CHEMBIOCHEM

Supporting Information

© Copyright Wiley-VCH Verlag GmbH & Co. KGaA, 69451 Weinheim, 2011

Peptide Structure Stabilization by Membrane Anchoring and its General Applicability to the Development of Potent Cell-Permeable Inhibitors

Liv Johannessen,^[a] Jarrett Remsberg,^[a] Vadim Gaponenko,^[b] Kristie M. Adams,^[c] Joseph J. Barchi, Jr.,^[c] Sergey G. Tarasov,^[d] Sheng Jiang,^[d] and Nadya I. Tarasova^{*[a]}

cbic_201000563_sm_miscellaneous_information.pdf



Supplementary Figure S1. Inhibition of SK-Mel2 melanoma cells growth by Hedgehog pathway inhibitor SMOi2-43 (Ac- $(\epsilon$ -Pal)-KLTYAWHTSFKAL-NH2). Cell number was determined with the help of MTT assay after 48 hours exposure to the compound. Negative values in cell number reflect cell death.



Supplementary Figure S2. Cytotoxic effects of IGF-1R JM analogs on MCF-7 cells grown in serum-free medium and stimulated to grow solely with human recombinant IGF-1 (10 μ g/ml). The cells were exposed to compound for 48 hours. Relative number of live cells number was determined with the help of MTT assay



Supplementary Figure S3. Toxicity assay in multiple cancer cell lines showed that sensitivity to IGFR1 JM analog 16 correlated well with IGF1 expression levels. Toxicity was determined by MTT assay after 48 hours exposure to the compound.



Supplementary

Supplementary Figure S4. Juxtamembrane domain analogs had low activity in inhibition of recombinant kinase domain of IFG1R in cell-free environment. Kinase activity was assayed with the help of OMNI recombinant kinase assay kit 4 (Biosource) that utilizes fluorogenic peptide substrate phosphorylated by IFG1R kinase. Recombinant human phosphorylated IGF1R kinase was a kind gift from Mrs. Carrie J. Saucedo (Molecular Target Development Program, NCI-Frederick).



Supplementary Figure S5. Lipopeptide inhibitors do not cause cell lysis. Cell membrane integrity was evaluated with the help of Promega CytoTox-ONETM assay that measures lactate dehydrogenase (LDH) released from cells with damaged membranes. Fluorogenic LDH substrate was added to MCF-7 cells seeded in black 96 wells plates and exposed to indicated concentrations of compounds for 1 hour. After 10 min incubation with the substrate and addition of the stop solution, fluorescence was measured on BMG FluoStar plate reader using 560 nm excitation and 590 nm emission. Presented data is an average of six measurements. Triton X-100 was used as positive control that causes complete release of intracellular LDH.









Supplementary Figure S6. Biodistribution of Hedgehog lipopeptide antagonist SMOi2-17 in mice. Distribution was evaluated with the help of ³H-labeled compound upon injections of 2μ Ci per animal or 1.1 mg/kg (610 nmol/kg). The tritiated compound was generated by AmbiosLabs

by catalytic substitution of protons of hydrocarbon chains of the peptide

(<u>http://ambioslabs.com/label.php</u>). Female wild type 8–12-wk-old BALB/c mice were provided by the Animal Production Area of the National Cancer Institute (NCI). Mice were sacrificed 30 min after administration. Organs were collected, tissue was homogenized with razor blades and radioactivity counted in scintillation counter. The data is an average for samples collected from groups of three animals.

NCI-Frederick is accredited by the American Association for the Accreditation of Laboratory Animal Care International and follows the Public Health Service Policy for the Care and Use of Laboratory Animals. Animal care was provided in accordance with the procedures outlined in the *Guide for Care and Use of Laboratory Animals* (National Research Council; 1996; National Academy Press; Washington, D.C.).

| Compound | Sequence | Calculated | Actual Molecular |
|----------|---|------------|------------------|
| - | - | Molecular | Weight |
| 1 | Dol HDEDNNEDI CNC NIL | Weight | 1615 0 |
| 1 | Pal-HRRENNSRLGNG-NH2 | 1043.9 | 1045.0 |
| 2 | | 14/4.8 | 14/4.6 |
| 3 | Pal-HRKRNNSRLGNGVLYASVN-NH ₂ | 2392.8 | 2392.5 |
| 4 | $Pal-HRKRNNSRLGNGVLYASVNPEYFSAA-NH_2$ | 3158.6 | 3158.2 |
| 5 | $Pal-HRKRNNSRLGNGVLYASV-NH_2$ | 2278.7 | 2277.8 |
| 6 | $Pal-VHRKRNNSRLGNGVLYASV-NH_2$ | 2377.8 | 2378.0 |
| 7 | $Pal-HRKRNNSRLGNGVLYAS-NH_2$ | 2179.6 | 2178.8 |
| 8 | Pal-HRKRNNSRLGNGVLYA-NH ₂ | 2092.5 | 2091.6 |
| 9 | Pal-HRKRNNSRLGNGVLY-NH ₂ | 2021.4 | 2021.6 |
| 10 | Pal-HRKRNNSRLGNGVL-NH ₂ | 1858.3 | 1858.2 |
| 11 | Pal-HRKRNNSRLGNGV-NH ₂ | 1746.1 | 1744.4 |
| 12 | Pal-RKRNNSRLGNGVLYASVN-NH ₂ | 2255.7 | 2255.2 |
| 13 | Pal-KRNNSRLGNGVLYASVN-NH ₂ | 2099.5 | 2100.8 |
| 14 | Pal-RNNSRLGNGVLYASVN-NH ₂ | 1971.3 | 1970.6 |
| 15 | Pal-NNSRLGNGVLY-NH ₂ | 1443.7 | 1443.2 |
| 16 | Pal-RNNSRLGNGVLY-NH ₂ | 1599.9 | 1599.4 |
| 17 | Pal-NSRLGNGVLY-NH ₂ | 1329.6 | 1329.2 |
| 18 | AC-YLVGNGLRSNNK-(ϵ -Pal)[All D] | 1770.1 | 1770.4 |
| 22 | Pal-YLVGNGLRSNNR-NH ₂ | 1599.6 | 1599.4 |
| 23 | Pal-HRKRNNSRLGNGVLYASVNP-NH ₂ | 2489.1 | 2489.2 |
| 24 | Pal-VFHRKRNNSRLGNGVLYASVN-NH ₂ | 2639.1 | 2639.0 |
| 27 | Pal-KRNNSRLGNGVLY-NH2 | 1728.1 | 1727.4 |
| IR-2 | Pal-RQPDGPLGPLY-NH ₂ | 1449.8 | 1450.0 |
| SMOi2-43 | Ac-(ϵ -Pal)-KLTYAWHTSFKAL-NH2 | 1845.2 | 1845.2 |
| SMOi2-9 | AC-LTYAWHTSFKAL-NH2 | 1477.6 | 1477.3 |
| SMOi2-27 | Pal-C(Rhod)LTYAWHTSFKAL-NH2 | 2259.0 | 2258.8 |
| | | | |

Table S1. Analytical data for compounds synthesized*.

*Molecular weight was determined by ion-spray mass spectrometry utilizing Agielent 1100 LC/MS

system.

















Figure S7. **HPLC analysis of the key compounds**. Chromatography was performed on Microsorb-MW 300A C8 column (Varian) in 0-100% 20 min gradient of 0.1 % (v/v) trifluoroacetic acid in water/acetonitrile containing 0.1% trifluoroacetic acid, flow rate 1 ml/min. Peptides were detected by UV monitoring at 228, 256 and 280 nm.