Supporting Information

for

Total synthesis and biological evaluation of fluorinated cryptophycins

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Full experimental procedures and detailed analytical data for the synthesis of all compounds

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1 General information

All chemicals were purchased from Sigma Aldrich (Hamburg, Germany), Acros (Geel, Belgium), Alfa Aesar (Ward Hill, USA) and VWR (Darmstadt, Germany), and were employed without additional purification. Moisture- and air-sensitive reaction steps were conducted in flame-dried glassware and under argon atmosphere. Dichloromethane and toluene were freshly distilled from CaH₂ and Na, respectively. Analytical RP-HPLC was performed on a Thermo Separation Products system equipped with a UV-6000 LP detector, a P-4000 pump, a Vydac high-performance guard column (C18) and a Phenomenex Jupiter 5 μ m (C18; 250 × 4.6 mm). A flow rate of 0.7 mL min⁻¹ using eluent A: H₂O/CH₃CN/TFA (95/5/0.1) and eluent B: CH₃CN/H₂O/TFA (95/5/0.1) was employed.

Method M1:

0 min	100% A	0% B
0–5 min	0% A	100% B
5–6 min	100% A	0% B
5–6.5 min	100% A	0% B

Preparative RP-HPLC was performed on a Thermo Separation Products system equipped with a UV-1000 detector, a P-4000 pump, a Vydac high-performance guard column (C18) and a Phenomenex Jupiter 10 μ m 300 Å column (C18; 250 × 21.20 mm). A flow rate of 7.5 mL min⁻¹ using eluent A: H₂O/CH₃CN/TFA (95/5/0.1) and eluent B: CH₃CN/H₂O/TFA (95/5/0.1) was employed.

Method M2:

0–3 min	70% A	30% B
3–20 min	30% A	70% B
20–50 min	30% A	70% B
50–55 min	70% A	30% B

In addition, a Hitachi Merck LaChrom system equipped with a UV-vis L-7420 detector, a L-7150 pump, a Vydac high-performance guard column (C18) and a Phenomenex Jupiter 10 μ m 300 Å column (C18; 250 × 21.20 mm) was used. Method M3:

0–3 min	80% A	20% B
3–13 min	0% A	100% B
13–35 min	0% A	100% B
35–50 min	80% A	20% B

For the chiral analytical HPLC a Thermo Separation Products system equipped with a UV-6000 detector, a P-4000 pump and a Daicel Chiralpak 10 μ m column (C18; 250 × 4.60 mm) was used. A flow rate of 1 mL min⁻¹ using an eluent consisting of hexane/isopropanol (9/1) was employed (method M4).

MALDI-TOF mass spectra were measured on a Voyager DE Instrument (PE Biosystems, Weiterstadt, Germany) mounted with a 1.2 m flight tube. 2,5-Dihydroxybenzoic acid was used as the matrix. Depending on the mass range the ions were accelerated at 15 to 25 kV with the option of detecting positive or negative ions. The instrument default calibration was used for calibrating the mass axis. ESI experiments were performed on a Fourier Transform Ion Cyclotron Resonance (FT-ICR) mass spectrometer APEX III (Bruker Daltonik, Bremen, Germany) equipped with a 7.0 T, 160 mm bore superconducting magnet (Bruker Analytik GmbH – Magnetics, Karlsruhe, Germany), infinity cell, and interfaced to an external (nano)ESI or MALDI ion source. Scan accumulation and Fourier transformation were done with XMASS NT (7.08) on a PC workstation, for further data processing DataAnalysis[™] 3.4 was used. Optical rotation was measured on a DIP-360 digital polarimeter (Jasco, Groß-Umstadt, Germany) at 20–23 °C. NMR spectra were recorded at 298 K in CDCl₃ on a DRX 500 (¹H: 500 MHz, ¹³C: 126 MHz, ¹⁹F: 470 MHz), an Avance III 500 (¹H: 500 MHz, ¹³C: 126 MHz, ¹⁹F: 470 MHz), an Avance III 500 (¹H: 500 MHz, ¹³C: 126 MHz, ¹⁹F: 470 MHz), and an Avance 600 spectrometer (¹H: 600 MHz, ¹³C: 151 MHz, ¹⁹F: 564 MHz) (Bruker Biospin, Rheinstetten, Germany). The acronyms uA, uB, uC and uD describe signals pertaining to cryptophycin-units A–D.

2 General procedures

General procedure GP1 – Condensation of units A-B with units C–D

A mixture of the units A–B (1.0 equiv), the units C–D (1.0–1.5 equiv) and DMAP (0.25 equiv) was dried for several hours under high vacuum and then dissolved in absolute THF (10 mL/mmol) under an argon atmosphere. After addition of absolute triethylamine (2.0 equiv) the solution was cooled to 0 °C and then 2,4,6-trichlorobenzoylchloride (1.50–2.50 equiv) was added. The reaction mixture was stirred for one further hour at 0 °C, and the conversion was monitored by TLC. Then, 10% citric acid solution (17 mL/mmol) was added, the mixture was allowed to warm up to room temperature, and the aqueous phase was extracted three times with EtOAc (3 × 55 mL/mmol). The combined organic extracts were washed with saturated NaHCO₃ solution (25 mL/mmol) and brine (25 mL/mmol), dried over MgSO₄ and evaporated to dryness in vacuum. The crude product was purified by flash chromatography or preparative RP-HPLC.

4

General procedure GP2 – Macrolactamization

After addition of absolute piperidine (5.0 equiv) at room temperature to a solution of the acyclic depsipeptide (1.0 equiv in absolute DMF, 30 mL/mmol), the reaction mixture was stirred overnight in the dark, and the conversion was monitored by TLC. Subsequently, the reaction mixture was diluted with ethyl acetate (300 mL/mmol) and washed with H_2O (400 mL/mmol). The aqueous phase was extracted with ethyl acetate (3 × 200 mL/mmol) and the combined organic layers were washed with brine (100 mL/mmol), dried over MgSO₄ and evaporated to dryness in vacuum. The residue was purified by flash chromatography.

General procedure GP3 – Cleavage of the acetonide

Trifluoroacetic acid (10 mL/mmol) and water (5 drops) were added at 0 °C to a solution of the acetonide (1.0 equiv) in absolute dichloromethane (10 mL/mmol). The reaction mixture was stirred at this temperature while the reaction progress was monitored by TLC. After complete conversion (2–3 h) the solvent was removed in vacuum without heating the bath, the residue was dissolved in EtOAc (500 mL/mmol), and saturated NaHCO₃ solution (750 mL/mmol) was added. After phase separation the aqueous layer was extracted with EtOAc (3 × 250 mL/mmol). The combined organic layers were dried over MgSO₄, filtered and evaporated to dryness in vacuum. The resulting diol was dried under high vacuum and subjected to further reactions without purification.

General procedure GP4 – Synthesis of the cyclic orthoformate

The diol resulting from GP3 and PPTS (2.5 equiv) were dried overnight under high vacuum. Under argon atmosphere absolute dichloromethane (30 mL/mmol) and absolute trimethyl orthoformate (10 mL/mmol) were added. The reaction mixture was

stirred for two hours at room temperature while the reaction progress was monitored by TLC or analytical RP-HPLC. The reaction mixture was then filtered through a thin plug of silica gel, which was washed with $EtOAc/CH_2Cl_2$ (300 mL, 1:1 v/v). The combined filtrates were evaporated to dryness in vacuum. The resulting cyclic orthoformate was subjected to further conversions after drying under high vacuum without purification.

General procedure GP5 – Synthesis of the bromohydrin formate

The orthoformate (1.0 equiv) was dissolved in absolute CH_2Cl_2 (15 mL/mmol) and a 0.5 M solution of acetyl bromide in absolute CH_2Cl_2 (2.5 equiv acetyl bromide) was added. The resulting solution was stirred for four hours at room temperature while the reaction progress was monitored by TLC or analytical RP-HPLC. Subsequently, the reaction mixture was diluted with absolute CH_2Cl_2 (10 mL) and then poured into an ice-cold mixture of saturated NaHCO₃ solution and ice water (50 mL, 1:1, v/v). The mixture was washed with absolute CH_2Cl_2 and after phase separation the aqueous layer was extracted with CH_2Cl_2 (3 × 20 mL). The organic layers were combined, dried over MgSO₄, filtered and evaporated to dryness. The resulting bromohydrin formate was dried under high vacuum and subjected without further purification to the next reaction step.

General procedure GP6 – Synthesis of the epoxide

To the bromohydrin formate was added under vigorous stirring an emulsion of 0.2 M K_2CO_3 in ethylene glycol/DME. The reaction mixture was stirred for 3 min and then rapidly diluted with absolute CH_2Cl_2 (20 mL). The solution was transferred to a separating funnel containing ice-cold 0.5% aqueous KHSO₄ solution (20 mL). The reaction flask was rinsed with absolute CH_2Cl_2 . The aqueous phase was extracted

with CH_2Cl_2 (2 × 20 mL) and the organic layers were combined and collected over solid MgSO₄. The organic layer should be separated from the aqueous phase as quickly as possible to avoid hydrolysis of the epoxide in aqueous acidic medium. For that reason, the formation of two clear phases was not awaited. After drying of the combined organic layers over MgSO₄ the solvent was evaporated in vacuum and the residue was purified by column chromatography.

3 Experimental procedures and compound characterization 3.1 Synthesis of the trifluoromethyl-substituted unit A building block Methyl 4-trifluoromethylphenylacetate (8)

 $M = 218.17 \text{ g mol}^{-1} \text{ C}_{10}\text{H}_9\text{F}_3\text{O}_2$

For the synthesis of **8** a procedure by Bodnar [1] was adapted. 4-Trifluoromethylphenylacetic acid (**7**) (5.00 g, 24.49 mmol, 1.0 equiv) was dissolved in MeOH p.a. (260 mL) and cooled to 0 °C. Thionylchloride (4.00 mL, 56.33 mmol, 2.3 equiv) was added during 30 min with cooling to 0 °C and the reaction mixture was stirred for further 30 min at the same temperature. The mixture was allowed to warm up to rt and was stirred overnight. Then, H₂O (50 mL) was added and stirring was continued for 10 min. Methanol was evaporated in vacuum (bath temperature = 30 °C) and the residue was taken up in H₂O (50 mL) and CH₂Cl₂ (200 mL). The phases were separated and the aqueous phase was extracted with CH₂Cl₂ (4 × 100 mL). The combined organic layers were washed with sat. NaHCO₃ solution (2 × 150 mL) and H₂O (100 mL) and subsequently dried over MgSO₄. The solvent was evaporated in vacuum and 4.37 g (82%) of the ester **8** was obtained as a colorless oil, which can be subjected to the next steps without further purification. The spectroscopic data were in agreement with literature values [1].

HPLC: $t_{\rm R}$ 4.380 min (method M1); ¹H NMR (500 MHz, CDCl₃, TMS) δ 3.69 (s, 2H, CH₂), 3.71 (s, 3H, CH₃), 7.40 (d, J = 8.1 Hz, 2H, C^{ar}H), 7.59 (d, J = 8.1 Hz, 2H, C^{ar}H).

(4-Trifluoromethylphenyl)acetaldehyde (9)

$$M = 188.15 \text{ g mol}^{-1} \text{ C}_9\text{H}_7\text{F}_3\text{O}$$

Adapting a procedure by Eißler et al. [2], methyl 4-trifluoromethylphenylacetate ($\mathbf{8}$, 3.08 g, 14.11 mmol, 1.0 equiv) was dissolved in abs. CH₂Cl₂ (85 mL) and the mixture

was cooled to -78 °C. A solution of DIBAL-H (1.0 M in CH₂Cl₂, 42.0 mL, 42.00 mmol, 3.0 equiv) was added dropwise at -78 °C during 60 min and the solution was stirred at this temperature for 3.5 h. Then, it was cooled to -100 °C and the reaction was quenched by the addition of MeOH (18 mL). After warming up to rt, 1 M AcOH (160 mL) and hexane (160 mL) was added to the solution. It was vigorously stirred, the phases were separated, and the aqueous phase was extracted with petrol ether (3 × 160 mL). The combined organic phases were washed with 1 M AcOH (100 mL) and brine (100 mL), dried over MgSO₄ and filtered through Celite[®]. The solvents were evaporated in vacuum to dryness and 2.51 g (94%) of the product **9** was obtained as a clear colorless liquid that can be subjected to further reactions without purification.

HPLC: t_R 3.752 min (method M1); ¹H NMR (500 MHz, CDCl₃, TMS) δ 3.79 (d, J = 1.3 Hz, 2H, CH₂), 7.34 (d, J = 8.1 Hz, 2H, C^{ar}H), 7.63 (d, J = 8.1 Hz, 2H, C^{ar}H), 9.79 (t, J = 2.0 Hz, 1H, CHO); ¹³C NMR (126 MHz, CDCl₃, TMS) δ 50.2 (CH₂), 124.0 (q, J = 272.5 Hz, CF₃), 125.8 (q, J = 3.8 Hz, <u>C</u>^{ar}HCCF₃), 129.8 (q, J = 32.7 Hz, <u>C</u>^{ar}CF₃), 130.0 (C^{ar}H), 135.9 (C^{ar}), 198.1 (CHO); ¹⁹F NMR (470 MHz, CDCl₃) δ -62.7 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 2832 w, 1723 s, 1618 m, 1418 m, 1320 s, 1160 s, 1108 s, 1065 s, 1018 s, 829 m, 732 w; HRMS–EI (C₉H₇F₃O): calcd 188.04490; found, 188.04420.

(E)-4-(p-Trifluoromethylphenyl)but-3-enoic acid (10)

$$F_{3}C$$
 $M = 230.18 \text{ g mol}^{-1} \text{ C}_{11}\text{H}_{9}\text{F}_{3}\text{O}_{2}$

Adapting a procedure by Eißler et al. [2], malonic acid (1.05 g, 10.08 mmol, 2.2 equiv) was dissolved in DMSO (3.8 mL). Acetic acid (5 μ L, 0.09 mmol, 0.2 equiv) in DMSO (110 μ L), and piperidine (9 μ L, 0.09 mmol, 0.2 equiv) in DMSO (110 μ L) was added to the solution, which was heated to 65 °C and (4-trifluoromethylphenyl)-acetaldehyde (9, 0.86 g, 4.56 mmol, 1.0 equiv), dissolved in DMSO (2 mL), was added dropwise during 30 min. The reaction mixture was stirred for 1.5 h at 65 °C and then allowed to cool to rt. H₂O (20 mL) was added and the aqueous phase extracted with Et₂O (4 × 5 mL). The combined organic phases were washed with 5% KHSO₄ solution (5 mL, w/v) and brine (5 mL), and dried over MgSO₄, and the solvent was evaporated in vacuum. The residue was purified by column chromatography over silica gel (petrol ether/EtOAc = 8/1 + 1% AcOH). 481 mg (2.09 mmol, 46%) of the free acid was isolated as a yellowish solid after repeated co-evaporation with toluene. The spectroscopic data were in agreement with the literature [3].

 $R_{\rm f}$ 0.34 (petrol ether/EtOAc = 5/1 + 1% AcOH); ¹H NMR (500 MHz, CDCl₃, TMS) δ 3.34 (dd, J = 7.1, 1.4 Hz, 2H, CH₂), 6.40 (dt, J = 15.9, 7.1 Hz, 1H, CH=C<u>H</u>CH₂), 6.56 (dm, J = 15.9 Hz, 1H, C<u>H</u>=CHCH₂), 7.47 (d, J = 8.2 Hz, 2H C^{ar}H), 7.57 (d, J = 8.2 Hz, 2H, C^{ar}H), 11.22 (br s, 1H, CO₂H).

Methyl (E)-4-[4-(trifluoromethyl)phenyl]but-3-enoate (11)

 $M = 244.21 \text{ g mol}^{-1} \text{ C}_{12}\text{H}_{11}\text{F}_3\text{O}_2$

Adapting a procedure by Eißler et al. [2], (*E*)-4-(*p*-trifluoromethylphenyl)but-3-enoic acid (**10**, 0.48 g, 2.08 mmol, 1.0 equiv) was dissolved in MeOH (22 mL) and the solution was cooled to 0 °C. Thionyl chloride (227 μ L, 3.12 mmol, 1.5 equiv) was

added dropwise. The reaction mixture was allowed to warm up to rt and stirred for 1 h at rt. It was then cooled to 0 °C and quenched with H_2O (16 mL). Stirring was continued for 10 min and then the aqueous phase was extracted with Et_2O . The combined organic phases were washed twice with sat. NaHCO₃ solution and H_2O , and dried over MgSO₄. The solvent was evaporated in vacuum and 428 mg (84%) of product **11** was obtained as a yellowish solid and subjected to the next step without further purification. The spectroscopic data were in agreement with literature values [4].

 $R_{\rm f}$ 0.33 (petrol ether/EtOAc = 1/1); ¹H NMR (500 MHz, CDCl₃, TMS) δ 3.29 (dd, J = 7.1, 0.6 Hz, 2H, CH₂), 3.73 (s, 3H, CH₃), 6.41 (dt, J = 15.9, 7.1 Hz, 1H, CH=C<u>H</u>CH₂), 6.53 (d, J = 15.9 Hz, 1H, C<u>H</u>=CHCH₂), 7.46 (d, J = 8.2 Hz, 2H, C^{ar}H), 7.56 (d, J = 8.2 Hz, 2H, C^{ar}H).

(4*R*,5*R*)-4-Hydroxy-5-[4-(trifluoromethyl)phenyl]dihydrofuran-2(3*H*)-one (12)



$$M = 246.18 \text{ g mol}^{-1} \text{ C}_{11}\text{H}_{9}\text{F}_{3}\text{O}$$

 $K_2OSO_4 \cdot 2H_2O$ (17 mg, 0.047 mmol, 1 mol %), $K_3[Fe(CN)_6]$ (3.87 g, 11.77 mmol, 2.5 equiv) and K_2CO_3 (1.95 g, 14.13 mmol, 3.0 equiv) were dissolved in H_2O (20.5 mL). *tert*-Butanol (20.0 mL), (DHQD)₂-PHAL (36.5 mg, 0.047 mmol, 1 mol %) and methanesulfonamide (448 mg, 4.71 mmol, 1.0 equiv) were added. The mixture was cooled to 0 °C and methyl ester **11** (1.15 g, 4.71 mmol, 1.0 equiv), dissolved in *tert*-butanol (0.8 mL), was added. The reaction progress was monitored by TLC. After vigorous stirring for 42 h at 0 °C conversion was usually complete. Na₂SO₃ (5.84 g, 47.10 mmol, 10.0 equiv) and H₂O (10 mL) were added and the mixture was allowed to warm up to rt and stirred for a further 2 h at rt. After separation of the phases the aqueous layer was extracted with Et₂O (4 × 50 mL). The combined organic layers were washed with H₂O (2 × 50 mL), dried over MgSO₄ and filtered through a plug of silica. The solvent was evaporated to dryness in vacuum, the residue was recrystallized (petrol ether/EtOAc = 10/1) and dried over several hours in vacuum to yield **12** (307 mg, 1.25 mmol, 27%, $[\alpha]_D^{21} = -29.7$) as light beige crystals. The residue of the mother liquor was purified by flash chromatography (petrol ether/EtOAc = 3/2) to give **12** (214 mg, 0.87 mmol, 18%, $[\alpha]_D^{21} = -27.7$) as a white solid. The product **12** was obtained in enantiomerically pure form as shown by chiral HPLC with a total yield of 521 mg (2.12 mmol, 45%).

*R*_f 0.21 (petrol ether/EtOAc = 1/1); $[\alpha]_D^{21}$ -29.7 (*c* 1.00 in MeOH); HPLC: *t*_R 12.1 min (method M4); ¹H NMR (500 MHz, CDCl₃, TMS) δ 1.72 (d, *J* = 1.5 Hz, 1H, OH), 2.73 (dm, *J* = 17.5 Hz, 1H, C^α<u>H</u>^AH^B), 2.93 (dd, *J* = 17.5, 5.0 Hz, 1H, C^αH^A<u>H</u>^B), 4.70 (m, 1H, C^βH), 5.54 (d, *J* = 3.5 Hz, 1H, C^γH), 7.50 (d, *J* = 8.1 Hz, 2H, C^{ar}H), 7.70 (d, *J* = 8.1 Hz, 2H, C^{ar}H); ¹³C NMR (126 MHz, CDCl₃, TMS) δ 38.6 (C^αH₂), 70.1 (C^βH), 84.4 (C^γH), 123.8 (q, *J* = 272.3 Hz, CF₃), 125.8 (q, *J* = 3.5 Hz, <u>C</u>^{ar}HCCF₃), 126.8 (C^{ar}H), 131.1 (q, *J* = 32.6 Hz, <u>C</u>^{ar}CF₃), 137.4 (C^{ar}), 175.0 (C=O); ¹⁹F NMR (470 MHz, CDCl₃): δ -62.7 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 3410 w, 2359 s, 2342 m, 2152 w, 2032 w, 1987 w, 1752 s, 1621 m, 1416 m, 1326 s, 1230 w, 1211 m, 1159 s, 1106 s, 1065 s, 1033 s, 1016 m, 987 m, 956 w, 901 w, 853 m, 792 s, 723 w, 703 m; HRMS–EI (*m*/*z*)⁻: [M + CI]⁻ calcd for [C₁₁H₉F₃O + CI]⁻ 281.01978; found, 281.01894.

(4S,5S)-4-Hydroxy-5-[4-(trifluoromethyl)phenyl]dihydrofuran-2(3H)-one (ent-12)



 $K_2OsO_4 \cdot 2H_2O$ (7.0 mg, 0.02 mmol, 1 mol %), $K_3[Fe(CN)_6]$ (1.55 g, 4.7 mmol, 2.5 equiv) and K_2CO_3 (0.78 g, 5.64 mmol, 3.0 equiv) were dissolved in H_2O (8.2 mL).

tert-Butanol (8.0 mL), (DHQ)₂-PHAL (15.0 mg, 0.02 mmol, 1 mol %) and methanesulfonamide (179 mg, 1.88 mmol, 1.0 equiv) were added. The mixture was cooled to 0 °C and methyl ester **11** (0.46 g, 1.88 mmol, 1.0 equiv), dissolved in *tert*-butanol (0.4 mL), was added. The reaction progress was monitored by TLC. After vigorous stirring for 42 h at 0 °C conversion was usually complete. Na₂SO₃ (2.37 g, 18.80 mmol, 10.0 equiv) and H₂O (4.0 mL) were added and the mixture was allowed to warm up to rt and stirred for further 2 h at rt.

After separation of the phases the aqueous layer was extracted with Et₂O (4 × 20 mL). The combined organic layers were washed with H₂O (2 × 25 mL), dried over MgSO₄ and filtered through a plug of silica. The solvent was evaporated to dryness in vacuum and the residue was purified by flash chromatography (petrol ether/EtOAc = 3/2). 226 mg (0.92 mmol, 49%) of the lactone *ent*-12 were obtained enantiomerically pure as a colorless crystalline solid. [α]_D²¹ +31.1 (*c* 1.00 in MeOH); HPLC: *t*_R 16.2 min (method M4). The other physical data of *ent*-12 were in agreement with those of the enantiomer 12.

(3*R*,4*R*,5*R*)-4-Hydroxy-5-[4-(trifluoromethyl)phenyl]-3-methyldihydrofuran-2(3*H*)one (13)



Diisopropylamine (0.45 mL, 3.05 mmol, 2.5 equiv) in abs. THF (7.6 mL) was cooled to -78 °C and *n*-BuLi (1.3 M in hexane, 2.21 mL, 3.05 mmol, 2.5 equiv) was added dropwise. The solution was stirred for 15 min at -78 °C and then for 30 min without any cold bath. It was then cooled to -78 °C and **12** (300 mg, 1.22 mmol, 1.0 equiv), dissolved in abs. THF (5.1 mL), was added with a syringe pump over 90 min. The reaction mixture was stirred overnight at -78 °C, cooled to -90 °C and methyl iodide (0.22 mL, 3.66 mmol, 3.0 equiv) was added with a syringe pump over 150 min at the same temperature. The reaction mixture was stirred for 3 d at -78 °C and then the reaction was quenched by addition of AcOH (0.12 mL) in abs. THF (0.25 mL). It was allowed to warm up to rt and the solvents were evaporated in vacuum (50 °C) to give a viscous, orange-brown suspension. This was transferred into a separating funnel with H₂O and Et₂O (10 mL each) and the phases were separated. The aqueous phase was extracted with Et₂O (4 × 10 mL). The combined organic phases were washed with 5% KHSO₄ solution (10 mL, w/v) and brine, and dried over MgSO₄, and the solvent was evaporated in vacuum. The residue was purified by column chromatography over silica gel (pentane/EtOAc = 1/1) to yield 154 mg (0.592 mmol, 48%) **13** as a light-brown solid in diastereomerically pure form according to ¹H NMR.

*R*_f 0.53 (pentane/EtOAc = 1/1); [α]_D²¹ -10.5 (*c* 1.00 in MeOH); ¹H NMR (500 MHz, CDCl₃) δ 1.41 (d, *J* = 7.7 Hz, 3H, CH₃), 1.58 (s, 1H, OH), 2.76, (dq, *J* = 7.7, 2.9 Hz, 1H, C^αH), 4.36 (m, 1H, C^βH), 5.64 (d, *J* = 4.4 Hz, 1H, C^γH), 7.50 (d, *J* = 8.1 Hz, 2H, C^{ar}H), 7.70 (d, *J* = 8.1 Hz, 2H, C^{ar}H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 13.0 (CH₃), 43.7 (C^αH), 75.9 (C^βH), 81.8 (C^γH), 123.8 (q, *J* = 272.3 Hz, CF₃), 125.8 (q, *J* = 3.5 Hz, <u>C</u>^{ar}HCCF₃), 126.7 (C^{ar}H), 131.1 (q, *J* = 32.7 Hz, <u>C</u>^{ar}CF₃), 137.5 (C^{ar}), 177.8 (C=O); ¹⁹F NMR (470 MHz, CDCl₃) δ -62.7 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 3735 w, 2360 vs, 2341 s, 1172 w, 1321 w, 1165 w, 1108 w, 1065 w, 999 w, 792 w, 669 m; HRMS–EI (C₁₂H₁₁F₃O₃): calcd 260.06603; found, 260.06390.

Methyl (*S*)-2-[(4*R*,5*R*)-5-(4-(trifluoromethyl)phenyl)-2,2-dimethyl-1,3-dioxolan-4yl]propanoate (14)



Abs. MeOH (0.26 mL) and abs. acetone dimethylketal (0.98 mL) were added to lactone **13** (206 mg, 0.793 mmol, 1.0 equiv) and Amberlyst-15[®] (16 mg). The reaction mixture was stirred for 8 d at rt, was then diluted with pentane and filtered through a pad of Celite. The residue was washed with pentane (200 mL) and the combined filtrates were washed with brine. After drying over MgSO₄ the solvent was evaporated in vacuum and the residue was purified by flash chromatography (pentane/EtOAc = 20/1) to yield 152 mg (2.12 mmol, 58%) **14** as a yellowish oil.

*R*_f 0.48 (pentane/EtOAc = 15/1); $[\alpha]_D^{2^1}$ -11.5 (*c* 1.00 in MeOH); ¹H NMR (500 MHz, CDCl₃) δ 1.28 (d, *J* = 7.0 Hz, 3H, C^{*a*}HC<u>H</u>₃), 1.48 (s, 3H, C(CH₃)₂), 1.56 (s, 3H, C(CH₃)₂), 2.70 (m, 1H, C^{*a*}H), 3.44 (s, 3H, CO₂CH₃), 4.09 (dd, *J* = 8.2, 6.1 Hz, 1H, C^{*β*}H), 4.80 (d, *J* = 8.2 Hz, 1H, C^{*γ*}H), 7.49 (d, *J* = 8.0 Hz, 2H, C^{*a*}H), 7.62 (d, *J* = 8.0 Hz, 2H, C^{*a*}H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 12.9 (C^{*α*}H<u>C</u>H₃), 27.1 (C(<u>C</u>H₃)₂), 27.2 (C(<u>C</u>H₃)₂), 41.5 (C^{*α*}H), 51.3 (CO₂<u>C</u>H₃), 80.6 (C^{*β*}H), 83.3 (C^{*γ*}H), 109.5 (<u>C</u>(CH₃)₂), 124.0 (q, *J* = 272.2 Hz, CF₃), 125.4 (q, *J* = 3.4 Hz, <u>C</u>^{*a*}HCCF₃), 127.6 (C^{*a*}H), 130.7 (q, *J* = 32.7 Hz, <u>C</u>^{*a*}CF₃), 141.7 (C^{*a*}), 173.6 (CO₂); ¹⁹F NMR (470 MHz, CDCl₃) δ -62.7 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 2987 w, 2941 w, 1735 s, 1621 w, 1456 m, 1435 w, 1422 w, 1372 m, 1322 s, 1236 m, 1162 m, 1162 m, 1121 m, 1063 s, 1016 s, 982 w, 888 m, 833 s, 812 m, 766 w; HRMS–EI (*m*/*z*)⁺: [M + Na]⁺ calcd for [C₁₆H₁₉F₃O₄ + Na]⁺ 355.11276; found, 355.11285.

(2*S*,3*S*)-2-[(4*R*,5*R*)-5-(4-(Trifluoromethyl)phenyl)-2,2-dimethyl-1,3-dioxolan-4yl]hex-5-en-3-ol (16)



 $M = 344.37 \text{ g mol}^{-1} \text{ C}_{18}\text{H}_{23}\text{F}_3\text{O}_3$

A solution of **14** (145 mg, 0.436 mmol, 1.0 equiv) in abs. CH_2Cl_2 (2.7 mL) was cooled to -78 °C and a solution of DIBAL-H (1.2 M in toluene, 0.84 mL, 1.00 mmol, 2.3 equiv) was added dropwise over 10 min. The reaction mixture was stirred for 4.5 h at the same temperature, and then cooled to -100 °C. The reaction was quenched by slow addition of abs. MeOH (0.55 mL). The solution was allowed to warm up to rt and diluted with 1 M AcOH (10 mL) and pentane (10 mL). After phase separation the aqueous phase was extracted with pentane (4 × 10 mL). The combined organic phases were washed with 1 M AcOH (6 mL) and brine (10 mL), dried over MgSO₄ and evaporated in vacuum. The crude aldehyde (*R*)-2-[(4*R*,5*R*)-5-(4-trifluoromethyl)phenyl)-2,2-dimethyl-1,3-dioxolan-4-yl]propanal (**15**) can be stored under argon atmosphere overnight at 5 °C. It was usually subjected to further conversions without purification.

Aldehyde **15** (0.436 mmol, 1.0 equiv) was dissolved in abs. CH_2Cl_2 (0.33 mL) and added dropwise at -78 °C to a solution of MgBr₂·Et₂O (171 mg, 0.664 mmol, 1.5 equiv) in abs. CH_2Cl_2 (0.37 mL). The mixture was stirred for 15 min, in which time it turned red. Subsequently, allyl-tri-*n*-butylstannane (205 µL, 0.660 mmol, 1.5 equiv) in abs. CH_2Cl_2 (0.90 mL) was added dropwise and the reaction mixture was stirred overnight at -78 °C. Et₂O (15 mL) was added and the solution was allowed to warm to rt. H₂O (10 mL) was added and the phases were separated. The aqueous phase was extracted with Et₂O (3 × 15 mL) and the combined organic phases were washed with brine (10 mL), and dried over MgSO₄, and the solvent was evaporated in vacuum. Flash chromatography (pentane/EtOAc = 12/1) and drying in high vacuum yielded 106 mg **16** (0.308 mmol, 71% over 2 steps) as a clear colorless oil.

*R*_f 0.26 (pentane/EtOAc = 10/1); $[α]_D^{21}$ -1.5 (*c* 1.00 in MeOH); ¹H NMR (500 MHz, CDCl₃) δ 1.09 (d, *J* = 7.0 Hz, 3H, CHC<u>H₃</u>), 1.50 (s, 3H, C(CH₃)₂), 1.56 (s, 3H, C(CH₃)₂), 1.75 (ddq, *J* = 7.0, 5.9, 2.0 Hz, 1H, C<u>H</u>CH₃), 2.13 (m, 1H, C<u>H</u>^AH^BCH=CH₂), 2.20 (br m, 1H, OH), 2.30 (m, 1H, CH^A<u>H</u>^BCH=CH₂), 3.59 (m, 1H, C<u>H</u>OH), 4.11 (dd, *J* = 8.8, 2.1 Hz, 1H, C⁴'H), 4.83 (d, *J* = 8.8 Hz, 1H, C^{5'}H), 4.96–5.07 (m, 2H, CH=C<u>H</u>₂), 5.72 (dddd, *J* = 16.9, 10.1, 7.7, 6.6 Hz, 1H, C<u>H</u>=CH₂), 7.48 (d, *J* = 8.0 Hz, 2H, C^{ar}H), 7.62 (d, *J* = 8.0 Hz, 2H, C^{ar}H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 10.5 (CH<u>C</u>H₃), 27.0 (C(<u>C</u>H₃)₂), 27.1 (C(<u>C</u>H₃)₂), 36.4 (<u>C</u>HCH₃), 39.5 (<u>C</u>H₂CH=CH₂), 73.2 (CHOH), 79.2 (C^{5'}H), 82.6 (C^{4'}H), 109.3 (<u>C</u>(CH₃)₂), 118.1 (CH=<u>C</u>H₂), 124.0 (q, *J* = 272.2 Hz, CF₃), 125.5 (q, *J* = 3.7 Hz, <u>C^{ar}</u>HCCF₃), 126.9 (C^{ar}H), 130.4 (q, *J* = 32.4 Hz, <u>C^{ar}</u>CF₃), 134.6 (<u>C</u>H=CH₂), 142.1 (C^{ar}); ¹⁹F NMR (470 MHz, CDCl₃) δ -62.6 (s, 3F, CF₃); IR (ATR) \bar{v} (cm⁻¹) 3453 br w, 2984 w, 2934 w, 1640 w, 1621 w, 1457 w, 1419 w, 1381 w, 1371 w, 1322 s, 1233 m, 1163 m, 1122 s, 1065 s, 1015 s, 994 m, 915 w, 887 m, 832 s, 767 w; HRMS–EI (*m*/*z*)⁻: [M + CI]⁻ calcd for [C₁₈H₂₃F₃O₃ + CI]⁻ 379.12933; found, 379.12961.

Fragment condensation and diol-epoxide-transformation

H-uA[Acetonide]- $C^{4'}$ -trifluoromethyl-uB-OTce (18)



A mixture of the unit B building block **17** (154 mg, 0.366 mmol, 1.2 equiv) and Grubbs II catalyst (13 mg, 0.015 mmol, 5 mol %) was dried for 1 h under high vacuum. Under argon atmosphere it was dissolved in abs. CH_2Cl_2 (2.0 mL) and a

solution of unit A building block **16** (105 mg, 0.305 mmol, 1.0 equiv) in abs. CH_2CI_2 (1.35 mL) was added at rt. The mixture was heated under reflux overnight in the dark and then allowed to cool to rt. The solvent was evaporated in vacuum (bath temperature ≤ 30 °C) and the residue was purified by flash chromatography (pentane/EtOAc = $2/1 \rightarrow 1/1 \rightarrow 6/4$). Drying in high vacuum yielded 170 mg of a mixture from 145 mg (85%) of the cross-metathesis product **18** and 25 mg (15%) of the homo-coupling product **23**. The latter could be separated after the next reaction step.

 $R_{\rm f}$ 0.24 (pentane/EtOAc = 6/4); ¹H NMR (500 MHz, CDCl₃) δ 1.04 (d, J = 6.9 Hz, 3H, uA-C[{]HCH₃), 1.46 (s, 3H, uA-C(CH₃)₂), 1.51 (s, 3H, uA-C(CH₃)₂), 1.72 (m, 1H, uA- $C^{\epsilon}H$), 2.23 (m, 1H, uA- $C^{\gamma}H^{A}H^{B}$), 2.34 (m, 1H, uA- $C^{\gamma}H^{A}H^{B}$), 2.36 (br m, 1H, uA-OH), 3.05 (dd, J = 14.2, 6.1 Hz, 1H, uB-C^{β}H^AH^B), 3.13 (dd, J = 14.4, 5.8 Hz, 1H, uB- $C^{\beta}H^{A}H^{B}$), 3.66 (br m, uA- $C^{\delta}H$), 3.83 (s, 3H, uB-OCH₃), 4.02 (dd, J = 8.9, 2.1 Hz, 1H, uA-C^ζH), 4.67 (d, J = 11.9 Hz, 1H, uB-C<u>H</u>^AH^BCCl₃), 4.74 (d, J = 11.9 Hz, 1H, uB- $CH^{A}H^{B}CCI_{3}$, 4.80 (d, J = 8.8 Hz, 1H, uA-C^{η}H), 4.99 (ddd, J = 7.7, 6.0, 5.9 Hz, 1H, uB-C^{α}H), 5.75 (dm, J = 15.3 Hz, 1H, uA-C^{α}H), 5.84 (d, J = 7.8 Hz, 1H, uB-NH), 6.79 $(ddd, J = 15.1, 8.0, 7.9 Hz, 1H, uA-C^{\beta}H)$, 6.80 (d, J = 8.4 Hz, 1H, uB- C^{5'}H), 6.98 (dd, J = 8.4, 2.4 Hz, 1H, uB-C⁶H), 7.12 (d, J = 2.2 Hz, 1H, uB-C²H), 7.43 (d, J = 7.9 Hz, 2H, C^{ar} H), 7.57 (d, J = 7.9 Hz, 2H, C^{ar} H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 10.6 (uA-C^εHCH₃), 27.0 (uA-C(CH₃)₂), 27.1 (uA-C(CH₃)₂), 36.5 (uA-C^γH₂), 36.9 (uA-C^εH), 37.7 $(uB-C^{\beta}H_{2}), 53.0 (uB-C^{\alpha}H), 56.1 (uB-OCH_{3}), 73.3 (uA-C^{\delta}H), 74.7 (uB-CH_{2}CCI_{3}), 79.1$ $(uA-C^{\eta}H)$, 82.7 $(uA-C^{\zeta}H)$, 94.2 $(uB-CCl_{3})$, 109.3 $(uA-C(CH_{3})_{2})$, 112.2 $(uB-C^{5}H)$, 122.5 $(uB-C^{3})$, 124.0 (q, J = 272.0 Hz, CF_{3}), 125.2 (uA- C^{α} H), 125.6 (q, J = 3.7 Hz, $C^{ar}HCCF_{3}$), 126.9 (uA- $C^{ar}H$), 128.4 (uB- $C^{6'}H$), 128.5 (uB- $C^{1'}$), 130.5 (q, J = 32.4 Hz, C^{ar}CF₃), 130.9 (uB-C²[·]H), 141.9 (uA-C^{ar}), 142.3 (uA-C^βH), 154.2 (uB-C⁴), 165.0 (uA-C=O), 170.1 (uB-C=O); ¹⁹F NMR (470 MHz, CDCl₃) δ -62.5 (s, 3F, CF₃); HRMS-

MALDI $(m/z)^+$: $[M + Na]^+$ calcd for $[C_{31}H_{34}Cl_4F_3NO_7 + Na]^+$ 752.09337; found, 752.09273.

seco-uA[Acetonide]-C⁴´-trifluoromethyl-cryptophycin-52 (20)



 $M = 1166.92 \text{ g mol}^{-1} \text{ C}_{57}\text{H}_{63}\text{C}\text{I}_4\text{F}_3\text{N}_2\text{O}_{12}$

The unit AB building block **18** (145 mg, 0.198 mmol, 1.0 equiv; in a mixture with 25 mg unit B homo-coupling product **23**) and unit CD building block **19** (135 mg, 0.296 mmol, 1.5 equiv) were esterified according to GP1. Purification by flash chromatography (pentane/EtOAc = 2/1) and drying in high vacuum yielded 151 mg (0.129 mmol, 65%) **20** as a colorless foam.

*R*_f 0.32 (pentane/EtOAc = 2/1); $[α]_{D}^{21}$ -13.2 (*c* 1.00 in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.89 (d, *J* = 6.2 Hz, 3H, uD-C^δH₃), 0.93 (d, *J* = 6.4 Hz, 3H, uD-C^δH₃), 1.13 (d, *J* = 7.5 Hz, 3H, uA-C^εHC<u>H₃</u>), 1.14 (s, 3H, uC-C(CH₃)₂), 1.22 (s, 3H, uC-C(CH₃)₂), 1.47 (s, 3H, uA-C(CH₃)₂), 1.49 (s, 3H, uA-C(CH₃)₂), 1.53 (m, 1H, uD-C^β<u>H</u>^AH^B), 1.70– 1.81 (m, 2H, uD-C^βH^A<u>H</u>^B and uD-C^γH), 1.95 (m, 1H, uA-C^εH), 2.46 (m, 1H, uA-C^γ<u>H</u>^AH^B), 2.57 (m, 1H, uA-C^γH^A<u>H</u>^B), 3.04 (dd, *J* = 14.2, 6.7 Hz, 1H, uB-C^β<u>H</u>^AH^B), 3.16 (dd, *J* = 14.2, 5.7 Hz, 1H, uB-C^βH^A<u>H</u>^B), 3.38 (d, *J* = 6.6 Hz, 2H, uC-CH₂NH), 3.79– 3.85 (m, 4H, uB-OCH₃ and uA-C^ζH), 4.21 (m, 1H, C<u>H</u>CH₂, Fmoc), 4.30–4.37 (m, 2H, CHC<u>H₂</u>, Fmoc), 4.64 (d, *J* = 11.9 Hz, 1H, uB-C<u>H</u>^AH^BCCl₃), 4.76 (d, *J* = 11.9 Hz, 1H, uB-C^αH), 5.02 (m, 1H, uB-C^αH), 5.08 (m, 1H, uA-C⁷H), 5.78 (dm, *J* = 15.7 Hz, 1H, uA-C^αH), 6.03 (m, 1H, NH), 6.41 (d, *J* = 8.0 Hz, 1H, NH), 6.71 (ddd, *J* = 15.5, 6.5, 6.5 Hz,

1H, uA-C^{β}H), 6.79 (d, J = 8.4 Hz, 1H, uB-C⁵H), 7.02 (dd, J = 8.4, 2.2 Hz, 1H, uB- C^{6} H), 7.14 (d, J = 2.2 Hz, 1H, uB- C^{2} H), 7.25–7.30 (m, 2H, C^{ar} H, Fmoc), 7.35–7.41 (m, 2H, $C^{ar}H$, Fmoc), 7.43 (d, J = 8.0 Hz, 2H, uA- $C^{ar}H$), 7.58 (d, J = 8.0 Hz, 2H, uA- $C^{ar}H$), 7.60–7.65 (m, 2H, $C^{ar}H$, Fmoc), 7.75 (d, J = 7.5 Hz, 2H, $C^{ar}H$, Fmoc); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 9.2 (uA-C^εH<u>C</u>H₃), 21.4 (uD-C^{δ}H₃), 22.2 $(uC-C(CH_3)_2)$, 22.7 $(uC-C(CH_3)_2)$, 23.1 $(uD-C^{\delta}H_3)$, 24.9 $(uD-C^{\gamma}H)$, 26.99 $(uA-C^{\delta}H_3)$ $C(\underline{C}H_3)_2)$, 27.01 (uA- $C(\underline{C}H_3)_2)$, 32.6 (uA- $C^{\gamma}H_2$), 35.5 (uA- $C^{\varepsilon}H$), 36.5 (uB- $C^{\beta}H_2$), 39.3 (uD-C^βH₂), 44.1 (uC-C(CH₃)₂), 47.2 (CHCH₂, Fmoc), 49.4 (uC-CH₂NH), 53.2 (uB-C^aH), 56.0 (uB-OCH₃), 66.8 (CHCH₂, Fmoc), 71.1 (uD-C^aH), 74.6 (uB-CH₂CCl₃), 75.7 $(uA-C^{\delta}H)$, 79.3 $(uA-C^{\eta}H)$, 81.9 $(uA-C^{\zeta}H)$, 94.2 $(uB-CCI_3)$, 109.5 $(uA-C(CH_3)_2)$, 112.0 $(uB-C^{5}H)$, 119.9 ($C^{ar}H$, Fmoc), 122.2 ($uB-C^{3}$), 123.9 (q, J = 272.1 Hz, CF_{3}), 125.3 $(C^{ar}H, Fmoc)$, 125.5 (uA-C^aH), 125.7 (q, J = 3.7 Hz, $C^{ar}HCCF_3$), 126.7 (uA-C^{ar}H), 127.6 (C^{ar}H, Fmoc), 128.4 (uB-C⁶H), 128.7 (uB-C¹), 130.6 (q, J = 32.5 Hz, <u>C</u>^{ar}CF₃), 131.1 (uB-C²H), 138.9 (uA-C^{β}H), 141.22, 141.24 (C^{*ar*}), 141.9 (uA-C^{*ar*}), 144.0 (C^{*ar*}), 154.0 (uB-C⁴), 156.9 (uC-NHCO₂), 165.1 (uA-C=O), 169.9 171.0, 176.7 (C=O) ; ¹⁹F NMR (470 MHz, CDCl₃) δ -62.6 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 3320 br w, 2960 br w, 2014 w, 1978 w, 1723 vs, 1676 m, 1644 m, 1521 m, 1502 s, 1449 m, 1371 w, 1323 s, 1255 br m, 1123 br s, 1065 s, 1015 br m, 889 w, 832 m, 809 m, 758 m, 740 m, 718 m; HRMS–MALDI $(m/z)^+$: [M + Na]⁺ calcd for [C₅₇H₆₃Cl₄F₃N₂O₁₂ + Na]⁺ 1187.29794; found, 1187.29628.

uA[Acetonide]-C⁴´-trifluoromethyl-cryptophycin-52 (21)



 $M = 795.28 \text{ g mol}^{-1} \text{ C}_{40}\text{H}_{50}\text{CIF}_3\text{N}_2\text{O}_9$

Macrolactamization of **20** (150 mg, 0.129 mmol) according to GP2, purification by flash chromatography (pentane/EtOAc = 1/2), and drying in high vacuum yielded 84 mg (0.106 mmol, 82%) **21** as a colorless amorphous solid.

 $R_{\rm f}$ 0.28 (pentane/EtOAc = 1/2); $[\alpha]_{\rm D}^{21}$ +10.6 (c 0.93 in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.78 (d, J = 6.6 Hz, 3H, uD-C⁵H₃), 0.88 (d, J = 6.6 Hz, 3H, uD-C⁵H₃), 1.14 $(d, J = 6.3 Hz, 3H, uA-C^{\ell}HCH_3)$, 1.15 (s, 3H, uC-C(CH₃)₂), 1.21 (s, 3H, uC-C(CH₃)₂), 1.25 (ddd, J = 13.6, 9.4, 3.6 Hz, 1H, uD-C^{β}H^AH^B), 1.47 (s, 3H, uA-C(CH₃)₂), 1.48 (s, 3H, uA-C(CH₃)₂), 1.62 (m, 1H, uD-C^{γ}H), 1.73 (ddd, J = 13.8, 10.5, 5.0 Hz, 1H, uD- $C^{\beta}H^{A}H^{B}$, 1.83 (m, 1H, uA- $C^{\epsilon}H$), 2.24 (ddd, J = 14.4, 11.0, 11.0 Hz, 1H, uA- $C^{\gamma}H^{A}H^{B}$), 2.57 (dm, J = 14.6 Hz, 1H, uA-C^{γ}H^A<u>H</u>^B), 3.03 (dd, J = 14.6, 7.6 Hz, 1H, uB-C^{β}<u>H</u>^AH^B), 3.10 (dd, J = 14.6, 5.0 Hz, 1H, uB-C^{β}H^AH^B), 3.13 (dd, J = 13.5, 3.6 Hz, 1H, uC- $CH^{A}H^{B}NH$), 3.38 (dd, J = 13.5, 8.4 Hz, 1H, uC- $CH^{A}H^{B}NH$), 3.77 (dd, J = 8.7, 1.6 Hz, 1H, uA-C^ζH), 3.87 (s, 3H, uB-OCH₃), 4.73 (ddd, J = 7.7, 5.1, 5.1 Hz, 1H, uB-C^αH), 4.75 (d, J = 8.3 Hz, 1H, uA-C^{η}H), 4.76 (dd, J = 10.4, 3.6 Hz, 1H, uD-C^{α}H), 5.04 (ddd, J = 11.4, 7.4, 2.1 Hz, 1H, uA-C⁵H), 5.57 (d, J = 7.8 Hz, 1H, NH), 5.67 (dm, J = 15.1Hz, 1H, uA-C^{α}H), 6.69 (ddd, J = 15.1, 10.6, 4.3 Hz, 1H, uA-C^{β}H), 6.83 (d, J = 8.4 Hz, 1H, uB-C⁵H), 7.05 (dd, J = 8.4, 2.2 Hz, 1H, uB-C⁶H), 7.18 (m, 1H, NH), 7.19 (d, J =2.1 Hz, 1H, uB-C²H), 7.45 (d, J = 8.0 Hz, 2H, uA-C^{ar}H), 7.63 (d, J = 8.0 Hz, 2H, uA-C^{ar}H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 9.5 (uA-C^εHCH₃), 21.2 (uD-C^δH₃), 22.7 (uC- $C(CH_3)_2)$, 22.8 (uC-C(CH_3)_2), 22.9 (uD-C^oH_3), 24.6 (uD-C^vH), 26.96 (uA-C(CH_3)_2), 27.03 (uA-C(CH₃)₂), 35.3 (uB-C^{β}H₂), 36.1 (uA-C^{γ}H₂), 36.5 (uA-C^{ϵ}H), 39.5 (uD-C^{β}H₂), 42.7 (uC-C(CH₃)₂), 46.4 (uC-CH₂NH), 54.3 (uB-C^αH), 56.1 (uB-OCH₃), 71.0 (uD- C^αH), 75.8 (uA-C^δH), 79.1 (uA-C^ηH), 82.1 (uA-C^ζH), 109.7 (uA-<u>C</u>(CH₃)₂), 112.3 (uB-C⁵'H), 122.5 (uB-C^{3'}), 123.9 (q, J = 272.5 Hz, CF₃), 124.4 (uA-C^αH), 125.7 (q, J = 3.5 Hz, <u>C</u>^{ar}HCCF₃), 126.5 (uA-C^{ar}H), 128.2 (uB-C^{6'}H), 129.5 (uB-C^{1'}), 130.6 (q, J = 32.5 Hz, <u>C</u>^{ar}CF₃), 130.9 (uB-C^{2'}H), 142.2 (uA-C^βH and uA-C^{ar}), 154.0 (uB-C^{4'}), 165.0 (uA-C(O)NH), 170.1 (C(O)NH), 170.2, 177.9 (C=O); ¹⁹F NMR (470 MHz, CDCl₃) δ -62.6 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 3411 br w, 2961 br w, 1747 m, 1719 m, 1655 br m, 1503 s, 1472 w, 1370 w, 1324 s, 1257 w, 1125 br s, 1066 s, 1015 m, 969 w, 869 w, 832 w; HRMS-EI (*m*/*z*)⁺: [M + Na]⁺ calcd for [C₄₀H₅₀ClF₃N₂O₉ + Na]⁺ 817.30491; found, 817.30490.

uA-C^{4'}-Trifluoromethyl-cryptophycin-52 (22)



 $M = 737.20 \text{ g mol}^{-1} \text{ C}_{37}\text{H}_{44}\text{CIF}_3\text{N}_2\text{O}_8$

The acetonide **21** (31 mg, 0.039 mmol) was converted into the epoxide according to GP3–6 and the crude product was purified by flash chromatography (pentane/EtOAc = 1/3). The purified product was dissolved in CH₃CN and H₂O and the mixture was lyophilized to give 11.3 mg (0.015 mmol, 39% over 4 steps) of epoxide **22** as voluminous, colorless solid.

 $R_{\rm f}$ 0.35 (pentane/EtOAc = 1/3); $[\alpha]_{\rm D}^{21}$ +16.8 (*c* 0.45 in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.82 (d, *J* = 6.6 Hz, 3H, uD-C^{{5}H}₃), 0.85 (d, *J* = 6.6 Hz, 3H, uD-C^{{5}H}₃), 1.15 (d, *J* = 7.0 Hz, 3H, uA-C^{{6}HCH}₃), 1.16 (s, 3H, uC-C(CH₃)₂), 1.22 (s, 3H, uC-C(CH₃)₂), 1.30 (ddd, *J* = 13.9, 8.8, 3.6 Hz, 1H, uD-C^{{6}H}^{A}H^{B}), 1.65 (m, 1H, uD-C^{7}H), 1.73 (ddd, *J* = 14.6, 10.6, 4.9 Hz, 1H, uD-C^{6}H^{A}H^{B}), 1.82 (m, 1H, uA-C^{{6}H}), 2.43 (ddd, *J* = 14.5, 11.1, 11.0 Hz, 1H, uA-C^{{7}H}^{A}H^{B}), 2.57 (ddd, *J* = 14.5, 2.2, 2.2 Hz, 1H, uA-C^{7}H^{A}H^{B}), 2.90 (dd, *J* = 7.4, 1.9 Hz, 1H, uA-C^{{5}H}), 3.03-3.13 (m, 3H, uB-C^{6}H^{A}H^{B}, uB-C^{6}H^{A}H^{B})

and uC-CH^AH^BNH), 3.42 (dd, J = 13.5, 8.6 Hz, 1H, uC-CH^AH^BNH), 3.74 (d, J =1.9 Hz, 1H, uA-C^{η}H), 3.87 (s, 3H, uB-OCH₃), 4.74 (ddd, J = 7.5, 7.4, 5.3 Hz, 1H, uB- $C^{\alpha}H$), 4.82 (dd, J = 10.3, 3.5 Hz, 1H, uD- $C^{\alpha}H$), 5.20 (ddd, J = 11.3, 5.0, 2.0 Hz, 1H, $uA-C^{\delta}H$), 5.49 (d, J = 7.8 Hz, 1H, NH), 5.72 (dm, J = 15.1 Hz, 1H, $uA-C^{\alpha}H$), 6.76 $(ddd, J = 14.9, 10.6, 4.2 Hz, 1H, uA-C^{\beta}H)$, 6.84 (d, $J = 8.3 Hz, 1H, uB-C^{5'}H)$, 7.04 (dd, J = 8.5, 2.2 Hz, 1H, uB-C⁶H), 7.19 (d, J = 2.3 Hz, 1H, uB-C²H), 7.20 (m, 1H, NH), 7.37 (d, J = 8.0 Hz, 2H, uA-C^{ar}H), 7.63 (d, J = 8.0 Hz, 2H, uA-C^{ar}H); ¹³C NMR (126 MHz, CDCl₃) δ 13.5 (uA-C^{{e}HCH₃}), 21.3 (uD-C^{{o}H₃}), 22.7 (uC- C(CH₃)₂), 22.8 $(uC-C(CH_3)_2)$, 22.9 $(uD-C^{\delta}H_3)$, 24.6 $(uD-C^{\gamma}H)$, 35.3 $(uB-C^{\beta}H_2)$, 36.9 $(uA-C^{\gamma}H_2)$, 39.4 (uD-C^βH₂), 40.5 (uA-C^εH), 42.7 (uC-C(CH₃)₂), 46.4 (uC-CH₂NH), 54.3 (uB-C^αH), 56.1 $(uB-OCH_3)$, 58.2 $(uA-C^{\eta}H)$, 63.5 $(uA-C^{\zeta}H)$, 71.1 $(uD-C^{\alpha}H)$, 75.7 $(uA-C^{\delta}H)$, 112.3 $(uB-C^{\delta}H)$ $C^{5}H$), 122.6 (uB- C^{3}), 123.9 (q, J = 271.8 Hz, CF₃), 124.7 (uA- $C^{\alpha}H$), 125.7 (q, J = 3.6 Hz, <u>C</u>^{ar}HCCF₃), 125.8 (uA-C^{ar}H), 128.3 (uB-C⁶H), 129.3 (uB-C¹), 130.7 (q, J = 32.7 Hz, $C^{ar}CF_3$, 130.9 (uB- $C^{2^{\iota}}H$), 140.9 (uA- C^{ar}), 141.6 (uA- $C^{\beta}H$), 154.1 (uB- $C^{4^{\iota}}$), 164.8 (uA-C(O)NH), 170.2 (C(O)NH), 170.4, 178.0 (C=O); ¹⁹F NMR (470 MHz, CDCl₃, TMS) δ -62.6 (s, 3F, CF₃); IR (ATR) \tilde{v} (cm⁻¹) 3851 w, 3733 w, 3674 w, 3415 w, 3272 w, 2958 br w, 2167 w, 1970 w, 1748 m, 1718 m, 1654 br m, 1504 br s, 1472 m, 1324 s, 1257 m, 1126 br s, 1066 s, 1017 m, 970 w, 894 w, 838 m; HRMS-EI $(m/z)^{+}$: $[M + Na]^{+}$ calcd for $[C_{37}H_{44}CIF_{3}N_{2}O_{8} + Na]^{+}$ 759.26305; found, 759.26191.

3.2 Synthesis of the pentafluorophenyl-substituted unit B building block

(R)-2,2,2-Trichloroethyl 2-(tert-butoxycarbonylamino)-3-(pentafluorophenyl)-

propanoate (25)

$$CI_{CI} \rightarrow 0^{O} \rightarrow 0^$$

By analogy to Nahrwold [5] Boc-pentafluoro-D-phenylalanine (24) (1.00 g, 2.82 mmol, 1.00 equiv) was dissolved in abs. CH₂Cl₂ (14 mL), and abs. pyridine (0.46 mL, 5.63 mmol, 2.00 equiv) and trichloroethanol (0.41 mL, 4.22 mmol, 1.50 equiv) were added sequentially at rt. Then, the reaction mixture was cooled to 0 °C, and DCC (612 mg, 2.96 mmol, 1.05 equiv), dried for 30 min in high vacuum and suspended in abs. CH₂Cl₂ (1.0 mL), was added dropwise at 0 °C to the reaction mixture. The suspension was stirred overnight at the same temperature. The reaction was monitored by TLC. If the conversion was not complete, abs. pyridine (0.46 mL, 5.63 mmol, 2.00 equiv), trichloroethanol (0.41 mL, 4.22 mmol, 1.50 equiv) and solid DCC (260 mg, 1.27 mmol, 0.45 equiv) were added at 0 °C. The mixture was stirred for a further 5.5 h at 0 °C. The solids were removed by filtration and the residue was washed with CH_2CI_2 (4 × 5 mL). The combined organic phases were washed with brine (10 mL), and dried over MgSO₄, and the solvent was evaporated to dryness in vacuum. The resulting white solid was purified by flash chromatography (petrol ether/EtOAc = 15/1) to yield 1.14 g (2.34 mmol, 83%) of product 25 as a colorless solid.

*R*_f 0.32 (hexane/EtOAc = 10/1); HPLC: t_R 5.6 min (method M1); $[\alpha]_D^{21}$ -7.8 (c 1.00 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, TMS) δ 1.40 (s, 9H, C(CH₃)₃), 3.12 (dd, *J* = 14.1, 8.2 Hz, 1H, C^{β}<u>H</u>^AH^B), 3.42 (dd, *J* = 14.1, 5.0 Hz, 1H, C^{β}H^A<u>H</u>^B), 4.73 (ddd, *J* = 8.4, 5.2, 5.1 Hz, 1H, C^{α}H), 4.79 (d, *J* = 12.0 Hz, 1H, C<u>H</u>^AH^BCCl₃), 4.82 (d, *J* = 12.0 Hz, 1H,

CH^A<u>H</u>^BCCl₃), 5.10 (d, *J* = 8.5 Hz, 1H, NH); ¹³C NMR (126 MHz, CDCl₃, TMS) δ 25.9 (C^βH₂), 28.1 (C(<u>C</u>H₃)₃), 52.6 (C^αH), 74.8 (<u>C</u>H₂CCl₃), 80.7 (<u>C</u>(CH₃)₃), 94.1 (CCl₃), 109.6 (tm, *J* = 18.2 Hz, C₆F₅<u>C</u>^{*ipso*}), 137.4 (dm, *J* = 251.3 Hz, C₆F₅<u>C</u>^{*meta*}), 140.5 (dm, *J* = 251.3 Hz, C₆F₅<u>C</u>^{*para*}), 145.5 (dm, *J* = 246.2 Hz, C₆F₅<u>C</u>^{*ortho*}), 154.8 (NCO₂), 169.6 (<u>C</u>O₂CH₂); ¹⁹F NMR (470 MHz, CDCl₃, TMS) δ -142.3 (dd, *J* = 22.4, 8.2 Hz, 2F, F^{*ortho*}), -155.0 (t, *J* = 20.8 Hz, 1F, F^{*para*}), -161.9 (td, *J* = 21.6, 8.2 Hz, 2F, F^{*meta*}); IR (ATR) \tilde{v} (cm⁻¹) 3481 w, 2357 w, 2341 w, 1763 m, 1693 s, 1523 s, 1501 s, 1444 w, 1369 m, 1270 m, 1250 w, 1157 s, 1123 w, 1061 m, 959 s, 889 w, 826 w, 776 m, 720 s; HRMS–EI (*m*/*z*)⁺: [M + Na]⁺ calcd for [C₁₆H₁₅Cl₃F₅NO₄ + Na]⁺ 507.98790; found, 507.98737.

(R)-2,2,2-Trichloroethyl 2-acrylamido-3-(pentafluorophenyl)propanoate (26)



Adapting a literature procedure [5], a solution of **25** (728 mg, 1.50 mmol, 1.0 equiv) in TFA (1.46 mL) was stirred for 2 h at rt. Then the volatiles were evaporated in vacuum to dryness. The residue was co-evaporated twice with toluene, dissolved in CH_2Cl_2 (18 mL) and washed with saturated NaHCO₃ solution (10 mL). The organic phase was dried over MgSO₄ and evaporated in vacuum to dryness. The residue was dried overnight under high vacuum and dissolved in abs. CH_2Cl_2 (10 mL). The solution was cooled to 0 °C, and abs. NEt₃ (0.83 mL, 5.98 mmol, 4.0 equiv) and acryloyl chloride (278 µL, 3.74 mmol, 2.5 equiv) were added. The reaction mixture was stirred for 7 h in the dark at 0 °C. Then, EtOAc (47 mL) and 5% KHSO₄ solution (10 mL, w/v) were added, and the phases were separated. After phase separation, the organic phase was washed with saturated NaHCO₃ solution and brine (10 mL each), dried over

MgSO₄, and evaporated in vacuum to dryness. Purification by flash chromatography $(CH_2CI_2/Et_2O = 25/1)$ yielded 373 mg (0.847 mmol, 57%) of **26** as a colorless solid. $R_{\rm f}$ 0.25 (hexane/EtOAc = 4/1); $[\alpha]_{\rm D}^{21}$ -29.6 (c 1.00 in CHCl₃); ¹H NMR (500 MHz, CDCl₃, TMS) δ 3.19 (dd, J = 14.2, 7.7 Hz, 1H, C^{β}H^AH^B), 3.48 (dd, J = 14.2, 5.3 Hz, 1H, $C^{\beta}H^{A}\underline{H}^{B}$), 4.80 (d, J = 11.9 Hz, 1H, $C\underline{H}^{A}H^{B}CCI_{3}$), 4.83 (d, J = 11.9 Hz, 1H, CH^AH^BCCl₃), 5.15 (ddd, *J* = 7.9, 5.4, 5.3 Hz, 1H, C^αH), 5.73 (dd, *J* = 10.4, 1.2 Hz, 1H, $CH=CH_2^{trans}$), 6.08 (m, 1H, NH), 6.10 (dd, J = 16.9, 10.4 Hz, 1H, $CH=CH_2$), 6.28 (dd, J = 17.0, 1.2 Hz, 1H, CH=CH₂^{cis}); ¹³C NMR (126 MHz, CDCl₃, TMS) δ 25.6 (C^{β}H₂), 51.2 (C^{α}H), 75.0 (<u>C</u>H₂CCl₃), 94.0 (CCl₃), 109.3 (tm, J = 18.2 Hz, C₆F₅C^{*ipso*}). 128.2 $(CH=CH_2)$, 129.5 $(CH=CH_2)$, 137.4 $(dm, J = 252.4 \text{ Hz}, C_6F_5C^{meta})$, 140.6 $(dm, J = 252.4 \text{ Hz}, C_6F_5C^{meta})$ 254.4 Hz, $C_6F_5C^{para}$), 145.5 (dm, J = 245.9 Hz, $C_6F_5C^{ortho}$), 165.2 (NCO₂), 169.3 (CO_2CH_2) ; ¹⁹F NMR (470 MHz, CDCl₃, TMS) δ -142.3 (dd, J = 21.9, 7.7 Hz, 2F, F^{ortho}), -154.3 (t, J = 20.8 Hz, 1F, F^{para}), -161.5 (td, J = 21.9, 8.1 Hz, 2F, F^{meta}); IR (ATR) v (cm⁻¹) 3339 w, 2358 m, 2340 m, 1739 s, 1660 m, 1629 w, 1519 s, 1502 s, 1449 w, 1374 w, 1311 m, 1218 m, 1190 m, 1126 w, 1051 m, 1023 w, 983 m, 956 s, 827 w, 807 s, 785 m, 720 s; HRMS-MALDI $(m/z)^+$: $[M + Na]^+$ calcd for $[C_{14}H_9CI_3F_5NO_3 + Na]^+$ 461.94604; found, 461.94609.

Fragment condensation and diol-epoxide-transformation

H-uA[Acetonide]-uB-pentafluorophenyl-OTce (28)



A mixture of the unit B building block **26** (100 mg, 0.227 mmol, 1.0 equiv) and Grubbs II catalyst (10.5 mg, 0.015 mmol, 5 mol %) was dissolved in abs. CH_2CI_2 (1.50 mL) and stirred for 45 min at rt. Then, the unit A building block **27** (75 mg,

0.272 mmol, 1.2 equiv), dissolved in abs. CH_2Cl_2 (0.75 mL), was added and the mixture was heated under reflux overnight in the dark. After cooling to rt the solvent was evaporated in vacuum and the residue was purified by chromatography (pentane/EtOAc = 3/1) to yield 67 mg (0.097 mmol, 43%) of **28** as a brownish viscous oil.

 $R_{\rm f}$ 0.56 (pentane/EtOAc = 2/1); $[\alpha]_{\rm D}^{21}$ -20.6 (c 0.98 in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 1.06 (d, J = 7.0 Hz, 3H, uA-C^εHCH₃), 1.49 (s, 3H, uA-C(CH₃)₂), 1.56 (s, 3H, $uA-C(CH_3)_2$, 1.77 (m, 1H, $uA-C^{\epsilon}H$), 2.23–2.36 (m, 2H, $uA-C^{\gamma}H_2$), 2.53 (br m, 1H, $uA-C^{\epsilon}H$), 2.23–2.36 (m, 2H, $uA-C^{\gamma}H_2$), 2.53 (br m, 2H, $uA-C^{\epsilon}H_2$), 2.54 (br m, 2H, $uA-C^{\epsilon}H_2$), 2.55 (br m, 2H, OH), 3.15 (dd, J = 14.1, 7.6 Hz, 1H, uB-C^{β}H^AH^B), 3.42 (dd, J = 14.3, 5.6 Hz, 1H, uB- $C^{\beta}H^{A}H^{B}$), 3.69 (br m, uA- $C^{\delta}H$), 4.05 (dd, J = 9.0, 2.3 Hz, 1H, uA- $C^{\zeta}H$), 4.78 (d, J = 11.9 Hz, 1H, uB-CH^AH^BCCl₃), 4.79 (d, J = 8.5 Hz, 1H, uA-C^{η}H), 4.81 (d, J = 11.9 Hz, 1H, uB-CH^A<u>H</u>^BCCI₃), 5.08 (ddd, J = 7.8, 5.5, 5.5 Hz, 1H, uB-C^{α}H), 5.67 (dm, J = 15.4 Hz, 1H, uA-C^{α}H), 5.95 (d, J = 8.1 Hz, 1H, uB-NH), 6.76 (ddd, J = 14.9, 7.4, 7.3 Hz, 1H, uA-C^{β}H), 7.32–7.38 (m, 5H, uA-C^{ar}H); ¹³C NMR (126 MHz, CDCl₃) δ 10.9 (uA-C^εHCH₃), 25.7 (uB-C^βH₂), 27.0 (uA-C(CH₃)₂), 27.2 (uA-C(CH₃)₂), 36.5 $(uA-C^{\varepsilon}H)$, 37.6 $(uA-C^{\gamma}H_2)$, 51.2 $(uB-C^{\alpha}H)$, 73.5 $(uA-C^{\delta}H)$, 74.9 $(uB-CH_2CCI_3)$, 79.9 $(uA-C^{\eta}H)$, 82.7 $(uA-C^{\zeta}H)$, 94.0 $(uB-CCl_{3})$, 109.0 $(uA-C(CH_{3})_{2})$, 109.4 (tm, J = 18.2 Hz)C₆F₅C^{*ipso*}), 124.7 (uA-C^αH), 126.8, 128.5, 128.7 (uA-C^{ar}H), 137.4 (uA-C^{ar}), 137.4 (dm, $J = 252.5 \text{ Hz}, C_6 F_5 C^{meta}$, 140.5 (dm, $J = 254.8 \text{ Hz}, C_6 F_5 C^{para}$), 142.8 (uA-C^βH), 145.5 $(dm, J = 245.9 \text{ Hz}, C_6F_5C^{ortho}), 165.2 (uA-C=O), 169.4 (uB-C=O); {}^{19}F \text{ NMR} (470 \text{ MHz}, 165.2 \text{ MHz}))$ CDCl₃) δ -142.2 (dd, J = 22.5, 8.0 Hz, 2F, F^{ortho}), -154.5 (t, J = 20.9 Hz, 1F, F^{para}), -161.6 (td, J = 21.6, 7.9 Hz, 2F, F^{meta}); IR (ATR) \tilde{v} (cm⁻¹) 2983 w, 2935 w, 2359 vs, 2341 s, 1736 m, 1670 m, 1637 m, 1520 s, 1503 s, 1453 w, 1371 m, 1235 m, 1167 m, 1123 w, 1040 m, 970 m, 882 w, 811 w, 755 m; HRMS-EI $(m/z)^+$: [M + Na]⁺ calcd for $[C_{29}H_{29}C]_{3}F_{5}NO_{6} + Na]^{+}$ 710.08728; found, 710.08810; $[M + K]^{+}$ calcd for $[C_{29}H_{29}CI_{3}F_{5}NO_{6} + K]^{+}$ 726.06122; found, 726.06204; [2 M + Na]^{+} calcd for $[(C_{29}H_{29}CI_{3}F_{5}NO_{6})_{2} + Na]^{+}$ 1397.18534; found, 1397.18624.

seco-uA[Acetonide]-uB-pentafluorophenyl-cryptophycin-52 (29)



$$M = 1124.41g \text{ mol}^{-1} \text{ C}_{55}\text{H}_{58}\text{CI}_3\text{F}_5\text{N}_2\text{O}_{11}$$

The unit AB building block **28** (67 mg, 97 μ mol, 1.0 equiv) and unit CD building block **19** (66 mg, 146 μ mol, 1.5 equiv) were esterified according to GP1. Purification by flash chromatography (pentane/EtOAc = 4/1), preparative RP-HPLC (method M3) and lyophilization yielded 49 mg (43.6 μ mol, 45%) of **29** as a voluminous colorless solid.

*R*_f 0.24 (pentane/EtOAc = 4/1); $[\alpha]_D^{21}$ -9.0 (*c* 0.95 in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.92 (d, *J* = 6.3 Hz, 3H, uD-C^δH₃), 0.97 (d, *J* = 6.2 Hz, 3H, uD-C^δH₃), 1.12 (d, *J* = 6.9 Hz, 3H, uA-C^εHC<u>H₃</u>), 1.16 (s, 3H, uC-C(CH₃)₂), 1.24 (s, 3H, uC-C(CH₃)₂), 1.45 (s, 3H, uA-C(CH₃)₂), 1.51 (s, 3H, uA-C(CH₃)₂), 1.58 (m, 1H, uD-C^γH), 1.67–1.83 (m, 2H, uD-C^βH₂), 1.94 (m, 1H, uA-C^εH), 2.35–2.52 (m, 2H, uA-C^γH₂), 3.14 (dd, *J* = 14.2, 7.6 Hz, 1H, uB-C^β<u>H</u>^AH^B), 3.36–45 (m, 3H, uB-C^βH^A<u>H</u>^B and uC-CH₂NH), 3.82 (dd, *J* = 8.8, 2.5 Hz, 1H, uA-C^ζH), 4.21 (m, 1H, C<u>H</u>CH₂, Fmoc), 4.30–4.39 (m, 2H, CHC<u>H₂</u>, Fmoc), 4.70 (d, *J* = 8.7 Hz, 1H, uA-C^ηH), 4.72 (d, *J* = 11.9 Hz, 1H, uB-C<u>H</u>^AH^BCCl₃), 4.74 (d, *J* = 11.9 Hz, 1H, uB-CH^A<u>H</u>^BCCl₃), 4.89 (dd, *J* = 10.0, 3.3 Hz, 1H, uD-C^αH), 5.02-5.15 (m, 2H, uB-C^αH and uA-C⁵H), 5.68 (dm, *J* = 15.5 Hz, 1H, uA-C^αH, 6.67 (m, 1H, NH), 6.61 (ddd, *J* = 15.4, 6.4, 6.4 Hz, 1H, uA-C^βH), 6.69 (d, *J* = 8.4 Hz, 1H, NH), 7.24–7.43 (m, 9H, uA-C^aH and C^{ar}H, Fmoc), 7.55–7.65 (m, 2H, C^{ar}H, Fmoc), 7.75 (d, *J* = 7.7 Hz, 2H, C^{ar}H, Fmoc); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 9.4 (uA-C^εH<u>C</u>H₃), 21.3 (uD-C⁵H₃), 22.1 (uC-C(<u>C</u>H₃)₂), 22.8 (uC-C(<u>C</u>H₃)₂), 23.1 (uD-

C^δH₃), 24.9 (uD-C^VH), 25.4 (uB-C^βH₂), 27.0 (uA-C(<u>C</u>H₃)₂), 27.2 (uA-C(<u>C</u>H₃)₂), 32.4 (uA-C^VH₂), 35.8 (uA-C^sH), 39.2 (uD-C^βH₂), 44.1 (uC-<u>C</u>(CH₃)₂), 47.2 (<u>C</u>HCH₂, Fmoc), 49.5 (uC-CH₂NH), 51.2 (uB-C^αH), 66.9 (CH<u>C</u>H₂, Fmoc), 71.3 (uD-C^αH), 74.8 (uB-<u>C</u>H₂CCl₃), 75.6 (uA-C⁵H), 80.3 (uA-C^ηH), 82.1 (uA-C⁵H), 94.1 (uB-CCl₃), 109.0 (uA-<u>C</u>(CH₃)₂), 119.9 (C^{ar}H, Fmoc), 124.9 (uA-C^αH), 125.2 (C^{ar}H, Fmoc), 126.7 (C^{ar}H), 126.96, 127.00, 127.6 (C^{ar}H, Fmoc), 128.5, 128.7 (C^{ar}H), 137.3 (uA-C^{ar}), 139.5 (uA-C^βH), 141.23, 141.25, 143.96, 144.02 (C^{ar}), 157.0 (uC-NHCO₂), 165.8 (uA-C=O), 169.8, 171.0, 176.9 (C=O). The signals of the C₆F₅ group were not visible in the ¹³C NMR spectrum because of their multiplicity and low intensity. ¹⁹F NMR (470 MHz, CDCl₃) δ -142.1 (dd, *J* = 22.9, 7.9 Hz, 2F, F^{ortho}), -154.9 (t, *J* = 20.9 Hz, 1F, F^{para}), -161.9 (td, *J* = 22.0, 8.0 Hz, 2F, F^{meta}); IR (ATR) \tilde{v} (cm⁻¹) 3368 br w, 2959 br m, 2249 w, 1721 br s, 1678 m, 1646 m, 1519 s, 1504 s, 1449 m, 1371 m, 1241 m, 1141 m, 1124 m, 1041 m, 1023 w, 969 m, 906 s, 813 w, 757 m, 726 s; HRMS–MALDI (*m*/z)⁺: [M + Na]⁺ calcd for [C₅₅H₅₈Cl₃F₅N₂O₁₁ + Na]⁺ 1145.29186; found, 1145.29277.

uA[Acetonide]-uB-pentafluorophenyl-cryptophycin-52 (30)



Macrolactamization of **29** (48 mg, 42.7 μ mol) according to GP2, purification by flash chromatography (pentane/EtOAc = 2/1) and drying in high vacuum yielded 26.5 mg (35.2 μ mol, 83%) of **30** as a colorless crystalline solid.

 $R_{\rm f}$ 0.21 (pentane/EtOAc = 2/1); $[\alpha]_{\rm D}^{21}$ +21.4 (*c* 0.98 in CHCl₃); ¹H NMR (500 MHz, CDCl₃) δ 0.84 (d, *J* = 6.5 Hz, 3H, uD-C^{δ}H₃), 0.93 (d, *J* = 6.6 Hz, 3H, uD-C^{δ}H₃), 1.14 (d, *J* = 6.9 Hz, 3H, uA-C^{ϵ}HCH₃), 1.18 (s, 3H, uC-C(CH₃)₂), 1.24 (s, 3H, uC-C(CH₃)₂),

1.36 (ddd, J = 13.8, 8.9, 3.5 Hz, 1H, uD-C^{β}<u>H</u>^AH^B), 1.46 (s, 3H, uA-C(CH₃)₂), 1.50 (s, 3H, uA-C(CH₃)₂), 1.66 (m, 1H, uD-C^{γ}H), 1.75 (ddd, J = 13.9, 10.5, 5.0 Hz, 1H, uD- $C^{\beta}H^{A}H^{B}$), 1.85 (m, 1H, uA-C^{{e}H}), 2.22 (ddd, J = 14.3, 10.8, 10.8 Hz, 1H, uA-C^{{e}H}^{A}H^{B}), 2.47 (dm, J = 14.2 Hz, 1H, uA-C^{γ}H^AH^B), 2.97 (dd, J = 14.5, 11.2 Hz, 1H, uB-C^{β}H^AH^B), 3.20 (dd, J = 13.5, 6.7 Hz, 1H, uC-CH^AH^BNH), 3.38 (dd, J = 13.5, 5.1 Hz, 1H, uC- $CH^{A}H^{B}NH$), 3.44 (dd, J = 14.5, 4.3 Hz, 1H, uB- $C^{\beta}H^{A}H^{B}$), 3.79 (dd, J = 8.8, 2.3 Hz, 1H, $uA-C^{\zeta}H$, 4.68 (m, 1H, $uB-C^{\alpha}H$), 4.71 (d, J = 8.7 Hz, 1H, $uA-C^{\eta}H$), 4.83 (dd, J = 10.4, 3.5 Hz, 1H, uD-C^{α}H), 5.02 (ddd, J = 11.3, 7.2, 2.0 Hz, 1H, uA-C^{δ}H), 5.68 (dm, J = 15.2 Hz, 1H, uA-C^{α}H), 5.80 (m, 1H, NH), 6.61 (ddd, J = 15.1, 10.2, 4.8 Hz, 1H, uA-C^βH), 7.09 (m, 1H, NH), 7.32–7.41 (m, 5H, uA-C^{ar}H); ¹³C{¹H} NMR (126 MHz, CDCl₃) δ 9.7 (uA-C^tH<u>C</u>H₃), 21.3 (uD-C^{δ}H₃), 22.8 (uC- C(<u>C</u>H₃)₂), 23.0 (uC-C(<u>C</u>H₃)₂ and uD- $C^{\delta}H_{3}$), 24.0 (uB- $C^{\beta}H_{2}$), 24.7 (uD- $C^{\nu}H$), 27.0 (uA- $C(\underline{C}H_{3})_{2}$), 27.2 (uA- $C(\underline{C}H_{3})_{2}$), 35.8 $(uA-C^{\gamma}H_2)$, 36.7 $(uA-C^{\epsilon}H)$, 39.5 $(uD-C^{\beta}H_2)$, 42.8 $(uC-\underline{C}(CH_3)_2)$, 46.8 $(uC-CH_2NH)$, 54.2 (uB-C^{α}H), 71.0 (uD-C^{α}H), 75.8 (uA-C^{δ}H), 80.1 (uA-C^{η}H), 82.3 (uA-C^{ζ}H), 109.1 $(uA-\underline{C}(CH_3)_2)$, 111.0 (tm, J = 18.6 Hz, $C_6F_5\underline{C}^{ipso}$), 124.4 (uA- C^{α} H), 126.6, 128.6, 128.8 (uA-C^{ar}H), 137.4 (dm, J = 252.0 Hz, C₆F₅C^{meta}), 137.5 (uA-C^{ar}), 140.2 (dm, J = 249.0Hz, $C_6F_5C^{para}$), 142.5 (uA- C^{β} H), 145.3 (dm, J = 242.1 Hz, $C_6F_5C^{ortho}$), 165.5 (uA-C(O)NH), 169.3 (C(O)NH), 170.3, 177.2 (C=O); ¹⁹F NMR (470 MHz, CDCl₃) δ -142.7 21.8, 7.9 Hz, 2F, F^{meta}); IR (ATR) v (cm⁻¹) 2931 br w, 2359 w, 2344 w, 1752 w, 1718 w, 1676 br m, 1519 s, 1501 s, 1473 w, 1369 w, 1302 w, 1229 w, 1187 w, 1151 m, 1122 m, 1060 w, 1009 w, 973 m, 882 w, 756 m, 700 m; HRMS-MALDI $(m/z)^{+}$: $[M + Na]^{+}$ calcd for $[C_{38}H_{45}F_5N_2O_8 + Na]^{+}$ 775.29883; found, 775.29718.



The acetonide **30** (26 mg, 34.5 μ mol) was converted into the epoxide according to GP3–6, the crude product was purified by preparative RP-HPLC (method M2), and the product was lyophilized to give 3.4 mg (4.9 μ mol, 14% over 4 steps) of epoxide **31** as a voluminous, colorless solid.

HPLC: $t_{\rm R}$ 37.9 min (method M2); $[\alpha]_{\rm D}^{21}$ +47.8 (c 0.50 in CHCl₃); ¹H NMR (600 MHz, CDCl₃) δ 0.85 (d, J = 6.0 Hz, 3H, uD-C⁵H₃), 0.86 (d, J = 6.0 Hz, 3H, uD-C⁵H₃), 1.16 $(d, J = 6.9 Hz, 3H, uA-C^{\ell}HCH_3)$, 1.19 (s, 3H, uC-C(CH₃)₂), 1.25 (s, 3H, uC-C(CH₃)₂), 1.35 (ddd, J = 13.6, 8.8, 3.3 Hz, 1H, uD-C^{β}H^AH^B), 1.69 (m, 1H, uD-C^{β}H^AH^B), 17.3 (m, 1H, uD-C^{γ}H), 1.81 (m, 1H, uA-C^{ϵ}H), 2.47 (ddd, J = 14.4, 10.9, 10.9 Hz, 1H, uA- $C^{\gamma}H^{A}H^{B}$), 2.58 (ddd, J = 14.4, 2.0, 2.0 Hz, 1H, uA- $C^{\gamma}H^{A}H^{B}$), 2.94 (dd, J = 7.6, 2.0 Hz, 1H, uA-C^ζH), 2.98 (m, 1H, uB-C^βH^AH^B), 3.24 (dd, J = 13.6, 7.1 Hz, 1H, uC- $CH^{A}H^{B}NH$), 3.35 (dd, J = 13.6, 4.8 Hz, 1H, uC- $CH^{A}H^{B}NH$), 3.45 (dd, J = 15.0, 4.3 Hz, 1H, uB-C^{β}H^AH^B), 3.70 (d, J = 1.8 Hz, 1H, uA-C^{η}H), 4.72 (ddd, J = 11.0, 8.5, 4.3 Hz, 1H, uB-C^{α}H), 4.88 (dd, J = 10.2, 3.4 Hz, 1H, uD-C^{α}H), 5.18 (ddd, J = 11.4, 5.1, 2.0 Hz, 1H, uA-C^{δ}H), 5.70 (d, J = 8.5 Hz, 1H, NH), 5.75 (dm, J = 15.1 Hz, 1H, uA- $C^{\alpha}H$), 6.75 (ddd, J = 15.1, 10.4, 4.7 Hz, 1H, uA- $C^{\beta}H$), 7.15 (m, 1H, NH), 7.25–7.28 (m, 2H, uA-C^{ar}H), 7.33–7.40 (m, 3H, uA-C^{ar}H); ¹³C NMR (151 MHz, CDCl₃) δ 13.6 $(uA-C^{\epsilon}HCH_3)$, 21.3 $(uD-C^{\delta}H_3)$, 22.8 $(uC-C(CH_3)_2)$, 22.9 $(uC-C(CH_3)_2)$, 23.0 $(uD-C^{\epsilon}HCH_3)$ $C^{\delta}H_{3}$), 24.0 (uB- $C^{\beta}H_{2}$), 24.6 (uD- $C^{\gamma}H$), 37.0 (uA- $C^{\gamma}H_{2}$), 39.4 (uD- $C^{\beta}H_{2}$), 40.7 (uA-C^εH), 42.7 (uC-C(CH₃)₂), 46.8 (uC-CH₂NH), 53.4 (uB-C^αH), 59.1 (uA-C^ηH), 63.0 (uA- $C^{\zeta}H$), 71.1 (uD-C^{α}H), 76.0 (uA-C^{δ}H), 110.9 (m, C₆F₅C^{*ipso*}), 124.7 (uA-C^{α}H), 125.6,

128.6, 128.8 (uA-C^{ar}H), 136.7 (uA-C^{ar}), 137.5 (dm, J = 249.4 Hz, $C_6F_5\underline{C}^{meta}$), 140.3 (dm, J = 245.0 Hz, $C_6F_5\underline{C}^{para}$), 142.1 (uA-C^βH), 145.4 (dm, J = 245.4 Hz, $C_6F_5\underline{C}^{ortho}$), 165.4 (uA-C(O)NH), 169.3 (C(O)NH), 170.6, 177.4 (C=O); ¹⁹F NMR (564 MHz, CDCl₃) δ -142.7 (dd, J = 22.6, 8.1 Hz, 2F, F^{ortho}), -155.3 (t, J = 20.9 Hz, 1F, F^{para}), -161.7 (td, J = 21.9, 8.0 Hz, 2F, F^{meta}); IR (ATR) \tilde{v} (cm⁻¹) 3289 w, 2958 br w, 2177 w, 2038 w, 2010 w, 1975 w, 1748 m, 1718 m, 1662 br m, 1520 s, 1503 s, 1473 m, 1371 w, 1322 w, 1305 w, 1189 m, 1147 s, 1125 s, 1067 w, 974 m, 887 w, 752 w, 698 m; HRMS–MALDI (m/z)⁺: [M + Na]⁺ calcd for [$C_{35}H_{39}F_5N_2O_7$ + Na]⁺ 717.25696; found, 717.25676.

4 References

- Bodnar, B. S.; Vogt, P. F. J. Org. Chem. 2009, 74, 2598–2600. doi:10.1021/jo802778z
- Eißler, S.; Nahrwold, M.; Neumann, B.; Stammler, H.-G.; Sewald, N. Org. Lett. 2007, 9, 817–819. doi:10.1021/ol0630321
- Thibonnet, J.; Abarbri, M.; Parrain, J.-L.; Duchêne, A. *Tetrahedron* 2003, *59*, 4433–4441. doi:10.1016/S0040-4020(03)00616-1
- Bulugahapitiya, P.; Landais, Y.; Parra-Rapado, L.; Planchenault, D.; Weber, V. J. Org. Chem. 1997, 62, 1630–1641. doi:10.1021/jo961952j
- 5. Nahrwold, M. ß2-Aminosäuren als Bausteine funktionalisierter Cryptophycin-Analoga, Dissertation, Universität Bielefeld, 2009.