

# **Solution Structure of Duplex DNA Containing a $\beta$ -Carba-Fapy-dG Lesion**

Mark Lukin, Tatiana Zaliznyak, Sivaprasad Attaluri, Francis Johnson and Carlos de los Santos\*

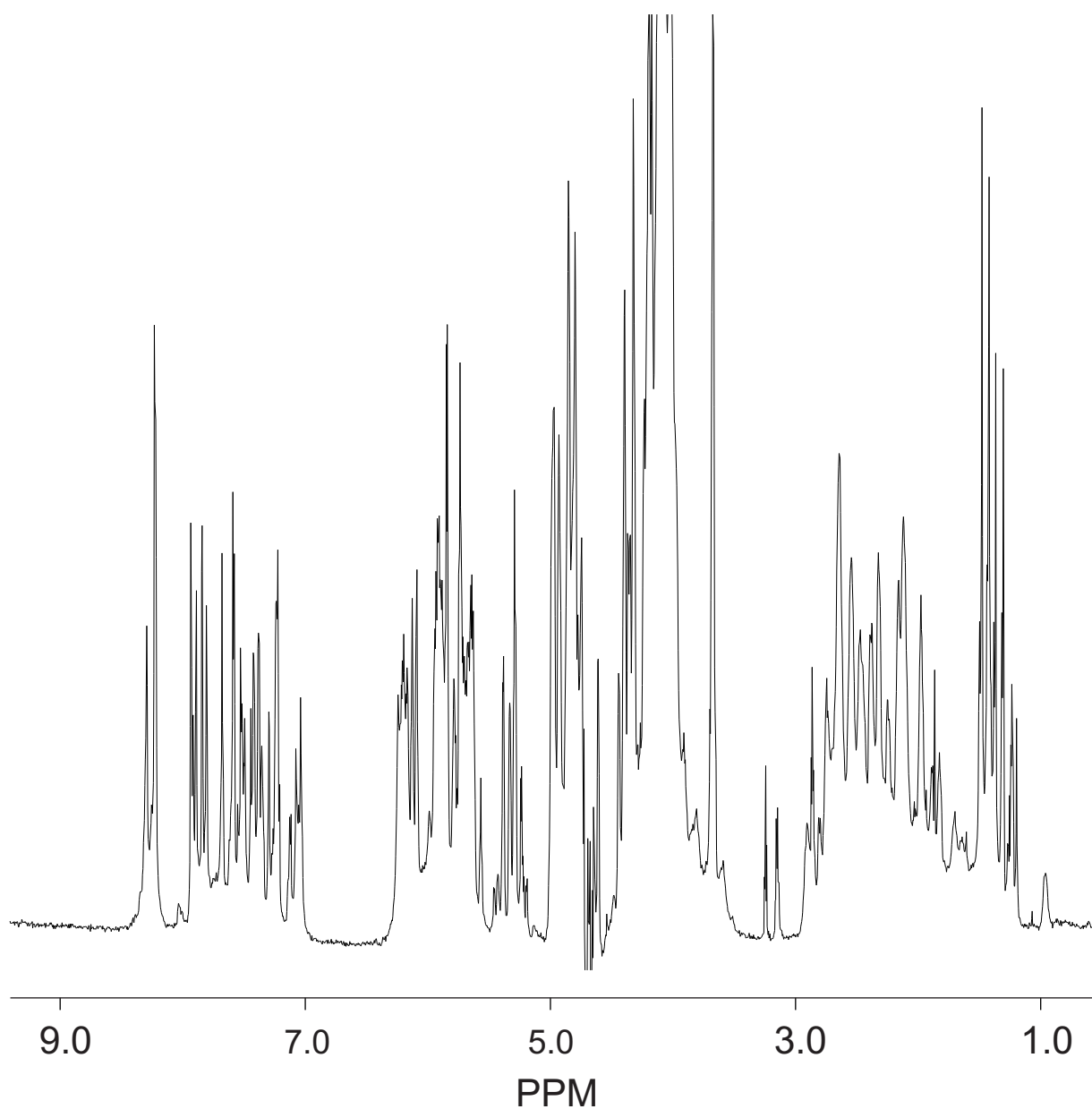
Department of Pharmacological Sciences. Stony Brook University-School of Medicine. Stony Brook, NY, 11794-8651

## **SUPPORTING INFORMATION**

Proton chemical shifts (Table 1S) on the  $\beta$ -cFapy-dG•dC duplex measured at 25 °C. One dimensional proton spectrum of the  $\beta$ -cFapy-dG•dC duplex recorded in 100% D<sub>2</sub>O buffer at 25 °C (Figure 1S), full 'finger print' region assignments on a 300 ms mixing time NOESY spectrum, recorded in 100% D<sub>2</sub>O buffer at 25 °C for the damaged (Figure 2SA) and undamaged (Figure 2SB) strands of the duplex, and expanded contour plot of the aromatic proton region on a 300 ms mixing time NOESY spectrum recorded in 100% D<sub>2</sub>O buffer at 25 °C (Figure 3S). Three-dimensional view of the twenty five *Z*- (Figure 4S) and *E*- (Figure 5S)  $\beta$ -cFapy-dG•dC duplex structures, and examples of water-mediated hydrogen bonds in the  $\beta$ -cFapy-dG•dC duplex (Figure 6S). UV<sub>260</sub> melting curves for the  $\beta$ -cFapy-dG•dC and dG•dC duplexes (Figure 7S).

Table 1S. Proton chemical shifts on the $\beta$ -cFapy-dG•dC duplex at 25 °C.									
	H6/H8	H1'	H2''	H2'	H3'	H4'	Me/H5/H2	G(H1)/T (H3)	CN <sub>4</sub> H
C1	7.59	5.74	2.39	1.98	4.66	4.03	5.85		
G2	7.93	5.95	2.76	2.64	4.94	4.33		12.81	
(Z)T3	7.23	5.71	2.47	2.11	4.85		1.48	13.50	
(E)T3	7.23	5.73	2.47	2.11	4.85		1.48	13.50	
(Z)A4	8.22	6.22	2.86	2.65	4.98	4.40	7.45		
(E)A4	8.22	6.21	2.86	2.64	4.97	4.40	7.42		
(Z)C5	7.35	5.90	2.37	2.10	4.80		5.29		8.28/6.64
(E)C5	7.27	5.89	2.30	2.00	4.76		5.20		8.22/6.76
(Z)F6	7.92	3.91	1.89	1.70	4.40	2.16		12.56	
(E)F6	7.21	3.83	1.90	1.72	4.41	2.17		12.63	
(Z)C7	7.50	5.91	2.53	2.14	4.87	4.23	5.65		8.59/6.90
(E)C7	7.54	5.86	2.50	2.13	4.85		5.69		8.39/6.76
(Z)A8	8.23	6.20	2.65	2.89	4.97	4.35	7.59		
(E)A8	8.26	6.20	2.64	2.90	4.97	4.35			
T9	7.08	5.72	2.32	1.94	4.82		1.37	13.57	
G10	7.81	5.88	2.55	2.65	4.92	4.32		12.74	
C11	7.38	6.13	2.13	2.16	4.44	4.01	5.33		8.15/6.62
G12	7.89	5.92	2.56	2.74	4.80	4.19		13.00	
C13	7.42	5.66	2.12	2.44	4.85	4.17	5.39		8.39/6.55
A14	8.30	6.24	2.66	2.92	4.99	4.39	7.68		
T15	7.04	5.68	1.98	2.37	4.81		1.43	13.55	
(Z)G16	7.68	5.91	2.52	2.71	4.86	4.33		12.59	
(E)G16	7.69	5.87	2.52	2.67	4.88	4.33		12.54	
(Z)C17	7.12	5.57	2.03	2.32	4.64	4.07	5.24		8.26/6.37
(E)C17	7.24	5.68	2.10	2.33	4.71		5.28		8.35/6.37
(Z)G18	7.53	5.84	2.65	2.47	4.75	4.24		12.54	
(E)G18	7.58	5.86	2.66	2.49	4.77	4.27		12.53	
(Z)T19	7.30	5.78	2.48	2.11	4.86		1.31	13.71	
(E)T19	7.30	5.79	2.48	2.11	4.86		1.31	13.71	
(Z)A20	8.23	6.17	2.81	2.63	4.98	4.37	7.52		
(E)A20	8.23	6.17	2.81	2.63	4.98	4.37	7.49		
C21	7.24	5.63	2.24	1.83	4.75		5.29		8.27/6.74
G22	7.84	6.09	2.32	2.54	4.61	4.12		13.01	

Chemical shifts are in ppm. Chemical shifts of the H6'/H6'' protons of cFapy are 0.97/1.98 ppm and 0.96/2.07 ppm for the Z and E isomers, respectively.



**Figure 1S.** One dimensional proton spectrum of the cFapy-dG·dC duplex recorded at 600 MHz in 100% D<sub>2</sub>O buffer, at 25 °C.

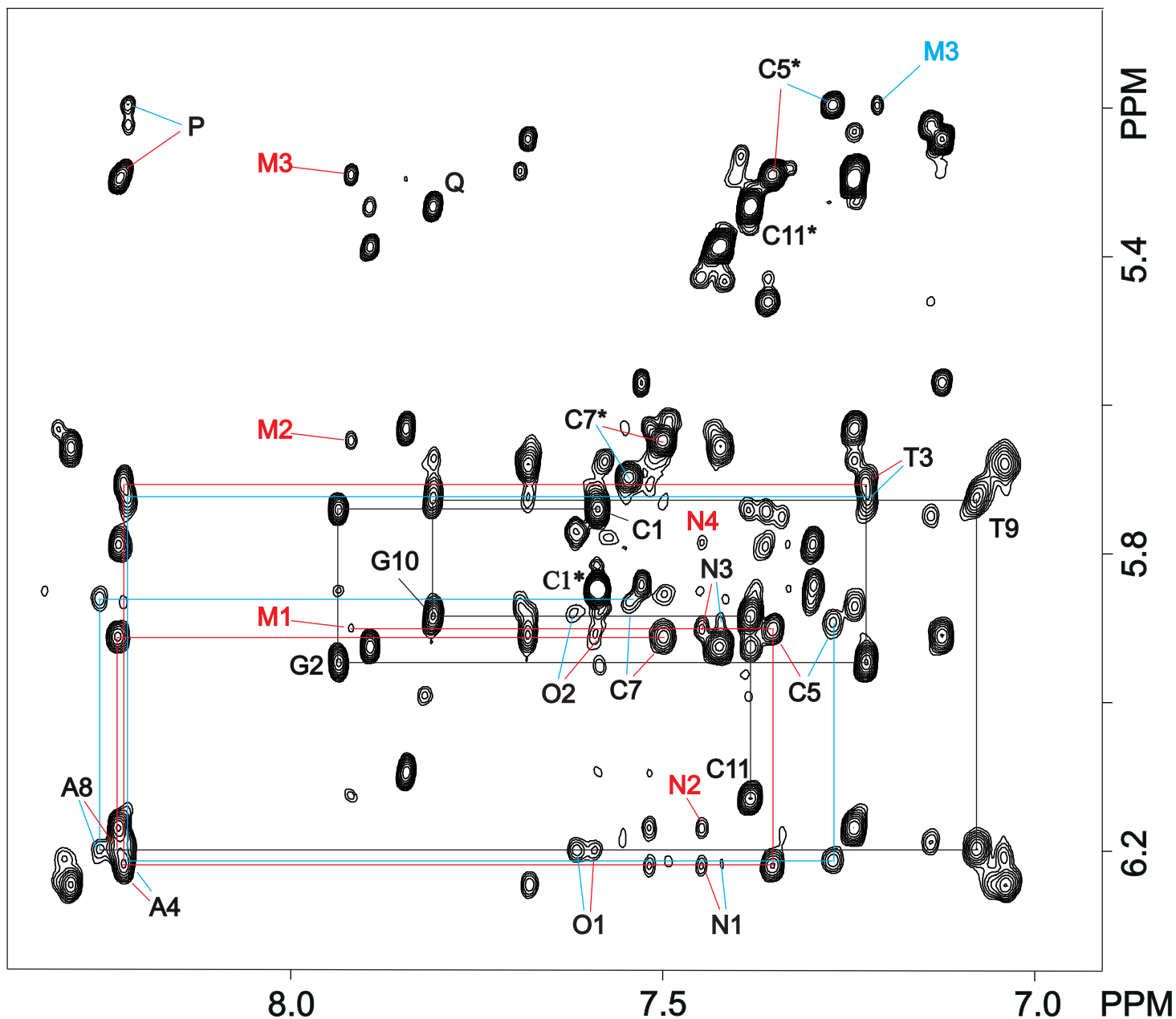


Figure 2SA. Expanded contour plot depicting NOE interactions in the base to H1' proton region of a 800 MHz NOESY spectrum (300 ms. mixing time) recorded in 100% D<sub>2</sub>O buffer, at 25 °C. Lines connect inter residue and sequential base-H1' NOE peaks seen in the modified strand of the damaged duplex. Red and blue colors trace these connectivities on the *Z* and *E* isomeric duplexes, respectively. Numbered letters label the intra residue NOE, and asterisks indicate cytosine(H5-H6) peaks. Other labels are assigned as follows: M1, F6H8-C5H1'; M2, F6H8-C7H5; M3, F6H8-C5H5; N1, A4H2-A4H1'; N2, A4H2-A20-H1'; N3, A4H2-C5H1'; N4, A4H2-T19H1'; O1, A8H2-A8H1'; O2, A8H2-G16H1'; P, A4H8-C5H5; Q, G10H8-C11H5.

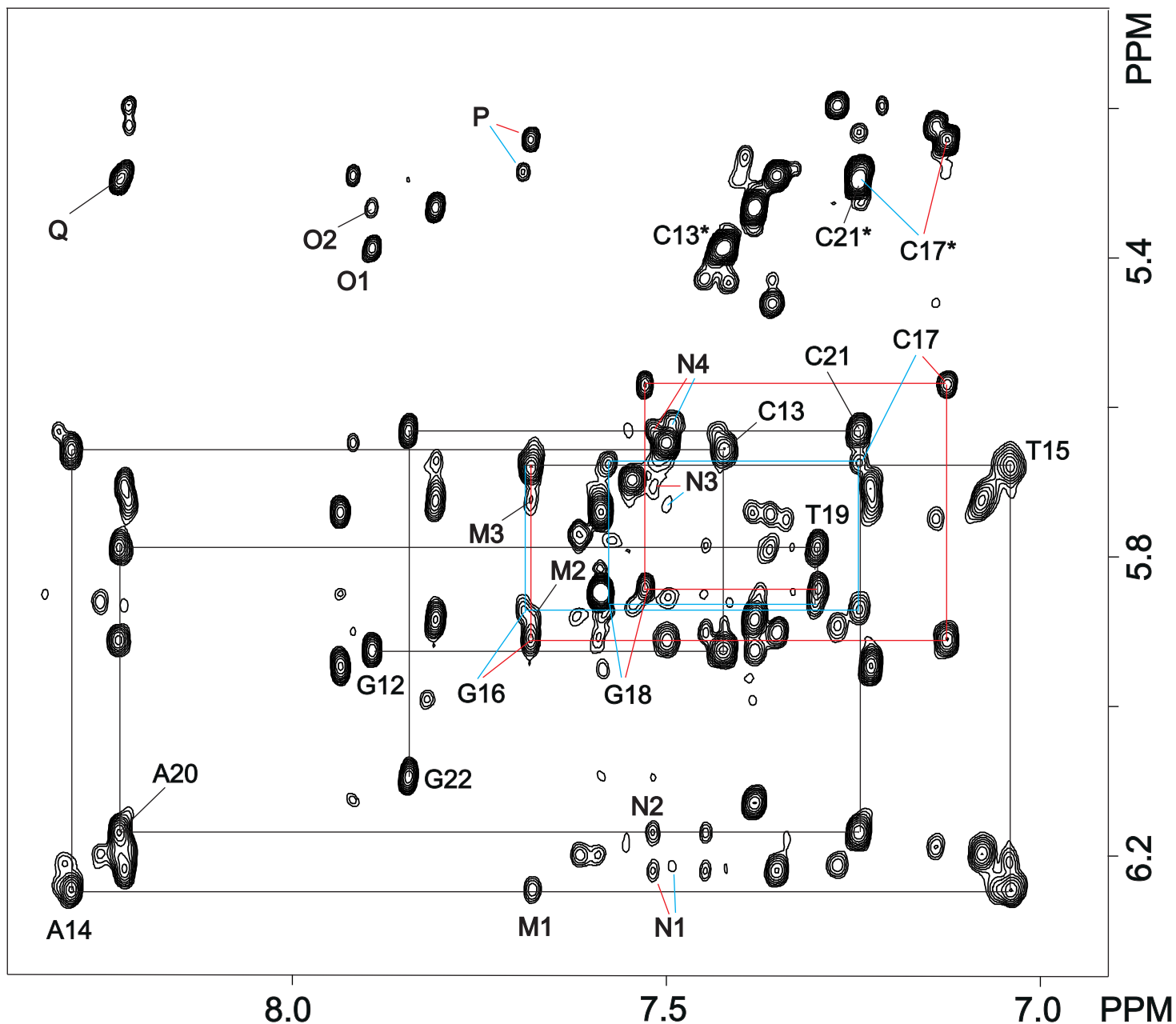


Figure 2SB. Expanded contour plot depicting NOE interactions in the base to H1' proton region of an 800 MHz NOESY spectrum (300 ms. mixing time) recorded in 100% D<sub>2</sub>O buffer, at 25 °C. Lines connect inter residue and sequential base-H1' NOE peaks seen in the unmodified strand of the damaged duplex. Red and blue colors trace these connectivities in the Z and E isomeric duplexes, respectively. Numbered letters label the intra residue NOE, and asterisks indicate cytosine(H5-H6) peaks. Other labels are assigned as follows: M1, A14H2-A14H1'; M2, A14H2-G10H1'; A3, A14H2-T9H1'; N1, A20H2-A4H1'; N2, A20H2-A20-H1'; N3, A20H2-T3H1'; N4, A20H2-C21H1'; O1, G12H8-C13H5; O2, G12H8-C11H5; P, G16H8-C17H5; Q, A20H8-C21H5.

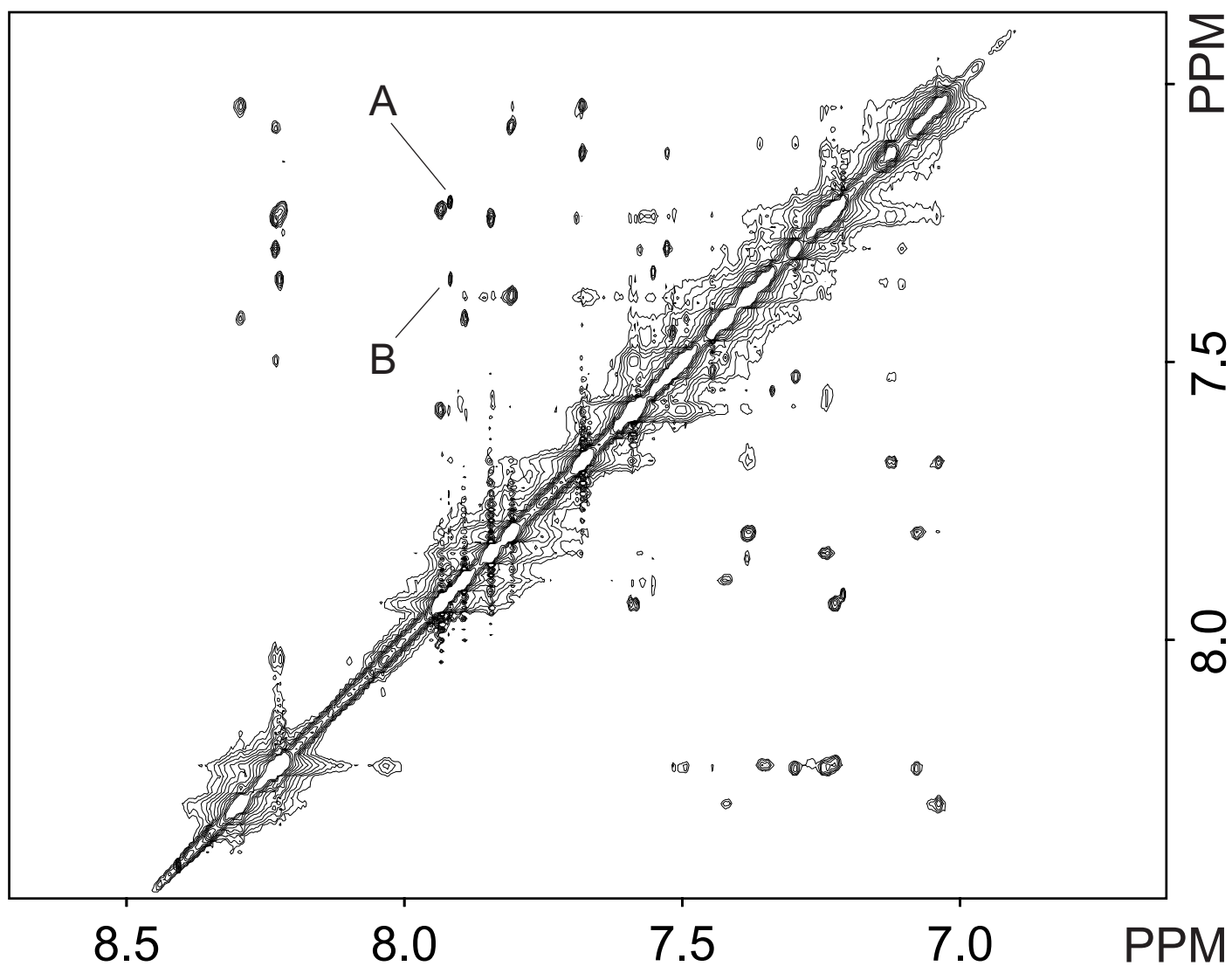


Figure 3S. Expanded contour plot showing the aromatic proton region of an 800 MHz NOESY spectrum (300 ms. mixing time) recorded in 100% D<sub>2</sub>O buffer at 25 °C. Labeled peaks are assigned as follows: A, Z-F6(HCO)-E-F6(HCO) exchange cross-peaks; B, Z-F6(HCO)-C5(H6).

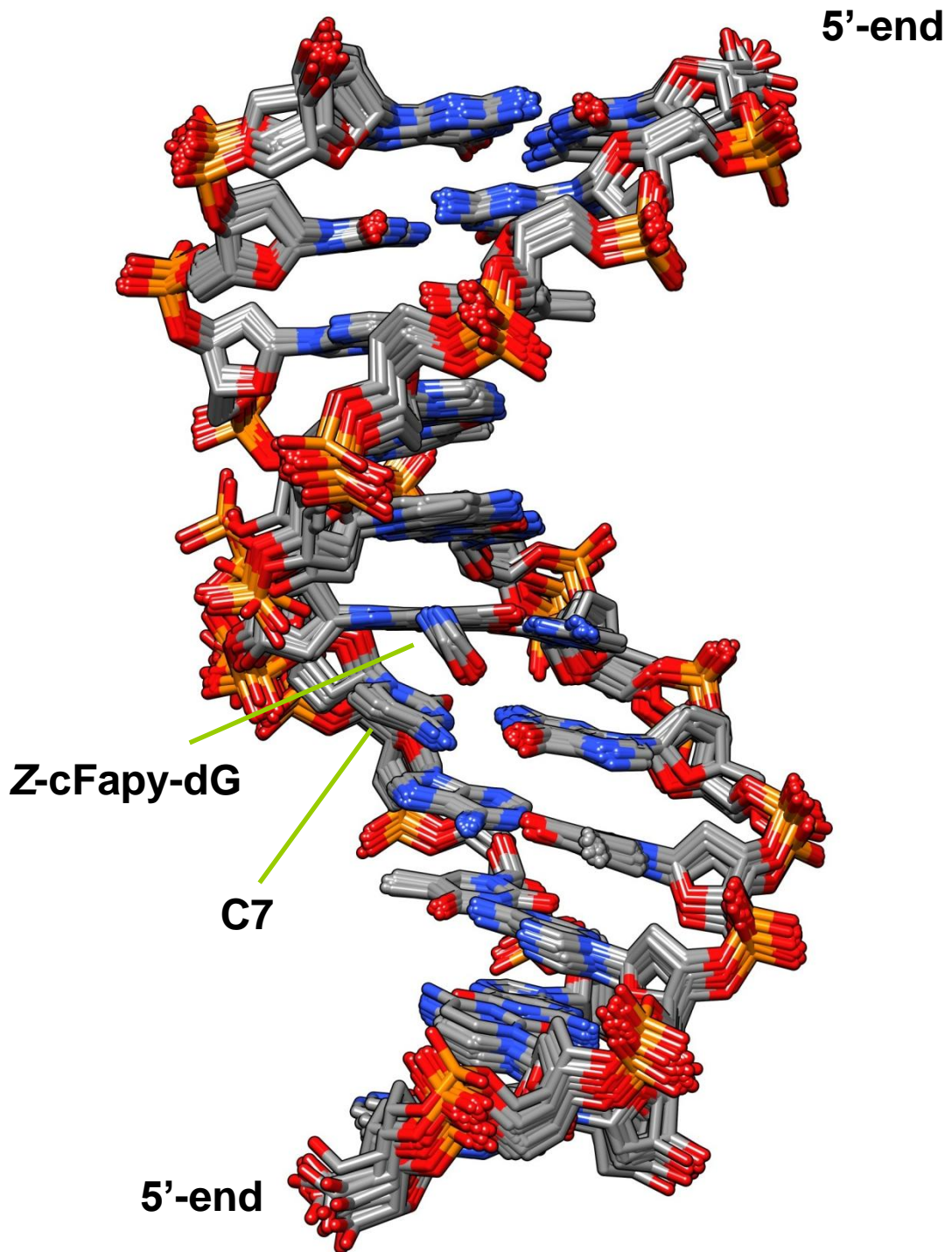


Figure 4S: Superposition of the final 25 structures of the Z-cFapy-dG•dC duplex seen with the major groove prominent.

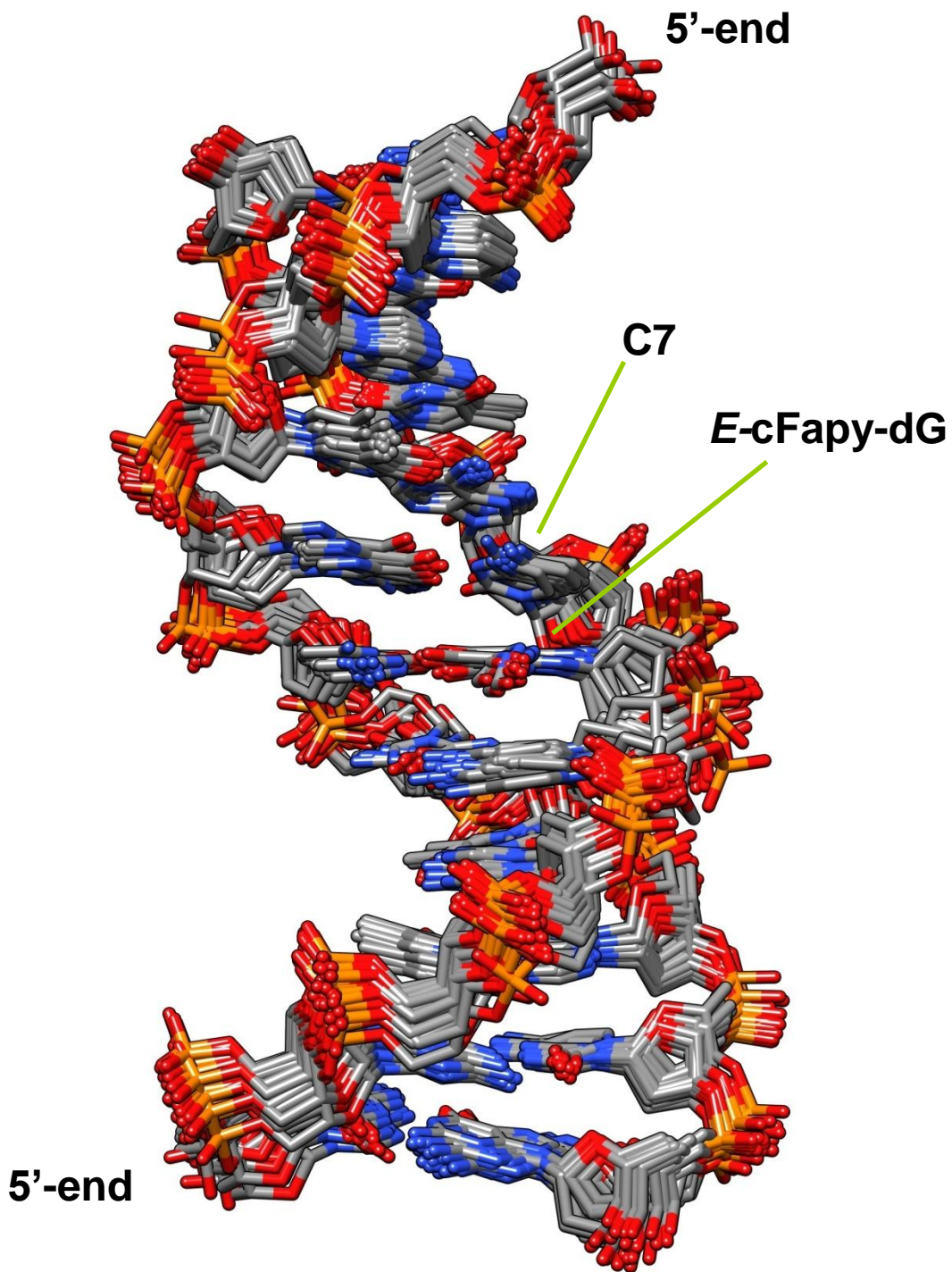
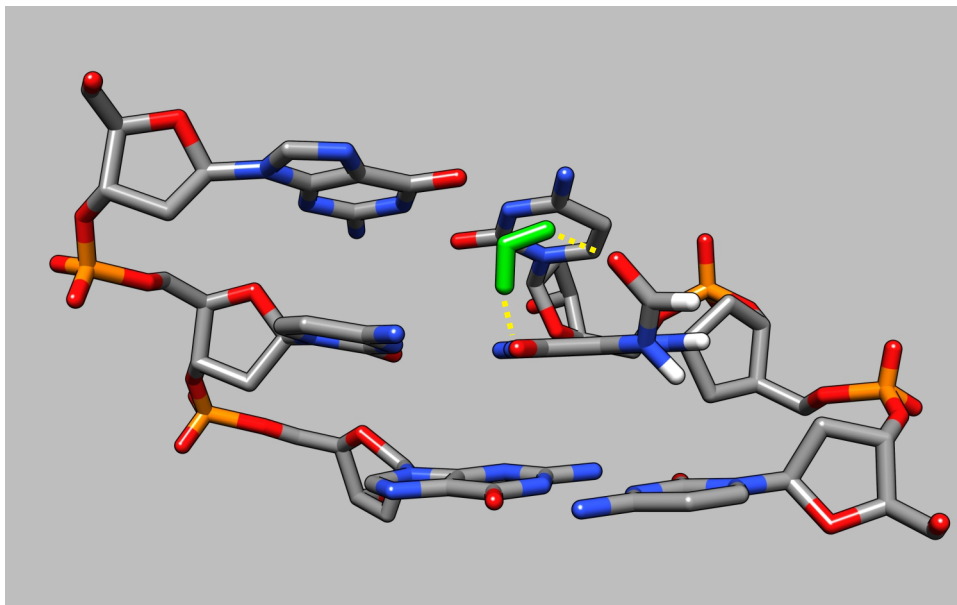
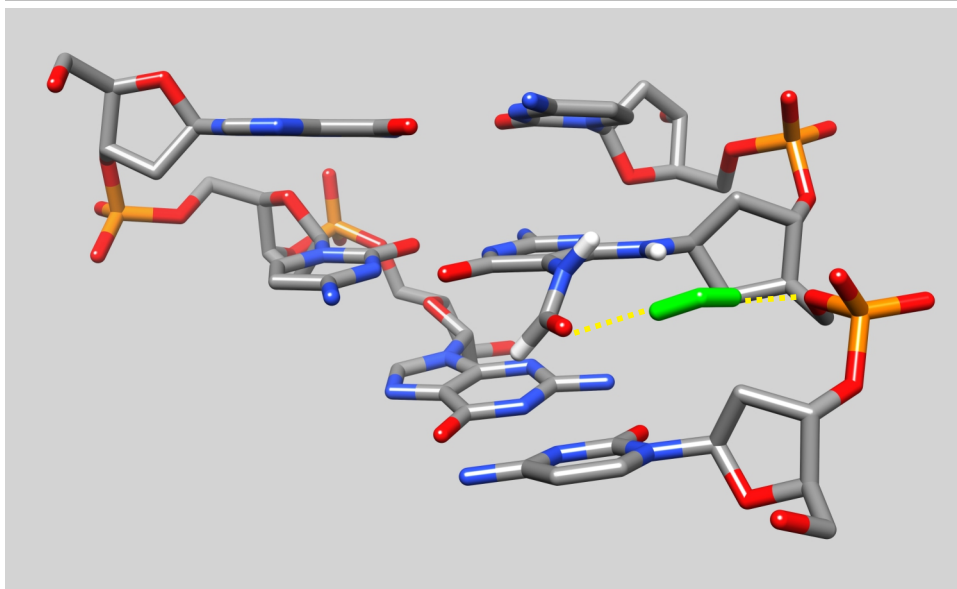


Figure 5S: Superposition of the final 25 structures of the *E*-cFapy-dG•dC duplex seen with the major groove prominent.

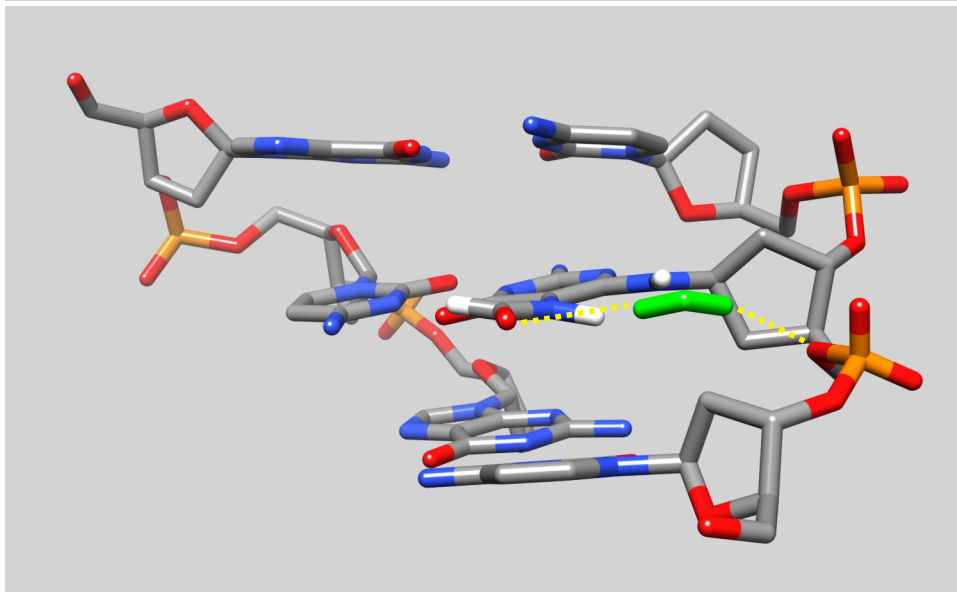




Z-cFapy-dG



*E*-cFapy-dG  
(major form)



*E*-cFapy-dG  
(minor form)

Figure 6S: Examples of water-mediated hydrogen bonds in the  $\beta$ -cFapy-dG•dC duplex. Water molecules are colored green.

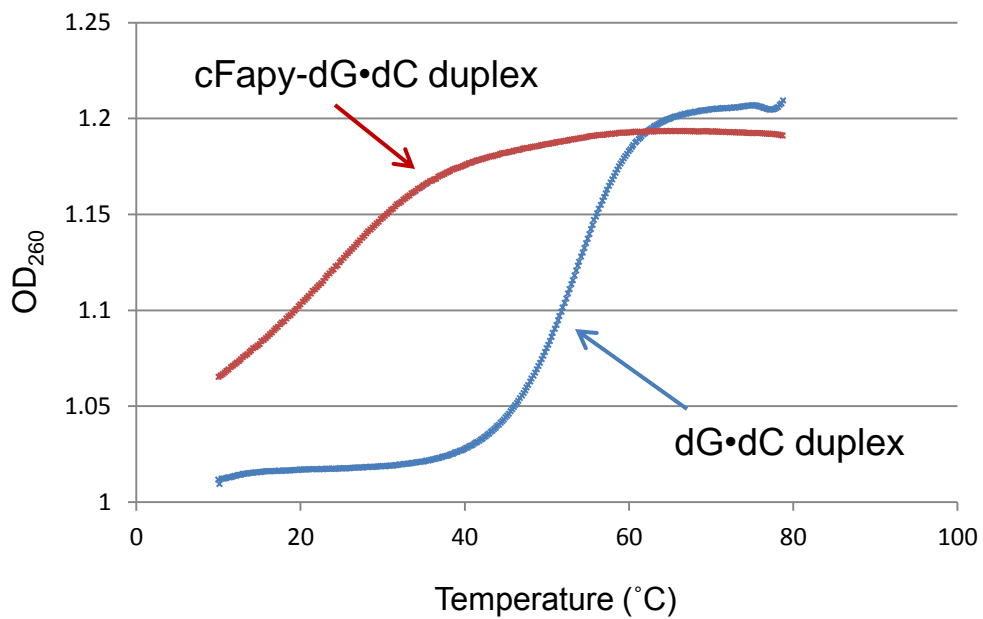


Figure 7S: UV melting profiles of the cFapy-dG•dC and dG•dC duplexes.