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#### Table S1

			Methylation Frequency in Cancer						
Name	ENSEMBL gene ID	CpG ID (Illumina)	Br	Col	Pro	Glio	Lu	AML	Ovr
CDH1	ENSG0000039068	cg11255163;cg23989635	0%	0%	0%	7%	0%	15%	0%
MLH1	ENSG0000076242	cg00893636	1%	10%	0%	0%	0%	0%	0%
BRCA1	ENSG0000012048	cg04658354;cg08993267	2%	0%	0%	0%	0%	0%	12%
RB1	ENSG00000139687	cg10552385;cg24937706	0%	0%	0%	0%	0%	0%	0%
APC	ENSG00000134982	cg16970232;cg20311501;cg21634602	28%	33%	51%	0%	44%	1%	9%

#### Table S1

#### Promoter hypermethylation frequencies at known tumour suppressor genes

We created a list of known tumour suppressor genes which when mutated in the germline are associated with pre-disposition to cancer. This was then limited to genes which had probes within CGIs and within 200bp of TSSs. Reported are these probe IDs and the % of tumours in each tissue which were methylated (mean beta > 0.3). Br=Breast, Col=Colorectal, Pro=Prostate, Glio=Glioblastoma, Lung=Lung, Ovr=Ovarian.



#### Methylation levels at hypermethylation prone genes vary between cancer types

A. Numbers of frequently hypermethylated genes vary between tumour types. Shown is a bargraph of the number of frequently hypermethylated genes found in each of the 7 tumour types analysed.

B. Methylation levels at hypermethylation-prone genes vary between tumour types. Shown is a boxplot of the median methylation levels found at the 1009 hypermethylation prone genes in the 7 tumour types analysed.



Tissue Specificity

# Genes frequently hypermethylated in multiple cancer types have regulated expression patterns in normal tissues

Histograms showing the distribution of tissue-specificity scores observed for different gene sets. Specificity scores for different gene sets were compared using a Wilcoxon rank sum test as indicated. (\*\*\* < 0.001). All as Figure 2A but using alternative gene sets.

A. Genes prone to hypermethylation in multiple cancer types. Gene sets are as Figure 2A but specificity scores were calculated from microarray expression data rather than RNA-seq.

B. Genes prone to hypermethylation as defined using alternative parameters. On the left the threshold used to define genes as hypermethylated was varied (see methods for details). On the right the frequency of hypermethylation required to be defined as frequently hypermethylated in a given cancer was varied.

C. Genes frequently hypermethylated in each of the individual cancer types examined in this study. Methylation resistant genes were defined as genes never methylated in that cancer type.

D. Genes aberrantly hypermethylated in colorectal cancer defined from datasets using alternative profiling methods [34-36]. Genes aberrantly hypermethylated in colorectal tumours were defined from either MBD-seq data (Illingworth *et al* and Xu *et al*) or whole-genome bisulfite sequencing (Berman *et al*). The Illingworth *et al* graph shows genes frequently hypermethylated in colon cancer, the Xu *et al* graph shows genes that are significantly hypermethylated in colon cancer and the Berman *et al* graph shows genes that are prone to hypermethylation in colon cancer (see methods for details).

E. Genes mutated or not mutated in breast cancer. Mutated genes were defined from the COSMIC database [69].



### Repeat densities and evolutionary conservation do not determine hypermethylation susceptibility in cancer

A. Transcriptional start sites are depleted of repetitive elements. Shown are graphs of the frequency of LINEs, SINEs and LTRs at 1Kb intervals around CGI or non-CGI TSSs (as Figure 3A).

B. Hypermethylation prone promoter regions are evolutionarily conserved. Shown are graphs of the level of conservation found in 500bp intervals around genes that are hypermethylated in colorectal tumours as defined by alternative profiling methods (as Figure 3B) [34-36]. The significance of observed differences between hypermethylated and non-hypermethylated genes was assessed using a Wilcoxon rank sum test for the scores -/+ 2Kb from the TSSs (\*\*\* p < 0.001).

C. Repeat densities do not determine hypermethylation susceptibility. Shown are heatmaps indicating the presence (red) or absence (white) of repeats in the region around hypermethylation prone and resistant TSSs. Repeat presence was assessed in 1Kb intervals - /+5Kb from each TSS and TSSs are ordered by their repeat density in this region.

D. Evolutionary conservation does not determine hypermethylation susceptibility. Shown are boxplots of the degree of evolutionary conservation found -/+2Kb from hypermethylation prone and resistant TSSs (measured as Figure 3B). The significance of differences between the distributions was assessed using Wilcoxon rank sum tests.



## Expression patterns in normal tissues explain differential susceptibility to hypermethylation in cancer

A. CM and VM gene promoters are both depleted in repetitive elements. Shown are graphs of the frequency of LINEs, SINEs and LTRs at 1Kb intervals around different sets of TSSs (as Figure 3A). The significance of the differences in densities were determined using Fisher's exact tests for the repeat counts in regions -/+ 2Kb from the TSSs (\*\*\* p < 0.001, \*\* p < 0.01 and \* p < 0.05).

B. CM and VM gene promoters are evolutionarily conserved. Shown are graphs of the level of conservation found in 500bp intervals around different sets of TSSs (as Figure 3B). The significance of observed differences were assessed using a Wilcoxon rank sum test based on the scores for the region -/+ 2Kb from the TSSs.

C. Specificity of expression in normal tissues is significantly correlated to the frequency of aberrant hypermethylation in cancers arising in different tissues. Shown is a boxplot of tissue-specificity scores for genes which are frequently aberrantly hypermethylated in different numbers of cancer types. A significant correlation exists between the two (Spearman's correlation, Rho=0.238, p= $5 \times 10^{-14}$ ).

D. Genes hypermethylated in colon cancer are repressed in normal colon tissue. Shown are boxplots of the expression levels in normal colon of genes which are found to be aberrantly methylated or not in colorectal tumour by either MDB-seq or whole-genome bisulfite sequencing. Differences between gene groups were tested using Wilcoxon rank sum tests.