## **Supporting Information**

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## **SI Materials and Methods**

**Two-Step Carcinogenesis.** A single dose of DMBA [7,12-Dimethylbenz(a)anthracene] (100 nmol in acetone/100  $\mu$ L acetone; Sigma-Aldrich) was topically applied on the shaved back skin, followed by biweekly TPA (12-O-Tetradecanoylphorbol-13-acetate) applications (100 nmol in DMSO/100  $\mu$ L acetone; Sigma-Aldrich) (1). Some mice cohorts were treated with a batch of DMBA with reduced mutagenic potency (indicated as "low dose DMBA" in Fig. 1*B* and Fig. S3). Tumor number was assessed once a week. A tumor was defined as a dome-forming lesion of at least 1-mm diameter that persisted for two consecutive weeks or more (2). Trials were terminated after about 6 mo or when disease reached the endpoint criteria. Tumor and tissue samples were collected, immediately frozen in liquid nitrogen, and kept frozen until use. For histological analyses, tissues were fixed with 4% paraformaldehyde in PBS, pH 7.4, and then embedded in paraffin.

**Preparation of Primary Murine Cells.** Tail skin was peeled off the underlying tissue and digested overnight at 4 °C in 8 mg/mL Dispase II (Roche). The epidermal sheets were separated from

 Gebhardt C, et al. (2008) RAGE signaling sustains inflammation and promotes tumor development. J Exp Med 205(2):275–285. the underlying dermis and both were digested in 0.05% trypsin, 1 mM EDTA (Gibco) for 10 min at 37 °C. The resulting dermal and epidermal cells were either prepared for Western blot analysis or the epidermal suspension was filtered through a 100-µm cell culture mesh and cultured in Cnt-57 basal keratinocyte medium (Cellntec) on collagen-coated dishes (Purecol; Nutacon).

Bone marrow-derived macrophages and dendritic cells were differentiated from tibial and femoral bone marrow cells, as described previously (3, 4)

**Immunohistochemistry.** For immunohistochemistry, 4- $\mu$ m sections were cut from the paraffin blocks of the paraformaldehyde-fixed human tissue samples obtained from University Hospital Tübingen after approval from the local ethics committee (University of Tuebingen; 547/2011BO2). After pretreatment with 10 mM citrate buffer (100 °C, 5 min) for epitope retrieval, sections were incubated with rabbit anti-apoptosis-associated speck-like protein containing a CARD (ASC) (2  $\mu$ g/mL; Adipogen) followed by HRP-labeled secondary antibody and visualized by HRP (DAKO). Nuclei were counterstained with hematoxylin.

- Didierlaurent A, et al. (2006) Tollip regulates proinflammatory responses to interleukin-1 and lipopolysaccharide. *Mol Cell Biol* 26(3):735–742.
- Gross O, et al. (2006) Card9 controls a non-TLR signalling pathway for innate antifungal immunity. Nature 442(7103):651–656.



**Fig. S1.** Tumors from ASC knock-out mice do show no difference to WT tumors in inflammatory cell infiltration and cell death. Histology was performed on tumors from WT and ASC<sup>-/-</sup> mice at trial termination. H&E and TUNEL stainings were performed to visualize cellular infiltrate into skin and apoptotic cells. (Scale bars, 100  $\mu$ m.)

<sup>1.</sup> Fürstenberger G, Kopp-Schneider A (1995) Malignant progression of papillomas induced by the initiation—Promotion protocol in NMRI mouse skin. *Carcinogenesis* 16 (1):61–69.



Fig. S2. Generation of floxed ASC mice. The genomic and the targeted locus are shown. Exons are highlighted as black boxes. The targeting vector contains a PGK-neomycin resistance cassette (gray boxes) and introduces loxP sites to excise the entire ASC gene.



**Fig. S3.** The phenotype of susceptibility to epithelial skin cancer of  $ASC^{-/-}$ ,  $K14-ASC^{-/-}$ , and  $LysM-ASC^{-/-}$  is reproducible in independent experiments. CD11c- $ASC^{-/-}$  do not show a phenotype in the susceptibility to epithelial skin cancer. (A)  $ASC^{-/-}$  (littermate WT n = 7,  $ASC^{-/-} n = 7$ ), (B)  $K14-ASC^{-/-}$  (littermate K14-WT n = 10,  $K14-ASC^{-/-} n = 9$ ), (C)  $LysM-ASC^{-/-}$  (littermate LysM-WT n = 8,  $LysM-ASC^{-/-} n = 6$ ), and (D)  $CD11c-ASC^{-/-}$  (littermate CD11c-WT n = 9,  $CD11c-ASC^{-/-} n = 9$ ) mice were treated with the DMBA/TPA protocol, and tumors were recorded. Tumors larger than 1 mm were counted. Data are expressed as mean  $\pm SEM$ ,  $*P \le 0.05$ ,  $**P \le 0.01$ ,  $***P \le 0.001$ .



**Fig. 54.** Caspase-1<sup>-/-</sup> keratinocytes do not show differences in BrdU staining. After 5 d in culture in the presence or absence of TPA, TNF, or EGF WT and caspase-1<sup>-/-</sup> primary murine keratinocytes were incubated for 30 min BrdU, stained with an anti-BrdU antibody, and analyzed by fluorescent microscopy. \* $P \le 0.05$ ; \*\* $P \le 0.01$ .



Fig. S5. ASC knock-down achieved through siRNA transfection in HaCaT cells. HaCaT cells were transfected with 20 nM, 50 nM, and 100 nM of siRNA against ASC and analyzed at time 72 h for ASC expression by Western blot. Tubulin was used as a loading control.