### **Supplemental Information**

## **LincRNA-p21 Suppresses Target mRNA Translation**

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#### INVENTORY of SUPPLEMENTAL INFORMATION

#### Supplemental text

Figure S1: Complements main Figure 1

Figure S2: Complements main Figure 1

Figure S3: Complements main Figure 2

Figure S4: Complements main Figures 3 and 4

**Legends of Supplemental Figures** 

Table S1

Table S2

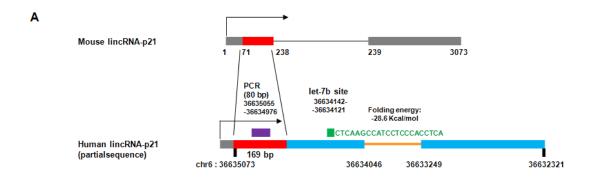
#### Supplemental text

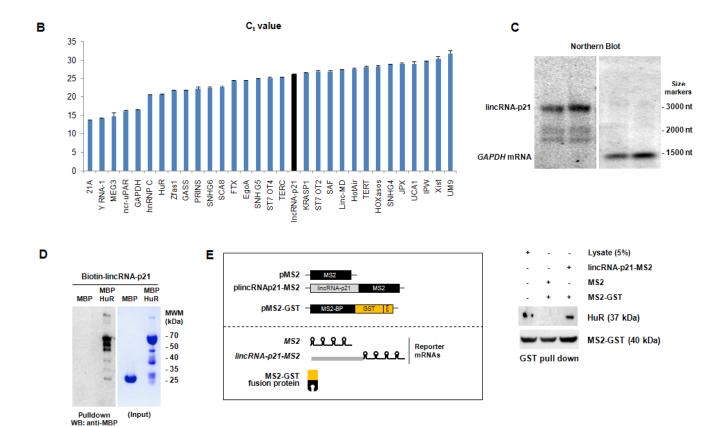
#### Human lincRNA-p21 partial sequence (1956 nt)

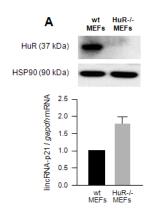
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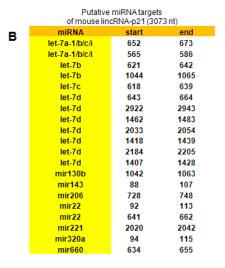
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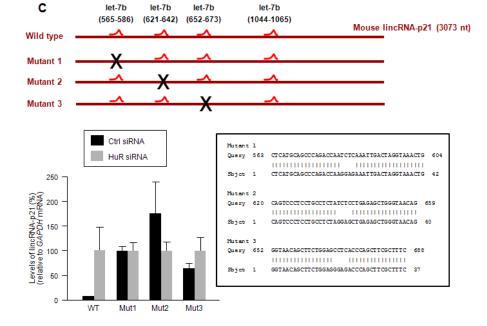
Rapid Amplification of cDNA Ends (RACE) was performed according to manufacturer's protocol (Invitrogen) using total RNA from HeLa cells.

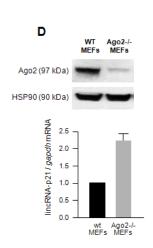




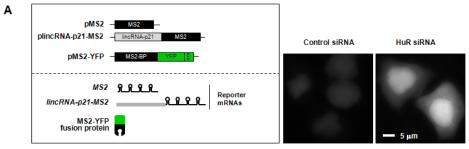


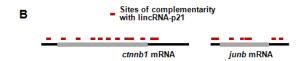


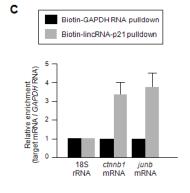


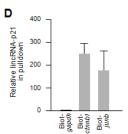


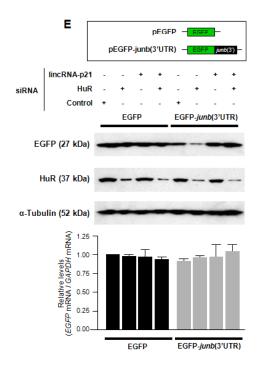
#### Yoon et al. Supplementary Fig. \$3

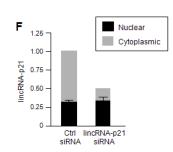


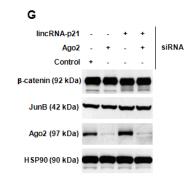


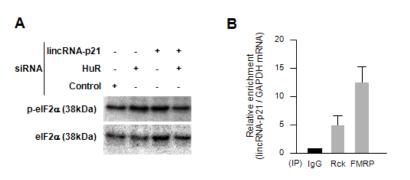




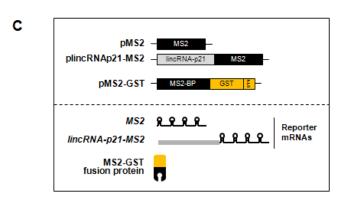


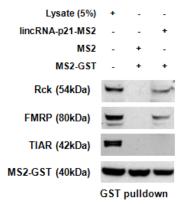


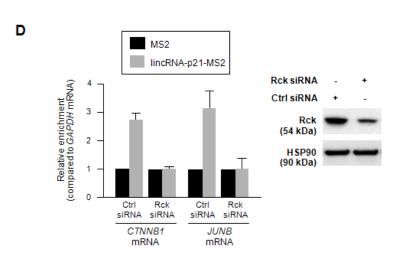


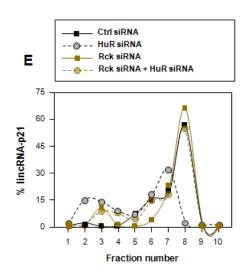


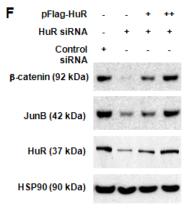
### Yoon et al. Supplementary Fig. S4

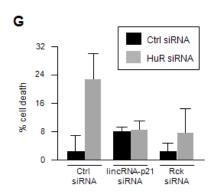












#### **Legends of Supplemental Figures**

# Figure S1 (complements Fig. 1). Additional characterization of mouse and human lincRNA-p21 and their interaction with HuR.

(A) Genomic localization of mouse (top) and human (bottom) lincRNA-p21. The human lincRNA-p21 was sequenced partially by RACE (Rapid Amplification of cDNA Ends; Invitrogen) using total RNA from HeLa cells (supplemental text). Red, region of homology between mouse and human lincRNA-p21. Purple, region used to detect human lincRNA-p21 by qPCR. Green, let-7b site, with the interaction sequence indicated in green text. (B) Relative abundance of lincRNA-p21 (black) as determined by calculating C<sub>t</sub> values among a subset of mRNAs (those encoding hnRNPC and HuR) and lincRNAs, as measured in HeLa cells. (C) Northern blot analysis of lincRNA-p21 expression in HeLa cells. Duplicate samples (20 and 30 µg of total RNA) were run in agarose/formaldehyde gels; after transfer to nylon membranes, lincRNA-p21 and GAPDH mRNA were detected using antisense oligomers (GGGTGGCTCACTCTTCTGGC and GCCCAATACGACCAAATCC respectively) that were end-labeled with <sup>32</sup>P and used as probes. Bands were visualized using a Typhoon scanner (GE Healthcare). (**D**) In vitro interaction of maltose-binding protein (MBP) or MBP-HuR (2 µg each) with biotinylated lincRNA-p21 was assessed by pulldown using streptavidin-coated beads and Western blotting (WB) detection using anti-MBP antibody. Input, 2 µg recombinant protein. As shown, MBP-HuR selectively interacted with biotinylated mouse lincRNA-p21, while MBP did not. MWM, protein molecular weight markers. (E) HeLa cells were transfected with the plasmids shown in the schematic (*left*). Forty-eight h later, lysates were used in IP reactions employing GSH-agarose beads. The presence of HuR in IP reactions from each transfection group (cells expressing MS2 RNA compared with cells expressing lincRNA-p21 RNA) was assessed by Western blot analysis using an anti-HuR antibody.

### Figure S2 (complements Fig. 1). Influence of HuR and let-7 on lincRNA-p21 levels.

(A) In wild-type (wt) and HuR-null (HuR-/-) MEFs, HuR levels were assessed by WB, and lincRNA-p21 by RT-qPCR. (B) Putative microRNAs interacting with mouse lincRNA-p21, as predicted using TargetScan. (C) Predicted let-7 sites on mouse lincRNA-p21. Using a plasmid vector that expressed the wild-type mouse lincRNA-p21 expressed from a CMV promoter (Huarte et al., 2010), site-directed mutagenesis (box) was used to generate the mutants indicated (Mut1, Mut2, Mut3). After transfection of each plasmid into MEFs expressing normal or silenced HuR, the levels of lincRNA-p21 (WT, Mut1, Mut2, Mut3) were quantified by RT-qPCR. Site-directed mutagenesis of mouse lincRNA-p21 was performed with the QuikChange® Site-Directed Mutagenesis Kit (Stratagene). (D) In wild-type (wt) and Ago2-null (Ago2-/-) MEFs, Ago2 levels were assessed by WB (top), and lincRNA-p21 by RT-qPCR (graph). Data in (A,C,D) represent the means and S.D. (error bars) from 3 independent experiments.

# Figure S3 (complements Fig. 2). Translational repression of $\beta$ -catenin and JunB translation by lincRNA-p21 in mouse cells.

(A) HeLa cells were transfected with the plasmids shown in the schematic. Forty-eight hours later, the localization of the MS2-tagged lincRNA-p21 was assessed by fluorescence microscopy. (B) Regions of predicted interaction between mouse lincRNA-p21 and mouse *ctnnb1* and *junb* mRNAs. (C) Following incubation of mouse biotin-lincRNA-p21 (or control biotin-*GAPDH* RNA) with MEF lysates, RNA was extracted and *ctnnb1* and *junb* mRNAs, as well as normalization control 18S rRNA were assessed by RT-qPCR and represented as fold difference relative to mRNA levels in biotin-*GAPDH* pulldown samples. (D) Biotinylated *GAPDH* mRNA, *ctnnb1* RNA (NM\_007614.3, nucleotides 288-2633), and *junb* RNA (NM\_008416.3, nucleotides 320-1354) were incubated with MEF lysates; after pulldown, the levels of lincRNA-p21 in beads were assessed by RT-qPCR. (E) MEFs were transfected with the plasmids indicated: pEGFP-junb(3'UTR), containing the entire mouse *junb* 3'UTR (amplified from mouse total RNA and cloned into pEGFP-C1), and control plasmid pEGFP, together with the siRNAs shown. Forty-eight h later, the levels of the reporter protein EGFP were detected by Western blot analysis and the levels of *EGFP* or *EGFP*-

junb(3'UTR) mRNAs by RT-qPCR analysis. (**F**) Forty-eight hours after siRNA transfection in HeLa cells, lincRNA-p21 levels were assessed by RT-qPCR in nuclear and cytoplasmic fractions as explained in main Fig. 2A. (**G**) Forty-eight hours after transfection with the siRNAs shown, the levels of Ago2, β-catenin, JunB and loading control HSP90 were assessed by Western blot analysis. Data in (C-F) represent the means and S.D. (error bars) from 3 independent experiments. Oligomer sequences are in Table 2.

<u>Microscopy</u>: forty-eight h after transfection of pMS2-YFP together with either pMS2 or pMS2-lincRNA-p21, HeLa cells were fixed and images were acquired with Axio Observer microscope (ZEISS) with Axio Vision 4.7 Zeiss image-processing software or with LSM 510 Meta (ZEISS).

# Figure S4 (complements Figs. 3 and 4). Translational inhibition by mouse lincRNA-p21 associated to the recruitment of translational repressor Rck.

(A) HeLa cells were transfected with the siRNAs shown and 48 h later, the levels of eIF2 $\alpha$  and phosphoeIF2α were assessed by Western blot analysis. (B) RIP analysis of lincRNA-p21 interaction with Rck and FMRP in MEFs. (C) Left, plasmids used to study the in vitro interaction of lincRNA-p21 with Rck, FMRP, and TIAR present in HeLa cell lysates, as detected by WB analysis in GST pulldown assays (right). (D) Mouse lincRNA-p21-MS2 and MS2-GST were expressed in HeLa cells using the plasmids shown in panel (C) in cells expressing normal Rck levels or cells in which Rck was silenced (right). RIP analysis was carried out to study if silencing Rck affected the interaction of lincRNA-p21 with endogenous CTNNB1 and JUNB mRNAs. (E) 48 h after transfecting HeLa cells with the siRNAs shown, polysomes were prepared as shown in the main Fig. 4E; the relative distribution of lincRNA-p21 is indicated. (F) In order to establish whether the effects of lincRNA-p21 were directly attributed to HuR, a rescue experiment was devised in which HeLa cells were transfected with siRNA directed to the 3'UTR of HuR or with a plasmid (1 and 2 µg) to express only the coding region of HuR (pFlag-HuR). Forty-eight h later, the levels of β-catenin and JunB were assessed by Western blot analysis. (G) Percentages of HeLa cell death (as assessed by trypan blue exclusion) 48 h after transfection of the siRNAs shown; cell number and viability were quantified using a TC10 automated cell counter (BioRad). Data in (B,D,G) represent the means and S.D. (error bars) from 3 independent experiments.

# Table S1. Putative Regions of interaction of lincRNA-p21 with human *CTNNB1* mRNA, *JUNB* mRNA, and (control) *GAPDH* mRNA.

I. Putative interaction regions of the CTNNB mRNA (query) with lincRNA-p21 (subject).

Matched sequence	Identity (%)	Mach Size	E-value
Query 3295 TTTTTAAGTCTCTCGTAGTGTTAA 3318 Sbjct 966 TTTTTAATTGTCCCTGTTAGTGTGAA 941 (intron)	76.92	26	54
Query 3441 TITTGTATAAAATAGAACAAATAGAA 3465 Sbjct 503 TTTTGACTAAACTACAAATAGAA 481	80	25	4.4
Query 879 ATACAAATGATGTAGAAACAG 899 Sbjct 195 ATGCAAAGCAAGTAGAAACAG 175	80.95	21	4.4
Query 3511 TGGATCTATTTCATGTTTT 3530 Sbjct 2192 TGGATCT-TCTCTTGGTTTT 2174	80	20	189
Query 1353 GAATGCAAGCTT-TAGGACT 1371 Sbjct 2307 GAAAGGAGGCTTCTAGGACT 2288	80	20	189
Query 3339 AGCAATTTCTAATTTTAA 3357 Sbjct 978 AGCCATTTGCAGTTTTTAA 960 (intron)	78.95	19	54
Query 297 TGGACATGGAAC 314 Sbjct 876 TGGACATGGAAC 859	83.33	18	15
Query 1509 TAAATGTGGTCACCTGTG 1526 Sbjct 1262 TAAATGTTCACCCGTG 1247 (intron)	83.33	18	54
Query 1350 GTGGAATGCAAGCTTTA 1366 Sbjct 806 GTGGAATACAGGCTGTA 790	82.35	17	54
Query 3060 AAACTTTTTGTTCTTGGT 3076 Sbjct 1042 AAACTTTTTCTTTGGT 1026 (intron)	82.35	17	54
Query 2885 CATTTGCTGTTTTAAA 2900 Sbjct 975 CATTTGCAGTTTTTAA 960 (intron)	87.5	16	4.4
Query 2173 AGCCACAGCTCCTCTG 2188 Sbjct 665 AGCCACAGCCTCTG 652	87.5	16	54
Query 247 TTTTGAAAA-TCCAG 260 Sbjct 217 TTTGAAAAATCCAG 203	93.33	15	15
Query 110 GGGAGGCGGACGC 124 Sbjct 1615 GGGAGGCTGAGGCGG 1601 (intron)	86.67	15	15
Query 1812 GAAATCTTGCCCTT 1826 Sbjct 2130 GAAATCTAGCCCCTT 2116	86.67	15	15
Query 2913 CCTTTCTCTTTAT 2927 Sbjet 2477 CCATTCTCTCTAT 2463	86.67	15	15
Query 1409 TGTCTT-TGGACTCT 1422 Sbjct 428 TGTCTTTTGGATTCT 414	86.67	15	189
Query 383 GCCACTA-CCACAGC 396 Sbjet 671 GCCACCAGCCAGC 657	86.67	15	189
Query 214 CCCTGAG-GGTATTT 227 Sbjct 2084 CCCTGGGTGGTATTT 2070	86.67	15	189
Query 1096 TGGTGGGCTGCAG 1108 Sbjct 2499 TGGTGGGCACTGCAG 2485	86.67	15	189
Query 384 CCACTAC-CACAGCT 397 Sbjet 2741 CCACTACGCCCAGCT 2727	86.67	15	189

# II. Putative interaction regions of the JUNB mRNA (query) with lincRNA-p21 (subject).

Matched Regions		Mach Size (bp)	E-value
Query 770 CCCCGGCTGGGCCCGGGGGGCGTCTACGCCGGCC 802 Sbjet 32 CCCGGCCAAGGCCAGCGGGAG-CTCTGCCGGCC 1	72.73	33	93
Query 1654 TGGGAAGGGGACCCCCCCCCCCCCCCCCCCCCCCCCCCC	80.77	26	0.18
Query 1559 AATATA-ATATAT-TTGTGTAT 1578	86.36	22	7.6
Query 956 CGGCGCAGCTGGGCTTGG 973 Sbjct 1767 CGGGCAGCTGGTCTGGG 1750	83.33	18	7.6
Query 1653 TTGGGAAGGGGACCCC 1669 Sbjct 2175 TTAGGAAGGGGCTCCC 2159	82.35	17	27
Query 11 GCCAGCCTCGGAGCCAG 27 Sbjct 2532 GCCAGCCTGGGGTCCAG 2516	82.35	17	27
Query 1564 AATATATTTGTGTATT 1579 Sbjct 1117 AATATATGTATATT 1102 (intron)	87.5	16	2.2
Query 1564 AATATATTTGTGTATT 1579 Sbjct 1898 AATATATTCTTGTATT 1883	87.5	16	2.2
Query 1053 CCCCATCA-ACATGGA 1067 Sbjct 2208 CCCCACCACACATGGA 2193	87.5	16	27
Query 1443 TGGACTCCGGCCCTC 1457 Sbjct 2672 TGGACTCCTGACCTC 2658	86.67	15	7.6

## III. Putative interaction regions of the GAPDH mRNA (query) with lincRNA-p21 (subject).

		Matched Regions	Identity (%)	Mach Size (bp)	E-value
Query Sbjct	431 2287	CTCATTTGCAGGGGGGGCC 450 CTCATTTGCATAGGGGTGCC 2268	85.00	20	0.44
Query Sbjct	98 1316	ACACCATGGGGAGG 112 ACAACATGGGGAGGG 1330 (intron)	86.67	15	5.4
Query Sbjct	506 2012	TGAACCATGAGAAGT 520 TGAACCATCATAAGT 2026	86.67	15	5.4

**Table S2. Primers used in qPCR reactions.** Bold text indicates T7 RNA polymerase promoter sequence.

Name	Species	Sequence	Reference
lincRNA-p21-F	human	GGGTGGCTCACTCTTCTGGC	Huarte et al., 201
lincRNA-p21-R	human	TGGCCTTGCCCGGGCTTGTC	Huarte et al., 201
GAPDH-F	human	AGCCACATCGCTCAGACAC	Huarte et al., 201
GAPDH-R	human	GCCCAATACGACCAAATCC	Huarte et al., 201
mLincRNA-p21-1 (F)		CCTGTCCACTCGCTTTC	
1 (7	mouse		Huarte et al., 201
mLincRNA-p21-1 (R)	mouse	GGAACTGGAGACGGAATGTC	Huarte et al., 201
mGapdh (F)	mouse	GGGAAATTCAACGGCACAGT	Huarte et al., 201
mGapdh (R)	mouse	AGATGGTGATGGGCTTCCC	Huarte et al., 201
JUNB human (F)	human	AGGCTCGGTTTCAGGAGTTT	
JUNB human (R)	human	GAACAGCCCTTCTACCACGA	
junb mouse (F)	mouse	CGATAGGGATCCGCCAGG	
junb mouse (R)	mouse	CGGGATACGGTCGGAGC	
b catenin human (F)	human	ATTGTCCACGCTGGATTTTC	
b catenin human (R)	human	TCGAGGACGGTCGGACT	
b catenin mouse (F)	mouse	ATGGAGCCGGACAGAAAAGC	
b catenin mouse (R)	mouse	GAATCCAAGTAAGACTGCTGCT	
18S rRNA (F)	human	CGAACGTCTGCCCTATCAACTT	
18S rRNA (R)	human	ACCCGTGGTCACCATGGTA	
T7 beta catenin (F)	mouse	CCAAGCTTCTAATACGACTCACTATAGGGAGAATGGCTACTCAAGCTGACCTG	
T7 beta catenin (R)	mouse	TTACAGGTCAGTATCAAACCAGGC	
T7 JunB (F)	mouse	CCAAGCTTCTAATACGACTCACTATAGGGAGAATGTGCACGAAAATGGAACA	
T7 JunB (R)	mouse	TCAGAAGGCGTGTCCCTT	
T7 HO1 (F)	mouse	CCAAGCTTCTAATACGACTCACTATAGGGAGAATGGAGCGTCCACAGCCC	
T7 HO1 (R)	mouse	TTACATGGCATAAATTCCCACTG	
T7 GAPDH ORF (F)	mouse	CCAAGCTTCTAATACGACTCACTATAGGGAGAATGGTGAAGGTCGGTGTGA	
T7 GAPDH ORF (R)	mouse	TTACTCCTTGGAGGCCATGT	
. ,			
Mut 2073-2094 (F)	mouse	ACTICTCAACATCTCTTGTGCTGTGTGTGTGTGTCACACACACA	
Mut 2073-2094 (R)	mouse	GCTGGTGTGTGTGTGTGACACACACACACACACACACACA	
Mut 2457-2496 (F)	mouse	AAACAGCTGTGGTGCAGGCCTAGGGGCCCGGCGCCCGAAGCTGGACACTGGCAGAGGCCG	
Mut2457-2496 (R)	mouse	GCGGCCTCTGCCAGTGTCCAGCTTCGGGCGCCCCGGGCCCCTAGGCCTGCACCACAGCTGTTT	
B-Catenin 5UTR Xho1 (F)	mouse	AAAACTCGAG GCGCGGCAACGCTCCGCGC	
B-Catenin 5UTR EcoR1 (R)	mouse	AAAAGAATTCTGTCCACGCAGCGGTGTCACA	
JunB 3UTR Kpn1 (F)	mouse	AAAAGGTACCTAAGAGCCTCCCTTGCCCCATACG	
JunB 3UTR BamH1 (R)	mouse	AAAAGGATCC ATTAATAAAGACAATTTTTT	
EGFP (F)		TAAACGGCCACAAGTTCAGCGT	
EGFP (R)		AAGTCGTGCTTCATGTGGT	
lincRNA-p21 mutant 1(F)	mouse	GCAGCCCAGACCAAGGAAAATTGACTAGGTAAAC	
lincRNA-p21 mutant 1(R)	mouse	GTTTACCTAGTCAATTTCTCCTTGGTCTGGGCTGC	
lincRNA-p21 mutant 2(F)	mouse	CCTCCTGCCTCTAGGAGCTGAGAGCTG	
lincRNA-p21 mutant 2(R)	mouse	CAGCTCTCAGCTCCTAGAGGCAGGAGG	
lincRNA-p21 mutant 3(F)	mouse	GTAACAGCTTCTGGAGGGAGACCCAGCTTCGCTTTCAC	
lincRNA-p21 mutant 3(R) 5'biotinylated sense (for pulldown)	mouse human	GTGAAAGCGAAGCTGGGTCTCCCTCCAGAAGCTGTTAC GGGTGGCTCACTCTTCTGGC	
5'biotinylated antisense (for pulldown)	human	GCCAGAAGAGTGAGCCACCC	