## DUOX2 BIDIRECTIONAL PROMOTER POLYMORPHISMS CONFER DIFFERENTIAL IMMUNE RESPONSES IN AIRWAY EPITHELIA

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## **ONLINE DATA SUPPLEMENT**

## SUPPLEMENTAL DATA

<u>Supplemental Table 1</u>: Distribution of SNP genotypes based on population data obtained from the 1000 Genomes project (<u>http://www.1000genomes.org/</u>). Number of individuals in each group (N) and percent of individuals carrying the specific genotype are shown.

Observed	African	Asian	European
Genotype	(246)	(286)	(381)
AA and TT	201 (81.71%)	286 (100%)	381 (100%)
AG and CT	44 (17.89%)	0	0
AG and TT	1 (0.41%)	0	0

<u>Supplemental Table 2</u>: Distribution of two-marker haplotypes (rs269855-rs2576089) in an African population. Overall haplotype frequencies are maximum likelihood estimates (42) based on the observed genotype frequencies (see Supplemental Table 1).

Haplotype	Overall frequency (%)		
A-T	90.85		
A-C	0		
G-T	0.2		
G-C	8.94		

Supplemental Figure 1: Variability in harvested plasmid levels for each time point in HBE1 cells after transfection. HBE1 cells in submerged cell culture conditions were transfected at 60-70% confluence with the bidirectional reporter plasmid and total cellular DNA was prepared at various time points after transfection. Cells were counted and underwent lysis with equal buffer volumes for each sample to ensure similar input total DNA prior to PCR amplification. Mean  $\pm$  SD of threshold cycles (Ct; inversely proportional to the log2 of plasmid DNA) measured by qRT-PCR of three separate samples per time point. Data from all experiments (three replicates for three different experiments) are shown.

Supplemental Figure 2: Chromosome 15 sequence encompassing the translation start codons (boxed) for both *DUOX2* (opposite strand) and *DUOXA2*. This region includes exon 1 and 2 of *DUOX2* and exon 1 of *DUOXA2* (black upper case) and the intervening intronic DNA (gray lower case). Various transcription start sites (TSS) for *DUOX2* (D2) or *DUOXA2* (DA2) are shown including the canonical sequence available in GenBank (www.ncbi.nlm.nih.gov/genbank) (bold arrows) or published by Puchucki or Grasberger (19, 34) (thin arrows). A CpG island region was identified using highly restrictive criteria including GC content > 60%, observed CpG/Expected CpG ratio > 0.7, and CpG island length > 200 bp. Two different CpG island prediction programs were used (31, 32) and both programs predicted highly similar CpG island regions (underline).

Supplemental Figure 3: Alignment of rat, mouse and human genomic sequence in two separate segments of the *DUOX2/A2* promoter demonstrating the conserved TATA box motif (box) for

*DUOXA2*, and the Inr (#)/DPE (\*) sites for *DUOX2* in all three species. Of note, two Inr sites were conserved in all three species. Several other DPE sites were identified within this genomic sequence (not shown), but did not align in all three species. Nucleotides are numbered starting with the ATG site of *DUOX2* and the transcription start site for each gene is bold underlined.

Supplemental Figure 4: Alignment of rat, mouse and human genomic sequence in two separate segments of the DUOX2/A2 promoter surrounding the two identified SNPs in the human genome (bold underline). Nucleotides preserved in all three species are denoted by (^). Nucleotides are numbered starting with the ATG site of DUOX2. Alignment of putative transcription factor binding sites for each SNP are shown (www.gene-regulation.com).

Supplemental Figure 5: Effect of rhinovirus (RV) infection or interferon-gamma ( $\gamma$ ) treatment on basal *DUOX2/DUOXA2* promoter activity. HBE1 cells were grown in submerged cell culture conditions and transfected with the full length promoter construct (FL) followed by infection with RV, interferon- $\gamma$ , or both for sequential measurements of *Renilla* and firefly luciferase activity. A  $\beta$ -galactosidase expression plasmid was co-transfected in all cells and used to normalize luciferase activity between experiments. (A) Cells were infected or treated for 4 hours prior to harvest at 12 hours after transfection. (B) Cells were infected or treated for 16 hours prior to harvest at 24 hours after transfection. Normalized data are presented as fold increase compared to the FL construct (mean ± SEM; n = 3).



Supp. Figure 1

1 cgctagagga gcctgatacT TGCCCGATGG ACCCAGGGAT CCAGTCAGAA GAGCTCCCAG GAGCATCAGT GCCTCTGGTC TTGCACGGAG CATGCCAACC CTGCAGCctg cggggtgagg gtgggggtgg taggtggtat gcgaaageca etgttaggge gteetetatg eeteeetet 91 181 tytteetaca getagtaetg gaggaggage accagetytt teeaettetg gaaggtaget gttagaagea teaeegagga eetteateea 271 aacgccactc tttccaggac aattggcagc tctggaggca ttgacactgt tccccatccg ccaccccatc agagagttaa cccccgacca 361 taggaaccca ctgggcagga gtcttctggg gcatgtcagt ccagggcagg actggtcaag cctcccaggg tgtgcccaat gtctccaatg 451 tttcacctcc accegeeece Aateegteag geteegette teeteeagga ggeagggaag ggaaaaaggt taetgaeetg ggagtgaggg 541 actgcagcac ccttccacaa tgaatccccc cttccccaat aaactcccct tctgcaatga acgcctgtgc atgatgggcg agggctaggg 631 tcagatccca aactctggtc taacCTGTGG TTTAGGGTGG TGTTGGGTTC AGATGTCTTC TTTCCTCTTA AAATCTTTGC TTCTGTGCTC 721 TACTTCTTGC CTTCACCCTC ACTCTTCCAG CTCCGCCGAT CCTCAGCCTC CCCGGCTGCA CTCTCACCTT TCTCTCTGGG TCCTTGGTCT  $\leftarrow$ D2 ר **D**2 D2 ר 811 CGCCACTGTG CAGGTGTCGG CTCAGGACAG ACCtgcgcca gtgtgagcat ctggacctag ggctcaccct cctgccgtgg aggtggggcc 901 cttatttgca taacctcttc cagetcagac cageceetgg getgggaeae eegtgtggea egtegeeeae getge**tataa a**aggggteee D2 ר 991 gcgcgacttc caaactCAGC GCCAACCCGC AGAACCAGGA AAGTAACGGC TACAGACAGT GAGAAATAGT TTCGCTCGCC GGCTAGAAAA DA2  $\rightarrow$ 1081 ACTCTGTCGG TACCAACCCC AGAGCGTTGA GAGCAGCCCA CCTCCACGCT TCCTTAACGG AGAGGTGCAG GACTCAGACT TCACCAGCCC 1171 ACTCGGTCCC AGCCTTGTAC GCAAAGAGAC GCCAAGGACG CGCTCTCCCG CGTCCAGGCA GCCCCAGCTT GCTGGCTTGC CTGCCCGCCT 1261 GCGTGCAGCA CTCGGCCGGC GTGCAGCATG ACCCTGTGGA ACGGCGTACT GCCTTTTTAC CCCCAGCCCC GGCATGCCGC AGGCTTCAGC 1351 GTTCCACTGC TCATCGTTAT TCTAGTGTTT TTGGCTCTAG CAGCAAGCTT CCTGCTCATC TTGCCGGGGGA TCCGTGGCCA CTCGqtaaqq

Supp. Figure 2

 $\leftarrow$  DUOX2

RATTCCCCTGCTTGCTGCCCACGCTCCTATAAAAG---AGGGTCTGCTTCTTGTGAACGC651MOUSEGCCCCTGCTTGCTGCCCACGCTCCTATAAAAGGAGGGGGTCTGCGTTTTGTGAACAC740

HUMAN CGTGTGGCACGTCGCCCACGCTGCTATAAAAG---GGGTCCCGCGCGTC-CAAACT $\underline{C}$  913



Supp. Figure 3

## ← DUOX2

RAT	ATTTCCCACATA			-CCGATTCTCC	AGGAG	265
MOUSE	ATTTCCCAAGTA			-CTGCTTCTCC	AGGAG	273
HUMAN	ATGTCTCCAATGTTTCACCTCC.	ACCCGCCCCC <b>A</b> AT	CCGTCAGGC	TCCGCTTCTCC	TCCAG	407
	^^			~ ~~~~~	~ ^^	
Put	tative TFs	===SP-1====	[A/G]			
		==Egr-1====	[A/G]			
		==Egr-2===	[G]			
		==AP-2α====	[G]			

RAT TAAGGACGGGATTTCCGTTATTTGCATGTCTCCGCCCCATCGTTTGCATGATCTCACCCA 567 MOUSE TAAGGACAGGATTTCAGTTATTTGCATGTCTCCGCCCCTTCGTTTGCATGACCTCAACCA 653 CGTGGAGGTGGGGCCC-TTATTTGCATA----ACCTCTTC----CA----GCTCA 836 HUMAN ~ ~ ~ ~ ~~ ~ ~ Putative TFs ===SRF==== [T/C] DUOXA2 ==Sox-9==[T]  $==C/EBP\alpha ===$ [T]  $=C/EBP\beta$ === [C]

Supp. Figure 4



Supp. Figure 5