Supplementary data

Removal of a frameshift between the *hsdM* and *hsdS* genes of the EcoKI Type IA DNA restrictionmodification (RM) system produces a new type of RM system and explains the origins of the different families of Type I RM systems.

Gareth A. Roberts^{1, +}, Kai Chen^{1, +}, Laurie P. Cooper^{1, +}, John H. White^{1, +}, Garry W. Blakely², David T.F. Dryden^{1, *}

¹ EastChem School of Chemistry, The University of Edinburgh, The King's Buildings, Edinburgh, EH9 3JJ, UK.

² Institute of Cell Biology, School of Biological Sciences, The University of Edinburgh, The King's Buildings, Edinburgh EH9 3JR, UK.

Supplementary Figure S1. *E. coli* NM679 (r^ms⁻) transformed with pJFMS and pBS17 (R subunit) or pMSFusion and pBS17. Phage $\lambda v.o$ at various dilutions was then spotted on each strain.

	<i>E. coli</i> NM679 (r ⁻ m ⁻ s ⁻) transformed with:				
Phage λ spot dilutions	pJFMS	pJFMS + pBS17	pMSFusion	pMSFusion + pBS17	
(-1) (-2) (-3) (-4)		00	000	00	
(-5) (-6) (-7) (-8)	20 C		4		

Supplementary Figure S2. *E. coli* NM679 (r^{-m-s-}) transformed with pJFMS or pMSFusion. Phage $\lambda v.k$ and $\lambda v.o$ were then spotted on these strains at various dilutions.

	<i>E. coli</i> NM679 (r m s) transformed with:				
	pJFMS		pMSFusion		
Phage λ					
spot	λv.k	λv.o	λv.k	λν.ο	
dilutions					
-1 -2	00	00		00	
-3 -4	00	00	90	00	
-5 -6		00		00	
-7 -8	and a	L.S.		Not .	

Supplementary Figure S3. *E. coli* NM679 (r⁻m⁻s⁻) transformed with plasmids pBS17 (HsdR subunit) and pJFMS, pBS17 and pMSFusion, and with pBS17 alone, was spotted with phage $\lambda v.k$. The eop was identical for pBS17+pJFMS, pBS17+pMSFusion and pBS17.

E. coli NM6			
pBS17 + pJFMS	pBS17 + pMSFusion	pBS17	Phage λ spot dilutions
00	00	00	(-1) (-2) (-3) (-4)
			(-5) (-6) (-7) (-8)

Supplementary Figure S4. Purification of MSFusion by column chromatography as visualised by Coomassie Blue staining of SDS-PAGE gels.

250,150,100,75,50,37,25,20,15,10

Purification step 1 - DEAE Column. M = Marker (250, 150, 100, 75, 50, 37, 25, 20, 15, 10 kDa), CE = Cell Extract, Lanes 1-12 are elution fractions from the salt gradient. Pooled fractions 3 and 4 for the heparin column.





Purification step 2 - Heparin Affinity Column. M = Marker (kDa as above), D = DEAE pooled sample, FT = unbound material from heparin column. Lanes 1-12 are elution fractions from the salt gradient. Pooled fractions 8 to 11 as pure MSFusion.



M D FT 1 2 3 4 5 6 7 8 9 10 11 12

Supplementary figure S5. a. Amino acid sequences of the C-terminus of M subunit fused to N-terminus of S subunit for EcoKI and EcoR124I. The sequence in red is from the M subunit and in black from the S subunit. b. BLAST output using the fused EcoKI sequence from part a. pulls out the central conserved region of an S subunit in Fusobacterium nucleatum. c. BLAST output using the fused EcoR124I sequence from part a. pulls out the central conserved region of several S subunits from different bacteria.

a.

```
>MS.EcoKI
DSEENKNTDQ HLATSRWRKF SREWIRTAKS DSLDISWLKD KDSIDADSLP EPDVLAAEAM GELVQALSEL DALMRELGAS DEADLQRQLL
EEAFGGVKE MSAGKLPEGW VIAPVSTVTT LIRGVTYKKE QAINYLKDDY LPLIRANNIQ NGKFDTTDLV FVPKNLVKES QKISPEDIVI
AMSSGSKSVV GKSAHQHLPF
```

>MS.EcoR124I QVFASKEDVA HLAKSVAFET VVANDYNLSV SSYVEAKDNR EIIDIAELNA ELKTTVSKID QLRKDIDAIV AEIEGCEVQK MSEMSYLEKL LDGVEVEWLP LGEITKYEQP TKYLVKAKDY HDTYTIPVLT AGKTFILGYT NETHGIYQAS KAPVIIFDDF TTANKWVDFD FKAKSSAMKM

b.

```
ref|ZP_02273165.1|type I restriction modification enzyme protein S [Fusobacterium
nucleatum subsp. polymorphum ATCC 10953]
Length=387
Score = 52.4 bits (124), Expect = 3e-05, Method: Compositional matrix adjust.
Identities = 47/201 (23%), Positives = 87/201 (43%), Gaps = 22/201 (10%)
           ENKNTDQHLATSRWRKFSREWIRTAKSDSLDISWLKDKDSIDADSLPEPDVLAAEAMGEL
                                                                     93
Ouerv 34
                                A S ++ I +K S + + P++
           +NKN D
                       +
                                                             + + +
Sbjct 107 KNKNVDLLYLYFYLKTIKNNLESIASSTTVKII---NKSSFEKIEISLPNLEIQKKISKK 163
Query 94
           VQAL-SELDALMRELGASDEADLQRQLLEEAFGGVKEMSAGKLPEGWVIAPVSTVTTLIR 152
           ++ L + +D +L E L + L FG +K
                                                + W T +
                                                                + T
Sbjct 164 LELLENNIDFRKNQLNYLKE--LNKSLFTRMFGDIKTND----KNWKIVKLEKYINIIG
                                                                     216
Query 153 GVTYKKEQAINYLKDDYLPLIRANNIQNGKFDTTDLVFVPKNLVKESQKISPEDIVIAMS
                                                                     212
           G +K K +PLIR NI +G+F +T+LVF+ +N E K+ P DI+I+++
Sbjct 217 GYAFKNID----FKSSGIPLIRIGNINSGQFKSTNLVFIEENKKFEKFKVFPNDILISLT 272
Query 213 S-----GSKSVVGKSAHQH 226
                  G+ ++G S ++
Sbjct 273 GTVGKDDYGNACILGDSYSEY 293
```

c.

```
ref[ZP_00682365.1] similar to Restriction endonuclease S subunits [Xylella fastidiosa
Ann-1]
       122 bits (306), Expect = 2e-26, Method: Compositional matrix adjust.
Score =
Identities = 78/158 (49%), Positives = 91/158 (57%), Gaps = 28/158 (17%)
          NDYNLSVSSYVEAKDNREIIDIAELNAELKTTVSKIDQLRKDIDAIVAEIEGCEVQKMSE 123
Query 64
                     +A NE DAEN T++ ID+L IA+ E
            D L
Sbjct 95
           RDALLRFGGSTDASGNGE--DGAERNQWKPTGINWIDEL---IAALCPE-----
                                                                      138
Query 124 MSYLEKLLDGVEVEWLPLGEITKYEQPTKYLVKAKDYHDTYTIPVLTAGKTFILGYTNET
                                                                      183
                  GVE + L GE+ YEQP KYLV + Y ++Y PVLTAG+TFILGYT+ET
Sbict 139 -----GVEFKML--GELLDYEOPGKYLVASTAYDNSYWTPVLTAGOTFILGYTDET 187
Query 184 HGIYQAS-KAPVIIFDDFTTANKWVDFDFKAKSSAMKM 220
            GIY AS + PVIIFDDFTTA KWVDF FKAKSSAMKM
Sbjct 188 SGIYAASPQEPVIIFDDFTTAFKWVDFPFKAKSSAMKM 225
Red is the M.EcoR124I sequence
```

Supplementary figure 5 c cont.

```
ref|YP_001181695.1| restriction modification system DNA specificity domain [Shewanella
putrefaciens CN-321
Score = 118 bits (296), Expect = 2e-25, Method: Compositional matrix adjust.
Identities = 74/149 (49%), Positives = 96/149 (64%), Gaps = 14/149 (9%)
Ouerv 83
          IDIAELNAELKTTVSKI-DQLRKDIDAIVAEIEGCEVQKMSEMSYL----EKLLD----G 133
           IDI L + K ++KI K I I+ + QK + M L ++LLD
Sbjct 157 IDIPPLPEQRK--IAKILSTWDKAISTTERLIDNSKQQKKALMQQLLTAKKRLLDDSGKP 214
Query 134 VEVEW--LPLGEITKYEQPTKYLVKAKDYHDTYTIPVLTAGKTFILGYTNETHGIYQASK 191
            E EW + LG++ Y+OPT YLVK+ DY + Y+IPVLTAGKTFILGY+NE GI++ +
Sbjct 215 FEGEWTKVELGKLLDYKQPTPYLVKSTDYSNEYSIPVLTAGKTFILGYSNENFGIFE-EE 273
Query 192 APVIIFDDFTTANKWVDFDFKAKSSAMKM 220
            P IIFDDFTTA+K+VDF FKAKSSAMK+
Sbjct 274 LPAIIFDDFTTASKFVDFPFKAKSSAMKI 302
Red is the M.EcoR124I sequence
ref|NP_858129.1| putative type i restriction hsds subunit [Lactobacillus delbrueckii
subsp. lactis]
Score = 111 bits (278), Expect = 3e-23, Method: Compositional matrix adjust.
Identities = 62/141 (43%), Positives = 90/141 (63%), Gaps = 9/141 (6%)
Ouerv 85
          IAELNAELKTTVSKIDQLRKDIDAIVAEIEGCEVQKM--SEMSYLEKLLDGVEVEW--LP 140
           IA +EL T ++ ++ ++ + + + + + + QKM + Y +G EW
Sbjct 171 IANFFSELDTAITLHEEKKQQLKCLKSAL----LQKMFAYKSGYPAIRFEGFSDEWEQCK 226
Query 141 LGEITKYEQPTKYLVKAKDYHDTYTIPVLTAGKTFILGYTNETHGIYQAS-KAPVIIFDD 199
           LGE+ YEQPTKY+VK+ +Y D + PVLTAGK+F+LGYT+E GI A+ + PV+IFDD
Sbjct 227 LGEVFNYEQPTKYIVKSTEYDDNFNTPVLTAGKSFLLGYTDEISGIKNATVENPVVIFDD 286
Query 200 FTTANKWVDFDFKAKSSAMKM 220
           FTT + +VDF FK KSSAMK+
Sbjct 287 FTTDSHYVDFPFKIKSSAMKL 307
```

Red is the M.EcoR124I sequence