Chemical Synthesis of Homogeneous Syndecan-1 Heparan Sulfate Glycopeptide

Supporting Information

Bo Yang, Keisuke Yoshida, Zhaojun Yin, Hang Dai, Herbert Kavunja, Mohammad H. El-Dakdouki, Suttipun Sungsuwan, Steven B. Dulaney, Xuefei Huang*

Department of Chemistry, Michigan State University, 578 S. Shaw Lane, East Lansing, Michigan 48824, USA <u>xuefei@chemistry.msu.edu</u>

Table of contents

General reaction procedures	S7
Building block preparation	S11
"3+2+3" assembly of octasaccharide 27	S24
Synthesis of syndecan-1 glycopeptide 1	S33
Probing the possibility of epimerization of glycosylated serine	S42
ESI-MS of 1	S45
HPLC chromatogram of 1	S46
¹ H-NMR (CDCl ₃ , 500 MHz) of 5	S47
¹³ C-NMR (CDCl ₃ , 125 MHz) of 5	S48
gCOSY (CDCl ₃ , 500 MHz) of 5	S49
¹ H-NMR (CDCl ₃ , 500 MHz) of 6	S50
¹³ C-NMR (CDCl ₃ , 125 MHz) of 6	S51
gCOSY (CDCl ₃ , 500 MHz) of 6	S52
¹ H-NMR (CDCl ₃ , 500 MHz) of 7	S53
¹³ C-NMR (CDCl ₃ , 125 MHz) of 7	S54
gCOSY (CDCl ₃ , 500 MHz) of 7	S55
gHMQC (CDCl ₃ , 500 MHz) of 7	S56
gHMQC (without ¹ H decoupling) (CDCl ₃ , 500 MHz) of 7	S57
gHMBC (CDCl ₃ , 500 MHz) of 7	S58
¹ H-NMR (CDCl ₃ , 500 MHz) of 8	S59
¹³ C-NMR (CDCl ₃ , 125 MHz) of 8	S60
gCOSY (CDCl ₃ , 500 MHz) of 8	S61
¹ H-NMR (CDCl ₃ , 500 MHz) of 9	S62
¹³ C-NMR (CDCl ₃ , 150 MHz) of 9	S63

gCOSY (CDCl ₃ , 500 MHz) of 9	S64
¹ H-NMR (CDCl ₃ , 500 MHz) of 13	S65
¹³ C-NMR (CDCl ₃ , 125 MHz) of 13	S66
gCOSY (CDCl ₃ , 500 MHz) of 13	S67
¹ H-NMR (CDCl ₃ , 500 MHz) of 14	S68
¹³ C-NMR (CDCl ₃ , 125 MHz) of 14	S69
gCOSY (CDCl ₃ , 500 MHz) of 14	S70
¹ H-NMR (CDCl ₃ , 500 MHz) of 18	S71
¹³ C-NMR (CDCl ₃ , 125 MHz) of 18	S72
gCOSY (CDCl ₃ , 500 MHz) of 18	S73
¹ H-NMR (CDCl ₃ , 500 MHz) of 19	S74
¹ H-NMR (CDCl ₃ , 500 MHz) of 21	S75
¹³ C-NMR (CDCl ₃ , 150 MHz) of 21	S76
gCOSY (CDCl ₃ , 600 MHz) of 21	S77
gHMQC (CDCl ₃ , 600 MHz) of 21	S78
gHMQC (without ¹ H decoupling) (CDCl ₃ , 600 MHz) of 21	S79
¹ H-NMR (CDCl ₃ , 500 MHz) of 24	S80
¹³ C-NMR (CDCl ₃ , 125 MHz) of 24	S81
gCOSY (CDCl ₃ , 500 MHz) of 24	S82
gHMQC (CDCl ₃ , 500 MHz) of 24	S83
gHMQC (without ¹ H decoupling) (CDCl ₃ , 500 MHz) of 24	S84
¹ H-NMR (CDCl ₃ , 600 MHz) of 25	S85
¹³ C-NMR (CDCl ₃ , 150 MHz) of 25	S86
gCOSY (CDCl ₃ , 600 MHz) of 25	S87
gHMQC (CDCl ₃ , 600 MHz) of 25	S88
gHMQC (without ¹ H decoupling) (CDCl ₃ , 600 MHz) of 25	S89
gHMBC (CDCl ₃ , 600 MHz) of 25	S90
¹ H-NMR (CDCl ₃ , 600 MHz) of 25a	S91
¹³ C-NMR (CDCl ₃ , 150 MHz) of 25a	S92
gCOSY (CDCl ₃ , 600 MHz) of 25a	S93
gHMQC (CDCl ₃ , 600 MHz) of 25a	S94
gHMQC (without ¹ H decoupling) (CDCl ₃ , 600 MHz) of 25a	S95
gHMBC (CDCl ₃ , 600 MHz) of 25a	S96
¹ H-NMR (CDCl ₃ , 600 MHz) of 26	S97
¹³ C-NMR (CDCl ₃ , 150 MHz) of 26	S98
gCOSY (CDCl ₃ , 600 MHz) of 26	S99
gHMQC (CDCl ₃ , 600 MHz) of 26	S100
gHMQC (without ¹ H decoupling) (CDCl ₃ , 600 MHz) of 26	S101
gHMBC (CDCl ₃ , 600 MHz) of 26	S102
¹ H-NMR (CDCl ₃ , 600 MHz) of 27	S103

¹³ C-NMR (CDCl ₃ , 150 MHz) of 27	S104
gCOSY (CDCl ₃ , 600 MHz) of 27	S105
gHMQC (CDCl ₃ , 500 MHz) of 27	S106
gHMQC (without ¹ H decoupling) (CDCl ₃ , 500 MHz) of 27	S107
gHMBC (CDCl ₃ , 500 MHz) of 27	S108
¹ H-NMR (CDCl ₃ , 500 MHz) of 28	S109
¹ H-NMR (CDCl ₃ , 500 MHz) of 29	S110
¹³ C-NMR (CDCl ₃ , 125 MHz) of 29	S111
¹ H-NMR (CDCl ₃ , 500 MHz) of 30	S112
¹³ C-NMR (CDCl ₃ , 125 MHz) of 30	S113
¹ H-NMR (CDCl ₃ , 500 MHz) of 31	S114
¹³ C-NMR (CDCl ₃ , 125 MHz) of 31	S115
HPLC chromatogram of crude peptide 32	S116
¹ H-NMR (CDCl ₃ , 500 MHz) of 34	S117
ESI-MS of 36	S118
ESI-MS of 37	S119
¹ H-NMR (CDCl ₃ , 500 MHz) of S2	S120
¹ H-NMR (CDCl ₃ , 500 MHz) of S3	S121
¹ H-NMR (CDCl ₃ , 500 MHz) of S4	S122
¹³ C-NMR (CDCl ₃ , 125 MHz) of S4	S123
gCOSY (CDCl ₃ , 500 MHz) of S4	S124
gHMQC (CDCl ₃ , 500 MHz) of S4	S125
gHMBC (CDCl ₃ , 500 MHz) of S4	S126
¹ H-NMR (CDCl ₃ , 500 MHz) of S5	S127
¹³ C-NMR (CDCl ₃ , 125 MHz) of S5	S128
¹ H-NMR (CDCl ₃ , 500 MHz) of S6	S129
¹³ C-NMR (CDCl ₃ , 125 MHz) of S6	S130
gCOSY (CDCl ₃ , 500 MHz) of S6	S131
¹ H-NMR (CDCl ₃ , 500 MHz) of S7	S132
¹³ C-NMR (CDCl ₃ , 125 MHz) of S7	S133
gCOSY (CDCl ₃ , 500 MHz) of S7	S134
¹ H-NMR (CDCl ₃ , 500 MHz) of S8	S135
¹³ C-NMR (CDCl ₃ , 125 MHz) of S8	S136
gCOSY (CDCl ₃ , 500 MHz) of S8	S137
¹ H-NMR (CDCl ₃ , 500 MHz) of S9	S138
¹³ C-NMR (CDCl ₃ , 125 MHz) of 89	S139
gCOSY (CDCl ₃ , 500 MHz) of S9	S140
¹ H-NMR (CDCl ₃ , 500 MHz) of S13	S141
¹ H-NMR (CDCl ₃ , 500 MHz) of S16	S142
¹³ C-NMR (CDCl ₃ , 125 MHz) of S16	S143

gCOSY (CDCl ₃ , 500 MHz) of S16	S144
¹ H-NMR (CD ₃ OD, 500 MHz) of S17	S145
¹³ C-NMR (CD ₃ OD, 125 MHz) of S17	S146
¹ H-NMR (CDCl ₃ , 500 MHz) of S18	S147
¹³ C-NMR (CDCl ₃ , 125 MHz) of S18	S148
gCOSY (CDCl ₃ , 500 MHz) of S18	S149
gHMQC (CDCl ₃ , 500 MHz) of S18	S150
gHMBC (CDCl ₃ , 500 MHz) of S18	S151
¹ H-NMR (CDCl ₃ , 500 MHz) of S19	S152
¹ H-NMR (CDCl ₃ , 500 MHz) of S20	S153
¹³ C-NMR (CDCl ₃ , 125 MHz) of S20	S154
gCOSY (CDCl ₃ , 500 MHz) of S20	S155
gHMQC (CDCl ₃ , 500 MHz) of S20	S156
¹ H-NMR (CDCl ₃ , 500 MHz) of S21	S157
¹³ C-NMR (CDCl ₃ , 125 MHz) of S21	S158
gCOSY (CDCl ₃ , 500 MHz) of S21	S159
¹ H-NMR (CDCl ₃ , 500 MHz) of S22	S160
¹³ C-NMR (CDCl ₃ , 125 MHz) of S22	S161
gCOSY (CDCl ₃ , 500 MHz) of S22	S162
¹ H-NMR (CDCl ₃ , 500 MHz) of S23	S163
¹³ C-NMR (CDCl ₃ , 125 MHz) of S23	S164
gCOSY (CDCl ₃ , 500 MHz) of S23	S165
¹ H-NMR (CDCl ₃ , 500 MHz) of S24	S166
¹³ C-NMR (CDCl ₃ , 125 MHz) of S24	S167
gCOSY (CDCl ₃ , 500 MHz) of S24	S168
¹ H-NMR (CDCl ₃ , 500 MHz) of S25	S169
¹³ C-NMR (CDCl ₃ , 125 MHz) of S25	S170
gCOSY (CDCl ₃ , 500 MHz) of S25	S171
¹ H-NMR (CDCl ₃ , 500 MHz) of S26	S172
¹³ C-NMR (CDCl ₃ , 125 MHz) of S26	S173
gCOSY (CDCl ₃ , 500 MHz) of S26	S174
¹ H-NMR (CDCl ₃ , 500 MHz) of S27	S175
¹ H-NMR (CDCl ₃ , 500 MHz) of S28	S176
¹ H-NMR (CDCl ₃ , 500 MHz) of S29	S177
¹ H-NMR (CDCl ₃ , 500 MHz) of S30	S178
¹³ C-NMR (CDCl ₃ , 125 MHz) of S30	S179
gCOSY (CDCl ₃ , 500 MHz) of S30	S180
gHMQC (CDCl ₃ , 500 MHz) of S30	S181
gHMBC (CDCl ₃ , 500 MHz) of S30	S182
¹ H-NMR (CDCl ₃ , 500 MHz) of S31	S183

¹³ C-NMR (CDCl ₃ , 150 MHz) of S31	S184
gCOSY (CDCl ₃ , 500 MHz) of S31	S185
gHMQC (CDCl ₃ , 600 MHz) of S31	S186
gHMQC (without ¹ H decoupling) (CDCl ₃ , 600 MHz) of S31	S187
gHMBC (CDCl ₃ , 600 MHz) of S31	S188
¹ H-NMR (CDCl ₃ , 500 MHz) of S32	S189
¹³ C-NMR (CDCl ₃ , 125 MHz) of S32	S190
gCOSY (CDCl ₃ , 500 MHz) of S32	S191
¹ H-NMR (CDCl ₃ , 500 MHz) of S33	S192
¹ H-NMR (CDCl ₃ , 600 MHz) of S34	S193
¹³ C-NMR (CDCl ₃ , 150 MHz) of S34	S194
gCOSY (CDCl ₃ , 600 MHz) of S34	S195
gHMQC (CDCl ₃ , 600 MHz) of S34	S196
gHMQC (without ¹ H decoupling) (CDCl ₃ , 600 MHz) of S34	S197
¹ H-NMR (CDCl ₃ , 600 MHz) of S36	S198
¹³ C-NMR (CDCl ₃ , 150 MHz) of S36	S199
gHMQC (CDCl ₃ , 600 MHz) of S36	S200
gHMQC (without ¹ H decoupling) (CDCl ₃ , 500 MHz) of S36	S201
¹ H-NMR (CDCl ₃ , 500 MHz) of S37	S202
gCOSY (CDCl ₃ , 500 MHz) of S37	S203
¹ H-NMR (CDCl ₃ , 500 MHz) of S38	S204
¹³ C-NMR (CDCl ₃ , 125 MHz) of S38	S205
gCOSY (CDCl ₃ , 500 MHz) of \$38	S206
¹ H-NMR (CDCl ₃ , 500 MHz) of S39	S207
¹³ C-NMR (CDCl ₃ , 125 MHz) of S39	S208
gCOSY (CDCl ₃ , 500 MHz) of 39	S209
¹ H-NMR (CDCl ₃ , 500 MHz) of S40	S210
¹³ C-NMR (CDCl ₃ , 125 MHz) of S40	S211
gCOSY (CDCl ₃ , 500 MHz) of S40	S212
¹ H-NMR (CDCl ₃ , 500 MHz) of S42	S213
¹³ C-NMR (CDCl ₃ , 125 MHz) of S42	S214
gCOSY (CDCl ₃ , 500 MHz) of S42	S215
¹ H-NMR (CDCl ₃ , 500 MHz) of S43	S216
¹³ C-NMR (CDCl ₃ , 125 MHz) of S43	S217
gCOSY (CDCl ₃ , 500 MHz) of S43	S218
¹ H-NMR (CDCl ₃ , 500 MHz) of S44	S219
¹³ C-NMR (CDCl ₃ , 125 MHz) of S44	S220
gCOSY (CDCl ₃ , 500 MHz) of S44	S221
gHMQC (CDCl ₃ , 500 MHz) of S44	S222
gHMQC (without ¹ H decoupling) (CDCl ₃ , 500 MHz) of S44	S223

gHMBC (CDCl ₃ , 500 MHz) of S44	S224
¹ H-NMR (CDCl ₃ , 600 MHz) of S45	S225
¹³ C-NMR (CDCl ₃ , 150 MHz) of S45	S226
gCOSY (CDCl ₃ , 600 MHz) of S45	S227
¹ H-NMR (CDCl ₃ , 500 MHz) of S46	S228
¹³ C-NMR (CDCl ₃ , 125 MHz) of S46	S229
gCOSY (CDCl ₃ , 500 MHz) of S46	S230
¹ H-NMR (CDCl ₃ , 500 MHz) of S50	S231
¹³ C-NMR (CDCl ₃ , 125 MHz) of S50	S232
¹ H-NMR (CDCl ₃ , 500 MHz) of S51	S233
¹³ C-NMR (CDCl ₃ , 125 MHz) of S51	S234
gCOSY (CDCl ₃ , 500 MHz) of S51	S235
MALDI-MS of S52	S236
MALDI-MS of S53	S237
¹ H-NMR (CDCl ₃ , 600 MHz) of S55	S238
¹³ C-NMR (CDCl ₃ , 150 MHz) of \$55	S239
¹ H-NMR (CDCl ₃ , 500 MHz) of S56	S240
¹³ C-NMR (CDCl ₃ , 125 MHz) of S56	S241
¹ H-NMR (CDCl ₃ , 500 MHz) of S60	S242
¹ H-NMR (CDCl ₃ , 500 MHz) of S61	S243
¹³ C-NMR (CDCl ₃ , 125 MHz) of S61	S244
¹ H-NMR (CDCl ₃ , 500 MHz) of S62	S245

General Experimental Procedures. All reactions were carried out under nitrogen with anhydrous solvents in flame-dried glassware, unless otherwise noted. Glycosylation reactions were performed in the presence of molecular sieves, which were flame-dried right before the reaction under high vaccum. Glycosylation solvents were dried using a solvent purification system and used directly without further drying. Chemicals used were reagent grade as supplied except where noted. Compounds were visualized by UV light (254 nm) and by staining with a yellow solution containing Ce(NH₄)₂(NO₃)₆ (0.5 g) and (NH₄)₆Mo₇O₂₄ 4H₂O (24.0 g) in 6% H₂SO₄ (500mL). Flash column chromatography was performed on silica gel 60 (230-400 Mesh). NMR spectra were referenced using residual CHCl₃ (δ ¹H-NMR 7.26 ppm, ¹³C-NMR 77.0 ppm). Peak and coupling constants assignments are based on ¹H-NMR, ¹H-¹H gCOSY and (or) ¹H-¹³C gHMQC and ¹H-¹³C gHMBC experiments.

Characterization of anomeric stereochemistry. The stereochemistries of the newly formed glycosidic linkages were determined by ${}^{3}J_{\rm H1,H1}$ through ¹H-NMR and/or ${}^{1}J_{\rm C1,H1}$ through gHMQC 2-D NMR (without ¹H decoupling). Smaller coupling constants of ${}^{3}J_{\rm H1,H2}$ (around 3 Hz) indicate α linkages and larger coupling constants ${}^{3}J_{\rm H1,H1}$ (7.2 Hz or larger) indicate β linkages. ${}^{1}J_{\rm C1,H1}$ around 170 Hz suggests α linkages and 160 Hz suggests β linkages.^[1]

General procedure for pre-activation based single-step glycosylation. A solution of donor (60 µmol) and freshly activated molecular sieve MS 4 Å (200 mg) in DCM (2 mL) was stirred at room temperature for 30 minutes, and cooled to -78 °C, which was followed by addition of AgOTf (47 mg, 180 µmol) dissolved in Et₂O (1 mL) without touching the wall of the flask. After 5 minutes, orange colored p-ToISCI (9.5 µL, 60 µmol) was added to the solution through a microsyringe. Since the reaction temperature was lower than the freezing point of p-TolSCl, p-TolSCl was added directly into the reaction mixture to prevent it from freezing on the flask wall. The characteristic yellow color of p-TolSCl in the reaction solution dissipated rapidly within a few seconds indicating depletion of p-TolSCI. After the donor was completely consumed according to TLC analysis (about 5 minutes at -78 °C), a solution of acceptor (54 μ mol) in DCM (0.2 mL) was slowly added dropwise via a syringe together with one equivalent of TTBP. The reaction mixture was warmed to 0 °C under stirring in 2 h. Then the mixture was diluted with DCM (20 mL) and filtered over Celite. The Celite was further washed with DCM until no organic compounds were observed in the filtrate by TLC. All DCM solutions were combined and washed twice with a saturated aqueous solution of NaHCO₃ (20 mL) and twice with water (10 mL). The organic layer was collected and dried over Na₂SO₄. After removal of the solvent, the desired oligosaccharide was purified from the reaction mixture via silica gel flash chromatography.

General procedure for protection of 6-OH with Lev. The compound containing 6-OH (1 equiv.) was dissolved in DCM (for 0.5 g of compound, 5 mL), followed by addition of levulinoyl acid (1.4 equiv.), EDC-HCl (1.6 equiv.) and DMAP (0.1 equiv.). The mixture was stirred at room temperature overnight and then was diluted with DCM (100 mL). The organic phase was washed with a saturated aqueous solution of NaHCO₃ and then dried

over Na₂SO₄. The solvent was concentrated in vacuo and the compound was purified by silica gel column chromatography.

General procedure for deprotection of Lev. The Lev-protected compound (1 equiv.) was dissolved in DCM/MeOH (for 150 mg of compound, 2.4 mL, 1:1) and acetic acid (0.2 mL). The mixture was cooled to 0 °C, followed by addition hydrazine monohydrate (5 equiv. for each Lev). The mixture was stirred at 0 °C for 2h and then was quenched by acetone (0.28 mL). The mixture was stirred at room temperature for another 1h and the acetone was evaporated under vaccum. The residue was diluted with EtOAc (50 mL) and washed with a saturated aqueous solution of NaHCO₃, 10% HCl and water and the organic phase was dried over Na₂SO₄. The solvent was concentrated in vacuo and the compound was purified by silica gel column chromatography.

General procedure for *O***-sulfation.** The mixture of OH-containing compound (for 20 mg of compound, 1 equiv.), DMF (1 mL) and SO₃-NEt₃ (20 equiv. per OH) was stirred at 55 °C for 24 h. The mixture was cooled to room temperature and then diluted with DCM/MeOH (1mL/1mL). The resulting solution was layered on the top of Sephadex LH-20 chromatography column that was eluted with DCM/MeOH (1/1, v/v). The fractions containing product were combined and evaporated to dryness under vacuo without further purification.

General procedure for global debenzylation. The mixture of the Bn/PMB-containing compound (for 3 mg of compound, 1 equiv.), DCM/MeOH (1 mL/1 mL) and Pd/C (15 mg) was stirred under H₂ at room temperature overnight and then filtered via PTFE membrane (pore size 0.22 μ m). The filtrate was concentrated to dryness under vacuum and then diluted with H₂O (15 mL). The aqueous phase was further washed with DCM (5 mL × 3) and MeOH (5 mL×3) and then the aqueous phase was dried under vacuum. The crude product was further purified by Sephadex LH-20 chromatography column. The fractions containing product were combined and evaporated to dryness under vacuo without further purification.

General procedure for solid phase peptide synthesis. Amino acids were purchased from Chem-impex. H-Gly-2-CTrt Resin was purchased from Advanced ChemTech (loading level: 0.6 mmol/g). Reaction vessel syringes (10 mL, disposable) and the Domino Block Synthesizer were purchased from Torvig.

(1) Pause Point:

If it is necessary to pause the synthesis, after each coupling-washing procedure, the resin is washed with DCM five times and dried with nitrogen gas at room temperature. The syringe was closed with a plunger and a cap, and stored at < 4 °C. Before resuming the synthesis, the sample was allowed to reach room temperature, and the dry resin was swelled as described in the following.

(2) Select the right syringe:

The optimal available volume for 6 mL syringe is < 4.8 mL;

The optimal available volume for 12 mL syringe is < 8.0 mL;

The concentration of amino acid used for coupling typically is $0.2 \sim 0.4$ M;

The final volume of swelling resin, solvent, coupling reagent need to be considered before choosig the right syringe. As an example for 200mg H-Gly-2-ClTrt-Resin with 10 eq amino acid, 9.9 eq HBTU, 20 eq DIPEA, the final reaction volume is about $6 \sim 7$ mL. Therefore, the 12 mL syringe should be used.

(3) Reagent:

HBTU: MW 379.24

223 mg (4.9 eq) for 200 mg resin (0.6 mmol/g);

451 mg (9.9 eq) for 200 mg resin (0.6 mmol/g);

DIPEA: MW 129.24 d 0.742

0.21 mL (10 eq) for 200 mg resin (0.6 mmol/g);

0.42 mL (20 eq) for 200 mg resin (0.6 mmol/g).

DIPEA usually is 2 eq;

20% piperidine in DMF:

20 mL piperidine + 80 mL DMF 10 mL/ g resin

Capping Reagent:

 $10 \text{ mL Ac}_2\text{O} + 10 \text{ mL DIPEA} + 80 \text{ mL DMF}$

Kaiser Reagent:

80% Phenol in EtOH (W/V);

- 2 mL 0.001M KCN + 98 mL Pyridine;
- 5% Ninhydrin in EtOH (W/V).
- (4) Standard Washing Procedure:
- This procedure is performed when Fmoc is removed or the coupling reaction is finished.
 - (a) Push the plunger to remove the reaction mixture, and then pull it back to the right position;
 - (b) Put the syringe in the plate;
 - (c) Fill the syringe with DMF;
 - (d) Take the plate out of the shaker, carefully wash the plunger and edge of the syringe with DMF, shake for 10s, and remove the solvent by filtration;
 - (e) Repeat steps a and b three times with DMF;
 - (f) Repeat steps a and b three times with DCM;
 - (g) Repeat steps a and b three times with DMF.

Section 1: Resin Swelling

- (a) Place the dry resin (200mg) in the syringe;
- (b) Fill the reactor with DCM until all the resin beads are immersed;
- (c) Shake for 30 min;
- (d) Remove DCM by vacuum filtration.

Section 2: Removal of Fmoc

(a) The swelled resin is washed once with DMF;

- (b) Fill the syringe with 20% piperidine in DMF (3 ml) and shake for 30 min;
- (c) Remove the DMF/piperidine solution by pushing the plunger;
- (d) Repeat steps b and c once;
- (e) Repeat the standard washing procedure once;

Section 3: Coupling Reaction

(a) Mix the Fmoc protected amino acid (5 eqiv.), DIPEA (10 eqiv.), and HBTU (4.9 eqiv.) with DMF in a dry vial. The reaction mixture is then transferred to syringe when all the compounds are completely dissolved;

- (b) Shake the reaction for 2 h;
- (c) Repeat standard washing procedure;
- (d) Check the resin with Kaiser Method to make sure no free amine is left; Repeat steps a and b if necessary.

Section 4: Capping

- (a) Fill the syringe with the capping reagent, and shake for 15 min;
- (b) Remove the capping solution by pushing the plunger and repeat step a for 15 min;
- (c) Repeat the standard washing procedure;

Section 5: Final Cleavage

- (a) Weigh the resin and place it in a round bottom flask. When the resin is dry, swell it as described in section 1;
- (b) Add 10 mL of cleavage cocktail (TFA/H₂O/Phenol/TIPS 8.5/0.5/0.5/0.5) per 100mg of resin, stir gently for 2h;
- (c) Filter the resin and wash it twice with fresh cleavage cocktail. Recover the filtrate in a rounD-bottom flask;
- (d) Concentrate the cleavage cocktail in vacuum to approximately ¹/₄ of its original volume;
- (e) Under vigorous stirring, add cold MTBE to precipitate the peptide. At least 10 times the initial TFA volume of MTBE should be added to precipitate the unprotected peptide. When the peptide does not precipitate, concentrate the solution in vacuum and go directly to step 8;
- (f) Filter out the precipitate;
- (g) Triturate and wash by filtration the precipitated peptide three times with MBTE;
- (h) Solubilize the peptide in CH₃CN/H₂O/TFA 50/50/0.1 and lyophilize. Solvent used for this step can be changed to increase solubility. The crude peptide is used for HPLC analysis.

General procedure for HPLC analysis

(1) Preparation of Sample:

- (a) Transfer a sample containing \sim 1- 2 mg dry peptide-resin to a small syringe (2 ml);
- (b) Add 300 μ l of the cleavage cocktail to the dried peptide resin, stir for 3h;
- (c) Collect the solution in a small HPLC vial, dilute with 400 μl ACN/H₂O 1/1 and mix;
- (d) At this point, the solution can be analyzed in an analytical HPLC system (inject 20 μ l) and /or further diluted (1/10) to be injected (2 μ l) in liquid chromatograph-mass spectrometer.
- (2) Preparation of HPLC Solvent:
 - (a) Eluent A: weak mobile phase solvent.
 Case A: 0.1% TFA in H₂O (1 ml TFA + 1 L H₂O)
 Case B: 0.12 % TFA in H₂O (1.2 ml TFA + 1 L H₂O)
 - (b) Eluent B: strong mobile phase solvent. Case A: 80% ACN / 0.1% TFA or 0.085% TFA/ACN (v/v)

(3) Choice of HPLC Column:

(a) low picomole amount of peptide: 0.21 cm \times 25 cm , 0.3 ml/min

- (b) < 1 mg peptide: 0.46 cm \times 25 cm , 1.0-1.5 ml/min
- (c) 1.0-10.0 mg peptide: $1.0 \text{ cm} \times 25 \text{ cm}$, 2.0 ml/min
- (d) >10 mg peptide: 2.2 cm \times 25 cm , 6.0-10.0 ml/min

(4) Column Preparation:

- (a) Eluent B, 2 ml/min, 20 min;
- (b) Decrease Eluent B to 0% over 10 min using a linear gradient;
- (c) Increase Eluent B to 100% over 10 min using a linear gradient;
- (d) Repeat 2;
- (e) Equilibrate the column with Eluent A for 20 min.
- (5) Detector: 220 nm

Building block preparation



Synthesis of galactose building block **S9**. Reagents and conditions: (a) BF_3/Et_2O , *p*-TolSH, r.t.; (b) NaOMe, DCM/MeOH, r.t.; (c) Bu_2SnO , toluene/THF, reflux, 3 h, then AllBr, Bu_4NBr , THF, reflux; (d) benzaldehyde dimethyl acetal, CSA, CH₃CN; (e) BzCl, DMAP, pyridine, 50 °C; (f) *p*-TsOH, DCM/MeOH, r.t.; (g) NaH, BnBr, DMF; (h) [Ir(COD)(PMePh_2)_2PF_6], THF, then H₂O, I₂, H₂, 0 °C-r.t..

2-O-benzoyl-4,6-di-O-benzyl-1-thio-β-D-galactopyranoside *p*-*Tolyl* **(S9)**. β-D-Galactopyranosyl pentaacetate S1 (10 g, 25.64 mol), p-toluenethiol (3.62 g, 29 mmol) were dissolved in DCM (100 mL) and boron trifluoride etherate (10.1 mL, 75 mmol) was added dropwise at room temperature. The mixture was stirred under N2 at room temperature for 20 hours and then diluted with DCM (200 mL). The organic phase was washed with a saturated aqueous solution of NaHCO3 until the pH reached 7 and then dried over Na₂SO₄, filtered and concentrated to afford crude product S2, which was recrystalized from hexanes/EtOAc. ¹H-NMR (500 MHz, CDCl₃): δ 1.95 (s, 3 H, COCH₃), 2.02 (s, 3 H, COCH₃), 2.07 (s, 3 H, COCH₃), 2.09 (s, 3 H, COCH₃), 2.32 (S, 3 H, SPhCH₃), 3.86-3.90 (m, 1 H), 4.07-4.18 (m, 2 H), 4.62 (d, 1 H, $J_{1,2} = 10$ Hz, H-1), 5.00-5.03 (m, 1 H), 5.19 (t, 1 H, J_{1,2} = 10 Hz), 5.38 (dd, 1 H, J = 1 Hz, J = 3.5 Hz), 7.09-7.11 (m, 2 H), 7.38-7.40 (m, 2 H). Compound S2 (9.89 g) was dissolved in MeOH (50 mL) and DCM (50 mL). 5.4 M NaOMe (19 mL, 0.1 mol) was added and the mixture was stirred at room temperature overnight. The mixture was neutralized by conc. HCl until the pH was around 7 and then concentrated and dried under vacuum. Silica gel column chromatography (9:1

DCM-MeOH) afforded p-tolyl 1-thio- β -D-galactopyranoside S3 as white solid (5.86 g, 94%). ¹H-NMR (500 MHz, CD₃OD): δ 2.29 (s, 3 H, SPhCH₃), 3.45-3.48 (m, 1 H), 3.51-3.53 (m, 1 H), 3.56 (t, 1 H, J = 4.5 Hz), 3.67-3.76 (m, 2 H), 3.87 (dd, 1 H, J = 1 Hz, J = 3.5Hz), 4.49 (d, 1 H, $J_{1,2} = 9.5$ Hz), 7.09-7.11 (m, 2 H), 7.43-7.45 (m, 2 H). Compound S3 with dibutyltin oxide (6.12 g, 24.6 mmol) in a flask equipped with a Dear-Stark device in anhydrous toluene and THF (200 mL) for 3 h and then concentrated. After cooling the reaction mixture down to room temperature, anhydrous THF (100 mL) was added followed by addition of Bu₄NBr (3.42 g, 22.5 mmol) and AllBr (2.69 mL, 22.5 mmol). The mixture was stirred for 4 h under reflux. After the reaction was complete, THF was removed under reduced pressure. The resulting residue was purified by silica gel column (1:1, hexanes-EtOAc) to afford p-tolyl 3-O-allyl-1-thio- β -D-galactopyranoside S4 (4.67 g, 70%). ¹H-NMR (500 MHz, CD₃OD): δ 2.30 (s, 3 H, SPhCH₃), 3.13-3.34 (m, 1 H), 3.65-3.78 (m, 3 H, H-2, H-6a, H-6b), 4.06-4.07 (m, 1 H, H-4), 4.13-4.25 (m, 2 H, CH_2CHCH_2O), 4.52 (d, 1 H, $J_{1,2} = 9.5$ Hz, H-1), 5.14-5.17 (m, 1 H, CH_2CHCH_2O), 5.30-5.35 (m, 1 H, CH₂CHCH₂O), 5.96-6.01 (m, 1 H, CH₂CHCH₂O), 7.10-7.12 (m, 2 H), 7.44-7.46 (m, 2 H). ¹³C-NMR (125 MHz, CD₃OD): δ 13.8, 20.8, 21.1, 26.8, 54.1, 62.6, 67.3, 70.0, 71.8, 80.3, 83.5, 90.5, 117.3, 130.5, 131.8, 133.0, 136.4, 138.4. HRMS: C₁₆H₂₂O₅S [M+NH₄]⁺ calcd: 344.1532, obsd: 344.1542. Compound **S4** (4.67 g, 14.34 mmol) was dissolved in CH₃CN (100 mL) followed by addition of camphorsulfonic acid (CSA, 999 mg, 4.302 mmol) and benzaldehyde dimethyl acetal (3.24 mL, 21.51 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was quenched by Et₃N and diluted with EtOAc (100 mL) and the organic phase was extracted by sat. NaHCO₃ solution and dried over Na₂SO₄. After concentration, the residue was purified by silica gel column (5:1:1, hexanes-DCM-EtOAc) to afford p-tolyl 3-Oallyl-4,6-O-benzylidene-1-thio- β -D-galactopyranoside S5 (4.87 g, 82%). ¹H-NMR (500 MHz, CDCl₃): δ 2.31 (s, 3 H, SPhCH₃), 2.43 (d, 1 H, J = 1.5 Hz, OH), 3.44-3.47 (m, 2 H, H-3, H-5), 3.81 (dt, 1 H, J = 1.5 Hz, J = 9.5 Hz, H-2), 3.98-4.01 (m, 1 H, H-6a), 4.13-4.21 (m, 3 H, H-4, CH_2CHCH_2O), 4.34-4.37 (m, 1 H, H-6b), 4.56 (d, 1 H, J = 9.5 Hz, H-1), 5.15-5.18 (m, 1 H, CH₂CHCH₂O), 5.25-5.29 (m, 1 H, CH₂CHCH₂O), 5.47 (s, 1 H, PhCH), 5.86-5.93 (m, 1 H, CH₂CHCH₂O), 7.02-7.04 (m, 2 H), 7.29-7.38 (m, 5 H), 7.55-7.57 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.4, 67.3, 69.7, 70.3, 71.1, 73.6, 80.3, 87.4, 101.4, 118.0, 126.8, 126.9, 128.2, 129.2, 129.9, 134.6, 135.0, 138.1, 138.6. HRMS: C₂₃H₂₆O₅S $[M+NH_4]^+$ calcd: 432.1845, obsd: 432.1831. Compound S5 (4.87 g, 11.76 mmol) was dissolved in dry pyridine (100 mL) followed by addition of DMAP (143 mg, 1.176 mmol) and benzovl chloride (2.05, 17.64 mmol). The resulting mixture was stirred under 50 °C overnight. After cooling down to room temperature, the reaction mixture was diluted with DCM and extracted with 10% HCl solution. The combined organic phase was washed with sat. NaHCO₃ solution and dried over Na₂SO₄. After concentration, the residue was purified by silica gel column (5:1:1, hexane-DCM-EtOAc) to afford p-tolyl 2-O-benzoyl-3-O-allyl-4, 6-O-benzylidene-1-thio- β -D-galactopyranoside S6 (5.48 g, 90%). ¹H-NMR (500 MHz, CDCl₃): δ 2.30 (s, 3 H, SPhCH₃), 3.52-3.54 (m, 1 H, H-5), 3.75 (dd, 1 H, J = 3.5 Hz, J =9.5 Hz, H-3), 3.97-4.08 (m, 3 H, H-6a, CH_2CHCH_2O), 4.28 (dd, 1 H, J = 1 Hz, J = 3.5 Hz, H-4), 4.37-4.40 (m, 1 H, H-6b), 4.78 (d, 1 H, J = 9.5 Hz, H-1), 5.02-5.05 (m, 1 H, CH₂CHCH₂O), 5.11-5.15 (m, 1 H, CH₂CHCH₂O), 5.43 (t, 1 H, J = 9.5 Hz, H-2), 5.67-5.76 (m, 1 H, CH₂CHCH₂O), 7.01-7.03 (m, 2 H), 7.30-7.34 (m, 3 H), 7.39-7.48 (m, 6 H), 7.55-7.59 (m, 1 H), 8.04-8.06 (m, 2H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.2, 69.3, 69.3, 70.1,

70.7, 73.8, 78.4, 85.5, 101.2, 117.4, 126.6, 127.5, 128.0, 128.3, 128.9, 129.4, 129.7, 130.3, 132.9, 134.4, 134.6, 137.6, 138.1, 164.8. HRMS: C₃₀H₃₀O₆S [M+NH₄]⁺ calcd: 536.2107, obsd: 530.2036. Compound **S6** (5.48 g, 10.58 mmol) was dissolved in DCM/MeOH (1:1. 100 mL) followed by addition of p-TsOH (685 mg, 3.98 mmol). The reaction mixture was kept under room temperature overnight and quenched with Et₃N. After concentration, the resulting residue was purified by silica gel column (2:1:2, hexanes-DCM-EtOAc) to afford *p*-tolyl 2-*O*-benzoyl-3-*O*-allyl-1-thio- β -D-galactopyranoside S7 (3.64 g, 80%). ¹H-NMR (500 MHz, CDCl₃): δ 2.10-2.13 (m, 1 H, OH), 2.29 (s, 3 H, SPhCH₃), 2.60 (br, 1 H, OH), 3.59-3.65 (m, 2 H, H-3, H-5), 3.79-3.85 (m, 1 H, H-6a), 3.96-4.03 (m, 2 H, CH₂CHCH₂O), 4.07-4.11 (m, 2 H, H-4, H-6b), 4.72 (d, 1 H, J = 10 Hz, H-1), 5.06-5.09 (m, 1 H, CH₂CHCH₂O), 5.13-5.17 (m, 1 H, CH₂CHCH₂O), 5.40 (t, 1 H, J = 10 Hz, H-2), 5.66-5.73 (m, 1 H, CH₂CHCH₂O), 7.04-7.06 (m, 2 H), 7.32-7.34 (m, 2 H), 7.43-7.47 (m, 2 H), 7.56-7.59 (m, 1 H), 8.04-8.06 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.2, 62.6, 67.2, 69.3, 69.8, 71.0, 78.4, 79.5, 86.8, 118.1, 128.4, 129.0, 129.6, 129.8, 129.9, 132.9, 133.1, 133.9, 138.1, 165.2. HRMS: $C_{23}H_{26}O_6S [M+H]^+$ calcd: 431.1528, obsd: 431.1508. Freshly activated MS AW 300 (2 g) was mixed with compound S7 (3.64 g, 8.47 mmol) in dry DMF. Under N₂, the resulting mixture was stirred under room temperature for 30 minutes, followed by addition of NaH (485 mg, 20.38 mmol) and BnBr (4.03 mL, 33.88 mmol). After the reaction was complete, it was guenched by 10% HCl and diluted with DCM. The organic phase was extracted with sat. NaHCO₃ and dried over Na₂SO₄. After concentration, the residue was purified by silica gel column (4:1, hexanes-EtOAc) to afford p-tolyl 2-Obenzoyl-3-O-allyl-4,6-di-O-benzyl-1-thio- β -D-galactopyranoside S8 (4.33 g, 84%). ¹H-NMR (500 MHz, CDCl₃): δ 2.26 (s, 3 H, SPhCH₃), 3.62-3.69 (m, 4 H, H-3, H-5, H-6a, H-6b), 3.94-3.99 (m, 2 H, CH₂CHCH₂O, H-4), 4.05-4.10 (m, 1 H, CH₂CHCH₂O), 4.40-4.47 (m, 2 H, CH_2Ph), 4.56-4.59 (m, 1 H, CH_2Ph), 4.73 (d, 1 H, J = 10 Hz, H-1), 4.95-4.97 (m, 1 H, CH₂Ph), 5.03-5.05 (m, 1 H, CH₂CHCH₂O), 5.13-5.17 (m, 1 H, CH₂CHCH₂O), 5.59 (t, 1 H, J = 10 Hz, H-2), 5.67-5.72 (m, 1 H, CH₂CHCH₂O), 6.97-6.99 (m, 2 H), 7.24-7.34 (m, 12 H), 7.43-7.46 (m, 2 H), 7.54-7.58 (m, 1 H), 8.05-8.07 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.2, 68.8, 70.6, 71.3, 73.1, 73.6, 74.3, 77.7, 81.4, 87.2, 117.2, 127.4, 127.9, 128.0, 128.1, 128.3, 128.4, 129.4, 129.7, 130.2, 132.6, 132.9, 134.3, 137.5, 137.9, 138.5, 165.2. HRMS: C₃₇H₃₈O₆S [M+NH₄]⁺ calcd: 628.2733, obsd: 628.2721. Compound S8 (4.33 g, 7.11 mmol) wais dissolved in THF (50 mL), followed by addition of 222 mg [Ir(COD)(Ph₂MeP)₂]PF₆. The resulting mixture was stirred under H₂ for 3 h, followed by addition of H_2O (60 mL) and I_2 (3.5 g). After the reaction was complete, the mixture was diluted with EtOAc and extracted with H₂O. The combined organic phase was dried over Na₂SO₄. After concentration, the residue was purified by silica gel column (4:1, hexane-EtOAc) to afford p-tolyl 2-O-benzoyl-4, 6-di-O-benzyl-1-thio- β -D-galactopyranoside S9 (3 g, 74%). ¹H-NMR (500 MHz, CDCl₃): δ 2.29 (s, 3 H, SPhCH₃), 2.47 (d, 1 H, J = 9.5Hz, OH), 3.68-3.81 (m, 4 H, H-3, H-5, H-6a, H-6b), 3.95-3.96 (m, 1 H, H-4), 4.46-4.53 (m, 2 H, CH_2Ph), 4.68-4.73 (m, 3 H, CH_2Ph , H-1, J = 9.5 Hz), 5.20 (t, 1 H, J = 9.5 Hz, H-2), 7.00-7.02 (m, 2 H), 7.26-7.34 (m, 12 H), 7.42-7.45 (m, 2 H), 7.54-7.58 (m, 1 H), 8.03-8.05 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.4, 68.6, 72.5, 73.8, 74.6, 75.5, 77.7, 86.5, 127.8, 128.0, 128.1, 128.6, 128.7, 128.7, 129.1, 129.8, 130.0, 130.2, 133.3, 133.4, 137.9, 138.2, 138.4, 165.5. HRMS: $C_{34}H_{34}O_6S[M+NH_4]^+$ calcd: 588.2420, obsd: 588.2412.



Synthesis of galactose building block **S13**. Reagents and conditions: (a) *p*-methoxybenzylidene dimethyl acetal, CSA, CH₃CN; (b) DCC, LevOH, DMAP, DCM; (c) BzCl, DMAP, DCM, 50 °C; (d) NH₂NH₂, HOAc, DCM/MeOH; (e) [Ir(COD)(PMePh₂)₂PF₆], H₂, THF, then H₂O, I₂, 0 °C-r.t..

p-Tolvl 2-O-benzovl-4,6-di-O-benzvlidene -1-thio-β-D-galactopyranoside (**S13**).^[2] *p*-Tolyl 1-thio- β -D-galactopyranoside S3 (5 g, 17.48 mmol) was dissolved in CH₃CN (100 mL) followed by addition of camphorsulfonic acid (1.21 g, 5.24 mmol) and benzaldehyde dimethyl acetal (3.95 mL, 26.22 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was quenched by Et₃N and diluted with EtOAc (100 mL) and the organic phase was extracted by sat. NaHCO₃ solution and dried over Na₂SO₄. After concentration, the residue was recrystalized from EtOH to afford *p*-tolyl 4, 6-*O*-benzylidene-1-thio- β -D-galactopyranoside **S10** (5.3 g, 81%). ¹H-NMR (500 MHz, CDCl₃): δ 2.34 (s, 3 H, SPhCH₃), 2.45-2.49 (m, 2 H), 3.53 (br, 1 H), 3.59-3.69 (m, 2 H), 3.99-4.02 (m, 2 H, CH₂Ph), 4.18-4.19 (m, 1 H), 4.35-4.37 (m, 1 H), 4.44 (d, 1 H, J = 9 Hz, H-1), 5.48 (s, 1 H, PhCH), 7.08-7.10 (m, 2 H), 7.33-7.38 (m, 5 H), 7.55-7.57 (m, 2 H). Compound S10 (5.3 g, 14.16 mmol) was dissolved in dry DCM (100 mL), followed by addition of dicyclohexylcarbodiimide (DCC) (4.53 g, 21.24 mmol), DMAP (878 mg, 7.1 mmol) and LevOH (1.98 g, 17 mmol). The resulting mixture was stirred under room temperature for 3 h. The reaction was diluted with DCM and washed with 10% HCl solution and saturated aqueous NaHCO3 solution sequentially. The combined organic phase was dried over Na₂SO₄ and purified by silica gel column (3:1. afford *p*-tolyl 3-O-levulinoyl-4,6-O-benzylidene-1-thio- β -Dhexanes-EtOAc) to galactopyranoside S11 (3.74 g, 56%). ¹H-NMR (500 MHz, CDCl₃): δ 2.04 (s, 3 H, $CH_3COCH_2CH_2O$), 2.32 (s, 3 H, SPh CH_3), 2.47 (d, 1 H, OH), 2.55-2.57 (t, 2 H, J = 6.5Hz, CH₃COCH₂CH₂O), 2.68-2.71 (t, 2 H, J = 6.5 Hz, CH₃COCH₂CH₂O), 3.56 (br, 1 H), 3.85-3.91 (m, 1 H), 3.97-4.00 (m, 1 H), 4.28-4.36 (m, 2 H), 4.52 (d, 1 H, J = 9.5 Hz, H-1), 4.92 (dd, 1 H, J = 3.5 Hz, J = 10 Hz), 5.44 (s, 1 H, PhCH), 7.05-7.07 (m, 3 H), 7.32-7.38 (m, 5 H), 7.55-7.57 (m, 2 H). Compound S11 (3.74 g, 7.93 mmol) was dissolved in dry DCM (100 mL) followed by addition of DMAP (97 mg, 0.793 mmol) and benzoyl chloride (1.38, 11.9 mmol). The resulting mixture was stirred under 50 °C overnight. After cooling down to room temperature, the reaction mixture was diluted with DCM and extracted with 10% HCl solution. The combined organic phase was washed with a saturated aqueous NaHCO₃ solution and dried over Na₂SO₄. After concentration, the residue was purified by silica gel column (5:1:1, hexanes-DCM-EtOAc) to afford *p*-tolyl 2-O-benzoyl-3-O-levulinoyl-4, 6-O-benzylidene-1-thio- β -D-galactopyranoside S12 (3.74 g,

82%). ¹H-NMR (500 MHz, CDCl₃): δ 1.84 (s, 3 H, CH₃COCH₂CH₂O), 2.31 (s, 3 H, SPhCH₃), 2.38-2.55 (m, 4 H, CH₃COCH₂CH₂O), 3.62 (br, 1 H), 4.01-4.04 (m, 1 H), 4.36-4.39 (m, 2 H), 4.79 (d, 1 H, J = 9.5 Hz, H-1), 5.15 (dd, 1 H, J = 3.5 Hz, J = 10 Hz), 5.46 (s, 1 H, PhCH), 5.15 (t, 1 H, J = 9.5 Hz, H-2), 7.03-7.05 (m, 2 H), 7.32-7.46 (m, 9 H), 7.55-7.58 (m, 1 H), 8.00-8.02 (m, 2 H). Compound S12 (3.74 g, 6.5 mmol) was dissolved in DCM/MeOH (1:1, 100 mL), followed by addition of HOAc (30 mL) and NH₂NH₂-H₂O (4 mL). The resulting reaction mixture was stirred under room temperature overnight and quenched by acetone, diluted with DCM. The organic phase was extracted with a saturated aqueous NaHCO₃ solution. After drying over Na₂SO₄ and concentration, the resulting rsidue was purified by silica gel column (4:1, hexanes-EtOAc) to afford p-tolyl 2-Obenzoyl-4, 6-O-benzylidene-1-thio- β -D-galactopyranoside S13 (2.8 g, 90%). Compound S6 (1 g, 1.93 mmol) wais dissolved in THF (30 mL), followed by addition of 60 mg [Ir(COD)(Ph₂MeP)₂]PF₆. The resulting mixture was stirred under H₂ for 3 h, followed by addition of H₂O (15 mL) and I₂ (948 mg). After the reaction was complete, the mixture was diluted with EtOAc and extracted with H₂O. The combined organic phase was dried over Na₂SO₄. After concentration, the residue was purified by silica gel column (4:1, hexanes-EtOAc) to afford *p*-tolyl 2-O-benzoyl-4,6-di-O-benzyl-1-thio-β-Dgalactopyranoside S13 (710 mg, 77%). ¹H-NMR (500 MHz, CDCl₃): δ 2.32 (s, 3 H, SPhCH₃), 2.54-2.56 (m, 1 H, OH), 3.59 (br, 1 H, H-5), 3.84-3.89 (m, 1 H, H-3), 4.03-4.06 (m, 1 H, H-6a), 4.23-4.25 (m, 1 H, H-4), 4.39-4.42 (m, 1 H, H-6b), 4.75 (d, 1 H, J=10 Hz, H-1), 5.17 (t, 1 H, J = 9.5 Hz, H-2), 5.51 (s, 1 H, PhCH), 7.05-7.07 (m, 2 H), 7.35-7.46 (m. 9 H), 7.55-7.59 (m, 1 H), 8.05-8.07 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.3, 69.1, 69.9, 70.7, 73.0, 75.7, 84.8, 101.5, 126.6, 126.9, 128.1, 128.3, 129.3, 129.5, 129.9, 133.1, 134.6, 137.4, 138.4, 165.5.

$$\begin{array}{c} HO \\ HO \\ HO \\ OH \\ S14 \\ HO \\ HO \\ HO \\ HO \\ HO \\ S17 \\ S18 \\ S18 \\ S19 \\ S10 \\ S1$$

Synthesis of xylose building block **S19**. Reagents and conditions: (a) Ac₂O, pyridine; (b) BF₃/Et₂O, *p*-TolSH, r.t.; (c) NaOMe, DCM/MeOH, r.t.; (d) Bu₂SnO, dioxane, then ClAcCl, DCM; (e) BzCl, DMAP, DCM, r.t..

p-Tolyl 2,3-di-O-benzoyl-4-chloroacetyl-1-thio-\beta-D-xylopyranose (**S19**). A solution of Dxylose **S14** (10 g, 66.67 mmol) in dry pyridine (50 mL) was added Ac₂O (44 mL, 457.6 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was diluted by DCM and washed with 10% HCl solution. The combined organic phase was further extracted with a saturate acquous solution of NaHCO₃ and dried over Na₂SO₄. After concentration, the crude product **S15** (19.7 g, 93%) was directly used for next step without further purification. D-Xylopyranosyl tetraacetate **S15** (19.7 g, 62 mol), *p*-toluenethiol (8.75 g, 70.4 mmol) were dissolved in DCM (100 mL) and boron trifluoride etherate (24.5 mL) was added dropwise at room temperature. The mixture was stirred under N₂ at room temperature for 20 hours and then diluted with DCM (200 mL). The organic phase was washed with saturated aqueous solution of NaHCO₃

until the pH is 7 and then dried over Na_2SO_4 , filtered and concentrated to afford crude product S16 (14.7 g, 62%), which was recrystalized from hexanes/EtOAc. ¹H-NMR (500 MHz, CDCl₃): δ 2.04 (s, 3 H, COCH₃), 2.05 (s, 3 H, COCH₃), 2.11 (s, 3 H, COCH₃), 2.36 (s, 3 H, SPhCH₃), 3.40 (dd, 1 H, J = 9 Hz, J = 11.5 Hz, H-5a), 4.27 (dd, 1 H, J = 5 Hz, J = 7 Hz, H-5b), 4.73 (d, 1 H, J = 8.5 Hz, H-1), 4.90-4.95 (m, 2 H, H-2, H-4), 5.18 (t, 1 H, J = 8.5 Hz, H-3), 7.13-7.15 (m, 2 H), 7.37-7.39 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 20.6, 20.6, 20.7, 21.1, 65.3, 68.4, 69.8, 72.2, 86.4, 128.1, 129.7, 133.4, 138.5, 169.2, 169.7, 169.9. HRMS: $C_{18}H_{22}O_7S [M+NH_4]^+$ calcd: 400.1430, obsd: 400.1442. Compound **S16** (14.7 g, 38.44 mmol) was dissolved in MeOH (50 mL) and DCM (50 mL). Freshly prepared NaOMe solution in MeOH was added to maintain pH above 12 and the mixture was stirred at room temperature overnight. The mixture was neutralized by conc. HCl until the pH is around 7 and then concentrated and dried under vacuum to afford p-tolyl 1thio- β -D-xylopyranose S17 as white solid (8.86 g, 90%) which was directly used without further purification. ¹H-NMR (500 MHz, CD₃OD): δ2.38 (s, 3 H, SPhCH₃), 3.15-3.22 (m, 2 H, H-2, H-5a), 3.32-3.35 (m, 1 H, H-5b), 3.42-3.47 (m, 1 H, H-4), 3.91-3.94 (dd, 1 H, J = 5 Hz, J = 11.5 Hz, H-3), 4.46 (d, 1 H, J = 9 Hz, H-1), 7.12-7.14 (m, 2 H), 7.40-7.42 (m, 2 H)2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 20.8, 70.4, 70.9, 73.6, 79.2, 90.3, 130.5, 130.8, 133.9, 139.0. HRMS: C₁₂H₁₆O₄S [M+H]⁺ calcd: 257.0848, obsd: 257.0857. Compound S17 (1 g, 3.91 mmol) was dissolved dixoane, followed by addition of Bu₂SnO (1.4 g, 5.62 mmol). The resulting mixture was boiled under reflux for 2 h and then was evaporated to dryness. The resulting residue was dissolved in 40 mL DCM, and a solution of chloroacetyl chloride (ClAcCl) (336 µL, 4.15 mmol) in DCM (4 mL) was added while stirring. After 80 minutes, the reaction was diluted with DCM and washed with sat. NaHCO₃ and dried over Na₂SO₄. After, concentration, the residue was purified by silica gel column (1:1, hexane-EtOAc) to afford p-tolyl 4-chloroacetyl-1-thio- β -D-xylopyranose **S18** (882 mg, 68%). ¹H-NMR (500 MHz, CDCl₃): δ 2.33 (s, 3 H, SPhCH₃), 2.74-2.94 (m, 2 H, OH), 3.27-3.35 (m, 2 H, H-2, H-5a), 3.72 (dd, 1 H, J = 9 Hz, H-3), 4.03-4.16 (m, H-5b, ClCH₂CO), 4.41 (d, 1 H, J = 9 Hz, H-1), 4.82-4.87 (m, 1 H, H-4), 7.11-7.13 (m, 2 H), 7.39-7.41 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 40.6, 66.1, 71.9, 72.3, 74.9, 88.5, 126.9, 129.9, 133.7, 138.9, 166.8. HRMS: $C_{14}H_{17}ClO_5S [M+Na]^+$ calcd: 355.0383, obsd: 355.0309. Compound S18 (882 mg, 2.66 mmol) was dissolved in DCM (20 mL), followed by addition of DMAP (32 mg, 0.266 mmol) and benzovl chloride (464 µL, 3.99 mmol). The resulting mixture was stirred under room temperature overnight and diluted with DCM. After extraction with 10% HCl solution, the combined organic phase was further washed with a saturated acquous solution of $NaHCO_3$ and dried over Na_2SO_4 . Silica gel column (4:1, hexanes/EtOAc) purification afforded p-Tolyl 2, 3-di-O-benzoyl-4chloroacetyl-1-thio- β -D-xylopyranose **S19** (72 mg, 5%). ¹H-NMR (500 MHz, CDCl₃): δ 2.36 (s. 3 H. SPhCH₃), 3.69-3.73 (m. 2 H. H-5a, H-5b), 3.97-4.05 (m. 2 H. ClCH₂CO), 4.12-4.16 (m, 1 H, H-4), 5.10 (d, 1 H, J = 6.5 Hz, H-1), 5.39 (t, 1 H, J = 7 Hz, H-2), 5.60 (t, 1 H, J = 7 Hz, H-3), 7.13-7.15 (m, 2 H), 7.40-7.44 (m, 6 H), 7.55-7.58 (m, 3 H), 8.01-8.03 (m, 3 H).



Synthesis of xylose building block **S24**. Reagents and conditions: (a) 2-methoxypropene, CSA, DMF, 60 °C; (b) NaH, PMBCl, DMF; (c) CSA, DCM/MeOH; (d) BzCl, DMAP, DCM, reflux; (e) DDQ, DCM/H₂O; (f) NaH, BnBr, DMF; (g) CSA, DCM/MeOH; (h) BzCl, DMAP, DCM, reflux; (i) DDQ, DCM/H₂O, reflux; (j) LevOH, EDC-HCl, DMAP, DCM; (k) CSA, DCM/MeOH; (l) BzCl, DMAP, DCM, reflux; (m) NH₂NH₂, HOAc, DCM/MeOH.

p-Tolyl 2, 3-di-O-benzoyl-4-p-methoxybenzyl-1-thio- β -D-xylopyranose (S23). A solution of p-tolyl 1-thio-*β*-D-xylopyranose S17 (7 g, 27.34 mmol) in dry DMF (50 mL) was added camphorsulfonic acid (953 mg, 4.1 mmol). The resulting mixture was stirred under 60 °C. 2-Methoxy propene (7.85 mL, 82.02 mmol) was added into the reaction mixture in portions. The reaction was stirred for another 2 h. After the reaction was complete, it was cooled back to room temperature and quenched with Et₃N. The resulting mixture was concentrated and purified by silica gel column (4:1:1, hexanes-DCM-EtOAc) to afford ptolyl 2, 3-isopropylidene-1-thio- β -D-xylopyranose S20 (6.3 g, 78%). ¹H-NMR (500 MHz, CDCl₃): δ 1.41 (s, 3 H, C(CH₃)₂), 1.46 (s, 3 H, C(CH₃)₂), 2.31 (s, 3 H, SPhCH₃), 2.53 (d, 1 H, J = 4 Hz, OH), 3.16-3.20 (m, 2 H, H-2, H-5a), 3.48 (t, 1 H, J = 9 Hz, H-3), 3.89-4.62 (m, 1 H, H-4), 4.06-4.10 (m, 1 H, H-5b), 4.69 (d, 1 H, J = 9.5 Hz, H-1), 7.09-7.11 (m, 2 H),7.42-7.44 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 26.4, 26.6, 68.9, 69.8, 75.0, 82.8, 85.5, 111.1, 127.7, 129.6, 133.6, 138.4. HRMS: $C_{15}H_{20}O_4S [M+H]^+$ calcd: 297.1161, obsd: 297.1160. Compound **S20** (6.3 g, 21.33 mmol) was dissolved in 40 mL DMF, followed by addition of NaH (1 g, 25.6 mmol) and PMBCl (3.76 mL, 27.73 mmol). After stirring under room temperature overnight, the reaction was quenched by 10% HCl solution and diluted with DCM. The organic phase was extracted with sat. NaHCO₃ and dried over Na₂SO₄. Silica gel column (4:1, hexane/EtOAc) purification afforded p-Tolyl 2, 3-isopropylidene-4-p-methoxybenzyl-1-thio- β -D-xylopyranose S21 (7.54 g, 85%). ¹H-NMR (500 MHz, CDCl₃): δ 1.43 (s, 3 H, C(CH₃)₂), 1.47 (s, 3 H, C(CH₃)₂), 2.31 (s, 3 H, SPhCH₃), 3.16-3.21 (m, 2 H, H-2, H-5a), 3.59 (t, 1 H, J = 9 Hz, H-3), 3.67-3.72 (m, 1 H, H-4), 3.77 (s, 3 H, CH₃OPhCH₂O), 4.01-4.04 (m, 1 H, H-5b), 4.47-4.49 (m, 1 H, CH₃OPhCH₂O), 4.69 (d, 1 H, J = 9.5 Hz, H-1), 4.69-4.71 (m, 1 H, CH₃OPhCH₂O), 6.83-6.86 (m, 2 H), 7.08-7.10 (m, 2 H), 7.22-7.24 (m, 2 H), 7.42-7.44 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 26.5, 26.7, 55.2, 68.4, 71.8, 75.2, 75.3, 82.4, 85.4, 111.0, 113.8, 127.9, 129.4, 129.5, 130.0, 133.5, 138.3, 159.3. HRMS: $C_{23}H_{28}O_5S$ [M+H]⁺ calcd: 417.1736, obsd: 417.1725.

Compound S21 (7.54 g, 18.1 mmol) was dissolved in DCM/MeOH (1:1, 60 mL), followed by addition of camphorsulfonic acid (4.23 g, 18.1 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was quenched by Et_3N and concentrated. The residue was purified by silica gel column (1:1, hexanes-EtOAc) to afford p-tolyl 4-p-methoxybenzyl-1-thio- β -D-xylopyranose S22 (6.68) g, 98%). ¹H-NMR (500 MHz, CDCl₃): δ2.32 (s, 3 H, SPhCH₃), 2.64 (br, 2 H, OH), 3.19-3.24 (m, 1 H, H-5a), 3.05-3.35 (m, 1 H, H-2), 3.39-3.44 (m, 1 H, H-4), 3.60-3.65 (m, 1 H, H-3), 3.78 (s, 3 H, CH_3OPhCH_2O), 4.01-4.04 (m, 1 H, H-5b), 4.44 (d, 1 H, J = 9 Hz, H-1), 4.53-4.59 (m, 2 H, CH₃OPhCH₂O), 6.83-6.86 (m, 2 H), 7.09-7.11 (m, 2 H), 7.21-7.23 (m, 2 H), 7.38-7.40 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 55.2, 67.1, 71.8, 72.6, 88.8, 113.9, 127.9, 129.4, 129.7, 129.9, 133.2, 138.4, 159.4. HRMS: $C_{20}H_{24}O_5S[M+NH_4]^+$ calcd: 394.1688, obsd: 394.1669. Compound S22 (6.68 g, 17.77 mmol) and DMAP (2.17 g, 17.77 mmol) were dissolved in DCM (100 mL). Benzoyl chloride (4.95 mL, 42.65 mmol) was added into the reaction mixture while stirring and the reaction was left under reflux overnight. After the reaction was complete, it was diluted with DCM and washed with 10% HCl solution. The combined organic phase was extracted with a saturate acquous solution of NaHCO₃ and dried over Na₂SO₄. Silica gel column (2:1, hexanes/EtOAc) 2,3-di-O-benzoyl-4-p-methoxybenzyl-1-thio-β-Dpurification afforded *p*-tolvl xylopyranose **S23** (10.2 g, 98%). ¹H-NMR (500 MHz, CDCl₃): δ 2.30 (s, 3 H, SPhCH₃). 3.48-3.52 (m, 1 H, H-5a), 3.70-3.74 (m, 1 H, H-4), 3.73 (s, 3 H, CH₃OPhCH₂O), 4.21-4.25 (m, 1 H, H-5b), 4.47-4.52 (m, 2 H, CH₃OPhCH₂O), 4.91 (d, 1 H, J = 8 Hz, H-1), 5.28 (t, 1 H, J = 8 Hz, H-2), 5.55 (t, 1 H, J = 8 Hz, H-3), 6.68-6.71 (m, 2 H), 7.06-7.11 (m, 4 H), 7.32-7.39 (m, 6 H), 7.47-7.52 (m, 2 H), 7.92-7.96 (m, 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 55.1, 66.4, 70.5, 72.3, 73.7, 73.9, 87.1, 113.8, 128.3, 128.3, 129.1, 129.4, 129.4, 129.6, 129.7, 129.8, 129.9, 133.0, 133.1, 138.2, 159.3, 165.2, 165.5. HRMS: C₃₄H₃₂O₇S $[M+NH_4]^+$ calcd: 602.2212, obsd: 602.2203.

p-Tolyl 2,3-*di-O-benzoyl-4-O-benzyl-1-thio-\beta-D-xylopyranose* (S27). *p*-Tolyl 2, 3isopropylidene-1-thio- β -D-xylopyranose S20 (600 mg, 2.03 mmol) was dissolved in 10 mL DMF, followed by addition of NaH (95 mg, 2.44 mmol) and BnBr (314 µL, 2.64 mmol). After stirring under room temperature overnight, the reaction was quenched by 10% HCl solution and diluted with DCM. The organic phase was extracted with a saturate aqueous solution of NaHCO₃ and dried over Na_2SO_4 . Silica gel column (4:1, hexanes/EtOAc) purification afforded p-tolyl 2,3-isopropylidene-4-benzyl-1-thio- β -Dxylopyranose **S25** (703 mg, 90%). ¹H-NMR (500 MHz, CDCl₃): δ 1.48 (s, 3 H, C(CH₃)₂), 1.53 (s, 3 H, C(CH₃)₂), 2.34 (s, 3 H, SPhCH₃), 3.22-3.29 (m, 2 H, H-2, H-4), 3.67 (t, 1 H, J = 9 Hz, H-3), 3.72-3.78 (m, 1 H, H-5a), 4.09-4.12 (m, 1 H, H-5b), 4.57-4.60 (m, 1 H, PhCH₂O), 4.75 (d, 1 H, J = 9.5 Hz, H-1), 4.80-4.83 (m, 1 H, PhCH₂O), 7.11-7.13 (m, 2 H), 7.26-7.35 (m, 5 H), 7.49-7.51 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 20.8, 26.3, 26.5, 68.0, 71.7, 75.0, 75.3, 82.2, 85.0, 113.2, 127.4, 127.4, 127.6, 129.3, 133.3, 137.7, 138.0. HRMS: C₂₂H₂₆O₄S [M+Na]⁺ calcd: 410.1528, obsd: 410.1547. Compound S25 (703 mg, 1.82 mmol) was dissolve in DCM/MeOH (1:1, 10 mL), followed by addition of camphorsulfonic acid (423 mg, 1.82 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was quenched by Et₃N and concentrated. The residue was purified by silica gel column (1:1, hexanes-EtOAc) to afford *p*-tolyl 4-benzyl-1-thio- β -D-xylopyranose S26 (581 mg, 92%). ¹H-NMR (500 MHz,

 $CDCl_3$: δ 2.34 (s, 3 H, SPh CH_3), 3.22-3.27 (m, 1 H, H-5a), 3.42-3.53 (m, 2 H, H-2, H-4), 3.71-3.75 (m, 1 H, H-3), 3.97-3.99 (m, 1 H, OH), 4.03-4.06 (m, 1 H, H-5b), 4.18 (br, 1 H, OH), 4.54 (d, 1 H, J = 9 Hz, H-1), 4.61-4.76 (m, 2 H, PhCH₂), 7.11-7.13 (m, 2 H), 7.29-7.36 (m, 5 H), 7.46-7.48 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.4, 67.1, 72.1, 73.0, 77.4, 88.8, 127.8, 127.9, 128.4, 128.6, 129.7, 133.0, 138.0. HRMS: C₁₉H₂₂O₄S [M+NH₄]⁺ calcd: 364.1583, obsd: 364.1582. Compound S26 (581 mg, 1.68 mmol) and DMAP (205 mg, 1.68 mmol) were dissolved in DCM (10 mL). Benzoyl chloride (468 µL, 4.03 mmol) was added into the reaction mixture while stirring and the reaction was left under reflux overnight. After the reaction was complete, it was diluted with DCM and washed with 10% HCl solution. The combined organic phase was extracted with a saturated aqueous solution of NaHCO₃ and dried over Na₂SO₄. Silica gel column (2:1, hexanes/EtOAc) afforded 2,3-di-O-benzoyl-4-p-methoxybenzyl-1-thio-B-Dpurification *p*-tolvl xylopyranose S27 (881 mg, 95%). ¹H-NMR (500 MHz, CDCl₃): δ 2.31 (s, 3 H, SPhCH₃), 3.53-3.57 (m, 1 H, H-5a), 3.74-3.78 (m, 1 H, H-4), 4.27-4.31 (m, 1 H, H-5b), 4.56-4.60 (m, 2 H, PhC H_2), 4.95 (d, 1 H, J = 9 Hz, H-1), 5.28-5.32 (m, 1 H, H-2), 5.58-5.62 (m, 1 H, H-3), 7.07-7.09 (m, 2 H), 7.19-7.24 (m, 5 H), 7.32-7.40 (m, 6 H), 7.47-7.53 (m, 2 H), 7.93-7.98 (m, 4 H). HRMS: $C_{33}H_{30}O_6S [M+NH_4]^+$ calcd: 572.2107, obsd: 572.2096.

p-Tolyl 2, 3-di-O-benzoyl-4-O-levulinoyl-1-thio-β-D-xylopyranose (S30). *p*-Tolyl 2, 3isopropylidene-1-thio- β -D-xylopyranose S20 (500 mg, 1.69 mmol) was dissolved in 10 mL DMF, followed by addition of LevOH (205 µL, 2.03 mmol), DMAP (206 mg, 1.69 mmol) and EDC-HCl (389 mg, 2.03 mmol). After stirring under room temperature overnight, the reaction was quenched by 10% HCl solution and diluted with DCM. The organic phase was extracted with sat. NaHCO₃ solution and dried over Na₂SO₄. Silica gel column (3:1, hexanes/EtOAc) purification afforded p-tolyl 2, 3-isopropylidene-4levulinoyl-1-thio- β -D-xylopyranose **S28** (586 mg, 88%). ¹H-NMR (500 MHz, CDCl₃): δ 1.40 (s, 3 H, C(CH₃)₂), 1.46 (s, 3 H, C(CH₃)₂), 2.15 (s, 3 H, CH₃COCH₂CH₂CO), 2.32 (s, 3 H, SPhCH₃), 2.51-2.81 (m, 4 H, CH₃COCH₂CH₂CO), 3.21-3.28 (m, 2 H, H-2, H-5a), 3.69-3.73 (m, 1 H, H-3), 4.20-4.24 (m, 1 H, H-5b), 4.76 (d, 1 H, J = 9.5 Hz, H-1), 4.91-4.96 (m, 1 H, H-4), 7.09-7.11 (m, 2 H), 7.42-7.44 (m, 2 H), HRMS: C₂₀H₂₆O₆S [M+NH₄]⁺ calcd: 412.2522, obsd: 412.2529. Compound S28 (586 mg, 1.49 mmol) was dissolved in DCM/MeOH (1:1, 8 mL), followed by addition of camphorsulfonic acid (346 mg, 1.49 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was quenched by Et₃N and concentrated. The residue was purified by silica gel column (1:1, hexane-EtOAc) to afford p-tolyl 4-benzyl-1-thio- β -Dxylopyranose **S29** (475 mg, 90%). ¹H-NMR (500 MHz, CDCl₃): δ 2.16 (s, 3 H, CH₃COCH₂CH₂CO), 2.31 (s, 3 H, SPhCH₃), 2.53-2.57 (m, 3 H, OH, CH₃COCH₂CH₂CO), 2.75-2.78 (m, 2 H, CH₃COCH₂CH₂CO), 2.87 (br, 1 H, OH), 3.26-3.37 (m, 2 H, H-2, H-5a), 3.68-3.72 (m, 1 H, H-3), 4.08-4.12 (m, 1 H, H-5b), 4.42 (d, 1 H, J = 9.5 Hz, H-1), 4.77-1004.82 (m, 1 H, H-4), 7.10-7.13 (m, 2 H), 7.40-7.42 (m, 2 H). HRMS: C₁₇H₂₂O₆S [M+NH₄]⁺ calcd: 372.1481, obsd: 372.1489. Compound S29 (475 mg, 1.34 mmol) and DMAP (163 mg, 1.34 mmol) were dissolved in DCM (8 mL). Benzoyl chloride (373 µL, 3.21 mmol) was added into the reaction mixture while stirring and the reaction was left under reflux overnight. After the reaction was complete, it was diluted with DCM and washed with 10% HCl solution. The combined organic phase was extracted with a saturated aqueous solution of NaHCO₃ and dried over Na₂SO₄. Silica gel column (3:1, hexanes/EtOAc)

purification afforded *p*-tolyl 2,3-di-*O*-benzoyl-4-*O*-levulinoyl-1-thio- β -D-xylopyranose **S30** (550 mg, 73%). ¹H-NMR (500 MHz, CDCl₃): δ 2.04 (s, 3 H, CH₃COCH₂CH₂CO), 2.31 (s, 3 H, SPhCH₃), 2.41-2.67 (m, 4 H, CH₃COCH₂CH₂CO), 3.59-3.63 (m, 1 H, H-5a), 4.43-4.46 (m, 1 H, H-5b), 5.03 (d, 1 H, *J* = 6 Hz, H-1), 5.08-5.11 (m, 1 H, H-4), 5.34 (t, 1 H, *J* = 6.5 Hz, H-2), 5.57 (t, 1 H, *J* = 6.5 Hz, H-3), 7.08-7.10 (m, 2 H), 7.36-7.39 (m, 6 H), 7.49-7.53 (m, 2 H), 7.97-7.99 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 14.1, 20.9, 21.0, 27.7, 29.5, 37.6, 60.2, 64.3, 68.4, 69.9, 71.5, 86.6, 128.3, 128.3, 128.6, 128.8, 129.2, 129.6, 129.7, 129.8, 133.2, 133.2, 133.3, 138.3, 164.9, 165.2, 171.5, 205.8. HRMS: C₃₁H₃₀O₈S [M+NH₄]⁺ calcd: 580.2005, obsd: 580.2008.

p-Tolyl 2, 3-di-O-benzoyl-1-thio-β-D-xylopyranose (S24). p-Tolyl 2, 3-di-O-benzoyl-4-O*p*-methoxybenzyl-1-thio- β -D-xylopyranose **S23** (10.2 g, 17.41 mmol) was dissolved in DCM/H₂O (10:1, 50 mL), followed by addition of DDQ (5.9 g, 26.11 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was diluted with DCM, washed with a saturate aqueous solution of NaHCO₃, dried over Na₂SO₄. Silica gel column (4:1, hexanes/EtOAc) purification afforded *p*-tolyl 2, 3-di-O-benzoyl-1-thio-β-D-xylopyranose S24 (6.54 g, 81%). p-Tolyl 2,3-di-O-benzoyl-4-*O*-benzyl-1-thio- β -D-xylopyranose **S27** (881 mg, 1.59 mmol) was dissolved in DCM/H₂O (10:1, 10 mL), followed by addition of DDQ (543 mg, 2.39 mmol). The resulting mixture was stirred under reflux overnight. After the reaction was complete, it was diluted with DCM, washed with a saturated aqueous solution of NaHCO₃, dried over Na₂SO₄. Silica gel column (4:1, hexanes/EtOAc) purification afforded p-tolyl 2,3-di-O-benzoyl-1-thio-B-Dxylopyranose **S24** (444 mg, 60%). *p*-Tolyl 2, 3-di-O-benzoyl-4-O-benzyl-1-thio-β-Dxylopyranose **S30** (550 mg, 0.98 mmol) was dissolved in DCM/MeOH (1:1, 8 mL), followed by addition of HOAc (6 mL) and NH₂NH₂-H₂O (570 µL). The resulting mixture was stirred under reflux overnight. After the reaction was complete, it was diluted with DCM, washed with a saturated aqueous solution of NaHCO₃, dried over Na₂SO₄. Silica gel column (4:1, hexanes/EtOAc) purification afforded p-tolyl 2,3-di-O-benzoyl-1-thio- β -Dxylopyranose **S24** (396 mg, 87%). ¹H-NMR (500 MHz, CDCl₃): δ 2.32 (s, 3 H, SPhCH₃), 3.02 (br, 1 H, OH), 3.53-3.58 (m, 1 H, H-5a), 3.94-3.98 (m, 1 H, H-4), 4.38-4.41 (m, 1 H, H-5b), 4.98 (d, 1 H, J = 7 Hz, H-1), 5.29 (t, 1 H, J = 7.5 Hz, H-3), 5.38 (t, 1 H, J = 7.5 Hz, H-2), 7.09-7.11 (m, 2 H), 7.36-7.40 (m, 6 H), 7.51-7.54 (m, 2 H), 7.96-8.02 (m, 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.2, 67.6, 68.2, 70.1, 76.0, 86.8, 128.4, 128.4, 128.7, 128.8, 129.2, 129.7, 129.7, 129.9, 133.2, 133.3, 133.5, 138.3, 165.0, 166.8. HRMS: C₂₆H₂₄O₆S $[M+NH_4]^+$ calcd: 482.1637, obsd: 482.1657.



Screening donor-acceptor pairs for Xylose-Serine synthesis. Reagents and conditions: (a) KHCO₃, BnBr, Bu₄NI, DMSO; (b) AgOTf, *p*-TolSCl, DCM, -78 °C, then **19**, TTBP, -78 °C-r.t.; (c) DDQ, DCM/H₂O, reflux; (d) DDQ, DCM/H₂O; (e) TBSOTf, 2, 6-lutidine, -40 °C-0 °C; (f) *p*-TsOH, THF/H₂O, 70% or HF/Pyridine, 72% or Tf₂O, THF/H₂O, 80%.

Fmoc-Ser-OBn (19).^[3] Dry DMSO (6 mL) was added to a 25 mL flask under nitrogen containing compound **S35** (0.962 g, 2.94 mmol), KHCO₃ (0.442 g, 4.40 mmol), and tetrabutylammonium iodide (0.1086 g, 0.294 mmol). The resulting white suspension was stirred under room temperature until a homogeneous solution was obtained. BnBr (1.048 mL, 8.80 mmol) was added to the reaction mixture and the reaction was kept for 8 h under room temperature. After the reaction was complete indicated by TLC, it was quenched by water and the mixture was extracted by EtOAc. The combined organic layer was washed by a saturated aqueous solution of NaHCO₃, saturate solution of Na₂S₂O₃ and brine sequentially. After drying over Na₂SO₄, the solution was concentrated to give yellow oil. The oil was cooled to -78 °C and triturated with hexanes. The obtained yellow solid was filtered and washed with hexanes under suction until a white solid was obtained (1 g, 82%). ¹H-NMR (500 MHz, CDCl₃): δ 1.95 (br, 1 H), 3.91-4.03 (m, 2 H), 4.20 (t, 1 H, *J* = 7 Hz), 4.39-4.49 (m, 3 H), 5.17-5.25 (m, 2 H), 5.64-5.68 (m, 1 H), 7.26-7.39 (m, 9 H), 7.57-7.59 (m, 2 H), 7.74-7.76 (m, 2 H).

N-Fluorenylmethyloxycarbonyl-O-(2, 3-di-O-benzoyl-4-O-benzyl -β-D-xylopyranosyl)-L-serine benzyl ester (**S31**). Compound **S31** was synthesized from donor **S13** and acceptor **19** in 83% yield following the general procedure of single step glycosylation. ¹H-NMR (500 MHz, CDCl₃): δ 3.37-3.42 (m, 1 H, H-5a), 3.64-3.70 (m, 1 H, H-4), 3.79-3.82 (m, 1 H, OCH₂CH), 3.89-3.92 (m, 1 H, H-5b), 4.11-4.14 (m, 1 H), 4.19-4.23 (m, 1 H), 4.28-4.35 (m, 2 H), 4.50-4.55 (m, 1 H, OCH₂CH), 4.59 (br, 2 H, PhCH₂), 4.64 (d, 1 H, *J* = 5.5 Hz, H-1), 5.08-5.22 (m, 3 H, H-2, PhCH₂), 5.55-5.59 (m, 2 H), 7.18-7.29 (m, 14 H), 7.35-7.42 (m, 5 H), 7.51-7.55 (m, 3 H), 7.74-7.76 (m, 2 H), 7.92-7.97 (m, 4 H). ¹³C-NMR (150 MHz, CDCl₃): δ 47.3, 54.5, 60.6, 62.6, 67.4, 67.6, 69.2, 71.0, 72.1, 72.7, 74.0, 76.1, 101.2, 120.1, 125.4, 127.3, 127.9, 128.0, 128.1, 128.5, 128.5, 128.6, 128.6, 128.7, 129.4, 129.6, 130.0, 130.1, 133.4, 133.5, 135.4, 137.6, 141.4, 143.9, 144.1, 156.1, 165.5, 165.7, 169.7. HRMS: C₅₁H₄₅NO₁₁ [M+NH₄]⁺ calcd: 865.3336, obsd: 865.3331.

N-Fluorenylmethyloxycarbonyl-O-(2,3-di-O-benzoyl-4-O-p-methoxybenzyl-\beta-D-

xylopyranosyl)-L-serine benzyl ester (**S33**). Compound **S33** was synthesized from donor **S32** and acceptor **19** in 76% yield following the general procedure of single step glycosylation. ¹H-NMR (500 MHz, CDCl₃): δ 3.32-3.28 (m, 1 H), 3.62-3.67 (m, 1 H), 3.73 (s, 1 H, CH₃OPh), 3.79-3.82 (m, 1 H), 3.86-3.91 (m, 1 H), 4.10-4.14 (m, 1 H), 4.18-4.22 (m, 1 H), 4.28-4.34 (m, 2 H), 4.49-4.54 (m, 2 H), 4.61 (d, 1 H, *J* = 6 Hz, H-1), 5.08-5.21 (m, 3 H), 5.52-5.59 (m, 2 H), 6.71-6.73 (m, 2 H), 7.11-7.14 (m, 2 H), 7.23-7.28 (m, 8 H), 7.35-7.42 (m, 6 H), 7.50-7.55 (m, 3 H), 7.74-7.76 (m, 2 H), 7.90-7.96 (m, 4 H). HRMS: C₅₂H₄₇NO₁₂ [M+NH₄]⁺ calcd: 895.3442, obsd: 895.3451.

N-Fluorenylmethyloxycarbonyl-O-(2,3-di-O-benzoyl-4-O-tert-butyldimethylsilyl- β -D-

xylopyranosyl)-L-serine benzyl ester (S34). Compound S24 (400 mg, 0.86 mmol) in DCM (5 mL) was cooled down to -40 °C, followed by sequential addition of 2, 6-lutidine (203 μ L, 1.74 mmol) and TBSOTf (299 μ L, 1.3 mmol). The resulting solution was warmed up very slowly to room temperature. The mixture was quenched by Et₃N and then diluted with DCM (100 mL). The organic phase was washed with a saturated aqueous solution of NaHCO₃ and then dried over Na₂SO₄, filtered and the solvents were removed in vacuo. Silica gel column chromatography (Hexanes-EtOAc) afforded p-Tolyl 2, 3-di-O-benzoyl-4-O-tert-butyldimethylsilyl-1-thio- β -D-xylopyranoside **18** as white solid (448 mg, 90%) (HRMS: C₃₂H₃₈O₆SSi [M+NH₄]⁺ calcd: 596.2502, obsd: 596.2480) which was used as donor to couple to acceptor 19 to produce compound S34 in 83% yield following the general procedure of single step glycosylation. ¹H-NMR (600 MHz, CDCl₃): δ -0.12 (s, 3) H, Si(CH₃)₂C(CH₃)₃), 0.02 (s, 3 H, Si(CH₃)₂C(CH₃)₃), 0.76 (s, 9 H, Si(CH₃)₂C(CH₃)₃), 3.31-3.35 (m, 1 H), 3.83-3.96 (m, 3 H), 4.10-4.13 (m, 1 H), 4.18-4.20 (m, 1 H), 4.30-4.34 (m, 2 H), 4.49-4.52 (m, 1 H), 4.57 (d, 1 H, J = 6.5 Hz, H-1), 5.11-5.17 (m, 2 H, PhCH₂), 5.22-5.25 (m, 1 H), 5.46-5.49 (m, 1 H), 5.54-5.56 (m, 1 H), 7.22-7.40 (m, 14 H), 7.47-7.55 (m, 3 H), 7.75-7.77 (m, 2 H), 7.90-7.95 (m, 4 H). ¹³C-NMR (150 MHz, CDCl₃): δ -5.0, -4.7, 17.7, 25.4, 47.0, 54.3, 65.9, 67.1, 67.3, 68.9, 69.1, 71.5, 74.7, 101.6, 119.9, 119.9, 125.1, 127.0, 127.6, 127.7, 128.1, 128.3, 128.3, 128.5, 129.2, 129.5, 129.7, 129.7, 133.0, 133.1, 135.2, 141.2, 141.2, 143.7, 143.8, 155.9, 165.3, 165.5, 169.4. HRMS: $C_{50}H_{53}NO_{11}Si[M+NH_4]^+$ calcd: 889.3732, obsd: 889.3686.

N-Fluorenylmethyloxycarbonyl-O-(2,3-di-O-benzoyl-β-D-xylopyranosyl)-L-serine benzyl ester (5). Compound S31 (300 mg, 0.354 mmol) was dissolved in 6 mL DCM/H₂O (10:1) and cooled down to 0 °C, followed by addition of 804 mg DDQ. The resulting mixture was stirred under reflux overnight. After cooling down to room temperature, the reaction mixture was diluted with DCM and washed with sat. NaHCO₃. The combined organic phase was dried over Na₂SO₄ and concentrated. Silica gel column purification afforded compound 5 (163 mg, 61%). Compound S33 (3 g, 3.42 mmol) was dissolved in 50 mL DCM/H₂O (10:1) and cooled down to 0 °C, followed by addition of 1.55 g DDO. The resulting mixture was stirred under room temperature overnight. After cooling down to room temperature, the reaction mixture was diluted with DCM and washed with sat. NaHCO₃. The combined organic phase was dried over Na₂SO₄ and concentrated. Silica gel column purification afforded compound 5 (2.2 g, 85%). Compound S34 (1 g, 1.15 mmol) was dissolved in 20 mL DCM/H₂O (10:1) and cooled down to 0 °C, followed by dropwise addition of 290 μ L Tf₂O. The resulting mixture was stirred under room temperature for another 2 h. After the reaction was complete, it was diluted with DCM and washed with sat. NaHCO₃. The combined organic phase was dried over Na₂SO₄ and concentrated.

Silica gel column purification afforded compound **5** (696 mg, 80%). ¹H-NMR (500 MHz, CDCl₃): δ 3.05 (d, 1 H, J = 5 Hz, OH), 3.38-3.45 (m, 4 H), 3.83 (dd, 1 H, J = 2.5 Hz, J = 8.5 Hz), 3.88-3.93 (m, 1 H), 4.05-4.14 (m, 2 H), 4.22-4.26 (m, 1 H), 4.30-4.35 (m, 2 H), 4.53-4.55 (m, 1 H), 4.65 (d, 1 H, J = 4.5 Hz, H-1), 5.08-5.18 (m, 2 H, PhCH₂), 5.24-5.32 (m, 2 H), 5.57 (d, 1 H, J = 7 Hz), 7.25-7.46 (m, 14 H), 7.51-7.55 (m, 3 H), 7.73-7.76 (m, 2 H), 7.94-7.99 (m, 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 47.0, 54.2, 54.3, 64.0, 64.1, 67.1, 67.3, 67.4, 67.6, 68.3, 68.4, 69.0, 69.1, 70.2, 70.3, 75.0, 75.1, 100.6, 100.7, 119.9, 120.0, 124.7, 125.1, 125.1, 127.1, 127.7, 128.2, 128.4, 128.5, 128.5, 128.8, 129.0, 129.7, 129.8, 129.9, 130.0, 132.6, 133.4, 133.4, 133.6, 133.7, 135.1, 141.2, 143.7, 143.8, 155.9, 165.1, 165.5, 167.0, 169.5. HRMS: C₄₄H₃₉NO₁₁ [M+NH₄]⁺ calcd: 775.2867, obsd: 775.2867.



Synthesis of trisaccharide **S36**. Reagents and conditions: (a) AgOTf, *p*-TolSCl, DCM, -78 °C, then **5**, TTBP, -78 °C-0 °C; (b) NH₂NH₂-H₂O, HOAc, DCM/MeOH.

N-Fluorenylmethyloxycarbonyl-O-[2-O-benzoyl-3-O-levulinoyl-4, 6-O-benzylidene -β-D-galactopyranosyl-(1→4)-2,3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine benzyl ester (**21**). Compound **21** was synthesized from donor **20** and acceptor **5** in 81% yield following the general procedure of single step glycosylation. ¹H-NMR (500 MHz, CDCl₃): δ 1.87 (s, 3 H, CH₃COCH₂CH₂), 2.41-2.61 (m, 4 H, CH₃COCH₂CH₂), 3.23-3.31 (m, 2 H), 3.71-3.83 (m, 4 H), 3.90-3.95 (m, 1 H), 4.09-4.13 (m, 1 H), 4.20-4.24 (m, 3 H), 4.29-4.33 (m, 1 H), 4.45-4.51 (m, 1 H), 4.55 (d, 1 H, *J* = 6 Hz), 4.76 (d, 1 H, *J* = 8 Hz), 5.00-5.17 (m, 4 H), 5.35 (s, 1 H, PhCH), 5.53-5.62 (m, 3 H), 7.20-7.48 (m, 21 H), 7.51-7.56 (m, 3 H), 7.74-7.76 (m, 2 H), 7.94-7.98 (m, 6 H). ¹³C-NMR (150 MHz, CDCl₃): δ 28.4, 29.6, 37.9, 38.1, 47.3, 54.5, 62.5, 66.9, 67.4, 67.5, 68.5, 69.2, 69.3, 69.6, 71.0, 72.0, 72.1, 73.4, 76.2, 91.5, 100.8, 101.2, 102.3, 120.2, 125.4, 126.5, 126.7, 127.3, 127.3, 127.9, 127.9, 128.2, 128.3, 128.4, 128.5, 128.5, 128.6, 128.6, 128.7, 128.8, 129.1, 129.3, 129.6, 129.7, 129.8, 130.0, 130.1, 130.1, 133.3, 133.6, 135.3, 137.7, 141.4, 143.9, 144.0, 156.1, 165.1, 165.3, 165.7, 169.6, 172.3, 206.3. HRMS: C₆₉H₆₃NO₁₉ [M+NH₄]⁺ calcd: 1227.4338, obsd: 1227.3872. *N-Fluorenylmethyloxycarbonyl-O-[2-O-benzoyl-4,6-O-benzylidene-β-D-galactopyranosyl-*

N-Fluorenylmethyloxycarbonyl-O-[2-O-benzoyl-4,6-O-benzylidene-β-D-galactopyranosyl-(*1*→4)-2, 3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine benzyl ester (**S36**). Compound **S36** was synthesized from compound **21** in 90% yield following the general procedure of Lev deprotection. ¹H-NMR (600 MHz, CDCl₃): δ 2.60 (d, 1 H, J = 9.5 Hz, OH), 3.26-3.34 (m, 2 H), 3.74-3.86 (m, 5 H), 3.93-3.97 (m, 1 H), 4.09-4.13 (m, 2 H), 4.21-4.33 (m, 2 H), 4.48-4.50 (m, 1 H), 4.57 (d, 1 H, J = 5 Hz), 4.71 (d, 1 H, J = 7 Hz), 5.02-5.10 (m, 2 H, COOCH₂Ph), 5.16-5.19 (m, 1 H), 5.27-5.30 (m, 1 H), 5.40 (s, 1 H, PhCH), 5.56 (d, 1 H, J = 7.5 Hz), 5.61 (t, 1 H, J = 6 Hz), 7.21-7.47 (m, 23 H), 7.52-7.54 (m, 3 H), 7.74-7.76 (m, 2 H), 7.95-8.01 (m, 5 H). ¹³C-NMR (150 MHz, CDCl₃): δ 47.0, 54.2, 62.2, 66.8, 67.2, 67.3, 68.3, 69.0, 70.7, 71.6, 71.8, 73.0, 75.3, 75.7, 100.5, 101.4, 101.7, 119.9, 125.1, 126.2, 126.5, 127.0, 127.7, 127.7, 127.9, 128.1, 128.2, 128.3, 128.3, 128.4, 128.4, 128.4, 128.5, 129.1, 129.1, 129.5, 129.6, 129.8, 129.9, 133.1, 133.2, 135.1, 137.2, 141.2, 143.7, 143.8, 155.9, 165.1, 165.5, 165.9, 169.4. HRMS: C₆₄H₅₇NO₁₇ [M+NH₄]⁺ calcd: 1129.3970, obsd: 1129.3920.

p-Tolyl 2,3-*di-O-levulinoyl-4*, 6-*O-benzylidene-β-D-galactopyranosyl-(1→3)-2-O-benzoyl-*4, 6-*O-benzylidene-1-thio-β-D-galactopyranoside* (**24**). Compound **24** was synthesized from donor **22**^[2] and acceptor **23** in 85% yield following the general procedure of single step glycosylation. ¹H-NMR (500 MHz, CDCl₃): δ 1.95 (s, 3 H, CH₃COCH₂CH₂), 2.00 (s, 3 H, CH₃COCH₂CH₂), 2.28 (s, 3 H, SPhCH₃), 2.10-2.69 (m, 8 H, CH₃COCH₂CH₂), 3.35 (br, 1 H), 3.52 (br, 1 H), 3.95-4.03 (m, 2 H), 4.13-4.19 (m, 3 H), 4.33-4.36 (m, 1 H), 4.43 (d, 1 H, J = 3 Hz), 4.65-4.68 (m, 2 H), 4.72 (d, 1 H, *J* = 9.5 Hz), 5.24-5.28 (m, 1 H), 5.42 (s, 1 H, PhCH), 5.48-5.53 (m, 1 H), 5.54 (s, 1 H, PhCH), 6.98-7.00 (m, 2 H), 7.29-7.34 (m, 6 H), 7.41-7.48 (m, 8 H), 7.57-7.61 (m, 1 H), 8.01-8.04 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 27.5, 28.1, 29.5, 29.5, 37.7, 37.7, 66.4, 68.2, 68.7, 69.0, 69.4, 70.3, 71.7, 73.1, 75.9, 86.1, 100.7, 100.8, 100.9, 126.3, 126.5, 127.9, 128.1, 128.2, 128.5, 128.7, 129.0, 129.4, 129.6, 130.0, 133.2, 133.7, 137.4, 137.8, 137.9, 164.7, 171.2, 172.1, 206.5, 206.6.

"3+2+3" assembly of octasaccharide 27



Synthesis of disaccharide **13**. Reagents and conditions: (a) AgOTf, *p*-TolSCl, DCM, -78 °C, then **11**, TTBP, -78 °C-0 °C; (b) Mg(OMe)₂, DCM, -20 °C-0 °C; (c) LevOH, EDC-HCl, DMAP, DCM; (d) DDQ, DCM/H₂O; (e) HF/Pyridine; (f) TBDPSCl, imidazole, DCM.

p-Tolyl 2-azido-3-O-benzyl-4-O-tert-butyl-dimethylsilyl-6-O-acetyl-2-deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl-6-O-p-methoxybenzyl-1-thio- α -L-idopyranoside (12). Compound 12 was synthesized from donor 10 and acceptor 11 in 80% yield following the general procedure of single step glycosylation. The identity of the compound was confirmed by comparison with literature data.^[4]

p-*Tolvl* 2-azido-3-O-benzyl-4-O-tert-butyl-dimethylsilyl-2-deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzovl-3-O-benzvl-6-O-p-methoxvbenzvl-1-thio- α -L-idopvranoside **(S37)**. Compound 12 (2.2 g, 2.13 mmol) was dissolved in 50 mL dry DCM and cooled down to -20 °C. Fresh methanolic Mg(OMe)₂ solution (8%) (21 mL) was added to the reaction mixture. The resulting mixture was left under N₂ and monitored by TLC. After the reaction was complete, it was neutralized by 1 M HOAc to pH 5 and diluted with DCM. After washing with saturated aqueous $NaHCO_3$ solution and drying over Na_2SO_4 , the solution was concentrated and purified by silica gel column to afford compound S37 (1.81 g, 86%). ¹H-NMR (500 MHz, CDCl₃): δ -0.14 (s, 3 H, Si(CH₃)₂), -0.01 (s, 3 H, Si(CH₃)₂), 0.85 (s, 9 H, C(CH₃)₃), 1.70-1.73 (m, 1 H), 2.28 (s, 3 H, SPhCH₃), 3.18 (m, 1 H, J = 3.5 Hz, J = 10Hz), 3.31-3.35 (m, 1 H), 3.44-3.48 (m, 1 H), 3.53-3.66 (m, 4 H), 3.73-3.75 (m, 2 H), 3.79 (s, 3 H, CH₃OPh), 3.99-4.02 (m, 1 H, CH₂Ph), 4.12-4.14 (m, 1 H), 4.24-4.27 (m, 1 H), 4.49 (s, 2 H), 4.58 (d, 1 H, J = 4.0 Hz), 4.70-4.73 (m, 1 H, CH_2 Ph), 4.88-4.91 (m, 1 H), 4.92-4.94 (m, 1 H, CH₂Ph), 5.31-5.33 (m, 1 H), 5.53 (br, 1 H), 6.85-6.87 (m, 2 H), 7.00-7.02 (m, 2 H), 7.08-7.10 (m, 2 H), 7.20-7.29 (m, 7 H), 7.32-7.36 (m, 4 H), 7.38-7.45 (m, 4 H), 8.08-8.10 (m, 2 H). HRMS: $C_{54}H_{65}N_{3}O_{11}SSi [M+NH_4]^+$ calcd: 1009.4453, obsd: 1009.4430.

p-Tolyl 2-azido-3-O-benzyl-4-O-tert-butyl-dimethylsilyl-6-O-levulinoyl-2-deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl-6-O-p-methoxybenzyl-1-thio- α -L-

idopyranoside (**S38**). Compound **S38** was synthesized from compound **S37** in 81% yield following the general procedure for protecting 6-OH with Lev. ¹H-NMR (500 MHz, CDCl₃): δ -0.09 (s, 3 H, Si(CH₃)₂), -0.01 (s, 3 H, Si(CH₃)₂), 0.88 (s, 9 H, C(CH₃)₃), 2.14 (s, 3 H, CH₃COCH₂CH₂), 2.31 (s, 3 H, SPhCH₃), 2.55-2.76 (m, 4 H, CH₃COCH₂CH₂), 3.27-3.29 (m, 1 H), 3.35-3.39 (m, 1 H), 3.50-3.54 (m, 1 H), 3.73-3.86 (m, 7 H), 4.02-4.15 (m, 2 H), 4.17 (br, 1 H), 4.24-4.27 (m, 1 H), 4.31-4.34 (m, 1 H), 4.50-4.56 (m, 2 H), 4.68-4.69 (m, 1 H), 4.75-4.77 (m, 1 H), 4.94-4.97 (m, 2 H), 5.37 (br, 1 H), 5.58 (br, 1 H), 6.88-6.90 (m, 2 H), 7.03-7.05 (m, 2 H), 7.14-7.16 (m, 2 H), 7.24-7.30 (m, 6 H), 7.34-7.39 (m, 4 H), 7.43-7.49 (m, 5 H), 8.12-8.14 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ -4.9, -3.9, 14.1, 17.8, 21.0, 25.8, 27.6, 29.7, 37.6, 55.1, 60.2, 62.9, 64.4, 67.1, 69.1, 70.0, 70.8, 71.2, 71.7, 72.5, 72.8, 74.4, 74.7, 80.4, 86.3, 98.2, 113.6, 126.9, 127.2, 127.8, 127.9, 128.0, 128.3, 128.3, 129.1, 129.5, 129.7, 129.8, 130.0, 131.7, 132.2, 133.1, 137.3, 137.4, 137.6, 159.0, 165.5, 172.3, 206.0. HRMS: C₅₉H₇₁N₃O₁₃SSi [M+NH₄]⁺ calcd: 1107.4821, obsd: 1107.4768.

p-*Tolyl* 2-azido-3-O-benzyl-4-O-tert-butyl-dimethylsilyl-6-O-levulinoyl-2-deoxy-α-Dglucopyranosyl-(1→4)-2-O-benzoyl-3-O-benzyl-1-thio-α-L-idopyranoside (**S39**). Compound **S38** (1.6 g, 1.48 mmol) was dissolved in DCM/H₂O (10:1, 30 mL), followed by addition of DDQ (500 mg, 2.25 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was diluted with DCM, washed with sat. NaHCO₃ solution, dried over Na₂SO₄. Silica gel column purification afforded compound **S39** (1.29 g, 90%). ¹H-NMR (500 MHz, CDCl₃): δ -0.17 (s, 3 H, Si(CH₃)₂), -0.03 (s, 3 H, Si(CH₃)₂), 0.84 (s, 9 H, C(CH₃)₃), 2.14 (s, 3 H, CH₃COCH₂CH₂), 2.31 (s, 3 H, SPhCH₃), 2.57-2.72 (m, 4 H, CH₃COCH₂CH₂), 3.22-3.25 (m, 2 H), 3.35-3.39 (m, 1 H), 3.68 (br, 1 H), 3.76-3.98 (m, 6 H), 4.06-4.10 (m, 1 H), 4.15 (br, 1 H), 4.40 (br, 1 H, *J* = 10.5 Hz), 4.52 (s, 1 H), 4.73-4.75 (m, 1 H, CH₂Ph), 4.82-4.85 (m, 1 H), 4.95-4.97 (m, 1 H, CH₂Ph), 5.36 (s, 1 H), 5.56 (s, 1 H), 7.03-7.10 (m, 4 H), 7.18-7.48 (m, 13 H), 8.10-8.12 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ -4.8, -3.9, 14.0, 17.8, 21.0, 25.7, 27.5, 29.7, 37.6, 61.3, 63.3, 64.4, 67.8, 69.7, 71.0, 71.1, 71.6, 72.3, 74.2, 75.9, 80.3, 86.3, 99.2, 126.7, 127.1, 127.9, 128.1, 128.2, 128.4, 129.6, 129.6, 129.9, 131.6, 132.2, 133.1, 137.2, 137.5, 137.6, 165.4, 172.3, 206.4. HRMS: $C_{51}H_{63}N_3O_{12}SSi[M+NH_4]^+$ calcd: 987.4245, obsd: 987.4199. *p*-Tolyl 2-azido-3-O-benzyl-4-O-tert-butyl-dimethylsilyl-2-deoxy-α-D-glucopyranosyl-

 $(1\rightarrow 4)$ -2-O-benzoyl-3-O-benzyl-1-thio- α -L-idopyranoside (S40). Compound S39 (1.29 g, 1.33 mmol) was dissolved in pyridine (10 mL) in a plastic flask followed by addition of 65-70% HF-pyridine solution (15 mL) under 0 °C. The solution was stirred overnight until complete disappearance of starting material as judged by TLC analysis. The reaction mixture was quenched by solid NaHCO₃ and diluted with DCM. The aqueous phase was extracted with DCM twice. The combined organic phase was further washed with a saturated aqueous solution of NaHCO₃ and dried over Na₂SO₄. Column purification afforded compound **S40** (1.08 g, 95%). ¹H-NMR (500 MHz, CDCl₃): δ 2.13 (s, 3 H, CH₃COCH₂CH₂), 2.26-2.29 (m, 1 H), 2.31 (s, 3 H, SPhCH₃), 2.54-2.72 (m, 4 H, $CH_3COCH_2CH_2$, 2.92 (br, 1 H), 3.21 (dd, 1 H, J = 3.5 Hz, J = 10 Hz), 3.31-3.35 (m, 1 H), 3.39-3.44 (m, 1 H), 3.69-3.71 (m, 1 H), 3.75-3.79 (m, 1 H), 3.85-3.90 (m, 2 H), 4.04-4.07 (m, 1 H, CH_2Ph), 4.12-4.14 (m, 1 H), 4.21 (dd, 1 H, J = 2 Hz, J = 12 Hz), 4.26-4.28 (m, 1 H, CH_2Ph), 4.35 (dd, 1 H, J = 5.5 Hz, J = 12 Hz), 4.55 (d, 1 H, J = 4 Hz), 4.73-4.76 (m, 1 H, CH₂Ph), 4.80-4.83 (m, 1 H), 4.95-4.98 (m, 1 H, CH₂Ph), 5.38 (s, 1 H), 5.55 (s, 1 H), 7.10-7.16 (m, 4 H), 7.24-7.30 (m, 4 H), 7.34-7.47 (m, 9 H), 8.13-8.15 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 14.1, 20.9, 21.0, 27.6, 29.7, 37.7, 60.3, 61.4, 63.2, 63.3, 68.0, 69.7, 70.5, 71.1, 71.2, 72.4, 74.8, 74.9, 80.2, 86.3, 98.6, 127.8, 127.9, 128.1, 128.3, 128.3, 128.4, 129.7, 129.9, 131.6, 132.4, 133.1, 137.2, 137.6, 137.7, 165.6, 173.1, 206.8. HRMS: $C_{45}H_{49}N_{3}O_{12}S[M+NH_4]^+$ calcd: 873.3381, obsd: 873.3345.

2-azido-3-O-benzvl-6-O-levulinovl-2-deoxv- α -D-glucopvranosvl- $(1 \rightarrow 4)$ -2-O*p*-*Tolvl* benzoyl-3-O-benzyl-6-O-tert-butyl-diphenylsilyl-1-thio- α -L-idopyranoside (13).Compound **S40** (1.08 g, 1.26 mmol) was dissolved in 10 mL DCM, followed by addition of imidazole (102 mg, 1.5 mmol) and TBDPSCI (487 µL, 1.88 mmol). The resulting mixture was stirred under room temperature overnight and diluted with DCM. The solution was washed with 10% HCl solution, sat. NaHCO₃ solution and dried over Na₂SO₄. After concentration, column purification afforded compound **13** (1.28 g, 93%). ¹H-NMR (500 MHz, CDCl₃): δ 1.08 (s, 9 H, C(CH₃)₃), 2.13 (s, 3 H, CH₃COCH₂CH₂), 2.29 (s, 3 H, $SPhCH_3$, 2.47-2.69 (m, 4 H, $CH_3COCH_2CH_2$), 2.76 (d, 1 H, J = 4.5 Hz), 3.24 (dd, 1 H, J= 3.5 Hz, J = 10 Hz, 3.31 - 3.36 (m, 1 H), 3.41 - 3.45 (m, 1 H), 3.56 - 3.61 (m, 2 H), 3.75 (br,1 H), 3.89-3.97 (m, 2 H), 4.14-4.17 (m, 1 H, CH₂Ph), 4.22-4.23 (m, 1 H), 4.33-4.36 (m, 1 H), 4.40-4.43 (m, 1 H, CH_2Ph), 4.68 (d, 1 H, J = 3.5 Hz), 4.75-4.81 (m, 2 H), 4.95-4.97 (m, 1 H, CH₂Ph), 5.39-5.41 (m, 1 H), 5.58-5.59 (m, 1 H), 6.99-7.01 (m, 2 H), 7.18-7.20 (m, 2 H), 7.26-7.50 (m, 19 H), 7.69-7.76 (m, 4 H), 8.12-8.15 (m, 2 H). ¹³C-NMR (125 MHz, $CDCl_3$: δ 14.1, 19.1, 21.0, 26.8, 27.6, 29.6, 37.8, 60.3, 62.5, 63.3, 63.5, 69.3, 69.4, 70.2, 70.3, 70.9, 72.5, 72.7, 74.9, 75.0, 79.9, 86.7, 98.5, 127.7, 127.7, 127.8, 127.9, 127.9, 128.3, 128.3, 128.4, 129.5, 129.7, 129.8, 129.9, 131.6, 132.7, 132.8, 132.9, 133.1, 135.5, 135.6, 137.5, 137.5, 137.8, 165.6, 173.4, 206.4. HRMS: $C_{61}H_{67}N_3O_{12}SSi [M+NH_4]^+$ calcd: 1111.4558, obsd: 1111.4517.



Synthesis of trisaccharide 7. Reagents and conditions: (a) TBDPSCl, imidazole, DCM; (b) NaOMe, DCM/MeOH; (c) LevOH, EDC-HCl, DMAP, DCM; (d) AgOTf, *p*-TolSCl, DCM, -78 °C, then **13**, TTBP, -78 °C-0 °C.

p-*Tolyl* 2-*O*-*benzoyl-3*-*O*-*benzyl-4*-*O*-*p*-*methoxybenzyl-6*-*O*-*tert*-*butyl*-*diphenylsilyl*-1-*thio*α-*L*-*idopyranoside* (**S42**). Compound **S41**^[5] (1.3 g, 2.16 mmol) was dissolved in 10 mL DCM, followed by addition of imidazole (176 mg, 2.59 mmol) and TBDPSCl (840 μL, 3.24 mmol). The resulting mixture was stirred under room temperature overnight and diluted with DCM. The solution was washed with 10% HCl solution, sat. NaHCO₃ solution and dried over Na₂SO₄. After concentration, column purification afforded compound **S42** (1.45 g, 80%). ¹H-NMR (500 MHz, CDCl₃): δ 1.13 (s, 9 H, C(CH₃)₃), 2.31 (s, 3 H, SPhCH₃), 3.74-3.76 (m, 1 H), 3.79 (s, 3 H, CH₃OPh), 3.96-4.05 (m, 2 H, H-6a, H-6b), 4.10-4.12 (m, 1 H, H-3), 4.36-4.48 (m, 2 H, PhCH₂), 4.73-4.76 (m, 2 H, H-5, PhCH₂), 4.92-4.95 (m, 1 H, PhCH₂), 5.48-5.49 (m, 1 H, H-2), 5.60 (dd, 1 H, *J* = 2.5 Hz, H-1), 6.72-6.74 (m, 2 H), 7.02-7.05 (m, 4 H), 7.32-7.53 (m, 16 H), 7.72-7.78 (m, 4 H), 8.02-8.04 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 19.1, 21.0, 26.8, 55.1, 63.0, 69.4, 70.1, 72.2, 72.4, 72.6, 73.7, 86.2, 113.5, 127.6, 127.7, 127.7, 128.1, 128.3, 129.3, 129.4, 129.6, 129.7, 129.9, 129.9, 131.8, 132.4, 132.9, 133.1, 133.2, 135.5, 135.6, 137.2, 137.6, 159.0, 165.5. HRMS: C₅₁H₅₄O₇SSi [M+NH₄]⁺ calcd: 856.3703, obsd: 856.3743.

3-O-benzyl-4-O-p-methoxybenzyl-6-O-tert-butyl-diphenylsilyl-1-thio- α -L*p*-*Tolvl* idopyranoside (S43). Compound S42 (2.7 g, 3.22 mmol) was dissolved in DCM/MeOH (1:1, 20 mL) and was added freshly prepared NaOMe in MeOH (5 M) to maintain the pH above 12. After the reaction was complete, the reaction was diluted with 10% HCl solution until the pH was around 6. The solution was washed with 10% HCl solution, a saturated aqueous NaHCO₃ solution and the organic phase was dried over Na₂SO₄. After concentration, column purification afforded compound S43 (2.1 g, 89%). ¹H-NMR (500 MHz, CDCl₃): δ 1.11 (s, 9 H, C(CH₃)₃), 2.30 (s, 3 H, SPhCH₃), 3.71-3.73 (m, 1 H, OH), 3.79 (s, 1 H, CH₃OPh), 3.82-3.84 (m, 1 H), 3.85-3.87 (m, 2 H), 4.03-4.10 (m, 2 H), 4.39-4.42 (m, 1 H), 4.52-4.55 (m, 1 H), 4.57-4.60 (m, 1 H), 4.75-4.81 (m, 2 H), 5.39 (br, 1 H, H-1), 6.80-6.82 (m, 2 H), 7.01-7.03 (m, 2 H), 7.09-7.11 (m, 2 H), 7.34-7.47 (m, 13 H), 7.70-7.73 (m, 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 19.1, 20.9, 26.8, 55.1, 62.6, 67.9, 69.4, 71.7, 71.9, 72.6, 73.4, 89.7, 113.8, 127.6, 127.6, 127.7, 128.4, 128.9, 129.4, 129.6, 129.6, 129.7, 131.9, 133.0, 133.1, 133.3, 135.5, 135.5, 136.9, 137.6, 159.4. HRMS: C₄₄H₅₀O₆SSi $[M+NH_4]^+$ calcd: 752.3441, obsd: 752.3485.

p-Tolyl 2-O-levulinoyl-3-O-benzyl-4-O-p-methoxybenzyl-6-O-tert-butyl-diphenylsilyl-1thio- α -L-idopyranoside (14). Compound S43 (2.1 g, 2.86 mmol) was dissolved in 10 mL DCM, followed by addition of LevOH (347 μ L, 3.43 mmol), DMAP (348 mg, 2.86 mmol) and EDC-HCl (657 mg, 3.43 mmol). After stirring under room temperature overnight, the reaction was quenched by 10% HCl solution and diluted with DCM. The organic phase was extracted with a saturated aqueous NaHCO₃ solution and dried over Na₂SO₄. Silica gel column purification afforded compound **14** (2.3 g, 97%). ¹H-NMR (500 MHz, CDCl₃): δ 1.05 (s, 9 H, C(CH₃)₃), 2.13 (s, 3 H, CH₃COCH₂CH₂), 2.27 (s, 3 H, SPhCH₃), 2.52-2.75 (m, 4 H, CH₃COCH₂CH₂), 3.59-3.61 (m, 1 H, H-4), 3.77 (s, 1 H, CH₃OPh), 3.88-3.91 (m, 3 H, H-3, H-6a, H-6b), 4.30-4.32 (m, 1 H), 4.45-4.47 (m, 1 H), 4.61-4.64 (m, 2 H), 4.82-4.85 (m, 1 H), 5.16-5.18 (m, 1 H, H-2), 5.42 (d, 1 H, *J* = 2.5 Hz, H-1), 6.75-6.77 (m, 2 H), 6.97-6.99 (m, 2 H), 7.05-7.07 (m, 2 H), 7.31-7.43 (m, 13 H), 7.64-7.69 (m, 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 14.1, 19.0, 20.9, 20.9, 26.7, 28.0, 29.7, 37.6, 55.1, 60.2, 62.9, 69.3, 70.0, 71.9, 72.5, 73.3, 86.0, 113.6, 127.6, 127.7, 127.8, 128.3, 129.4, 129.4, 129.5, 129.9, 131.8, 132.0, 133.1, 133.2, 135.5, 135.5, 137.1, 137.7, 159.1, 165.4, 171.7, 206.1. HRMS: C₄₉H₅₆O₈SSi [M+NH₄]⁺ calcd: 850.3809, obsd: 850.3805.

p-Tolyl 2-O-levulinoyl-3-O-benzyl-4-O-*p*-methoxybenzyl-6-O-tert-butyl-diphenylsilyl- α -L-idopyranosyl- $(1 \rightarrow 4)$ -2-azido-3-O-benzyl-6-O-levulinoyl-2-deoxy- α -D-glucopyranosyl-

 $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl-6-O-tert-butyl-diphenylsilyl-1-thio- α -L-idopyranoside (7). Compound 7 was synthesized from donor 14 and acceptor 13 in 52% yield following the general procedure of single step glycosylation. ¹H-NMR (500 MHz, CDCl₃): δ 1.03 (s, 9 H, C(CH₃)₃), 1.06 (s, 9 H, C(CH₃)₃), 2.01 (s, 3 H, CH₃COCH₂CH₂), 2.02 (s, 3 H, CH₃COCH₂CH₂), 2.27 (s, 3 H, SPhCH₃), 2.31-2.68 (m, 8 H, CH₃COCH₂CH₂), 3.23 (dd, 1 H, J = 4 Hz, J = 10.5 Hz), 3.41 (t, 1 H, J = 9 Hz), 3.53-3.55 (m, 1 H), 3.66-3.71 (m, 2 H), 3.77 (s, 1 H, CH₃OPh), 3.82-3.95 (m, 7 H), 3.98-4.02 (m, 3 H), 4.15-4.17 (m, 1 H), 4.23 (d, 1 H, J = 10.5 Hz), 4.29-4.31 (m, 1 H), 4.33-4.36 (m, 1 H), 4.58-4.62 (m, 4 H), 4.78-4.83 (m, 2 H), 4.87-4.89 (m, 2 H), 4.93-4.95 (m, 1 H), 5.37 (t, 1 H, <math>J = 4.5 Hz), 5.58 (d, 1 H, J)= 3.5 Hz), 6.73-6.75 (m, 2 H), 6.96-6.99 (m, 4 H), 7.12-7.17 (m, 6 H), 7.22-7.40 (m, 23 H), 7.45-7.49 (m, 1 H), 7.56-7.61 (m, 4 H), 7.65-7.67 (m, 2 H), 7.72 (m 2 H), 8.06-8.08 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 14.1, 19.1, 19.1, 21.0, 26.8, 26.9, 27.7, 27.9, 29.5, 29.6, 29.6, 34.6, 37.6, 55.2, 60.3, 62.9, 63.9, 64.0, 70.1, 70.4, 70.8, 71.4, 72.2, 73.0, 73.6, 74.3, 74.9, 75.2, 75.4, 75.6, 75.9, 78.9, 86.4, 97.9, 98.7, 113.6, 113.9, 127.1, 127.6, 127.6, 127.7, 127.7, 127.7, 127.8, 127.8, 128.0, 128.1, 128.2, 128.3, 128.6, 129.4, 129.4, 129.5, 129.6, 129.7, 129.7, 129.8, 129.8, 129.9, 131.3, 132.0, 132.8, 132.9, 133.1, 133.3, 135.6, 135.6, 135.6, 135.6, 137.5, 137.7, 137.9, 138.1, 159.2, 165.3, 171.7, 172.1, 206.0, 206.2. MALDI-MS: $C_{103}H_{115}N_{3}O_{20}SSi_{2}[M+Na]^{+}$ calcd: 1826.26, obsd: 1826.33.



Synthesis of disaccharide **8**. Reagents and conditions: (a) DDQ, DCM/H₂O; (b) HF/Pyridine; (c) TBDPSCl, imidazole, DCM.

p-Tolyl 2-azido-3,6-di-O-benzyl-4-O-tert-butyl-dimethylsilyl-2-deoxy-β-D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzovl-3-O-benzvl-1-thio- β -D-glucopyranoside (S44). Compound 17 (300) mg, 0.277 mmol) was dissolved in DCM/H₂O (10:1, 5 mL), followed by addition of DDQ (95 mg, 0.42 mmol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was diluted with DCM, washed with sat. NaHCO₃, dried over Na₂SO₄. Silica gel column purification afforded compound S44 (189 mg, 71%). ¹H-NMR (500 MHz, CDCl₃): δ -0.10 (s, 3 H, Si(CH₃)₂), -0.08 (s, 3 H, Si(CH₃)₂), 0.78 (s, 9 H, C(CH₃)₃), 2.30 (s, 3 H, SPhCH₃), 3.04 (d, 1 H, J = 6 Hz), 3.15 (dd, 1 H, J = 5.5 Hz, J =8.5 Hz), 3.40 (dd, 1 H, J = 5.5 Hz, J = 8.5 Hz), 3.46-3.52 (m, 2 H), 3.61-3.65 (m, 2 H), 3.70-3.72 (m, 1 H), 3.88-3.94 (m, 2 H), 4.02 (t, 1 H, J = 7.5 Hz), 4.11 (t, 1 H, J = 7.5 Hz), 4.45-4.47 (m, 1 H), 4.62-4.81 (m, 6 H), 5.31 (t, 1 H, J = 8 Hz), 5.61 (d, 1 H, J = 3.5 Hz), 7.06-7.08 (m, 2 H), 7.11-7.18 (m, 5 H), 7.23-7.35 (m, 12 H), 7.54-7.57 (m, 1 H), 8.05-8.07 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ -4.8, -3.9, 14.0, 17.8, 21.0, 21.0, 25.7, 60.2, 61.3, 63.2, 68.5, 71.3, 72.6, 73.0, 73.1, 73.3, 74.4, 74.8, 79.1, 80.2, 84.9, 86.5, 97.9, 114.2, 127.1, 127.3, 127.4, 127.5, 127.7, 127.8, 128.1, 128.1, 128.3, 128.3, 128.8, 129.5, 129.6, 129.7, 133.1, 133.1, 137.2, 137.3, 137.7, 138.0, 164.9. HRMS: $C_{53}H_{63}N_3O_{10}SSi[M+NH_4]^+$ calcd: 979.4347, obsd: 979.4371.

p-Tolyl 2-azido-3,6-di-O-benzyl-4-O-tert-butyl-dimethylsilyl-2-deoxy-a-D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzovl-3-O-benzvl-1-thio- β -D-glucopyranoside (S45). Compound S44 (1.3 g. 1.35 mmol) was dissolved in pyridine (10 mL) in a plastic flask followed by addition of 65-70% HF-pyridine solution (15 mL) under 0 °C. The solution was stirred overnight until complete disappearance of starting material as judged by TLC analysis. The reaction mixture was quenched by solid NaHCO₃ and diluted with DCM. The aqueous phase was extracted with DCM twice. The combined organic phase was further washed with a saturated aqueous solution of NaHCO₃ and dried over Na₂SO₄. Column purification afforded compound **S45** (800 mg, 70%). ¹H-NMR (600 MHz, CDCl₃): δ 2.31 (s, 3 H, SPhCH₃), 2.43-2.46 (m, 1 H), 2.51-2.53 (m, 1 H), 3.16-3.19 (m, 1 H), 3.50-3.53 (m, 1 H), 3.58-3.66 (m, 3 H), 3.72-3.77 (m, 2 H), 3.79-3.83 (m, 1 H), 3.90-3.94 (m, 1 H), 4.00-4.06 (m, 2 H), 4.50-4.58 (m, 2 H, CH₂Ph), 4.67-4.69 (m, 1 H, CH₂Ph), 4.74-4.81 (m, 3 H), 4.88-4.90 (m, 1 H), 5.30 (t, 1 H, J = 7.5 Hz), 5.58-5.59 (m, 1 H), 7.07-7.09 (m, 2 H), 7.12-7.20 (m, 5 H), 7.26-7.39 (m, 12 H), 7.43-7.47 (m, 2 H), 7.56-7.59 (m, 1 H), 8.07-8.09 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 21.1, 21.1, 60.3, 61.7, 62.4, 69.4, 71.0, 72.0, 72.3, 73.0, 73.6, 74.6, 75.0, 78.9, 79.6, 84.9, 86.5, 97.8, 113.7, 127.6, 127.7, 127.8, 127.8, 128.0, 128.2, 128.4, 128.5, 128.6, 128.7, 129.6, 129.8, 133.1, 133.3, 137.3, 137.3, 137.9, 138.3, 165.1, 171.7, 172.1, 206.0, 206.2. HRMS: $C_{47}H_{49}N_3O_{10}S[M+NH_4]^+$ calcd: 865.3482, obsd: 865.3478.

p-Tolyl 2-azido-3,6-di-O-benzyl-2-deoxy- α -D-glucopyranosyl-(1 \rightarrow 4)-2-O-benzoyl-3-Obenzyl-6-O-tert-butyl-diphenylsilyl-1-thio- β -D-glucopyranoside (**8**). Compound **S45** (800 mg, 0.945 mmol) was dissolved in 10 mL DCM, followed by addition of imidazole (102 mg, 1.5 mmol) and TBDPSCl (487 µL, 1.88 mmol). The resulting mixture was stirred under room temperature overnight and diluted with DCM. The solution was washed with 10% HCl solution, a saturated aqueous NaHCO₃ solution and dried over Na₂SO₄. After concentration, column purification afforded compound **8** (930 mg, 92%). ¹H-NMR (500 MHz, CDCl₃): δ 1.09 (s, 9 H, C(CH₃)₃), 2.28 (s, 3 H, SPhCH₃), 2.31-2.32 (m, 1 H), 3.13-3.19 (m, 2 H), 3.25-3.28 (m, 1 H), 3.43-3.47 (m, 1 H), 3.60-3.67 (m, 3 H), 3.92-3.97 (m, 2 H), 4.00-4.04 (m, 2 H), 4.20-4.35 (m, 2 H, CH₂Ph), 4.68-4.76 (m, 2 H, CH₂Ph), 4.82-4.87 (m, 3 H), 5.34-5.38 (m, 1 H), 5.59 (d, 1 H, J = 3.5 Hz), 6.98-7.00 (m, 2 H), 7.12-7.20 (m, 7 H), 7.27-7.41 (m, 16 H), 7.44-7.47 (m, 2 H), 7.56-7.59 (m, 1 H), 7.70-7.72 (m, 4 H), 8.08-8.10 (m, 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 19.3, 21.0, 26.9, 62.4, 63.8, 69.0, 70.8, 72.0, 72.9, 73.1, 73.5, 74.4, 74.9, 79.2, 79.6, 85.0, 87.3, 97.6, 127.6, 127.7, 127.8, 127.9, 128.2, 128.3, 128.4, 128.5, 129.6, 129.6, 129.7, 129.8, 130.4, 131.9, 133.1, 133.3, 133.6, 235.5, 135.8, 137.3, 137.5, 137.5, 138.0, 165.2. HRMS: C₆₃H₆₇N₃O₁₀SSi [M+NH₄]⁺ calcd: 1103.4660, obsd: 1103.4486.

N-Fluorenylmethyloxycarbonyl-O-[2,3-di-O-levulinoyl-4,6-O-benzylidene-\beta-D-galactopyranosyl-(1\rightarrow3)-2-O-benzoyl-4,6-O-benzylidene-\beta-D-galactopyranosyl-(1\rightarrow4)-

2,3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine benzyl ester (25). Compound 25 was synthesized from donor 24 and acceptor 5 in 43% yield following the general procedure of single step glycosylation. ¹H-NMR (600 MHz, CDCl₃): δ 1.96 (s, 3 H, CH₃COCH₂CH₂), 2.02 (s, 3 H, CH₃COCH₂CH₂), 2.13-2.40 (m, 3 H, CH₃COCH₂CH₂), 2.51-2.70 (m, 5 H, CH₃COCH₂CH₂), 3.19 (s, 1 H), 3.23-3.27 (m, 1 H), 3.35 (s, 1 H), 3.70-3.76 (m, 3 H), 3.80-3.83 (m, 1 H, CH₂Ph), 3.88-3.92 (m, 1 H), 3.95-3.98 (m, 1 H), 4.08-4.13 (m, 2 H), 4.16-4.24 (m, 4 H), 4.29-4.33 (m, 2 H), 4.46-4.48 (m, 1 H), 4.53 (d, 1 H, J = 6.0 Hz), 4.68-6.04.71 (m, 3 H), 4.99-5.08 (m, 2 H, CH₂Ph), 5.12-5.15 (m, 1 H), 5.26-5.30 (m, 1 H), 5.52-5.44 (m, 2 H), 5.51-5.60 (m, 3 H), 7.15-7.47 (m, 27 H), 7.51-7.57 (m, 3 H), 7.74-7.76 (m, 2 H), 7.93-8.01 (m, 6 H). ¹³C-NMR (125 MHz, CDCl₃): δ 27.4, 28.1, 29.5, 29.6, 37.6, 37.7, 47.0, 54.2, 60.3, 62.3, 66.4, 67.1, 67.1, 67.2, 68.2, 68.2, 68.7, 69.0, 70.8, 71.2, 71.6, 71.6, 73.1, 75.5, 75.6, 75.7, 100.5, 100.6, 100.9, 102.2, 119.9, 125.1, 126.3, 126.3, 127.0, 127.0, 127.6, 127.7, 127.8, 128.1, 128.2, 128.3, 128.3, 128.3, 128.4, 128.5, 128.7, 129.0, 129.1, 129.4, 129.5, 129.7, 129.9, 133.0, 133.1, 133.4, 135.1, 137.4, 137.7, 141.2, 141.2, 143.6, 143.8, 155.8, 164.6, 165.1, 165.5, 169.4, 171.1, 172.1, 206.5, 206.5. HRMS: $C_{87}H_{83}NO_{26}[M+NH_4]^+$ calcd: 1576.5625, obsd: 1576.5555.

N-Fluorenylmethyloxycarbonyl-O-[2,3-di-O-levulinoyl-4,6-O-benzylidene-β-D-

galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4,6-O-benzylidene- α -D-galactopyranosyl- $(1 \rightarrow 4)$ -2,

3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine benzyl ester (25a). Compound 25a was a side product generated in 45% yield during the synthesis of compound 25. ¹H-NMR (600 MHz, CDCl₃): δ 1.94 (s, 3 H, CH₃COCH₂CH₂), 1.95 (s, 3 H, CH₃COCH₂CH₂), 1.96-2.15 (m, 3 H, CH₃COCH₂CH₂), 2.41-2.66 (m, 5 H, CH₃COCH₂CH₂), 3.44 (s, 1 H), 3.46-3.49 (m, 1 H), 3.77 (s, 1 H), 3.81-3.84 (m, 1 H), 3.96-4.12 (m, 6 H), 4.14-4.17 (m, 1 H), 4.23-4.36 (m, 6 H), 4.50-4.52 (m, 2 H), 4.61 (d, 1 H, J = 5.5 Hz), 4.68 (dd, 1 H, J = 3 Hz, J = 8.5 Hz), 4.73 (d, 1 H, J = 6.5 Hz), 5.10-5.17 (m, 3 H), 5.25-5.29 (m, 1 H), 5.40-5.41 (m, 1 H), 5.45-5.49 (m, 2 H), 5.54-5.61 (m, 3 H), 7.14-7.46 (m, 26 H), 7.50-7.55 (m, 6 H), 7.66-7.68 (m, 2 H), 7.73-7.76 (m , 2 H), 7.85-7.87 (m 2 H). 13 C-NMR (125 MHz, CDCl₃): δ 14.1, 20.9, 27.1, 28.0, 29.5, 29.5, 37.5, 37.6, 47.0, 54.2, 60.3, 63.8, 64.0, 66.4, 67.2, 67.3, 68.3, 68.8, 68.9, 70.0, 71.6, 71.7, 72.0, 72.8, 73.2, 75.1, 76.2, 98.6, 100.4, 100.8, 101.2, 101.3, 119.9, 125.1, 126.1, 126.2, 127.0, 127.0, 127.6, 127.6, 128.0, 128.1, 128.1, 128.2, 128.2, 128.3, 128.3, 128.5, 128.6, 128.8, 128.9, 129.0, 129.4, 129.4, 129.6, 133.0, 133.1, 133.2, 135.1, 137.4, 137.6, 141.2, 141.2, 143.6, 143.7, 155.8, 165.1, 165.2, 165.3, 169.3, 170.9, 172.0, 206.2, 206.5. ESI-MS: $C_{87}H_{83}NO_{26}$ calcd: $[M+NH_4]^+$ calcd: 1575.5, obsd: 1575.9. HRMS: $[M+NH_4]^+$ calcd: 1575.5625, obsd: 1575.5486.

N-Fluorenylmethyloxycarbonyl-O-[4,6-O-benzylidene-\beta-D-galactopyranosyl-(1\rightarrow3)-2-<i>O-benzoyl-4,6-O-benzylidene-\beta-D-galactopyranosyl-(1\rightarrow4)-2,3-<i>di-O-benzoyl-\beta-D-

xylopyranosyl]-*L*-serine benzyl ester (**9**). Compound **9** was synthesized from compound **25** in 72% yield following the general procedure for Lev deprotection. ¹H-NMR (500 MHz, CDCl₃): δ 2.53-2.55 (m, 1 H), 2.77 (s, 1 H), 3.25-3.36 (m, 4 H), 3.64-3.68 (m, 1 H), 3.73-3.76 (m, 2 H), 3.80-3.84 (m, 2 H), 3.95-4.06 (m, 4 H), 4.10-4.14 (m, 1 H), 4.17-4.34 (m, 6 H), 4.48-4.50 (m, 1 H), 4.56 (d, 1 H, *J* = 5.5 Hz), 4.78 (d, 1 H, *J* = 8 Hz), 5.01-5.09 (m, 2 H, C*H*₂Ph), 5.15-5.18 (m, 1 H), 5.44-5.47 (m, 2 H), 5.54-5.63 (m, 3 H), 7.20-7.54 (m, 29 H), 7.73-7.76 (m, 2 H), 7.94-7.97 (m, 6 H), 8.33-8.37 (m 1 H). ¹³C-NMR (125 MHz, CDCl₃): δ 47.0, 54.2, 62.2, 66.7, 67.0, 67.2, 67.3, 68.3, 69.0, 70.7, 71.0, 71.3, 71.5, 72.0, 75.0, 75.5, 75.9, 77.9, 100.4, 101.1, 101.2, 101.9, 104.1, 119.9, 125.1, 126.2, 126.6, 127.0, 127.0, 127.6, 127.7, 128.0, 128.2, 128.2, 128.3, 128.3, 128.4, 128.4, 128.5, 128.7, 128.8, 129.1, 129.2, 129.5, 129.5, 129.6, 129.8, 129.9, 130.8, 133.1, 133.1, 133.2, 135.1, 137.4, 137.6, 141.2, 141.2, 143.6, 143.8, 155.9, 165.1, 165.5, 165.6, 169.4. MALDI-MS: C₇₇H₇₁NO₂₂ [M+Na]⁺ calcd: 1385.38, obsd: 1385.57.

p-Tolyl 2-O-levulinoyl-3-O-benzyl-4-O-*p*-methoxybenzyl-6-O-tert-butyl-diphenylsilyl- α -L $idopyranosyl-(1\rightarrow 4)-2-azido-3-O-benzyl-6-O-levulinoyl-2-deoxy-\alpha-D-glucopyranosyl (1 \rightarrow 4)$ -2-O-benzovl-3-O-benzvl-6-O-tert-butyl-diphenylsilyl- α -L-idopyranosvl- $(1 \rightarrow 4)$ -2azido-3, 6-di-O-benzyl-2-deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzyl-3-O-benzyl-6-Otert-butyl-diphenvlsilyl-1-thio- β -D-glucopyranoside (26). Compound 26 was synthesized from donor 7 and acceptor 8 in 93% yield following the general procedure of single step glycosylation. ¹H-NMR (600 MHz, CDCl₃): δ 0.99 (s, 9 H, C(CH₃)₃), 1.00 (s, 9 H, C(CH₃)₃), 1.04 (s, 9 H, C(CH₃)₃), 2.00 (s, 3 H, CH₃COCH₂CH₂), 2.01 (s, 3 H, CH₃COCH₂CH₂), 2.25 (s, 3 H, SPhCH₃), 2.30-2.65 (m, 8 H, CH₃COCH₂CH₂), 3.05-3.10 (m, 1 H), 3.13-3.15 (m, 2 H), 3.18-3.20 (m, 1 H), 3.40-3.44 (m, 2 H), 3.47-3.59 (m, 3 H), 3.64-3.69 (m, 2 H), 3.72-3.76 (m, 6 H), 3.82-4.03 (m, 10 H), 4.08-4.20 (m, 6 H), 4.28-4.31 (m, 2 H), 4.53-4.57 (m, 2 H), 4.60-4.64 (m, 2 H), 4.69-4.79 (m, 6 H), 4.84-4.89 (m, 2 H), 4.97 (d, 1 H, J = 9.5 Hz), 5.16-5.19 (m, 2 H), 5.27-5.31 (m, 1 H), 5.46-5.48 (m, 1 H), 6.71-6.73 (m, 2 H), 6.93-6.97 (m, 4 H), 7.02-7.38 (m, 52 H), 7.42-7.49 (m, 4 H), 7.53-7.76 (m, 12 H), 8.03-8.06 (m 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 19.2, 19.2, 19.2, 21.0, 26.8, 26.9, 27.0, 27.7, 27.9, 29.6, 29.6, 37.6, 55.2, 62.2, 62.6, 63.0, 63.0, 63.7, 63.9, 68.1, 69.6, 69.9, 70.6, 70.9, 71.9, 72.0, 72.9, 73.2, 73.2, 74.2, 74.3, 74.5, 74.6, 74.7, 74.9, 74.9, 75.0, 78.4, 78.9, 79.7, 84.6, 87.0, 97.7, 97.7, 98.3, 113.6, 127.1, 127.3, 127.4, 127.5, 127.6, 127.6, 127.7, 127.7, 127.8, 127.8, 127.9, 128.0, 128.0, 128.1, 128.2, 128.2, 128.3, 128.3, 128.4, 128.6, 129.4, 129.6, 129.6, 129.6, 129.7, 129.7, 129.8, 129.8, 129.9, 130.3, 132.0, 132.9, 133.2, 133.3, 133.5, 135.6, 135.6, 135.6, 135.6, 135.7, 137.4, 137.5, 137.5, 137.8, 137.8, 138.1, 138.2, 159.2, 165.2, 165.3, 171.7, 172.1, 206.1, 206.1, MALDI-MS: $C_{159}H_{174}N_6O_{30}SSi_3[M+Na]^+$ calcd: 2768.43, obsd: 2790.93.

Octasaccharide 27: A mixture of donor 26 (39 mg, 0.014 mmol) and freshly activated molecular sieves 4Å (200 mg) in CH₂Cl₂ (2 mL) was stirred at room temperature for 30 minutes and then cooled to -78 °C, which was followed by the addition of AgOTf (11 mg, 0.042 mmol) dissolved in acetonitrile (0.02 mL) without touching the wall of the flask. After 5 minutes, orange coloured *p*-TolSCl (2.2 μ L, 0.014 mmol) was added through a microsyringe. Since the reaction temperature was lower than the freezing point of *p*-TolSCl, *p*-TolSCl was added directly into the reaction mixture to prevent it from freezing on the flask wall. The characteristic yellow colour of *p*-TolSCl in the reaction solution

dissipated rapidly within a few seconds indicating its depletion. After the donor was completely consumed according to TLC analysis (about 5 minutes at -78 °C), a solution of acceptor 9 (19 mg, 0.014 mmol) and TTBP (3.5 mg, 0.01 mmol) in CH₂Cl₂ (1 mL) was slowly added dropwise via a syringe. The reaction mixture was stirred for 1.5 h until the temperature reached 0 °C, at which point sat. NaHCO₃ solution was added to quench the reaction. The reaction mixture was diluted with CH₂Cl₂ (50 mL) and filtered through Celite. The Celite was washed extensively with CH₂Cl₂ until TLC showed no products in the filtrate. The filtrate was combined and extracted with a saturate NaHCO₃ solution and dried with Na₂SO₄. After filtration, the reaction mixture was purified by flash column chromatography (hexanes : EtOAc = 1 : 1) to give compound 27 (47 mg, 83%).¹H-NMR (600 MHz, CDCl₃): δ 0.92 (s, 9 H, C(CH₃)₃), 1.00 (s, 9 H, C(CH₃)₃), 1.03 (s, 9 H, C(CH₃)₃), 1.99 (s, 3 H, CH₃COCH₂CH₂), 2.00 (s, 3 H, CH₃COCH₂CH₂), 2.20 (s, 1 H), 2.27-2.66 (m, 8 H, CH₃COCH₂CH₂), 3.02-3.06 (m, 2 H), 3.09-3.15 (m, 2 H), 3.18-3.22 (m, 2 H), 3.30-3.46 (m, 5 H), 3.47-3.49 (m, 1 H), 3.53-3.57 (m, 1 H), 3.64-3.87 (m, 19 H), 3.94-4.33 (m, 18 H), 4.48-4.77 (m, 12 H), 4.84-4.88 (m, 2 H), 4.96-5.03 (m, 3 H), 5.14-5.18 (m, 3 H), 5.22-5.27 (m, 1 H), 5.31 (s, 2 H), 5.42-5.44 (m, 1 H), 5.47-5.51 (m, 1 H), 5.53-5.55 (m, 1 H), 5.58-5.62 (m, 1 H), 6.71-6.73 (m, 2 H), 6.94-6.96 (m, 4 H), 7.01-7.03 (m, 2 H), 7.04-7.07 (m, 3 H), 7.11-7.38 (m, 78 H), 7.43-7.64 (m, 18 H), 7.42-7.46 (m, 2 H), 7.94-8.02 (m 10 H). ¹³C-NMR (125 MHz, CDCl₃): δ 19.2, 19.2, 19.3, 26.7, 26.7, 26.9, 27.0, 27.6, 27.9, 29.6, 29.6, 29.6, 37.6, 47.0, 54.2, 55.2, 62.2, 62.3, 62.6, 62.9, 62.9, 63.7, 64.2, 66.8, 67.0, 67.2, 67.3, 68.0, 68.2, 69.0, 69.5, 68.7, 69.9, 70.6, 70.7, 71.3, 71.5, 71.8, 72.0, 73.2, 73.2, 73.3, 73.9, 74.1, 74.2, 74.3, 74.4, 74.7, 74.8, 74.9, 75.0, 75.0, 75.2, 75.5, 75.5, 75.8, 76.5, 78.1, 78.4, 78.9, 83.3, 97.6, 97.7, 98.3, 100.1, 100.5, 101.0, 101.4, 102.0, 104.0, 113.6, 119.9, 125.1, 126.0, 126.6, 127.0, 127.1, 127.3, 127.4, 127.5, 127.6, 127.7, 127.8, 127.8, 127.8, 127.9, 127.9, 127.9, 128.0, 128.0, 128.1, 128.2, 128.2, 128.3, 128.3, 128.3, 128.4, 128.5, 128.6, 128.7, 129.1, 129.4, 129.5, 129.6, 129.6, 129.7, 129.7, 129.9, 130.0, 132.9, 133.1, 133.1, 133.1, 133.2, 133.2, 133.3, 133.5, 135.1, 135.4, 135.4, 135.6, 135.6, 135.6, 137.5, 137.5, 137.6, 137.7, 138.1, 138.2, 141.2, 143.6, 143.8, 155.9, 159.1, 165.1, 165.2, 165.3, 165.3, 165.5, 169.4, 171.7, 172.1, 206.1, 206.2. MALDI-MS: $C_{222}H_{231}N_7O_{25}Si_3$: [M+NH₄]⁺ calcd: 3931.48, obsd: 3930.60.

Synthesis of syndecan-1 glycopeptide 1



Synthesis of octasaccharide 6. Reagents and conditions: (a) HF/Pyridine; (b) TEMPO, BAIB, DCM, H₂O, *t*-BuOH; (c) MeI, K₂CO₃, DMF.

N-Fluorenylmethyloxycarbonyl-O-[2-O-levulinoyl-3-O-benzyl-4-O-p-methoxybenzyl- α -L $idopyranosyl-(1\rightarrow 4)-2-azido-3-O-benzyl-6-O-levulinoyl-2-deoxy-\alpha-D-glucopyranosyl (1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- α -L-idopyranosyl- $(1 \rightarrow 4)$ -2-azido-3, 6-di-O-benzvl-2deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -4, 6-O-benzylidene- β -D-galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-O-benzylidene-β-Dgalactopyranosyl- $(1 \rightarrow 4)$ -2, 3-di-O-benzoyl- β -D-xylopyranosyl]-L-serine benzyl ester (S46). Compound 27 (715 mg, 0.178 mmol) was dissolved in pyridine (6 mL) in a plastic flask followed by addition of 65-70% HF-pyridine solution (4.2 mL) under 0 °C. The solution was stirred overnight until complete disappearance of starting material as judged by TLC analysis. The reaction mixture was quenched by solid NaHCO₃ and diluted with DCM. The aqueous phase was extracted with DCM twice. The combined organic phase was further washed with sat. NaHCO3 and dried over Na2SO4. Column purification afforded compound **S46** (520 mg, 90%). ¹H-NMR (500 MHz, CDCl₃): δ 2.11 (s, 3 H, CH₃COCH₂CH₂), 2.14 (s, 3 H, CH₃COCH₂CH₂), 2.43-2.78 (m, 8 H, CH₃COCH₂CH₂), 3.20-3.32 (m, 7 H), 3.36-3.42 (m, 2 H), 3.46-3.53 (m, 3 H), 3.56-3.64 (m, 4 H), 3.68-4.35 (m, 41 H), 4.41-4.44 (m, 1 H), 4.47-4.53 (m, 4 H), 4.56-4.58 (m, 1 H), 4.63-4.65 (m, 1 H), 4.69-4.93 (m, 11 H), 5.02-5.11 (m, 4 H), 5.16-5.20 (m, 2 H), 5.25-5.29 (m, 1 H), 5.35-5.40 (m, 2 H), 5.52-5.64 (m, 4 H), 6.81-6.84 (m, 2 H), 7.12-7.47 (m, 66 H), 7.53-7.58 (m, 2 H), 7.96-8.01 (m, 8 H), 8.07-8.09 (m 2 H). ¹³C-NMR (125 MHz, CDCl₃): δ 14.1, 20.9, 27.7, 27.8, 29.6, 29.7, 37.6, 37.7, 46.9, 54.1, 55.1, 60.3, 60.8, 61.1, 61.6, 62.2, 62.3, 63.3, 63.6, 66.7, 66.9, 67.1, 67.2, 67.5, 68.0, 68.1, 68.7, 69.0, 69.2, 69.3, 69.8, 69.9, 70.0, 70.6, 71.3, 71.5, 71.8, 72.1, 72.3, 72.7, 72.8, 73.1, 73.4, 73.8, 74.1, 74.6, 74.8, 75.0, 75.1, 75.3, 75.4,

75.6, 76.5, 78.7, 78.8, 83.2, 97.0, 97.3, 97.5, 97.5, 100.1, 100.4, 100.9, 101.8, 103.8, 113.7, 119.9, 125.0, 125.8, 126.5, 127.0, 127.0, 127.5, 127.6, 127.6, 127.7, 127.7, 127.8, 127.8, 127.9, 128.0, 128.0, 128.1, 128.1, 128.2, 128.2, 128.2, 128.3, 128.3, 128.4, 128.4, 128.5, 128.7, 129.0, 129.4, 129.5, 129.5, 129.6, 129.7, 129.7, 129.8, 129.8, 133.0, 133.1, 133.1, 133.2, 135.0, 137.1, 137.4, 137.4, 137.5, 137.6, 137.8, 141.1, 141.1, 143.6, 143.7, 155.8, 159.3, 165.0, 165.1, 165.3, 165.4, 165.7, 169.4, 171.0, 171.8, 172.1, 206.7, 206.8. MALDI-MS: $C_{181}H_{183}N_7O_{52}$ [M+Na]⁺ calcd: 3311.41, obsd: 3311.30. *N-Fluorenylmethyloxycarbonyl-O-[methyl* 2-O-levulinoyl-3-O-benzyl-4-O-pmethoxybenzyl- α -L-idopyranosyluronate- $(1 \rightarrow 4)$ -2-azido-3-O-benzyl-6-O-levulinoyl-2 $deoxv-\alpha$ -D-glucopvranosvl- $(1 \rightarrow 4)$ -methyl 2-O-benzovl-3-O-benzvl-a-L $idopyranosyluronate-(1 \rightarrow 4)-2-azido-3,$ 6-di-O-benzyl-2-deoxy-a-D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -4, 6-O-benzvlidene-β-Dgalactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-O-benzylidene- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -2, 3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine benzyl ester (6). Compound S46 (115 mg. 0.035 mmol) was dissolved in DCM/tBuOH/H₂O (4:4:1, 4.5 mL), followed by addition of TEMPO (4 mg) and BAIB (90 mg). The resulting mixture was stirred under room temperature overnight. After the reaction was complete indicated by TLC analysis, it was neutralized by 1 M HCl solution to adjust pH around 6. The solution was first diluted with DCM, then extracted with H₂O. The combined organic phase was dried over Na₂SO₄. After concentration, the crude product S47 was confirmed by MALDI-MS analysis. MALDI-MS: $C_{181}H_{177}N_7O_{55}$ calcd: $[M+Na]^+$ calcd: 3351.00, obsd: 3351.13. The crude compound was dissolved in dry DMF (5 mL), to which was added MeI (17 µL, 0.263 mmol) and K_2CO_3 (73 mg, 0.525 mmol). The resulting mixture was stirred under room temperature overnight. The reaction mixture was diluted with DCM and H₂O. The aqueous phase was extracted with DCM twice. The combined organic phase was further washed with sat. NaHCO₃ and dried over Na₂SO₄. Column purification afforded compound 6 (110 mg, 93%). ¹H-NMR (500 MHz, CDCl₃): δ 2.04 (s, 3 H, CH₃COCH₂CH₂), 2.11 (s, 3 H, CH₃COCH₂CH₂), 2.43-2.74 (m, 8 H, CH₃COCH₂CH₂), 3.18-3.59 (m, 17 H), 3.62-3.99 (m, 19 H), 4.06-4.25 (m, 11 H), 4.29-4.78 (m, 19 H), 4.85-4.89 (m, 2 H), 4.93-4.95 (m, 1 H), 5.02-5.09 (m, 2 H), 5.13-5.17 (m, 2 H), 5.21-5.28 (m, 2 H), 5.33-5.38 (m, 2 H), 5.45-5.61 (m, 4 H), 6.76-6.81 (m, 1 H), 7.09-7.48 (m, 68 H), 7.51-7.56 (m, 2 H), 7.74-7.76 (m, 1 H), 7.85-7.89 (m, 2 H), 7.93-7.99 (m, 7 H), 8.07-8.09 (m 1 H). ¹³C-NMR (125 MHz, CDCl₃): δ 14.0, 21.4, 22.6, 27.7, 27.8, 27.9, 29.3, 29.5, 29.6, 29.7, 31.8, 37.6, 37.9, 47.0, 51.7, 51.9, 52.1, 54.2, 55.2, 61.9, 62.2, 62.7, 63.0, 66.8, 66.9, 67.1, 67.3, 68.2, 68.7, 69.0, 69.8, 69.9, 70.2, 70.7, 70.8, 71.2, 71.3, 71.5, 72.3, 73.1, 73.4, 73.7, 73.8, 74.2, 74.4, 74.6, 75.0, 75.4, 75.4, 75.5, 75.7, 77.8, 77.9, 82.6, 97.2, 97.7, 97.9, 99.2, 100.0, 100.5, 100.6, 101.0, 101.8, 103.8, 113.7, 119.9, 125.1, 125.2, 125.8, 126.6, 127.0, 127.0, 127.3, 127.4, 127.6, 127.6, 127.7, 127.7, 127.8, 127.8, 127.9, 127.9, 128.0, 128.0, 128.1, 128.1, 128.2, 128.2, 128.3, 128.3, 128.3, 128.4, 128.6, 128.8, 129.0, 129.0, 129.2, 129.3, 129.4, 129.5, 129.6, 129.6, 129.7, 129.8, 133.1, 133.2, 133.4, 135.1, 137.2, 137.4, 137.4, 137.6, 137.6, 137.8, 137.9, 141.2, 143.6, 143.8, 155.8, 159.3, 164.9, 165.1, 165.1, 165.4, 165.5, 168.9, 169.4, 169.4, 169.9, 171.8, 172.3, 206.1, 206.5. MALDI-MS: C₁₈₄H₁₈₃N₇O₅₅ [M+Na]⁺ calcd: 3395.44, obsd: 3395.40.



Synthesis of dipeptide **29**. Reagents and conditions: (a) BOP, DIPEA, DCM/THF; (b) TFA, DCM.

N-(Acetyl)-O-(benzyl)-L-serglycine-t-butyl-ester (**S50**). Serine **S48** (1.329 g, 5.6 mmol), glycine **S49** (940 mg, 5.6 mmol) were dissolved in DCM/THF (1:1, 20 mL), followed by addition of BOP (4.95 g, 11.2 mmol) and DIPEA (1.85 mL, 11.2 mol). The resulting mixture was stirred under room temperature overnight. After the reaction was complete, it was diluted with DCM. The solution was washed with 10% HCl solution, sat. NaHCO₃ solution and dried over Na₂SO₄. After concentration, column purification afforded compound **S50** (2.05 g, 82%). ¹H-NMR (500 MHz, CDCl₃): δ 1.44 (s, 9 H, C(CH₃)₃), 1.99 (s, 3 H, CH₃CONH), 3.52 (dd, 1 H, *J* = 7 Hz, *J* = 9.5 Hz), 3.86-3.90 (m, 3 H), 4.51-4.60 (m, 3 H), 6.44-6.45 (m, 1 H, NH), 6.95-6.98 (m, 1 H, NH), 7.13-7.23 (m, 1 H), 7.24-7.33 (m, 4 H). ¹³C-NMR (125 MHz, CDCl₃): δ 23.1, 27.9, 42.1, 52.3, 69.1, 73.4, 82.2, 125.2, 127.8, 127.9, 128.1, 128.4, 128.9, 137.3, 168.4, 170.0, 170.2. HRMS: C₁₈H₂₆N₂O₅ [M+H]⁺ calcd: 351.1920, obsd: 351.1895.

N-(Acetyl)-O-(benzyl)-L-serglycine (**29**). Compound **S50** (2.05 g, 1.86 mmol) was dissolved in 4 mL DCM, followed by addition of TFA (4 mL). The resulting mixture was stirred under room temperature until the reaction was complete indicated by TLC analysis. The solution was concentrated to dryness to afford compound **29** which was used directly for next step without further purification. ¹H-NMR (500 MHz, CD₃OD): δ 2.00 (s, 3 H, CH₃CONH), 3.70-3.77 (m, 2 H), 3.91-3.93 (m, 2 H), 4.52-4.57 (m, 2 H), 6.63 (t, 1 H, *J* = 4.5 Hz), 7.24-7.27 (m, 1 H), 7.29-7.34 (m, 4 H), 8.21-8.24 (m, 1 H, COOH). ¹³C-NMR (125 MHz, CD₃OD): δ 22.5, 41.9, 54.7, 70.7, 74.2, 128.7, 128.9, 128.9, 129.3, 139.2, 172.5, 173.4. HRMS: C₁₄H₁₈N₂O₅ [M+H]⁺ calcd: 295.1294, obsd: 295.1287.



Synthesis of octasaccharide **28**. Reagents and conditions: (a) Ac_2O , pyridine, 50 °C; (b) piperidine, DCM.

 $\label{eq:sphere:product} N-Fluorenylmethyloxycarbonyl-O-[methyl 2-O-levulinoyl-3-O-benzyl-4-O-p-methoxybenzyl-a-L-idopyranosyluronate-(1 \rightarrow 4)-2-azido-3-O-benzyl-6-O-levulinoyl-2-deoxy-a-D-glucopyranosyl-(1 \rightarrow 4)-methyl 2-O-benzoyl-3-O-benzyl-a-L-idopyranosyluronate-(1 \rightarrow 4)-2-azido-3, 6-di-O-benzyl-2-deoxy-a-D-glucopyranosyl-$

6-0- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-acetyl-4, benzvlidene- β -D-galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-O-benzylidene-β-Dgalactopyranosyl- $(1 \rightarrow 4)$ -2, *3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine* benzyl ester (S51). Compound 6 (30 mg, 0.0089 mmol) was dissolved in 2 mL pyridine, followed by addition of 1 mL Ac₂O. The resulting mixture was stirred under 50 °C overnight. After cooling back to room temperature, it was diluted with DCM, washed with 10% HCl, sat. NaHCO₃. The combined organic phase was dried over Na₂SO₄. Column purification afforded compound **S51** (27 mg, 88%). ¹H-NMR (500 MHz, CDCl₃): δ 2.04 (s, 3 H, CH₃COCH₂CH₂), 2.11 (s, 3 H, CH₃COCH₂CH₂), 2.33 (s, 3 H, COCH₃), 2.44-2.75 (m, 8 H, CH₃COCH₂CH₂), 3.10-3.36 (m, 9 H), 3.44-3.57 (m, 9 H), 3.65-3.97 (m, 18 H), 4.06-4.22 (m, 10 H), 4.27-4.42 (m, 4 H), 4.45-4.70 (m, 13 H), 4.74-4.79 (m, 3 H), 4.84-4.89 (m, 3 H), 4.99-5.07 (m, 2 H), 5.11-5.15 (m, 3 H), 5.17-5.22 (m, 2 H), 5.27-5.33 (m, 3 H), 5.36-5.38 (m, 1 H), 5.44-5.48 (m, 1 H), 5.52-5.58 (m, 3 H), 6.79-6.81 (m, 2 H), 7.07-7.55 (m, 69 H), 7.73-7.77 (m, 2 H), 7.88-8.00 (m, 9 H). ¹³C-NMR (125 MHz, CDCl₃): δ 20.1, 21.4, 27.8, 27.9, 29.5, 29.7, 37.6, 37.9, 47.0, 51.7, 51.9, 52.1, 54.2, 55.2, 62.3, 62.7, 63.0, 66.7, 67.0, 67.1, 67.2, 68.1, 69.0, 69.8, 70.8, 70.9, 71.2, 71.3, 71.3, 71.7, 72.3, 72.9, 73.4, 73.7, 73.8, 73.9, 74.5, 74.7, 75.0, 75.2, 75.4, 75.4, 75.7, 77.9, 82.6, 97.2, 97.7, 97.9, 99.2, 100.1, 100.4, 100.5, 102.2, 113.7, 119.9, 125.1, 125.2, 126.0, 126.4, 127.0, 127.0, 127.4, 127.6, 127.6, 127.6, 127.7, 127.7, 127.8, 127.8, 127.9, 127.9, 128.0, 128.0, 128.0, 128.1, 128.2, 128.2, 128.3, 128.3, 128.3, 128.4, 128.4, 128.5, 128.6, 128.7, 129.0, 129.1, 129.2, 129.3, 129.5, 129.5, 129.7, 129.9, 129.9, 133.1, 133.4, 135.1, 137.0, 137.3, 137.5, 137.7, 137.8, 137.8, 137.9, 141.2, 143.6, 143.8, 155.9, 159.3, 164.5, 164.6, 165.0, 165.1, 165.5, 168.8, 168.9, 169.4, 169.9, 171.8, 172.3, 206.1, 206.5. MALDI-MS: $C_{186}H_{185}N_7O_{56}$ [M+Na]⁺ calcd: 3435.18, obsd: 3435.70.

N-(Acetyl)-O-(benzyl)-L-seryl-glycyl-O-[methyl] 2-O-levulinoyl-3-O-benzyl-4-O-pmethoxybenzyl- α -L-idopyranosyluronate- $(1 \rightarrow 4)$ -2-azido-3-O-benzyl-6-O-levulinoyl-2-2-O-benzoyl-3-O-benzyl-α-L $deoxy-\alpha$ -D-glucopyranosyl- $(1 \rightarrow 4)$ -methyl $idopvranosvluronate-(1 \rightarrow 4)-2-azido-3,$ 6-di-O-benzyl-2-deoxy-a-D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-acetyl-4, 6-0benzvlidene- β -D-galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-O-benzylidene-β-Dgalactopyranosyl- $(1 \rightarrow 4)$ -2, 3-di-O-benzoyl- β -D-xylopyranosyl]-L-serine benzyl ester (30). Compound S51 (31 mg, 0.009 mmol) was dissolved in DCM (0.6 mL), followed by addition of 46 µL piperidine. After 3 h, the mixture was diluted with DCM and extracted with H₂O. The combined organic phase was dried over Na₂SO₄. Column purification afforded compound **28** (20 mg, 70%). ¹H-NMR (500 MHz, CDCl₃): δ 2.04 (s, 3 H, CH₃COCH₂CH₂), 2.11 (s, 3 H, CH₃COCH₂CH₂), 2.33 (s, 3 H, COCH₃), 2.43-2.77 (m, 8 H, CH₃COCH₂CH₂), 3.07 (br, 1 H), 3.16-3.19 (m, 2 H), 3.22-3.34 (m, 8 H), 3.45-3.58 (m, 10 H), 3.64-4.02 (m, 20 H), 4.04-4.20 (m, 8 H), 4.35-4.42 (m, 3 H), 4.45-4.78 (m, 16 H), 4.84-4.88 (m, 3 H), 4.98 (s, 2 H), 5.09-5.15 (m, 3 H), 5.17-5.22 (m, 2 H), 5.31 (d, 1 H, J= 11 Hz), 5.36-5.38 (m, 1 H), 5.41-5.46 (m, 1 H), 5.52-5.57 (m, 2 H), 6.79-6.81 (m, 2 H), 7.07-7.54 (m, 62 H), 7.88-7.98 (m, 10 H). Compound 28 (22 mg, 0.0069 mmol) and compound 29 (4 mg, 0.0138 mmol) were dissolved in 0.6 mL DMF, followed by addition of HATU (5.2 mg, 0.0138 mmol) and DIPEA (4.8 µL, 0.0276 mmol). The resulting mixture was stirred under room temperature overnight and diluted with DCM. The solution was washed with 10% HCl, sat. NaHCO₃. The combined organic phase was dried
over Na₂SO₄. Column purification afforded compound **30** (18 mg, 77%). ¹H-NMR (500 MHz, CDCl₃): δ 1.95 (s, 3 H, CH₃COCH₂CH₂), 2.04 (s, 3 H, CH₃COCH₂CH₂), 2.33 (s, 3 H, COCH₃), 2.37-2.73 (m, 8 H, CH₃COCH₂CH₂), 2.78-2.93(m, 6 H), 3.12-3.38 (m, 8 H), 3.44-3.57 (m, 8 H), 3.69-3.97 (m, 18 H), 3.06-4.20 (m, 9 H), 4.38-4.89 (m, 23 H), 4.95-5.27 (m, 7 H), 5.30-5.55 (m, 6 H), 6.45-6.47 (m, 1 H), 6.67-6.71 (m, 1 H), 6.79-6.81 (m, 2 H), 6.93-6.96 (m, 1 H), 7.09-7.54 (m, 67 H), 7.88-7.99 (m, 10 H). ¹³C-NMR (125 MHz, CDCl₃): δ 20.1, 21.4, 23.0, 27.8, 27.9, 29.5, 29.7, 37.6, 37.9, 38.5, 42.8, 51.7, 51.9, 52.1, 52.5, 52.7, 55.2, 61.9, 62.2, 62.8, 63.1, 66.7, 67.1, 67.3, 68.3, 69.1, 69.1, 69.8, 70.8, 71.0, 71.2, 71.3, 71.5, 72.3, 72.9, 73.3, 73.4, 73.6, 73.7, 73.8, 74.5, 74.5, 74.6, 75.0, 75.3, 75.4, 75.4, 75.5, 75.5, 77.8, 77.9, 82.6, 97.2, 97.7, 98.0, 99.2, 100.1, 100.1, 100.4, 100.5, 102.2, 113.7, 125.2, 126.0, 126.3, 127.3, 127.4, 127.6, 127.6, 127.7, 127.7, 127.8, 127.8, 127.8, 127.9, 127.9, 127.9, 128.0, 128.0, 128.1, 128.2, 128.2, 128.2, 128.3, 128 128.4, 128.4, 128.5, 128.6, 128.7, 128.9, 129.0, 129.2, 129.3, 129.3, 129.5, 129.5, 129.5, 129.7, 129.7, 129.9, 130.0, 133.1, 133.1, 133.3, 133.4, 135.0, 137.0, 137.3, 137.4, 137.5, 137.7, 137.7, 137.8, 137.8, 137.8, 138.0, 159.4, 164.5, 164.6, 165.0, 165.1, 165.7, 168.3, 168.8, 169.0, 169.3, 169.9, 170.3, 170.4, 171.8, 172.3, 206.1, 206.5. MALDI-MS: $C_{185}H_{191}N_9O_{58}[M+Na]^+$ calcd: 3489.23, obsd: 3489.22.

N-(Acetyl)-O-(benzyl)-L-seryl-glycyl-O-[methyl] 2-O-levulinovl-3-O-benzyl-4-O-pmethoxybenzyl- α -L-idopyranosyluronate-(1 \rightarrow 4)-2-azido-3-O-benzyl-6-O-levulinoyl-2 $deoxy-\alpha$ -D-glucopyranosyl- $(1 \rightarrow 4)$ -methyl 2-O-benzovl-3-O-benzvl-α-L $idopyranosyluronate-(1 \rightarrow 4)-2-azido-3,$ 6-di-O-benzyl-2-deoxy-a-D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-acetyl-4, 6-0benzylidene- β -D-galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-O-benzylidene-β-Dgalactopyranosyl- $(1 \rightarrow 4)$ -2, 3-di-O-benzoyl- β -D-xylopyranosyl]-L-serine (31). Compound **30** (32 mg, 0.009 mmol) was dissolved in DCM/MeOH (1:1, 2 mL), followed by addition of Pd/C (3 mg) and NH₄OAc (3 mg, 0.032 mmol). The resulting mixture was stirred under H_2 atomosphere. The reaction was carefully monitored by TLC. After the complete disappearance of starting material, the reaction was diluted with DCM and filtered. After concentration, the residue was purified by silica gel column to afford compound 31 (25 mg, 82%). ¹H-NMR (500 MHz, CDCl₃): δ 1.14 (s, 3 H), 1.18 (s, 3 H), 1.97 (s, 3 H, CH₃COCH₂CH₂), 2.03 (s, 3 H, CH₃COCH₂CH₂), 2.38-2.63 (m, 8 H, CH₃COCH₂CH₂), 3.12-3.18 (m, 5 H), 3.20 (s, 3 H), 3.24-3.27 (m, 6 H), 3.29 (s, 3 H), 3.35-3.42 (m, 9 H), 3.54-3.58 (m, 4 H), 3.63-3.72 (m, 8 H), 3.75-4.19 (m, 20 H), 4.24-4.69 (m, 22 H), 4.76-4.78 (m, 3 H), 4.91-4.96 (m, 2 H), 5.01-5.10 (m, 3 H), 5.20-5.28 (m, 4 H), 5.39-5.44 (m, 2 H), 6.71-6.73 (m, 2 H), 6.96-7.23 (m, 48 H), 7.25-7.49 (m, 14 H), 7.75-7.88 (m, 10 H). ¹³C-NMR (125 MHz, CDCl₃): δ 27.6, 29.2, 29.4, 37.7, 51.6, 55.0, 55.1, 70.8, 72.1, 73.1, 73.6, 74.5, 75.3, 76.5, 97.6, 100.4, 113.5, 113.6, 125.7, 125.9, 126.1, 127.6, 127.7, 128.2, 128.9, 129.0, 129.2, 129.4, 129.5, 129.5, 133.3, 136.8, 137.1, 137.2, 137.4, 137.5, 137.6, 137.7, 159.2, 164.6, 164.9, 165.2, 165.3, 166.0, 168.7, 169.3, 169.8, 171.8, 172.3, 206.6, 207.2. MALDI-MS: C₁₇₈H₁₈₅N₉O₅₈ [M+Na]⁺ calcd: 3401.40, obsd: 3401.65.

Peptide **32**: Compound **32** was synthesized following the general procedure for solid phase peptide synthesis. MALDI-MS: $C_{63}H_{78}N_{10}O_{16}[M+Na]^+$ calcd: 1254.35, obsd: 1254.80. *Peptide* **33**: Compound **32** (6 mg, 0.00488 mmol) was dissolved in 1 mL dry DMF, followed by addition of BnBr (1.2 μ L, 0.00975 mmol) and DIPEA (1.7 μ L, 0.00975

mmol). The resulting mixture was stirred under room temperature overnight. The product was precipitated from *tert*-butyl methyl ether and purified by silica gel column to afford benzylated compound (4 mg, 60%). MALDI-MS: $C_{70}H_{84}N_{10}O_{16}$ [M+Na]⁺ calcd: 1344.47, obsd: 1344.10. This compound was dissolved in 0.5 mL DMF, followed by addition of 13 μ L piperidine. The resulting mixture was stirred under room temperature for 2 h and the product was precipitated from *tert*-butyl methyl ether and used directly for next step without further purification. MALDI-MS: $C_{55}H_{74}N_{10}O_{14}$ [M+H]⁺ calcd: 1099.54, obsd: 1099.90.

Glycopeptide **34**: Peptide **33** (13 mg, 0.011 mmol) and glycopeptide **31** (13 mg, 0.00385 mmol) were dissolved in 0.6 mL dry DMF, to which were added 2.2 mg HATU. The resulting mixture was stirred under room temperature overnight and diluted with DCM. The solution was washed with a saturated aqueous NaHCO₃ solution. The combined organic phase was dried over Na₂SO₄ and concentrated, purified by silica gel column to afford compound **34** (14 mg, 75%). ¹H-NMR (500 MHz, CDCl₃): δ 0.75-0.94 (m, 15 H), 1.91 (br, 37 H), 2.04 (s, 3 H), 2.11 (s, 3 H), 2.46-2.74 (m, 8 H, CH₃COCH₂CH₂), 3.19-3.34 (m, 8 H), 3.44-3.62 (m, 15 H), 3.68-3.98 (m, 23 H), 4.09-4.23 (m, 10 H), 4.35-4.78 (m, 22 H), 4.85-4.89 (m, 3 H), 5.02-5.21 (m, 6 H), 5.33-5.39 (m, 2 H), 5.54-5.57 (m, 1 H), 6.79-6.81 (m, 3 H), 7.01-7.48 (m, 80 H), 7.79-8.01 (m, 6 H). MALDI-MS: C₂₃₃H₂₅₇N₁₉O₇₁ [M+Na]⁺ calcd: 4479.71, obsd: 4479.45.



Synthesis of glycopeptide **35**. Reagents and conditions: (a) Zn, CuSO₄ (sat.), Ac₂O/THF/HOAc; (b) NH₂NH₂-H₂O, HOAc, DCM/MeOH; (c) SO₃-Et₃N, DMF, 55 °C. (d) H₂, Pd/C, DCM/MeOH.

Glycopeptide **S52**: Compound **34** (7 mg, 0.0016 mmol) was dissolved in THF/Ac₂O/HOAc (3:2:1, 1.5 mL), followed by addition of Zn (100 mg), CuSO₄ (saturated solution, 10 μ L). The resulting mixture was stirred under room temperature overnight. After filtration, the mixture was diluted with DCM and washed with sat. NaHCO₃. The combined organic phase was dried over Na₂SO₄ and purified by silica gel column to afford compound **S52** (5 mg, 62%). MALDI-MS: C₂₃₇H₂₆₅N₁₅O₇₃ [M+Na]⁺ calcd: 4514.70, obsd: 4514.91.

Glycopeptide **S53**: Compound **S53** was synthesized from compound **S52** following the general procedure for Lev deprotection. MALDI-MS: $C_{227}H_{253}N_{15}O_{69}$ [M+Na]⁺ calcd: 4318.50, obsd: 4318.75.

Glycopeptide **S54**: Compound **S54** was synthesized from compound **S53** following the general procedure for *O*-sulfation. ESI-MS: $C_{227}H_{251}N_{15}O_{75}S_2^{2^-}$ [M+Li-3H]²⁻ calcd: 2228.28, obsd: 2228.35.

Glycopeptide **35**: Compound **35** was synthesized from compound **S116** following the general procedure for global debenzylation. ESI-MS: $C_{135}H_{173}N_{15}O_{74}S_2^{2^-}[M-3H]^{3^-}$ calcd: 1084.32, obsd: 1084.93, $[M-4H]^{4^-}$ calcd: 812.99, obsd: 813.22.



Synthesis of Glycopeptide **36**. Reagents and conditions: (a) Zn, CuSO₄ (sat.), Ac₂O/THF/HOAc; (b) NH₂NH₂-H₂O, HOAc, DCM/MeOH; (c) SO₃-Et₃N, DMF, 55 °C; (d) Pd/C, H₂, DCM/MeOH; (e) NaOMe, MeOH, pH = 9.5.

6-di-O-benzyl-2-deoxy-α-D-glucopyranosyl $idopyranosyluronate-(1 \rightarrow 4)-2-N-acetyl-3$, $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-acetyl-4, 6-0benzylidene- β -D-galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-O-benzylidene-β-Dgalactopyranosyl- $(1 \rightarrow 4)$ -2, 3-di-O-benzovl- β -D-xylopvranosyl]-L-serine benzyl ester (S55). Compoud 30 (33 mg, 0.0095 mmol) was dissolved in THF/Ac₂O/HOAc (3:2:1, 6 mL), followed by addition of Zn (480 mg), CuSO₄ (saturated solution, 30 μ L). The resulting mixture was stirred under room temperature overnight. After filtration, the mixture was diluted with DCM and washed with sat. NaHCO₃. The combined organic phase was dried over Na₂SO₄ and purified by silica gel column to afford compound S55 (25 mg, 76%). ¹H-NMR (600 MHz, CDCl₃): δ 1.28 (s, 3 H), 1.30 (s, 3 H), 1.34 (s, 3 H), 1.92 (s, 3 H, CH₃COCH₂CH₂), 2.09 (s, 3 H, CH₃COCH₂CH₂), 2.41-2.72 (m, 8 H, CH₃COCH₂CH₂), 2.74 (s, 3 H), 2.82 (br, 1 H), 2.90 (br, 1 H), 3.12 (br, 1 H), 3.17-3.24 (m, 3 H), 3.26-3.93 (m, 66 H), 3.97-4.65 (m, 43 H), 4.72-4.78 (m, 2 H), 4.89-4.97 (m, 5 H), 5.02-5.07 (m, 5 H), 5.14-5.19 (m, 1 H), 5.24-5.30 (m, 5 H), 5.37-5.42 (m, 2 H), 5.49-5.52 (m, 1 H), 6.74-6.76 (m, 2 H), 6.97-7.51 (m, 66 H), 7.80-7.95 (m, 11 H). ¹³C-NMR (125 MHz, CDCl₃): δ 19.9, 20.6, 22.2, 22.6, 27.8, 29.5, 29.6, 36.5, 37.6, 37.8, 38.4, 42.4, 51.6, 51.9, 52.4, 52.6, 53.3, 55.1, 62.0, 65.0, 66.5, 66.9, 67.2, 68.2, 68.5, 69.2, 69.6, 70.1, 70.7, 71.2, 71.5, 72.0, 72.8, 72.9, 73.1, 73.2, 73.2, 73.9, 74.5, 74.9, 75.5, 78.3, 81.6, 97.6, 97.8, 98.6, 100.0, 100.4, 102.1, 113.6, 125.9, 126.2, 126.9, 127.1, 127.5, 127.6, 127.7, 127.7, 127.7, 127.8, 127.9, 128.0, 128.0, 128.1, 128.2, 128.2, 128.3, 128.3, 128.4, 128.4, 128.8, 129.4, 129.6, 129.8, 133.1, 133.3, 134.9, 136.1, 136.9, 137.3, 137.5, 137.6, 137.7, 138.2, 159.2, 162.9, 164.6, 165.1, 165.8, 168.3, 168.7, 169.0, 169.1, 169.6, 170.4, 170.8, 171.0, 171.9, 172.3, 174.5, 206.7, 207.1. MALDI-MS: $C_{189}H_{199}N_5O_{60}$ [M+Na]⁺ calcd: 3523.60, obsd: 3523.87.

N-(Acetyl)-O-(benzyl)-L-seryl-glycyl-O-[methyl] 3-O-benzyl-4-O-p-methoxybenzyl-a-Lidopyranosyluronate- $(1 \rightarrow 4)$ -2-N-acetyl-3-O-benzyl-2-deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ methyl 2-O-benzoyl-3-O-benzyl- α -L-idopyranosyluronate- $(1 \rightarrow 4)$ -2-N-acetyl-3, 6-di-Obenzyl-2-deoxy- α -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-benzoyl-3-O-benzyl- β -D-glucopyranosyl- $(1 \rightarrow 4)$ -2-O-acetyl-4, 6-O-benzylidene- β -D-galactopyranosyl- $(1 \rightarrow 3)$ -2-O-benzoyl-4, 6-Obenzylidene- β -D-galactopyranosyl- $(1 \rightarrow 4)$ -2, 3-di-O-benzoyl- β -D-xylopyranosyl]-L-serine benzyl ester (S56). Compound S56 was synthesized from compound S55 in 84% yield following the general procedure of Lev deprotection. ¹H-NMR (500 MHz, CDCl₃): δ 1.22 (s, 3 H), 1.28 (s, 3 H), 1.37 (s, 3 H), 2.00 (s, 3 H), 2.39-2.48 (m, 2 H), 3.13-3.43 (m, 21 H), 3.50-3.61 (m, 10 H), 3.65-3.88 (m, 30 H), 3.97-4.12 (m, 12 H), 4.19-4.70 (m, 38 H), 4.76-4.86 (m, 3 H), 4.88-4.90 (m, 1 H), 4.92-5.01 (m, 5 H), 5.03-5.21 (m, 8 H), 5.27-5.33 (m, 5 H), 5.40-5.54 (m, 3 H), 6.58-6.60 (m, 1 H), 6.76-6.79 (m, 3 H), 6.99-7.54 (m, 61 H), 7.84-7.97 (m, 14 H). ¹³C-NMR (125 MHz, CDCl₃): δ 17.8, 19.9, 22.3, 22.7, 22.8, 24.8, 25.8, 42.5, 51.7, 51.8, 52.0, 52.4, 52.4, 52.6, 55.2, 61.0, 62.0, 66.5, 67.0, 67.2, 68.1, 68.2, 68.5, 69.1, 70.1, 70.7, 71.2, 71.4, 72.2, 72.5, 72.6, 72.8, 72.9, 73.1, 73.2, 73.5, 74.0, 74.4, 74.6, 74.8, 75.2, 75.4, 75.5, 75.7, 78.0, 78.6, 81.5, 96.1, 97.8, 98.8, 100.0, 100.4, 100.9, 102.1, 113.7, 125.9, 126.3, 126.6, 126.9, 127.0, 127.1, 127.5, 127.6, 127.7, 127.8, 127.8, 127.9, 128.0, 128.0, 128.1, 128.1, 128.2, 128.3, 128.3, 128.4, 128.4, 128.5, 128.7, 128.8, 128.9, 129.0, 129.3, 129.4, 129.5, 129.6, 129.7, 129.8, 133.2, 133.4, 133.7, 134.9, 136.1, 136.8, 137.3, 137.3, 137.4, 137.6, 137.8, 138.1, 138.2, 159.4, 164.5, 164.6, 165.1, 165.8, 168.2,

168.5, 169.0, 169.1, 169.6, 170.0, 170.2, 170.5, 170.7. MALDI-MS: $C_{179}H_{187}N_5O_{56}$ [M+Na]⁺ calcd: 3327.40, obsd: 3327.95.

Glycopeptide **S57.** Compound **S57** was synthesized from compound **S56** in 90% yield following the general procedure of *O*-sulfation. ESI-MS: $C_{179}H_{185}N_5O_{62}S_2^{2^-}[M-2H]^{2^-}$ calcd: 1730.05, obsd: 1730.84.

Glycopeptide **S58.** Compound **S58** was synthesized from compound **S57** in 95% yield following the general procedure of global debenzylation. ESI-MS: $C_{101}H_{120}N_5O_{61}S_2^{3-}$ [M-2H]²⁻ calcd: 1221.79, obsd: 1222.28, [M-3H]³⁻ calcd: 814.19, obsd: 814.58.

Glycopeptide **36**: Compound **S58** (1 mg) was dissolved in freshly dried MeOH (0.5 mL). 0.5 M NaOMe in MeOH was freshly prepared and added dropwisely to the reaction to maintian pH around 9.5. The reaction was left under N₂ atomosphere for 50 h and quenched by 1 M HOAc. The mixture was loaded onto LH-20 column. Glycopeptide containing fractions were combined and concentrated to afford compound **36**. ESI-MS: $C_{59}H_{90}N_3O_{52}S_2^{3-}$ [M-2H]²⁻ calcd: 868.69, obsd: 868.68.

Glycopeptide **37**: Peptide **33** (4 mg, 0.004 mmol) and compound **36** (4 mg) were dissolved in 0.5 mL dry DMF, to which were added 2 mg HATU and 2 μ L 2, 4, 6-collidine. The resulting mixture was stirred under room temperature overnight. The solution was loaded onto LH-20 column. Glycopeptide containing fractions were combined and concentrated to afford compound **37**. ESI-MS: C₁₁₉H₁₇₁N₁₅O₆₈S₂²⁻ [M-2H]²⁻ calcd: 1480.99, obsd: 1481.28.

Glycopeptide 1: Compound **37** was treated following the general procedure for global debenzylation. ESI-MS: $C_{98}H_{151}N_{15}O_{68}S_2^{4-}$ [M-3H]³⁻ calcd: 896.94, obsd: 897.30. [M-4H]⁴⁻ calcd: 672.45, obsd: 672.99. The product was dissolved in MeOH/H₂O (1:1, 0.5 mL), to which 0.5 M NaOH solution was added to maintain pH around 9.5. The resulting mixture was left under room temperature for 3 h. The solution was loaded onto LH-20 column. Glycopeptide containing fractions were combined and concentrated to afford compound 1. ¹H-NMR (600 MHz, D₂O): δ 0.88-0.94 (m, 10 H), 1.24-1.43 (m, 20 H), 1.51-1.79 (m, 13 H), 2.03 (s, 3 H), 2.06 (s, 3 H), 2.11 (s, 3 H), 2.12-2.38 (m, 14 H), 3.32-4.69 (m, 63 H), 5.12-5.42 (m, 11 H). ESI-MS: $C_{95}H_{142}N_{15}O_{68}S_2^{7-}$ [M-6H+2NH₄]⁴⁻ calcd: 666.19, obsd: 665.45. [M-4H+NH₄]⁴⁻ calcd: 888.58, obsd: 888.19.

Probing the possibility of epimerization of glycosylated serine

A potential concern is the possible epimerization^[6] of the glycosylated serine in compound **31** during the most basic step in the synthesis, i.e., transesterification. Due to the complexity of **31**, it was extremely challenging to test whether epimerization occurred during this step. To probe this possibility, we carried out a model reaction using xylosylserine **S61** containing two *O*-benzoyl groups. **S61** was subjected to the same transesterification condition (pH 9.5, NaOMe/MeOH). Even after 75 hours, no epimerization product of **S62** was detected, which suggests the epimerization of **31** likely did not occur to a significant extent.



N-O-[2, 3-di-O-benzoyl-4-O-p-methoxybenzyl-\beta-D-xylopyranosyl]-L-serine benzyl ester (**S59**). Compound **S33** (101 mg, 0.115 mmol) was dissolved in DCM (4 mL), followed by addition of 471 µL piperidine. After 3 h, the mixture was diluted with DCM and extracted with H₂O. The combined organic phase was dried over Na₂SO₄. Column purification afforded compound **S59** (55 mg, 73%). ESI-MS: C₃₇H₃₇NO₁₀ [M+H]⁺ calcd: 656.24, obsd: 656.14.

N-*Acetyl-O-[2,3-di-O-benzoyl-4-O-p-methoxybenzyl-β-D-xylopyranosyl]-L-serine benzyl ester* (**S60**). Compound **S59** (106 mg, 0.162 mmol) was dissolved in pyridine (3 mL), followed by addition of 153 µL Ac₂O. After overnight, the mixture was diluted with DCM and extracted with 10% HCl, sat. NaHCO₃ solution and dried over Na₂SO₄. Column purification afforded compound **S60** (97 mg, 86%). ¹H-NMR (500 MHz, CDCl₃): δ 1.71 (s, 3 H, NHC*H*₃), 3.34-3.38 (m, 1 H), 3.63-3.67 (m, 1 H), 3.72-3.73 (m, 1 H), 3.74 (s, 3 H, PhOC*H*₃), 3.88-3.91 (m, 1 H), 4.29 (dd, 1 H, *J* = 2.5 Hz, *J* = 10 Hz), 4.47-4.53 (m, 2 H), 4.58 (d, 1 H, *J* = 6.5 Hz), 4.73-4.76 (m, 1 H), 5.09-5.18 (m, 3 H), 5.52 (t, 1 H, *J* = 8 Hz), 6.14 (d, 1 H, *J* = 8.5 Hz), 6.71-6.73 (m, 2 H), 7.11-7.13 (m, 2 H), 7.29-7.42 (m, 9 H), 7.48-7.54 (m, 2 H), 7.92-7.95 (m, 4 H).

N-Acetyl-O-[2, 3-di-O-benzoyl-β-D-xylopyranosyl]-L-serine (**S61**). Synthesis of **S61** from compound **S60** was achieved in quantitative yield followed the general procedure of global debenzylation. ¹H-NMR (500 MHz, CD₃OD): δ 1.59 (s, 3 H, NHC*H*₃), 3.36-3.38 (m, 1 H), 3.50-3.55 (m, 1 H), 3.94-4.01 (m, 2 H), 4.06-4.11 (m, 1 H), 4.14-4.18 (m, 1 H), 4.34-4.37 (m, 1 H), 4.79 (d, 1 H, *J* = 7.5 Hz), 5.17-5.21 (m, 1 H), 5.45-5.59 (m, 1 H), 7.39-7.45 (m, 4 H), 7.53-7.57 (m, 2 H), 7.93-7.96 (m, 4 H). ¹³C-NMR (125 MHz, CD₃OD): 22.4, 55.8, 66.8, 69.3, 70.4, 73.4, 76.9, 78.9, 79.2, 79.4, 102.0, 102.1, 129.4, 129.5, 130.6, 130.6, 130.7, 130.8, 134.3, 134.5, 166.8, 167.4, 172.7, 176.8. ESI-MS: C₂₄H₂₅NO₁₀ [M-H]⁺ calcd: 486.15, obsd: 486.00.

N-Acetyl-O-β-D-xylopyranosyl-L-serine (**S62**). Compound **S61** (4 mg) was dissolved in 1 mL dry MeOH followed by addition of freshly prepared 0.1 M NaOMe until the pH was 9.5. The resulting solution was stirred under room temperature for 75 h and quenched by 1 M HOAc. The mixture was concentrated in vacuo to afford compound **S62** in quantitative yield. ¹H-NMR (500 MHz, D₂O): δ 2.08 (s, 3 H, NHCH₃), 3.39-3.38 (m, 2 H), 3.46 (t, 1 H, J = 9 Hz), 3.61-3.67 (m, 1 H), 3.89-3.93 (m, 1 H), 3.96-4.00 (m, 1 H), 4.18-4.22 (m, 1 H), 4.40-4.45 (m, 2 H). ESI-MS: C₁₀H₁₇NO₈ [M-H]⁺ calcd: 278.10, obsd: 278.10.

References:

- [1] K. Bock, C. Pedersen, J. Chem. Soc., Perkin Trans. 2 1974, 293.
- [2] X. Ye, Y. Wang, L. Zhang, (Peking University, Peop. Rep. China). Chinese Patent Application: CN, **2006**, CN 1715286 A20060104, p. 18 pp.
- [3] Y. Huang, S. Dey, X. Zhang, F. Soennichsen, P. Garner, J. Am. Chem. Soc. 2004, 126, 4626.
- [4] G. Tiruchinapally, Z. Yin, M. El-Dakdouki, Z. Wang, X. Huang, *Chem. Eur. J.* **2011**, *17*, 10106.
- [5] Z. Wang, Y. Xu, B. Yang, G. Tiruchinapally, B. Sun, R. Liu, S. Dulaney, J. Liu, X. Huang, *Chem. Eur. J.* **2010**, *16*, 8365.
- [6] Y. Zhang, S. M. Muthana, D. Farnsworth, O. Ludek, K. Adams, J. J. Barchi, J. C. Gildersleeve, J. Am. Chem. Soc. 2012, 134, 6316-6325.

ESI-MS of 1





HPLC Chromatogram of glycopeptide 1

HPLC mobile phase: gradient 40% to 100% B in A over 30 min (solvent A: H_2O ; solvent B: CH_3CN). Flow rate: 1 mL/min. Detection Wavelength: 220 nm. HPLC column: SupelCOSIL LC-18, 25 cm x 4.6 mm, 5 μ m particle size.



















¹³C-NMR (CDCl₃, 125 MHz) of **6**









































gCOSY (CDCl₃, 500 MHz) of 9





1 H-NMR (CDCl₃, 500 MHz) of **13**



















gCOSY (CDCl₃, 500 MHz) of 14












gCOSY (CDCl₃, 500 MHz) of 18





¹H-NMR (CDCl₃, 500 MHz) of **19**





¹³C-NMR (CDCl₃, 150 MHz) of **21**











¹H-NMR (CDCl₃, 500 MHz) of **24**





¹³C-NMR (CDCl₃, 125 MHz) of **24**

















¹H-NMR (CDCl₃, 600 MHz) of **25**





¹³C-NMR (CDCl₃, 150 MHz) of **25**





gCOSY (CDCl₃, 600 MHz) of 25





gHMQC (CDCl₃, 600 MHz) of $\mathbf{25}$







gHMBC (CDCl₃, 600 MHz) of $\mathbf{25}$







¹³C-NMR (CDCl₃, 150 MHz) of **25a**





gCOSY (CDCl₃, 600 MHz) of 25a





gHMQC (CDCl₃, 600 MHz) of 25a









gHMBC (CDCl₃, 600 MHz) of 25a





 1 H-NMR (CDCl₃, 600 MHz) of **26**



¹³C-NMR (CDCl₃, 150 MHz) of **26**











¹H-NMR (CDCl₃, 600 MHz) of **27**





¹³C-NMR (CDCl₃, 150 MHz) of **27**











gHMBC (CDCl₃, 500 MHz) of 27




¹H-NMR (CDCl₃, 500 MHz) of **28**















¹³C-NMR (CDCl₃, 125 MHz) of **30**



¹H-NMR (CDCl₃, 500 MHz) of **31**





¹³C-NMR (CDCl₃, 125 MHz) of **31**





HPLC Chromatogram of crude peptide **32** HPLC mobile phase: gradient 5% to 100% B in A over 60 min (solvent A: 0.1% TFA in H₂O; solvent B: 0.1% TFA in CH₃CN). Flow rate: 1 mL/min. Detection Wavelenth: 220 nm. HPLC column: SupelCOSIL LC-18, 25 cm x 4.6 mm, 5 μ m particle size.













ESI-MS of 37





















gCOSY (CDCl₃, 500 MHz) of S4





gHMQC (CDCl₃, 500 MHz) of $\mathbf{S4}$





gHMBC (CDCl₃, 500 MHz) of S4

































gCOSY (CDCl₃, 500 MHz) of S7




















































gHMQC (CDCl₃, 500 MHz) of $\mathbf{S18}$

























































gCOSY (CDCl₃, 500 MHz) of **S23**















 1 H-NMR (CDCl₃, 500 MHz) of **S25**









gCOSY (CDCl₃, 500 MHz) of $\mathbf{S25}$













 1 H-NMR (CDCl₃, 500 MHz) of **S27**






































gHMQC (without ¹H decoupling) (CDCl₃, 600 MHz) of **S31**



gHMBC (CDCl₃, 600 MHz) of $\mathbf{S31}$









 $^{13}\text{C-NMR}$ (CDCl₃, 125 MHz) of S32





gCOSY (CDCl₃, 500 MHz) of $\mathbf{S32}$

















gCOSY (CDCl₃, 600 MHz) of **S34**







gHMQC (without ¹H decoupling) (CDCl₃, 600 MHz) of $\mathbf{S34}$















g COSY (CDCl_3, 500 MHz) of S37
































































gCOSY (CDCl₃, 500 MHz) of $\mathbf{S44}$



gHMQC (CDCl₃, 500 MHz) of $\mathbf{S44}$





gHMQC (without 1 H decoupling) (CDCl₃, 500 MHz) of S44



gHMBC (CDCl₃, 500 MHz) of S44

∠OBn







$g \text{COSY}\xspace(\text{CDCl}_3,\,600\ \text{MHz})$ of S45





¹H-NMR (CDCl₃, 500 MHz) of **S46**





¹³C-NMR (CDCl₃, 125 MHz) of **846**





gCOSY (CDCl₃, 500 MHz) of S46













¹H-NMR (CDCl₃, 500 MHz) of **S51**





¹³C-NMR (CDCl₃, 125 MHz) of **S51**





gCOSY (CDCl₃, 500 MHz) of S51











MALDI-MS of S53



 1 H-NMR (CDCl₃, 600 MHz) of **S55**



¹³C-NMR (CDCl₃, 150 MHz) of **S55**





 1 H-NMR (CDCl₃, 500 MHz) of **S56**





¹³C-NMR (CDCl₃, 125 MHz) of **S56**



















