

## Supplemental information:

### Supplemental figure legends:

**Supplemental Figure 1:** Phylogenetic tree constructed using the program clustalw (<http://clustalw.genome.jp/>) showing the similarity between the different classes of chitin synthases among Basidiomycetes and Ascomycetes. Abbreviations: Af=*Aspergillus fumigatus*; An=*Aspergillus nidulans*; Ao=*Aspergillus oryzae*; Um=*Ustilago maydis*; Cn=*Cryptococcus neoformans*; Bf=*Botryotinia fuckeliana*; Fo=*Fusarium oxysporum*; Ed=*Exophiala dermatitidis*.

**Supplemental Figure 2:** Targeted replacement of *A. fumigatus* *CSMA* and *CSMB* genes. **(A)**. Physical maps of the genomic regions and strategy for the construction of the different disruption vectors designated as pCJ-E1, pCJ-Eb2, pCJ-D2 and pCJ-F4. **(B)**. Analysis of the transformants A30 ( $\Delta csmA$ ), B1 and B7 ( $\Delta csmB$ ), D22 and D78 ( $\Delta csmA/\Delta csmB$ ) by Southern blotting.

**Supplemental Figure 3:** Construction of the  $\Delta csmA/\Delta csmB$  strain by using the self excising  $\beta$ -rec/six blaster cassette in a  $\Delta akuB^{KU80}$  background: **(A)**. Schematic representation of genotypes after *CSMA* replacement, subsequent marker excision and *CSMB* replacement. Restriction sites and hybridizing probe (black bar) are schematically indicated; **(B)**. Southern analysis of the genomic DNA isolated from corresponding  $\Delta csmA$  and  $\Delta csmAx$  strains. The calculated sizes of hybridization signals by using the probe 1 are specified of DNA fragments are specified; **(C)**. Southern analysis of the genomic DNA isolated from corresponding  $\Delta csmAx/\Delta csmB$  strains. The calculated sizes of hybridization signals by using the probe 2 are specified.

**Supplemental Figure 4:** Ectopic integration of the *CSMA* and *CSMB* genes in the single and double mutants. R1: mutant  $\Delta csmA$  transformed with *CSMA*; S5: mutant  $\Delta csmB$  transformed with *CSMB*; pB1, pB4 y pB7: double mutant  $\Delta csmA/\Delta csmB$  transformed with *CSMB* (genotype  $\Delta csmA/CSMB$ ); pA1, pA3 double mutant  $\Delta csmA/\Delta csmB$  transformed with *CSMA* (genotype  $CSMA/\Delta csmB$ ). Panel A is hybridized with a *CSMB* probe and Panel B with a *CSMA* probe after digestion of the genomic DNA with *XhoI*, *EcoRI* and *SpeI*.

**Supplemental Figure 5:** Expression of all chitin synthase genes in the vegetative mycelium grown for 16 h in YG medium.

**Supplemental Figure 6:** Expression of all chitin synthase genes in the conidiating mycelia (grown for 24 h in aerial conditions) of the parental,  $\Delta csmA$  and  $\Delta csmA/\Delta csmB$  strains.

**Supplemental Figure 7:** **(A)**. Sizes of the colonies of the  $\Delta csmA$ ,  $\Delta csmB$ ,  $\Delta csmA/\Delta csmB$  and parental strains when 10-months old (1-3) or 2-weeks old conidia (4-6) were spotted on the malt-agar at three different concentrations (per spot: 1 & 4 – 5000, 2 & 4 – 2500 and 3 & 6 – 1250 conidia, respectively). Note that the colony diameter of the  $\Delta csmA/\Delta csmB$  double mutant was highly reduced with 10-month old conidia due to the loss of viability of the conidia over storage **(B)**. Survival of the conidia during storage of the slants over time at RT.

**Supplemental Figure 8:** **(A)**. Ten-fold conidial dilutions ( $2 \times 10^6$ ) of the  $\Delta csm$  mutants and the parental strains were spotted on YG plates with 0.1  $\mu\text{g/ml}$  of caspofungin (CAS), anidulafungin (ANF) and micafungin (MCF). Plates were incubated 3-days at 28°C; **(B)**.

Sensitivity of the  $\Delta csm$  mutants and the parental strains to other drugs (itraconazole, voriconazole and amphotericin B (AMB)) and calcofluor white. Selected concentrations showing an inhibitory effect are presented. YG plates were incubated 48 h at 37°C.

**Supplemental Figure 9:** Lack of correlation between the phenotypes of the *CHS* mutant and their affiliation to a family defined by BLAST homologies. The bar-graph showed the number of  $\Delta chs$  mutants generated in the Ascomycetes, Basidiomycetes, yeasts and these mutants with phenotypes.

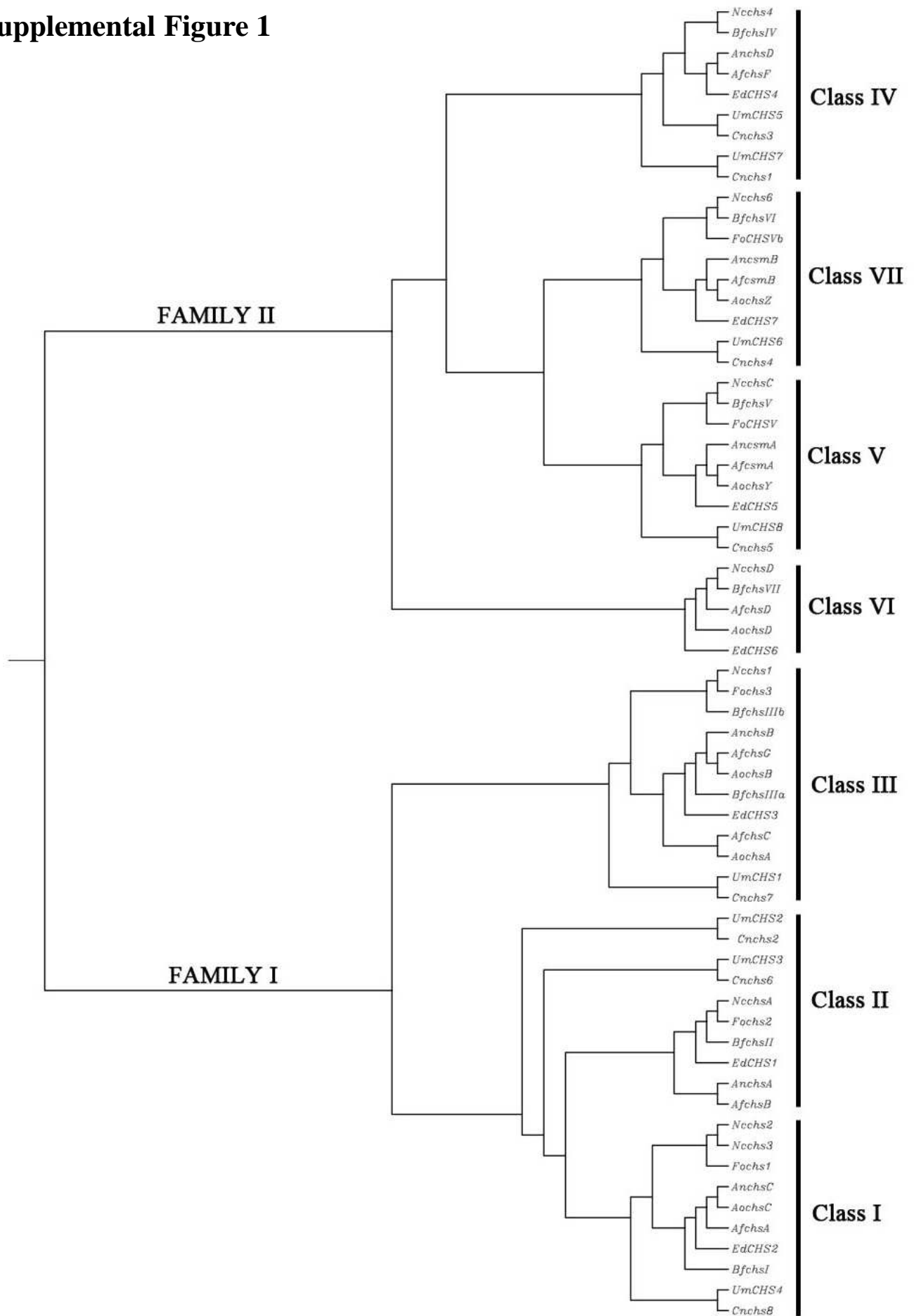
**Supplemental Table 1:** Primers used to construct the  $\Delta cms$  mutant strains and to verify the complementation of the  $\Delta cmsA/\Delta csmB$  double mutant strain

<b>Primers used for the deletion of CSMA and CSMB (Strategy 1)</b>	
AfV-A1	5'-TAGCGGCCGCGTCCCTAGCACCTAATATTCC-3' <i>NotI</i>
AfV-A2	5'-TCGATGGATCCTATCTAGAGAGAAATGCCGCGTAAGATAG-3' <i>BamHI</i> <i>XbaI</i>
AfV-A5	5'-GCTAAGGATCCATGTTAACGATTCCAATGCAACAGCTTTG-3' <i>BamHI</i> <i>HpaI</i>
AfV-A6	5'-AAGCGGCCGCGGATACATTAGCCGCTGC-3' <i>NotI</i>
AfVB-B1	5'-TTGCGGCCGCGACCTAGGTCTGAAGCGTTTGG-3' <i>NotI</i>
AfVB-B2	5'-CGATCGGATCCAGTCTAGAGTACCTGATCTACCGTGTCGC-3' <i>BamHI</i> <i>XbaI</i>
AfVB-B5	5'-GCTAAGGATCCATGTTAACGAGGACGTCACAAGATA-3' <i>BamHI</i> <i>HpaI</i>
AfVB-B6	5'-AAGCGGCCGCGCAAAGTCTGGTACTGTCC-3' <i>NotI</i>
<b>Primers used for the deletion of CSMA and CSMB (Strategy 2)</b>	
5'csmAforw	5'-AATTCGAGCTCGGTACTGCGCACCTAATATTCCTAAGAGTTC-3'
5'csmArev	5'-GGACCTGAGTGATGCTTCAAATCTGAGTACGAATAATG-3'
csmA3'forw	5'-TGGTCCATCTAGTGCATCGCCTCATCCACTGAGAATGG-3'
csmA3'rev	5'-GCCAAGCTTGCATGCCTGCGCAGAATGGAAAGATGACATCGTCC-3'
forw csmA	5'TCAGATTTGAAGCACCTCCG3'
5'csmBforw	5'AATTCGAGCTCGGTACTGCGCACGATCCGCACCCCAAAC3'
5'csmBrev	5'GGACCTGAGTGATGCTTGGCTTTCACGATGACCC3'
csmB3'forw	5'TGGTCCATCTAGTGCCATGCGATTTGTGGTACTGG3'
csmB3'rev	5'GCCAAGCTTGCATGCCTGC GCAGCCGTTGTTCCACC3'
forw csmB	5'TGTCTATTCAGTCATTCAGC3'
Sv630	5'-TCGGAGAACACCTTGCTGACG3'
<b>Primers used to verify complementation of the <math>\Delta cmsA/\Delta csmB</math> mutant</b>	
sE_F	GATACCTACAACCTTCCTACC
sE_R	CATCTCTAGCTCCAGCGG
sEb_F	CAGCCGTGATCTCGATGG
sEb_R	GTTTCATAGAACCAGTCAGG

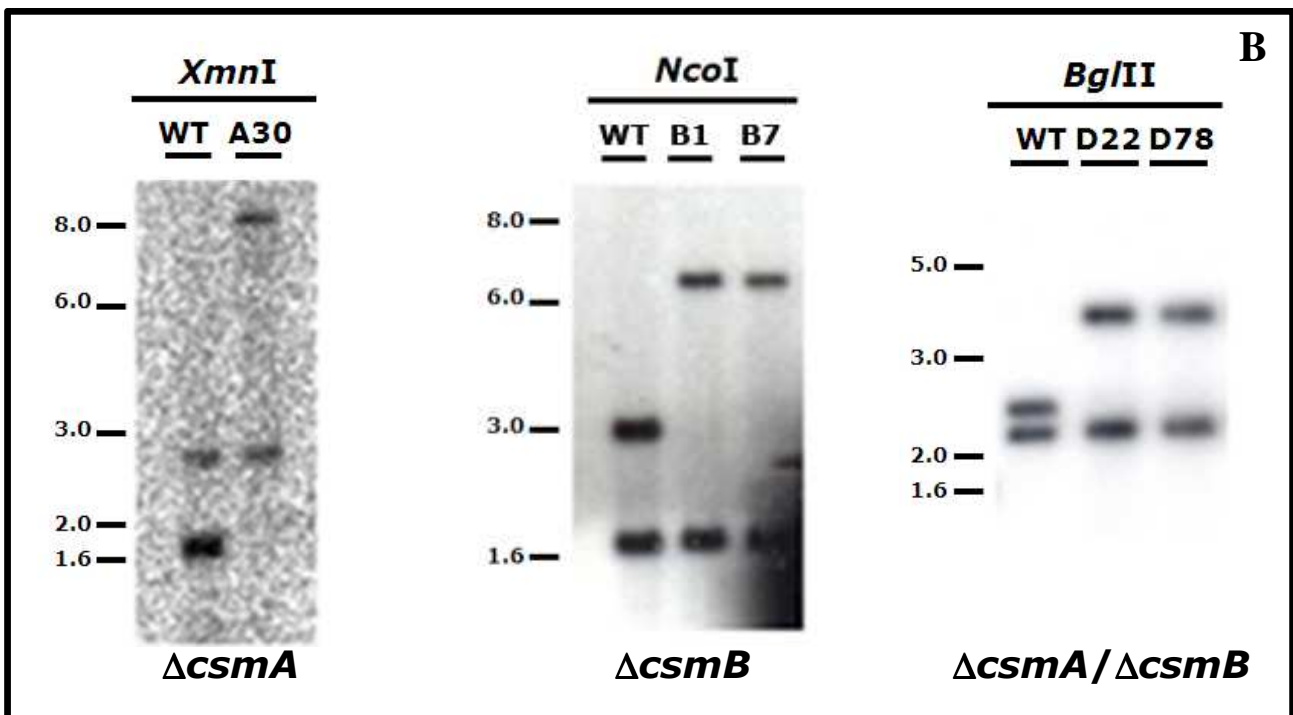
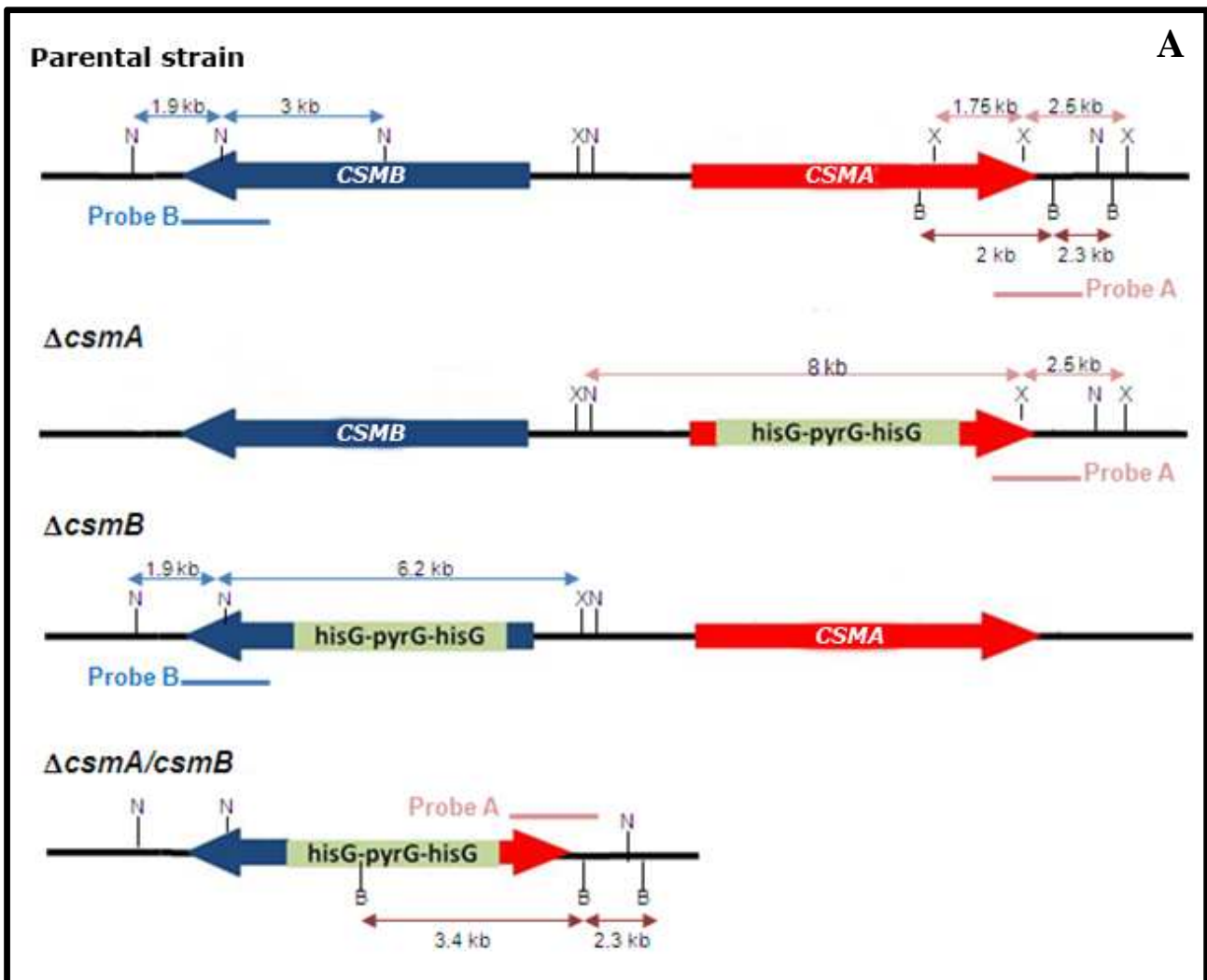
**Supplemental Table 2:** Primers used for Quantitative RT-PCR

<b>Name</b>	<b>Sequence</b>
<i>CHSAa</i> (AFUA_2G01870)	ATGCGACGGATGATGACAGG
<i>CHSAb</i> (AFUA_2G01870)	ACGACCAGGAACCACATTGC
<i>CHSBa</i> (A FUA_4G04180)	GCGGCGGACTGGACAAGG
<i>CHSBb</i> (A FUA_4G04180)	ACAATGGAGCAGGTGGCAAGG
<i>CHSCa</i> (AFUA_5G00760)	GGTGCCGAGTGCGATTCAG
<i>CHSCb</i> (A FUA_5G00760)	CGTAGGTTGTAGCCGTTGCG
<i>CHSDa</i> (A FUA_1G12600)	GGACCGAGAGCCGATGCC
<i>CHSDb</i> (A FUA_1G12600)	GCCTTGAGCCTTAGCCAGTTC
<i>CSMAa</i> (A FUA2G13440)	TGTCATTGTCAAGGTCGGAAAGC
<i>CSMAb</i> (A FUA_2G13440)	CACGGTTCAGGAATCTCATCAGC
<i>CSMBa</i> (A FUA_2G13430)	AGGAGGGCGGAGGATGGATG
<i>CSMBb</i> (A FUA_2G13430)	CAAGTGCGGTGAAGGCTATGC
<i>CHSFa</i> (A FUA_8G05630)	ACTTTGACCTTCTGAACTGGCTTG
<i>CHSFb</i> (A FUA_8G05630)	TCCTCTTATCTTCTCCCGCTTGG
<i>CHSGa</i> (A FUA_3G14420)	TGGTGCGTGCCTTCAGTGG
<i>CHSGb</i> (A FUA_3G14420)	ACCGAATGTAGCAGCGAGAGC
<i>TEFa</i> (A FUA_1G06390)	CCATGTGTGTCGAGTCCTTC
<i>TEFb</i> (A FUA_1G06390)	GAACGTACAGCAACAGTCTGG

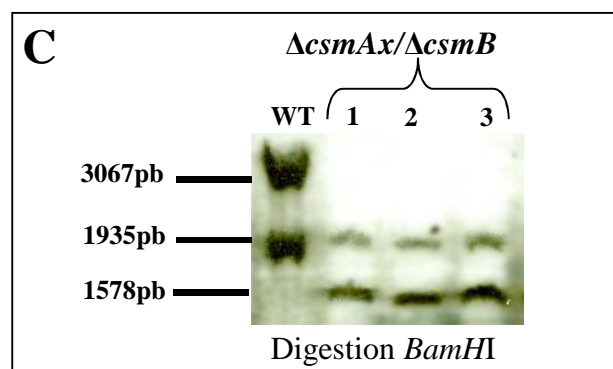
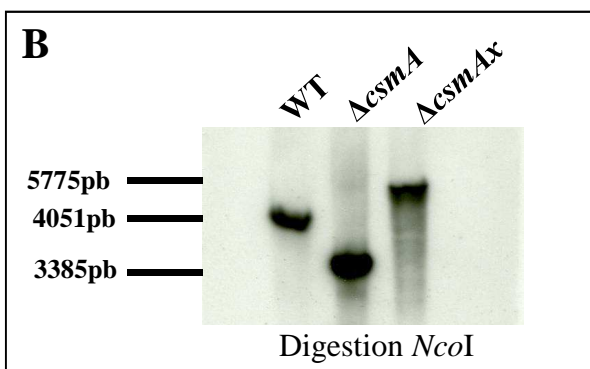
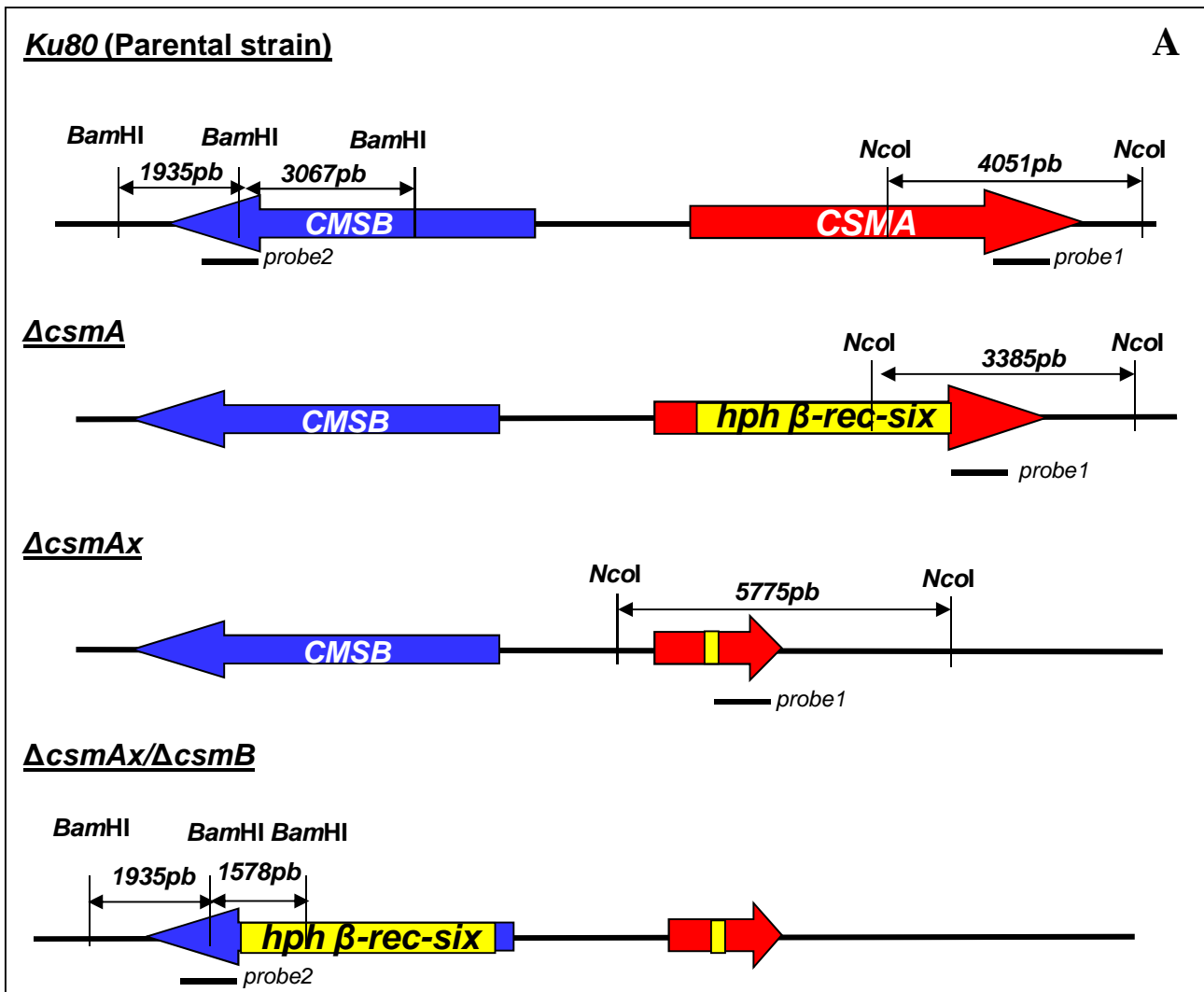
**Supplemental Figure 1**



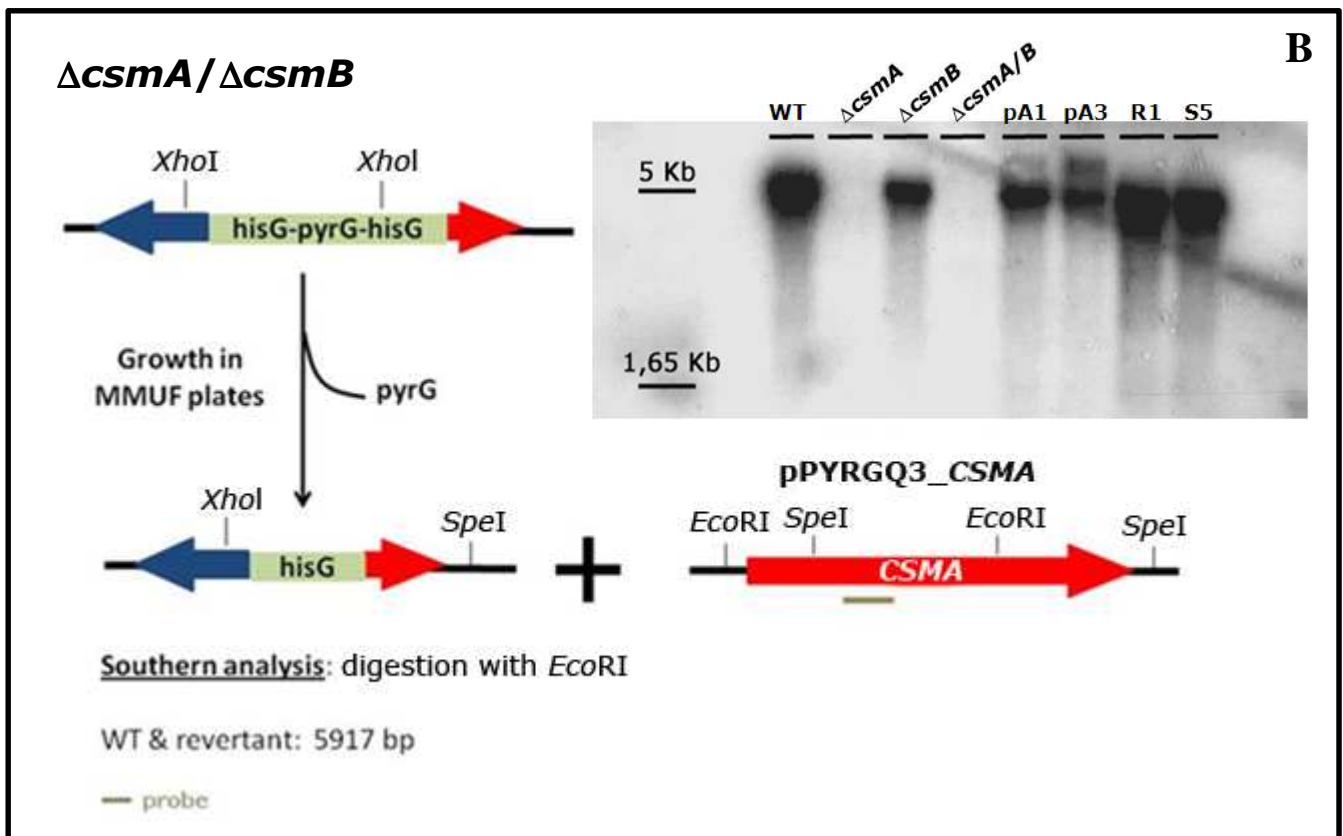
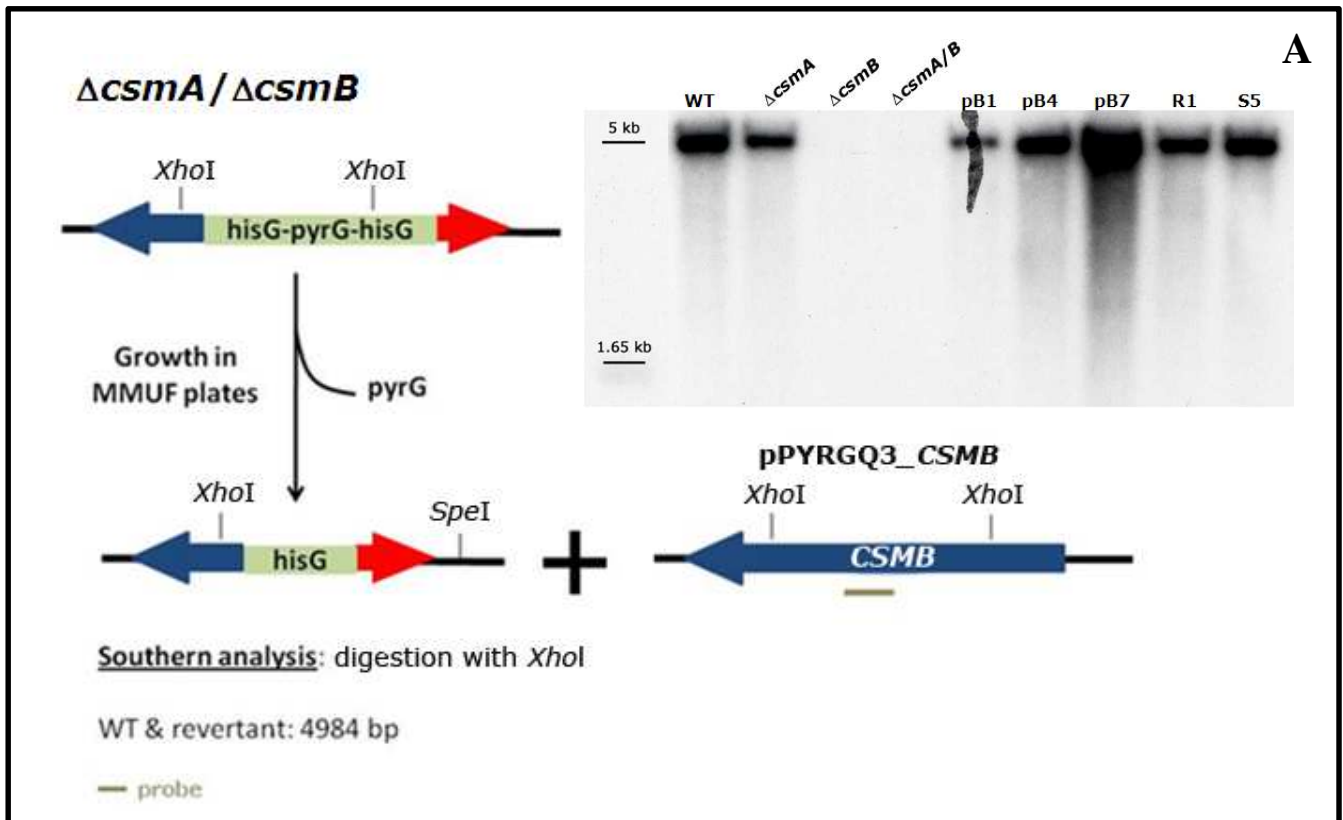
Supplemental Figure 2



# Supplemental Figure 3

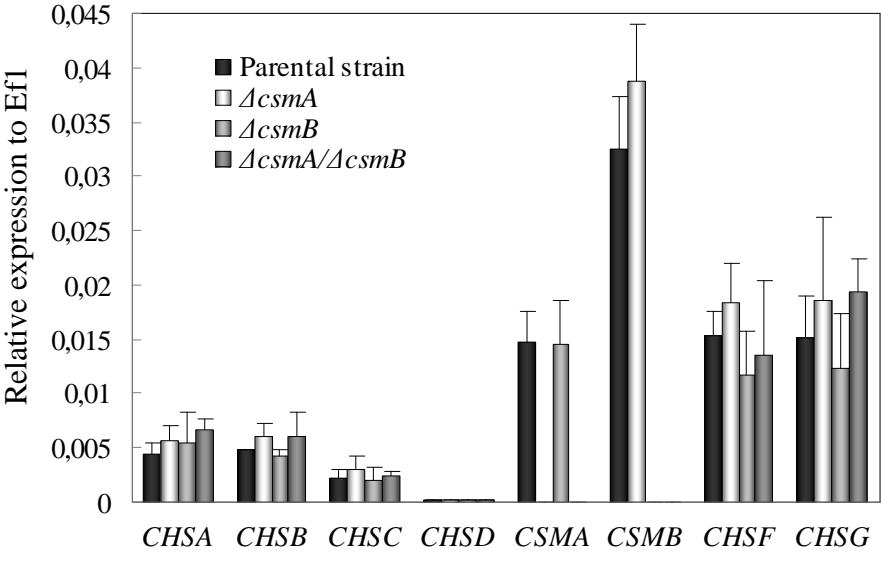


Supplemental Figure 4

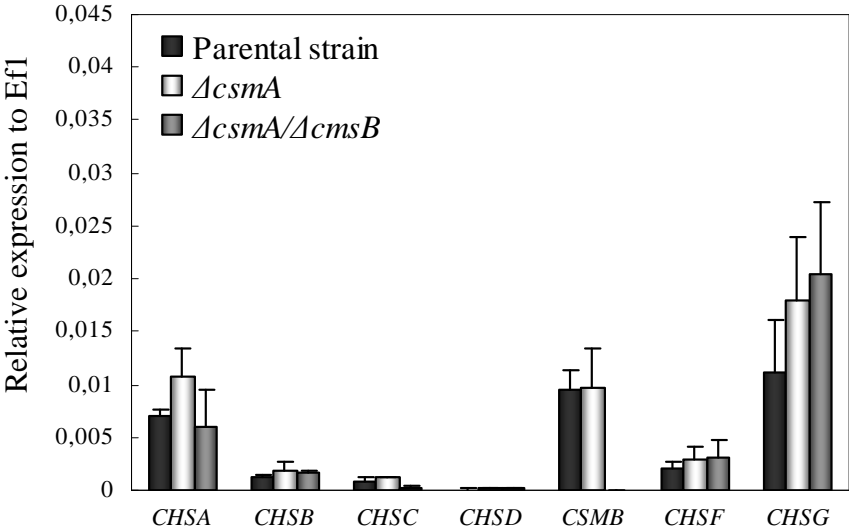




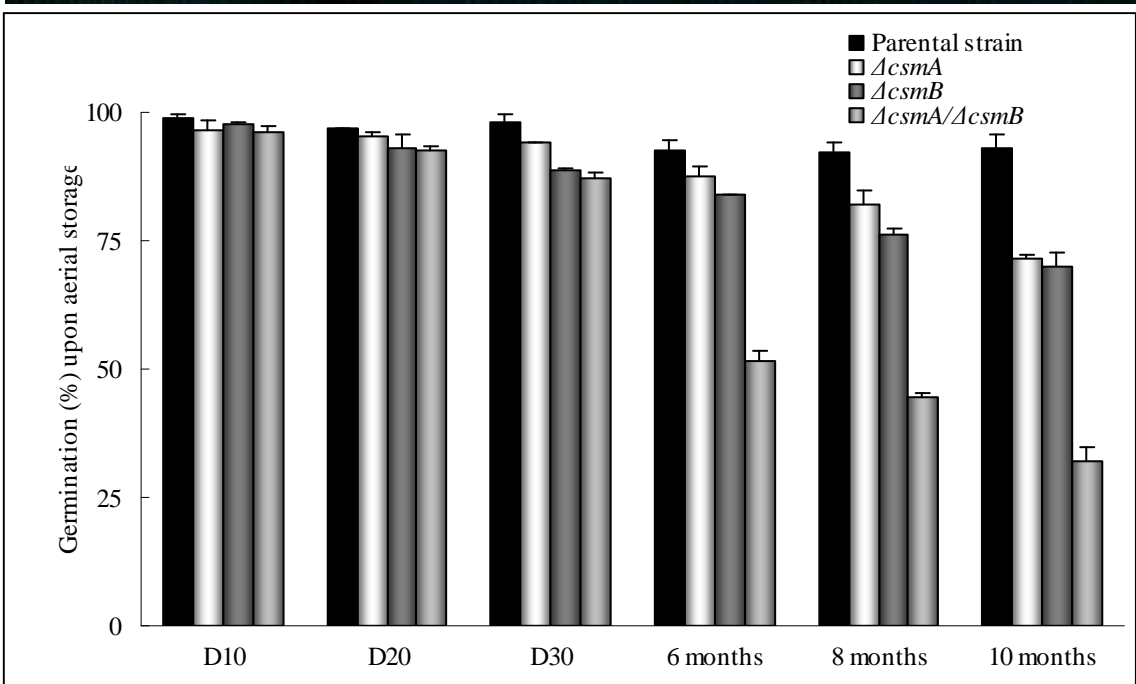
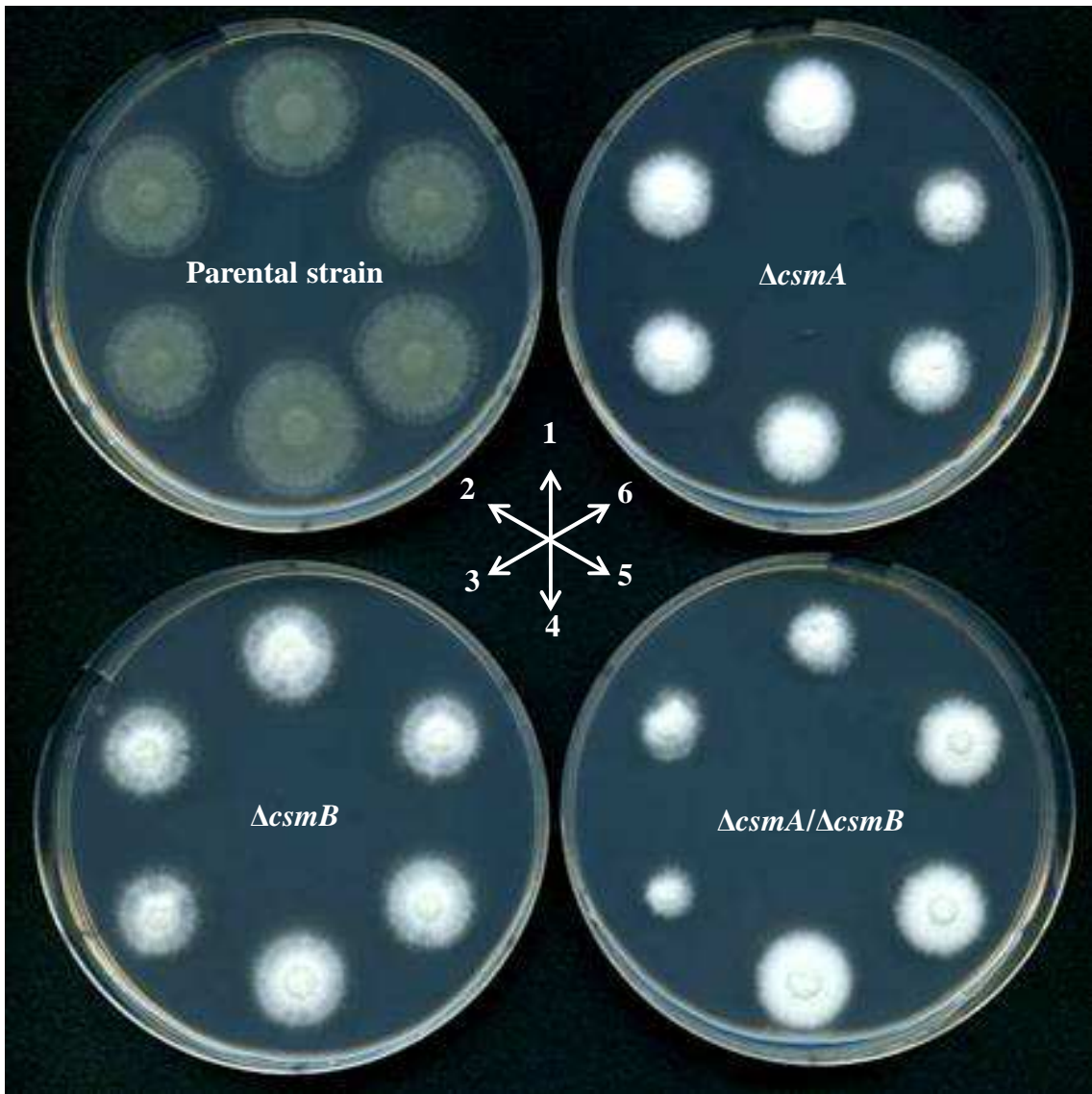
# Supplemental Figure 5



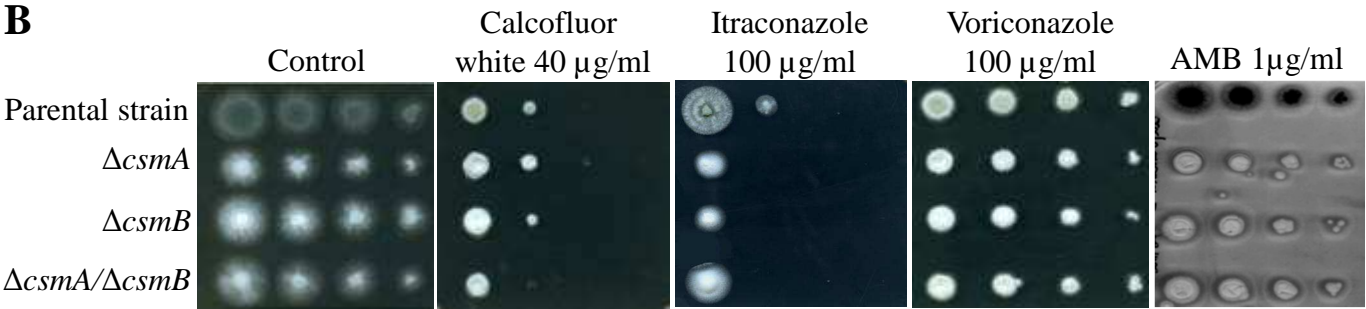
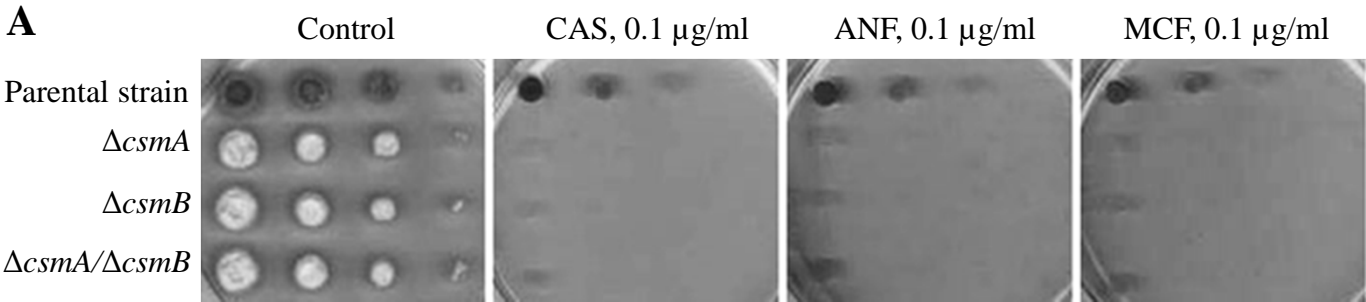
# Supplemental Figure 6



Supplemental Figure 7



# Supplemental Figure 8



# Supplemental Figure 9

