MKK6\_RP*: CAAAAGCTTCGTTAAAGCTCTCTC

MKK9 (Salk\_146400)

*MKK9.2\_LP*: GAAACTCAACGTTCTCGGATG

*MKK9.2\_RP*: CCCAAAACCTTATGTACACGATTG

#### **T-DNA screening primers (SAIL)**

*T-DNA\_LB*: GCTTCCTATTATTCTTCCCAAATTACCAATACA

MKK2 (SAIL\_511\_H01)

*MKK2\_LP*: TCACTGGAAGGTAAAACAAGAAATC

*MKK2\_RP*: GTTAAAGCCATCCCTGACTCC

MKK4 (SAIL\_565\_A12)

*MKK4\_LP*: CATCAACCTGATTAGGTTTGAG

*MKK4\_RP*: CAGAGAGAACCGGGGAAAAG

MKK9 (SAIL\_060\_H06)

*MKK9.1\_LP*: TCCCCTAACATTCTGGAGTATA

*MKK9.1\_RP*: TCAAACCGGCGAATCTTCTTC

#### **T-DNA screening primers (SM)**

*T-DNA\_LB*: TACGAATAAGAGCGTCCATTTTAGAGTGA

MKK7 (SM\_3\_17177)

*MKK7\_LP*: AAGCACACGACGCACATTAAC

*MKK7\_RP*: CAGGCTCCTCACTTAAATCCC

#### **RT-PCR primers**

*MKK9\_F1*: ATGGCTTTAGTACGTGAACG

*MKK9\_R1*: AAGATCTTCCCGGAGAAAA

*TS\_r*: CGTATGGGTAACCAGCGTA

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