

Magnetic field-induced increase in chlorophyll *a* delayed fluorescence of photosystem II: A 100- to 200-ns component between 4.2 and 300 K

(photosynthesis/primary acceptor W/luminescence/radical pair mechanism/primary donor P-680)

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ABSTRACT At room temperature the delayed fluorescence (luminescence) of spinach chloroplasts, in which the acceptor Q is prereduced, consists of a component with a lifetime of 0.7 μ s and a more rapid component, presumably with a lifetime of 100–200 ns and about the same integrated intensity as the 0.7- μ s component. Between 4.2 and 200 K only a 100- to 200-ns luminescence component was found, with an integrated intensity appreciably larger than that at room temperature. At 77 K the 150-ns component approached 63% of saturation at roughly the same energy as the variable fluorescence of photosystem II at room temperature. At 77 K the emission spectra of prompt fluorescence but not that of the 150-ns luminescence had a preponderant additional band at about 735 nm. The 150-ns emission also occurred in the photosystem I-lacking mutant FL5 of *Chlamydomonas*. These experiments indicate that the 150-ns component originates from photosystem II. At room temperature a magnetic field of 0.22 T stimulated the 0.7- μ s delayed fluorescence by about 10%. At 77 K the field-induced increase of the 150-ns component amounted to 40–50%, being responsible for the observed \approx 2% increase of the total emission; the magnetic field increased the lifetime about 20%. In order to explain these phenomena a scheme for photosystem II is presented with an intermediary acceptor W between Q and the primary donor chlorophyll P-680; recombination of P-680⁺ and W⁻ causes the fast luminescence. The magnetic field effect on this emission is discussed in terms of the radical pair mechanism.

Since the discovery of delayed chlorophyll *a* fluorescence (in the following we use the term "luminescence" for the sake of simplicity) by Strehler and Arnold in 1951 (1) in the green alga *Chlorella pyrenoidosa*, it has been suggested by many workers that the emission of luminescence is caused by back reactions, which result in the reversal of the primary photochemical reaction, albeit with a low efficiency, and thus in the reexcitation of chlorophyll *a* and in light emission. For photosystem II (PS II) reexcitation of chlorophyll *a* takes place by the recombination of the oxidized primary donor P⁺ and reduced acceptor. For a discussion of the so-called recombination hypothesis and evidence that PS II is the source of luminescence in algae and higher plants, we may refer to reviews of Lavorel (2), Malkin (3), and Ames and van Gorkom (4).

For spinach chloroplasts and the green alga *Chlorella vulgaris* a 1- μ s component was reported (5, 6), which mainly seemed to originate from reaction centers which were in the so-called "closed" state P Q⁻. Q is an acceptor, a plastoquinone, formerly believed to be the primary acceptor. In the presence of reduced Q, Q⁻, the fluorescence yield of chlorophyll *a* is high (7). In the light of the recombination hypothesis another acceptor W different from Q was postulated and the luminescence was postulated to be due to the charge recombination P⁺ W⁻ Q⁻ \rightarrow P* W Q⁻ \rightarrow P W Q⁻ + $h\nu$. W might be an intermediary acceptor between P and Q analogous to I in photosynthetic bacteria, or W might be on a side path. In bacterial species in which the noniron complex X is prereduced, a 10-ns

luminescence component has been found due to the recombination of P-870⁺ and I⁻, the oxidized bacteriochlorophyll dimer and the reduced bacterioopheophytin, respectively (8–10). This luminescence slightly but progressively decreases from 300 K down to 77 K (8, 9, 11). A magnetic field of the order of 1–100 mT decreased the reaction center triplet yield and induced a luminescence increase, which was determined by measuring the fractional increase of the emission (the sum of fluorescence and luminescence) (12–14). This phenomenon was explained in terms of the radical pair mechanism (see ref. 15 for an introduction to the theory of this mechanism).

We have investigated the temperature dependence of the (sub) microsecond luminescence of PS II of spinach chloroplasts in which the acceptor Q was prereduced. Lowering the temperature revealed a strong luminescence component with a lifetime of 100–200 ns. Surprisingly, upon cooling we found, in contrast to bacteria mentioned above, an increase of the total luminescence. We also studied the influence of magnetic fields up to 0.3 T on the chlorophyll *a* luminescence emission yield. These experiments were carried out in two different ways. First by measuring exclusively luminescence after a short laser flash and second by determining the total emission (the sum of fluorescence and luminescence), using a continuous lamp as excitation source, both in the absence and presence of a magnetic field.

MATERIALS AND METHODS

Chloroplasts were prepared from spinach leaves as described in ref. 16. *Chlorella vulgaris* strain W.T. and *Chlamydomonas reinhardtii* strains W.T. and FL5 were grown as described in refs. 17 and 18, respectively. The chloroplast suspension was diluted with isolation medium [*N*-tris(hydroxymethyl)methylglycine, pH 7.8/0.4 M sucrose/10 mM KCl/5 mM MgCl₂]. The suspensions of algae were diluted with growth medium. The diluted suspensions were mixed with a solution of glycerol, which was almost saturated with sucrose, in a ratio of 40:60 (vol/vol) in order to prevent crystallization upon cooling. The final absorbance of all suspensions was adjusted to 0.1/mm at 680 nm, corrected for scattering. The sample was contained in a Perspex cell of 1 mm thickness. In the case of chloroplasts a few grains of sodium dithionite (final concentration about 10 mM) were added 5 min prior to the measurements to reduce Q.

Luminescence measurements were performed with the apparatus and methods described in ref. 19. The sample was

Abbreviations: PS II, photosystem II of photosynthesis; PS I, photosystem I of photosynthesis; P, primary donor P-680 of PS II, probably a chlorophyll dimer; W, primary acceptor of PS II; Q, electron acceptor of PS II, a plastoquinone molecule; Z, secondary donor of PS II of unknown identity; B, magnetic field in tesla units; F, chlorophyll emission (fluorescence plus luminescence); L, chlorophyll luminescence or delayed fluorescence; a.u., energy/s in arbitrary units L and F.

placed between the poles of a small home-built electromagnet. In order to prevent orientation of the luminescing species in the magnetic field, the magnet was fed from the 50-Hz mains. The effect of the alternating magnetic field up to 0.6 T on the photomultiplier was much smaller than the accuracy of the measurement ($<10^{-2}$ of the luminescence).

RESULTS

Fig. 1A (curve a) shows the decay kinetics of the 0.7- μ s luminescence component of spinach chloroplasts with prerduced Q at room temperature. Analysis of the curve indicates that a more rapid (100- to 200-ns) component also is present, for which further evidence will be given in *Discussion*. Curve b in this figure shows the luminescence decay kinetics under the same conditions at 77 K. It can be observed that at 77 K the lifetime of the luminescence is about 150 ns. Its amplitude is about one order of magnitude higher than at room temperature. In Fig. 1B the temperature dependence of the luminescence (lifetime and total yield) is shown for the whole region 77–300 K. Be-

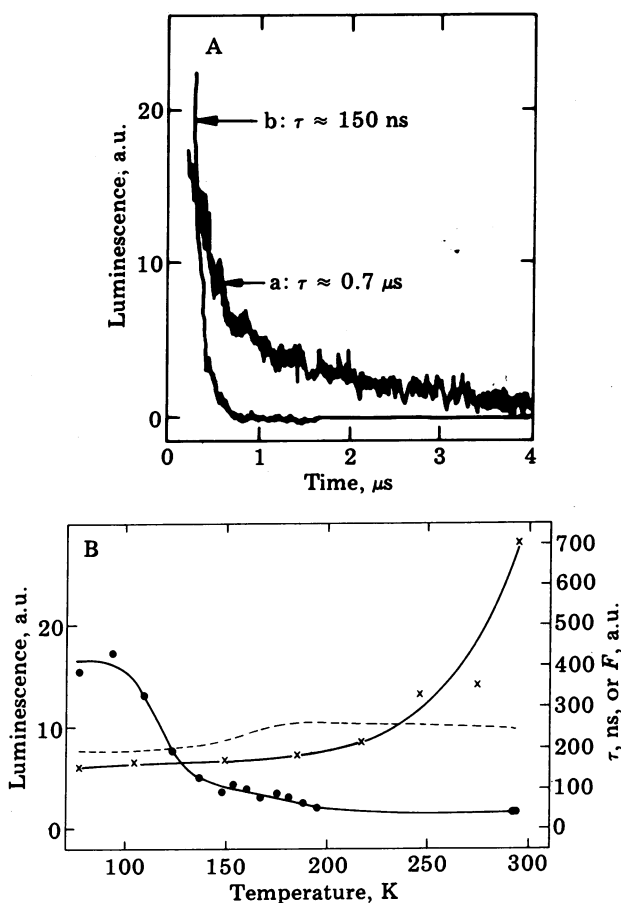


FIG. 1. (A) Typical decay kinetics after a just-saturating laser flash of spinach chloroplasts in which Q was prerduced, at room temperature (curve a) and at 77 K (curve b); τ is the lifetime of luminescence. For reasons of presentation the 0.7- μ s luminescence at room temperature (a) is amplified by a factor of about 8. Luminescence is plotted as energy/s in arbitrary units (a.u.). (B) The integrated luminescence [= (amplitude at time zero) \times τ] (●) and τ (×) as a function of temperature. The lifetime τ was obtained by a best-fit method assuming one exponential component, which presumably is not correct for temperatures larger than 200 K (see *Discussion*). Conditions were as given for A. The broken line represents the temperature dependence of the relative fluorescence yield, F , measured under the same conditions. The measuring wavelength was 685 nm for both luminescence and fluorescence. Preliminary results indicate that at 4.2 K the decay time is about 200 ns and the integrated luminescence is roughly 8 a.u.

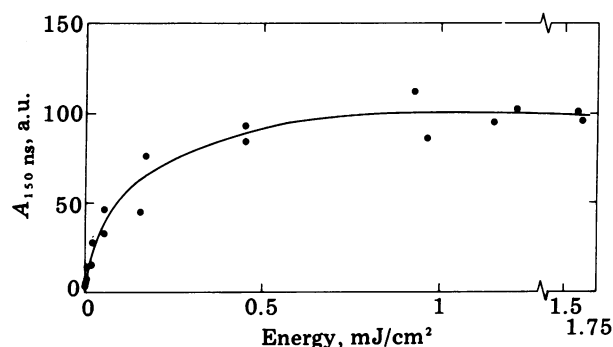


FIG. 2. Amplitude of the 150-ns luminescence at 77 K of spinach chloroplasts, in which Q was prerduced, as function of the laser flash energy. Exciting wavelength was 530 nm; measuring wavelength was 685 nm.

tween 293 and about 200 K the 0.7- μ s component disappears, and at 200 K only the 150-ns component remains. Below 200 K the lifetime of the latter component hardly changed, but the integrated luminescence increased by a factor of about 8 down to 77 K. Sometimes we found an increase by a factor of about 4 in the integrated luminescence that was accompanied by a flatter course in the region 77–150 K. Under the same conditions the fluorescence yield in this temperature range hardly changed, in agreement with ref. 20.

Fig. 2 shows the amplitudes at time zero (obtained by extrapolation) of the 150-ns luminescence component of spinach chloroplasts at 77 K as function of the energy of the laser flash. As shown in this figure, the luminescence is saturated at energies higher than 500 μ J/cm². The energy that produced 63% of the maximal luminescence amplitude is about 150 μ J/cm². At room temperature the energy for 63% saturation of the variable fluorescence of untreated chloroplasts in the same medium is also about 150 μ J/cm² when the same source of excitation is used (data not shown).

In Fig. 3 the 150-ns luminescence emission spectrum at 77 K of spinach chloroplasts is shown. The fluorescence emission spectrum is also displayed in this figure.

Fig. 4A shows the influence of a magnetic field B on the amplitude of the 150-ns luminescence component at about 80

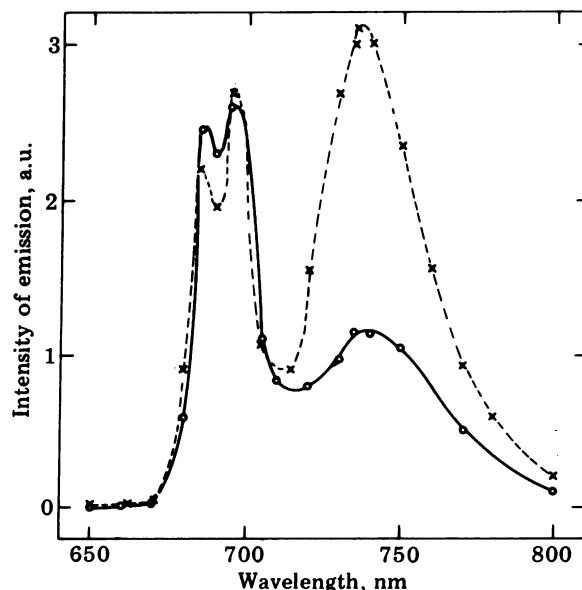


FIG. 3. The 150-ns luminescence (O—O) and fluorescence (×---×) emission spectra of spinach chloroplasts, in which Q was prerduced, at 77 K.

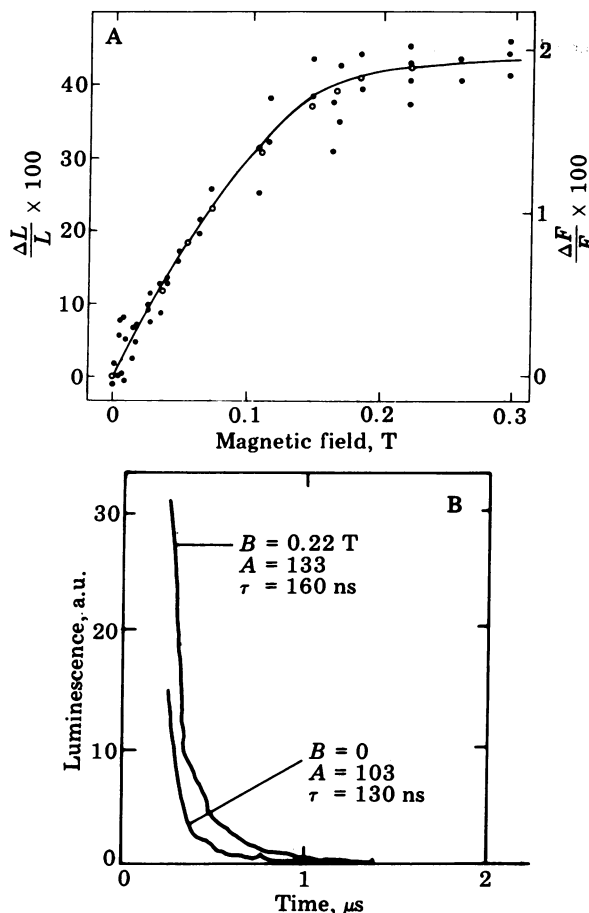


FIG. 4. (A) Fractional increase, $\Delta L/L$, of the 150-ns luminescence component, measured at 685 nm, as a function of external magnetic field B (●) for spinach chloroplasts, in which Q was prereduced. Temperature is about 80 K, L is the integrated luminescence (= amplitude $A \times$ lifetime τ) and $\Delta L = L - L(B = 0)$. The corresponding fractional increase of fluorescence, $\Delta F/F$, is given by ○. (B) Typical decay kinetics in the absence and presence of a magnetic field of 0.22 T. A is the amplitude of the luminescence at time zero (obtained by extrapolation). Note that for the luminescence the same units as for Fig. 1A are used. Conditions are as given for Fig. 4A.

K. A considerable increase of the amplitude and lifetime (see Fig. 4B) is observed with an increase in the value of B . In addition, the magnetic field-induced fractional increase of fluorescence, $\Delta F/F$, under the same conditions is displayed in this figure. When Q was in the oxidized state ΔF was found to be zero. This measurement was performed at very low light intensity of continuous illumination to prevent the formation of Q^- . The luminescence, L , and the magnetic field-induced luminescence increase, ΔL , were also much smaller (<10%) under these conditions than in the case of Q prereduced. One might expect a negligible L and ΔL , but because repeated flashing was necessary to measure the luminescence, small amounts of Q^- may have been formed, which may have caused the weak luminescence observed.

The magnetic field-induced increase of the amplitude of the 0.7- μs luminescence at room temperature was found to be about 10% at 0.22 T. Within the limits of accuracy of determination of the decay time ($\pm 5\%$) we did not observe a change in the lifetime of the 0.7- μs component upon the application of a magnetic field of 0.22 T. These effects are much smaller than the increase in these parameters found at 77 K for the 150-ns component caused by a magnetic field of the same strength.

The results obtained for *Chlorella vulgaris* strain W.T. and *Chlamydomonas reinhardtii* strains W.T. and FL5 were sim-

ilar to the results shown above for spinach chloroplasts. For algae Q was not prereduced with sodium dithionite, because this compound failed to reduce Q in the cells. In this case Q was reduced by a number of laser flashes. Because of the low temperature (77 K), Q^- was not reoxidized on the time scale of our measurements. When Q in spinach chloroplasts was reduced by the laser flashes, the amplitude of the 150-ns luminescence component was smaller than 20% of the amplitude in chloroplasts in which Q was prereduced by sodium dithionite. An analogous effect has also been observed at room temperature for the 0.7- μs luminescence in *Chlorella* (6).

DISCUSSION

A few previous studies showed that the yield of luminescence above 0.1 ms is almost negligible when Q is in the reduced state before the flash (21, 22). At room temperature we found that micro- and millisecond luminescence components were absent, except for the 0.7- μs luminescence, which shows an appreciable increase when Q is reduced prior to the luminescence-inducing flash. At low temperature a strong component of about 150 ns was found with prereduced Q. Analogously with the hypothesis proposed for the 0.7- μs luminescence component at room temperature (see Introduction), we attribute the 150-ns luminescence to recombination of P^+ and W^- , which results in the excited state P^*W . The following arguments indicate that the 150-ns component originates in PS II. The saturation of the 150-ns component at 77 K (see Fig. 2) occurs at roughly the same energy as that for the variable fluorescence of PS II at room temperature. This indicates that reaction center chlorophyll of this photosystem is the source of the luminescence. It has been proposed that the fluorescence emission of chloroplasts and algae at 77 K has maxima at 685 and 695 nm belonging to PS II and that PS I mainly contributes to the emission around 735 nm, where PS II has a minor contribution (23–26). The spectrum of the variable fluorescence of spinach chloroplasts at 77 K (26) is very similar to the 150-ns luminescence spectrum, displayed in Fig. 3, which also does not contain the peak around 735 nm. From these data we conclude that the latter spectrum probably originates from the chlorophyll of PS II. This conclusion is strengthened by the presence of the 150-ns luminescence component in the PS I-lacking mutant FL5 of *Chlamydomonas*.

A scheme for energy and electron transfer reactions occurring in the antenna and the reaction center complex of PS II, consistent with the experiments discussed and the magnetic field-induced increases (Fig. 4) in luminescence and fluorescence, is given in Fig. 5. This scheme is similar to schemes proposed for photosynthetic bacteria (cf. refs. 12–14). It should be mentioned that the state P^T , the reaction center chlorophyll in the lowest triplet state, has not been observed (possibly because of rapid decay) in PS II but was experimentally observed in bacterial mutants lacking carotenoid.

The marked magnetic field-induced increase of the amplitude and lifetime of the 150-ns luminescence component as shown in Fig. 4 may be explained by the radical pair mechanism: In a high magnetic field B , only the triplet sublevel T_0 (not T_1 and T_{-1}) is sufficiently close to the singlet level for the occurrence of the interconversion $(P^+W^-)^S \rightarrow (P^+W^-)^T$. The field-reduced interconversion causes a lowering of the ratio $(P^+W^-)^T/(P^+W^-)^S$ and thus an increase of the concentration $(P^+W^-)^S$. Therefore the concentration of P^*W , and thus the luminescence intensity, is increased by the magnetic field. If the triplet formation $(P^+W^-)^T \rightarrow P^*W$ is the reaction that mainly determines the decay rate of P^*W^- , then the magnetic field will increase the luminescence decay time. If the decay were mainly caused by k_g (or by k_s , but this is presumably rel-

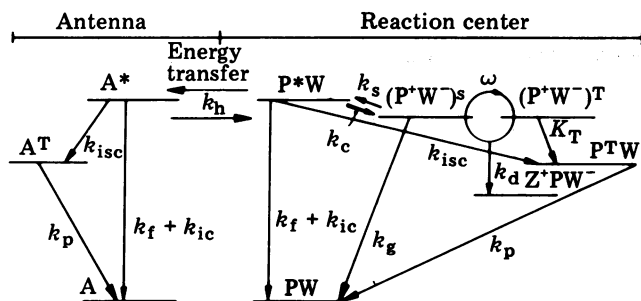


FIG. 5. Hypothetical scheme for energy and electron transfer occurring in the antenna chlorophyll and the reaction center complex of PS II, in the presence of reduced acceptor Q. k_h , k_f , k_{ic} , and k_{isc} are the rate constants for deexcitation of the lowest excited singlet states A* and P* by energy transfer, fluorescence, internal conversion, and intersystem crossing to the triplet states A^T and P^T. The decay of the triplet states to the ground state occurs via k_p . k_c is the rate constant for charge separation. k_r , k_b , and k_g are the rate constants for the recombination of P⁺W⁻ to the triplet state P^T, the excited singlet state P*, and the ground state, respectively. ω is the angular frequency for interconversion between the singlet and triplet states of P⁺W⁻. Z is the secondary donor, which reduces P⁺ with a rate constant k_d . For further explanation see text.

atively small), then the magnetic field would decrease the decay time because the magnetic field increases the concentration of (P⁺W⁻)^S. The observed increase in luminescence lifetime thus indicates that k_T is rate determining, because the magnetic field reduces the amount of (P⁺W⁻)^T.

The fractional increase of fluorescence, $\Delta F/F$, as a function of external magnetic field closely follows the relative field-induced change of the 150-ns luminescence (Fig. 4A). We conclude that ΔF is mainly if not completely caused by a change in the 150-ns luminescence: In a magnetic field of 0.15 T we measured an increase of 40% of the total emission of the 150-ns luminescence (obtained by extrapolation to zero). The yield of the 150-ns component was established to be 20 times smaller than the fluorescence yield, which means that $\Delta F/F$ should be $0.05 \times 40\% = 2\%$ at $B = 0.15$ T, in agreement with the value of 1.7% for $\Delta F/F$ obtained by direct measurement.

At room temperature the magnetic field-induced ($B = 0.22$ T) increase of the 0.7 μ s luminescence, measured at 0.4–4 μ s, was about 10%. The yield of this luminescence (measured in flashes of low intensity) is about 1/160th of the fluorescence yield under the same conditions. If the 0.7- μ s luminescence component were the only one present, a value of $1/160 \times 10\% = 0.06\%$ would be expected for $\Delta F/F$ at room temperature. We measured a value of about 0.2% for $\Delta F/F$. This difference can be explained by the presence at room temperature of a second, more rapid, luminescence component with a somewhat larger integrated luminescence than that of the 0.7- μ s component. The observation of an at least 15% increase in luminescence integrated from 0.25–0.8 μ s indeed shows that such a component is present. Because of the presence of the 0.7- μ s component and the rapidity of this component we could not measure it with sufficient precision with the apparatus as it is at the moment. We estimate that the amplitude of this fast component was roughly 1/10th of that at 77 K, and its decay time was similar.

Corresponding curves of $\Delta F/F$ against B for photosynthetic bacteria approach saturation at about $B = 150$ mT; half the effect is attained at $B_{1/2} = 25$ –35 mT (14), as compared to 65 mT for half-saturation in chloroplasts (Fig. 4A). At room temperature we observed a value of $B_{1/2}$ that was about half the value of $B_{1/2} = 65$ mT at 77 K. Preliminary experiments also showed a considerable decrease (to $1/2$ to $1/3$) of $B_{1/2}$ at 80 and

290 K when the chloroplasts were suspended in isolation medium brought to pH 3.

Magnetic field modulation of the luminescence has been found *in vitro* not only when the luminescence is produced by radical pair recombination but also when it is produced in a process known as triplet exciton fusion (for an introduction and review see refs. 27–29). It seems to us that this mechanism might be consistent with our results if the two triplets are generated in one reaction center by heterofission P*C → P^TC^T in which C is a carotenoid molecule and C^T is the lowest carotenoid triplet state. Luminescence is generated by the reverse reaction. Such luminescence may be increased by a magnetic field (see also ref. 30). In an analogous way as for the radical pair mechanism one might assume that P^T (or C^T) disappears in 150 ns independent of temperature. Normally the triplet decay to the ground state requires milliseconds (chlorophyll triplet) or microseconds (carotenoid triplet), but the triplet might be transferred to another molecule. For this mechanism we do not understand the observed increase in luminescence lifetime caused by a magnetic field.

Finally, we will discuss the temperature behavior of integrated luminescence and luminescence lifetime as displayed in Fig. 1. Lowering the temperature may be expected to cause a decrease in the integrated luminescence and an increase in lifetime, because of the activation energy needed for the back reaction P⁺W⁻ → P*W. To our surprise the opposite was observed: when the temperature was lowered from 290 to 80 K the integrated luminescence increased by a factor of 8, the 0.7- μ s component disappeared, and a 150-ns component increased in amplitude but not in decay time. Even at 4.2 K this fast component exists, with a decay time of about 200 ns.

A possible explanation for the approximate constancy of the decay time of the 150-ns component with the decreasing temperature is the following. The decay time of this component may be assumed to be equal to the decay time of the radical pair P⁺W⁻, which decay time is mainly determined by the most rapid reaction, removing the state P⁺W⁻. At low temperatures the reduction of P⁺ by the secondary donor Z has a small rate constant k_d of the order of 0.1–20 ms⁻¹ (31, 32) and therefore k_d probably does not affect markedly the decay rate of the radical pair. Because a magnetic field of 0.22 T increases the luminescence lifetime by about 20% the reaction with rate constant k_T (see scheme of Fig. 5) seems to be rate determining. Recombination of P⁺W⁻ to the reaction center triplet state P^TW may be a temperature-independent electron tunneling. Then the luminescence decay time will be essentially independent of temperature because, as already discussed, the magnetic field effects suggest that the reaction center triplet formation is the most rapid reaction. The luminescence intensity is determined by the rate of the back reaction P⁺W⁻ → P*W (→ PW + $h\nu$). On the basis of the yield at 80 K of the 150-ns component, which is 0.5% (0.05 times the yield of fluorescence, which is about 10%) the rate constant k_s for this back reaction may be of the order of 10^5 s⁻¹ at 80 K. The sign of the magnetic field effect indicates that the reaction of P⁺W⁻ to the ground state (rate constant k_g in the scheme of Fig. 5) is also rather slow. For photosynthetic bacteria k_g was also found to be relatively small in comparison with the other rate constants for the decay of the radical pair (11, 33).

The 0.7- μ s component at room temperature might be explained by the assumptions that at room temperature part of Z is oxidized by P⁺ in a time longer than 150 ns. Z⁺ is reduced by a tertiary donor in 0.7 μ s and at a smaller rate by the back reaction Z⁺P W⁻ → Z P⁺W⁻, which causes the 0.7- μ s luminescence component. The ΔL and ΔF measurements at room temperature show that a fast component, which may be the

150-ns emission, is also present at room temperature, but with an appreciably smaller amplitude than at 77 K. This suggests that at room temperature the luminescence originates for a part directly from P^+W^- recombination with the same decay time as at low temperatures, and partly from $Z P^+W^-$, which is reoxidized by $Z^+P W^-$. At lower temperatures the 0.7- μ s component is absent because the reaction $Z P^+ \rightarrow Z^+P$ is much slower than 150 ns (31, 32). Preliminary measurements at room temperature with the reduction of P^+ blocked by low pH or the addition of hydroxylamine (10 mM) indicate that only a fast component (lifetime about 150 ns and amplitude 1/10th to 1/20th of that at 77 K) is present and that the 0.7- μ s component has disappeared. This strengthens the suggestion about the twofold origin of the luminescence at room temperature discussed above. Additional experiments are required to establish whether this or other mechanisms are responsible for the disappearance of the 0.7- μ s component with decreasing temperature.

The fact that at 4.2 K appreciable 150-ns luminescence emission occurs shows that the energy of the state causing the emission is not much less than that of the state $P^+W Q^-$. On the other hand, between 200 and 300 K the amplitude of the 150-ns component is rather low. If the state causing the emission is $P^+W^-Q^-$, as shown in Fig. 5, the energy of this state must be appreciably lower than that of $P^+W Q^-$. An explanation of the increase in 150-ns luminescence upon lowering of the temperature may be that at room temperature P^+W^- decays through various (conformational) states of progressively lower energies and with nonzero activation energies. Somehow this process does not affect the rate-determining decay of P^+W^- via k_T . At low temperature—e.g., 80 K—the system remains (at least during 0.15 μ s) at one of the higher energy states. In this high energy state the luminescence-generating back reaction proceeds at a relatively high rate. Computer simulation for the schemes proposed should be informative. The working hypotheses proposed about details of PS II reactions can be tested by various methods, specifically absorption difference spectroscopy.

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