

Supporting Information

Chemical Synthesis

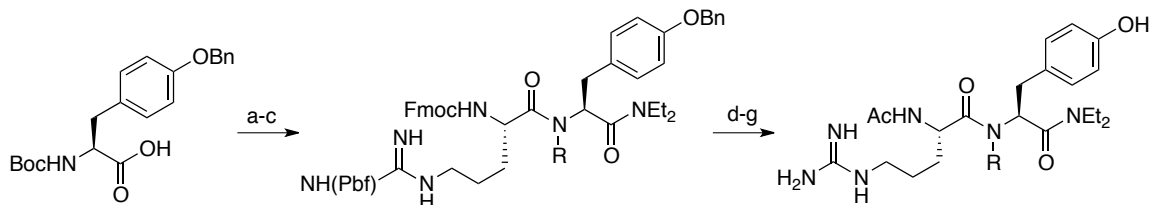
All reagents were obtained from Merck (Novabiochem), Sigma Aldrich or Fisher Scientific. Thin-layer chromatography was carried out on Merck silica gel aluminium-backed plates with a fluorescence indicator (254 nm). Visualisation of TLC plates was carried out by UV light, then by either a cerium sulphate/ammonium molybdate or KMnO_4 stain. Flash chromatography was performed with 35–70 μm silica gel (Fisher Davisil). Infrared spectra were recorded using a FTIR spectrometer fitted with an ATR accessory. Absorption peaks were recorded in cm^{-1} and were described as strong (s), medium (m), weak (w) or broad (br). ^1H NMR and ^{13}C NMR were recorded using a Bruker DPX400 (400 and 100 MHz) in $(\text{CD}_3)_2\text{SO}$ with DMSO (2.50 ppm ^1H , 39.51 ppm ^{13}C) as an internal reference. Chemical shifts δ are given in ppm; multiplicities are indicated as s (singlet), d (doublet), t (triplet), q (quartet), quin (quintet), m (multiplet), and br (broad); coupling constants, J, are reported in Hz. ^{13}C signal arising from the quaternary carbon bearing the boronic acid group was not observed and therefore were not listed, this was accordance with work by Hall et al. in "Mothana, S.; Grassot, J.; Hall, D. G. *Angew. Chem. -Int. Edit.* (2010), 49, 2883-2887".

Cyclic peptide synthesis

The linear counterparts of the CRYTFNV alanine-scanning analogues were synthesized at the Hershey Macromolecular Core Facility of the Pennsylvania State University. The linear peptides were cyclized and purified as previously described for CRYFNV in "A. Tavassoli, S. J. Benkovic, *Angew. Chem. Int. Ed. Engl.* (2005), 44, 2760-2763".

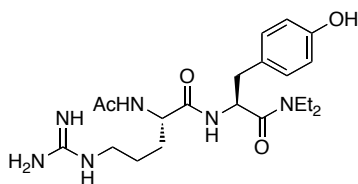
dipeptide synthesis

All capped dipeptides were synthesized on a similar scale as detailed below for compound 8, starting from the appropriate amino acids (from Merck Novabiochem or SigmaAldrich).



Scheme 1 Reagents and conditions: a) EDC, HOBT, Et₂NH, CH₂Cl₂; b) TFA/CH₂Cl₂; c) Fmoc-Arg(Pbf)-OH, EDC, HOBT, CH₂Cl₂; d) DBU, EtOAc; e) Ac₂O, Et₃N, CH₂Cl₂; f) 10% Pd/C, H₂, MeOH; g) TFA/TIS/H₂O.

Ac-Arg-Tyr-NEt₂



Chemical Formula: C₂₁H₃₄N₆O₄
 Exact Mass: 434.2642
 Molecular Weight: 434.5325

Synthesis of diethyl amide

To a stirred solution of N- α -t-Boc-O-benzyl-L-tyrosine (Novabiochem, Cat. No. 853065, 2.0 g, 5.38 mmol, 1 eq) dissolved in CH₂Cl₂ (30 ml) at room temperature, HOBT (730 mg, 5.38 mmol, 1 eq) and EDC.HCl (1.03 g, 5.38 mmol, 1 eq) were added, followed by diethyl amine (1.17 ml, 11.30 mmol, 2.1 eq). The reaction was stirred at room temperature for 4 hours, after which 10% citric acid (20 ml) was added. The organic layer was collected and the aqueous layer was re-extracted with CH₂Cl₂ (3 x 20 ml). The combined organic layers were washed with 10% citric acid (20 ml), saturated NaHCO₃ (20 ml) and brine (20 ml). The organic layer was dried (MgSO₄) and concentrated *in vacuo* to give a yellow oil (2.18 g, 5.12 mmol, 95 % crude yield) that was used without further purification.

Boc deprotection and peptide coupling

TFA (6 mL) was added to a stirred solution of Boc amine (1.0 g, 3.06 mmol) in CH₂Cl₂ (6 mL) at room temperature. The reaction was stirred at room temperature, and after 30 minutes the reaction was concentrated *in vacuo* to give a yellow oil, to which was added to a solution of N- α -Fmoc-N^G-(2,2,4,6,7-

pentamethyldihydrobenzofuran-5-sulfonyl)-L-arginine (NovaBiochem, Cat. No. 852067, 1.98 g, 3.06 mmol, 1 eq) in CH_2Cl_2 (25 ml), followed by HOBt (454 mg, 3.3 mmol, 1.1 eq) and EDC.HCl (632 mg, 3.3 mmol, 1.1 equiv). The reaction was stirred at room temperature, after 4 hours 10% citric acid (20 ml) was added. The organic layer was collected and the aqueous layer was re-extracted with CH_2Cl_2 (3 x 20 ml). The combined organic layers were washed with 10% citric acid (25 ml), saturated NaHCO_3 (25 ml) and brine (25 ml). The organic layer was dried (MgSO_4) and concentrated in vacuo to give a yellow oil that was purified by column chromatography (SiO_2 , 75 % EtOAc-hexane) to give a white solid (1.36 g, 1.42 mmol, 46 % yield) that was characterized by mass spectroscopy.

Fmoc deprotection and acetylation

At room temperature, to a stirred solution of Fmoc protected amine (300 mg, 0.31 mmol) dissolved in EtOAc was added DBU (1.05 eq). The reaction was stirred at room temperature, after 30 minutes the reaction was concentrated in vacuo to give a yellow oil. Purification by SiO_2 (short- path) eluting with MeOH 1-10%: CH_2Cl_2 gave a yellow oil that was dissolved in CH_2Cl_2 (10 ml). To the stirred solution at room temperature, Ac_2O (47 μl , 0.34 mmol, 1.1 eq) and triethylamine (38 μl , 0.34 mmol, 1.05 eq) were added. After 15 hours the reaction was concentrated in vacuo and purified by column chromatography (SiO_2 , 7% MeOH- CH_2Cl_2) to give a to give a yellow oil (209 mg, 0.27 mmol, 87 % yield).

Debenzylation

At room temperature under an atmosphere of H_2 , a suspension of the protected dipeptide (100 mg, 0.13 mmol) and 10% Pd/C (18 mg, 0.012 mmol, 0.1 eq) in MeOH (10 ml) was stirred for 15 hours. The reaction mixture was filtered through celite and concentrated in vacuo to give a yellow oil (77 mg) that was used in the next step without further purification.

Side chain deprotection

At room temperature, a solution containing TFA (4.75 ml), TIS (0.125 ml) and H_2O 0.125 ml) at a % ratio of 95:2.5:2.5 was added to the dipeptide (77mg, 0.11 mmol). The reaction was stirred at room temperature,

after 2 hours the reaction was concentrated in vacuo followed by diethyl ether precipitation to give a white solid. Purification by RP-HPLC using a Waters Alantis Prep T3 OBD 5 μ m 19 x 100 mm column at a 17 mL min⁻¹ elution with a 95% A 5% B \rightarrow 30% A 70% B gradient over 10 minutes (A = water with 1% TFA. B = acetonitrile with 1% TFA) gave the desired dipeptide after lyophilisation as a white solid (25 mg, 52%).

$[\alpha]_{26D} -28.5$ (MeOH, c 0.53).

IR ν_{max} (neat) 3272 (m), 3192 (m), 1614 (s), 1541 (s), 1514 (s) cm⁻¹.

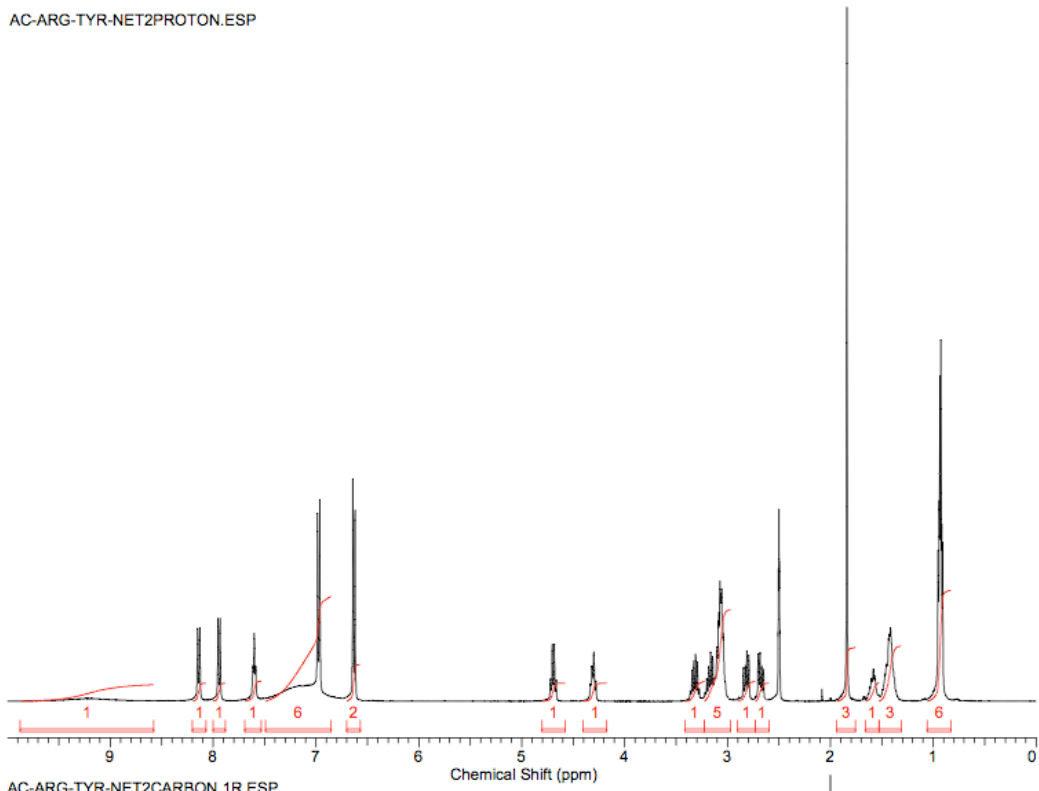
¹H NMR (400 MHz, DMSO-d₆) δ 9.85 - 8.66 (br s, 1H), 8.14 (d, J = 7.5 Hz, 1H), 7.94 (d, J = 8.1 Hz, 1H), 7.60 (t, J = 5.6 Hz, 1H), 6.97 (d, J = 8.3 Hz, 2H), 7.54 - 6.86 (m, 3H), 6.63 (d, J = 8.3 Hz, 2H), 4.69 (dt, J = 6.8, 7.5 Hz, 1H), 4.31 (dt, J = 6.1, 8.1 Hz, 1H), 3.32 (dd, J = 7.0, 13.4 Hz, 1H), 3.23 - 2.99 (m, 5H), 2.82 (dd, J = 7.5, 13.2 Hz, 1H), 2.68 (dd, J = 6.8, 13.2 Hz, 1H), 1.84 (s, 3H), 1.65 - 1.51 (m, 1H), 1.51 - 1.33 (m, 3H), 0.94 (t, J = 7.0 Hz, 3H), 0.93 (t, J = 7.0 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 170.9, 169.9, 169.1, 156.7, 156.0, 130.2, 127.2, 114.9, 51.6, 50.2, 41.0, 40.4, 39.7, 37.4, 29.3, 24.9, 22.5, 14.0, 12.7.

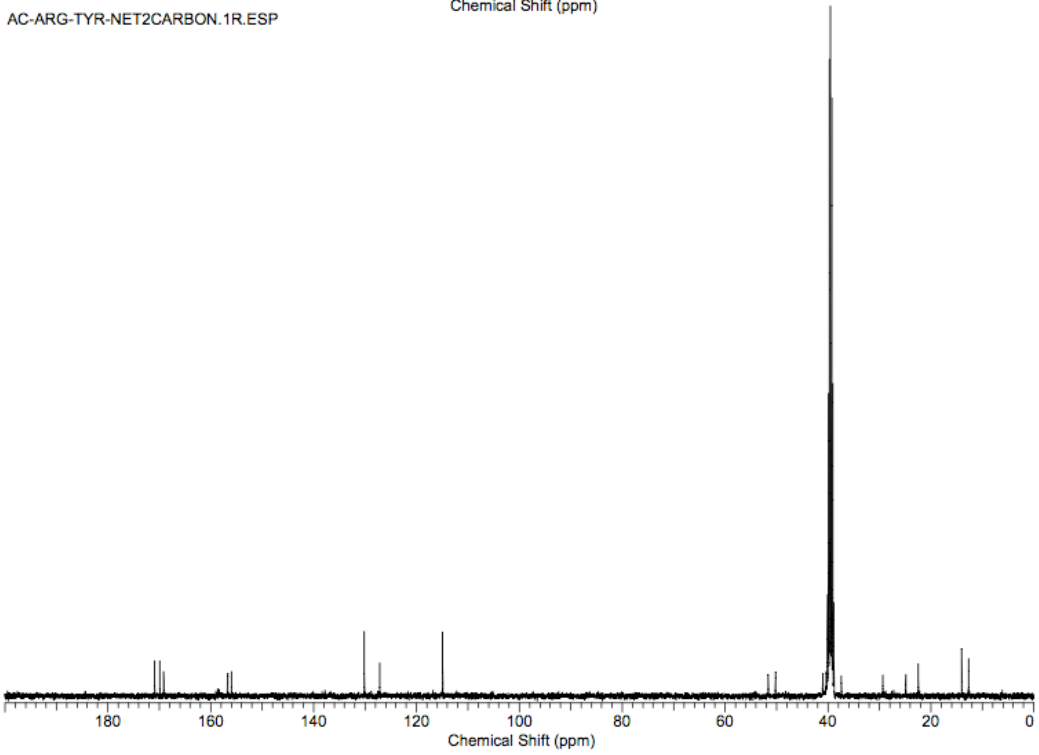
LRMS [M+H]⁺ 435 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 435.2714, [M+H]⁺ found = 435.2711.

AC-ARG-TYR-NET2PROTON.ESP

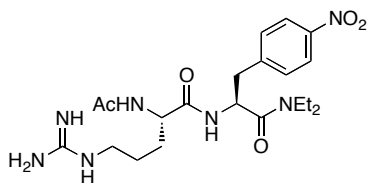


AC-ARG-TYR-NET2CARBON.1R.ESP



Ac-Arg-Phe(4-NO₂)-NEt₂

The protocol outlined above was repeated on a similar scale, with the appropriate amino acids, with an overall yield of 23%.



Chemical Formula: C₂₁H₃₃N₇O₅
Exact Mass: 463.2543
Molecular Weight: 463.5306

[α]_D²⁷ -31.6 (MeOH, c 0.55).

IR ν_{max} (neat) 3278 (m), 3193 (m), 2979 (w), 2943 (w), 1621 (s), 1519 (s) cm⁻¹.

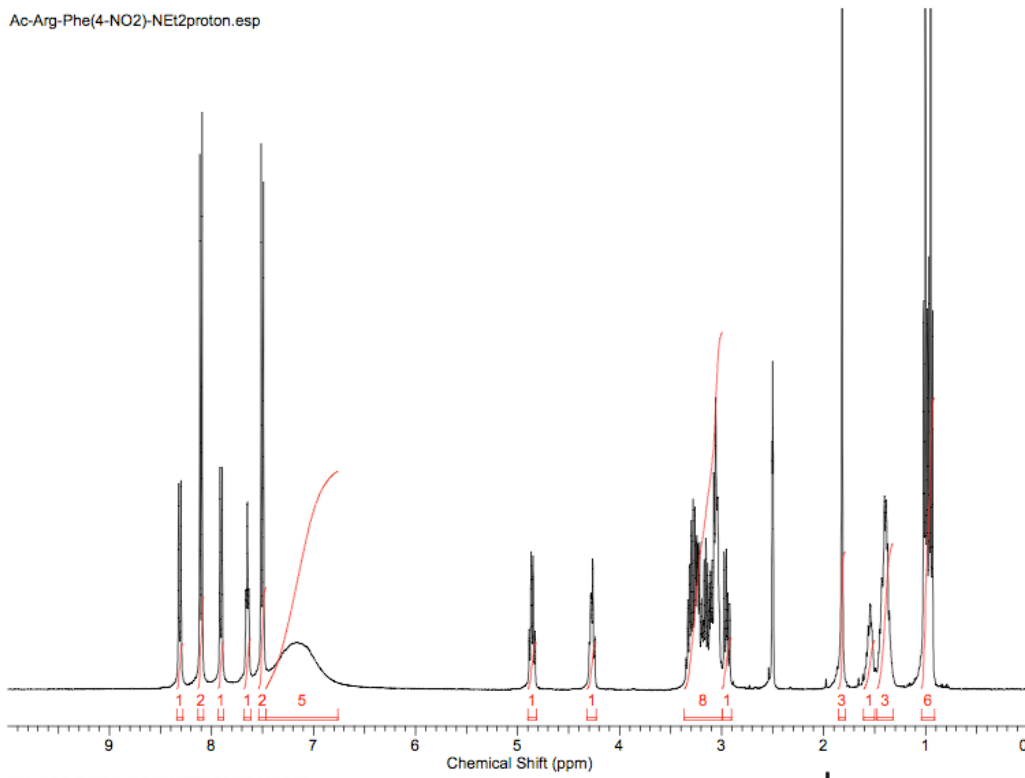
¹H NMR (400 MHz, DMSO-d₆) δ 8.31 (d, J = 8.6 Hz, 1H), 8.10 (d, J = 8.6 Hz, 2H), 7.91 (d, J = 8.1 Hz, 1H), 7.65 (t, J = 5.6 Hz, 1H), 7.50 (d, J = 8.6 Hz, 2H), 7.17 (br. s., 3H), 4.90 - 4.82 (m, 1H), 4.32 - 4.23 (m, 1H), 3.37 - 3.00 (m, 7H), 2.95 (dd, J = 8.1, 13.6 Hz, 1H), 1.82 (s, 3H), 1.61 - 1.50 (m, 1H), 1.48 - 1.32 (m, 3H), 1.00 (t, J = 7.1 Hz, 3H), 0.95 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.1, 169.1, 169.1, 156.8, 146.3, 145.8, 130.8, 123.1, 51.8, 49.4, 41.1, 40.4, 39.8, 37.6, 29.2, 25.0, 22.4, 14.2, 12.7.

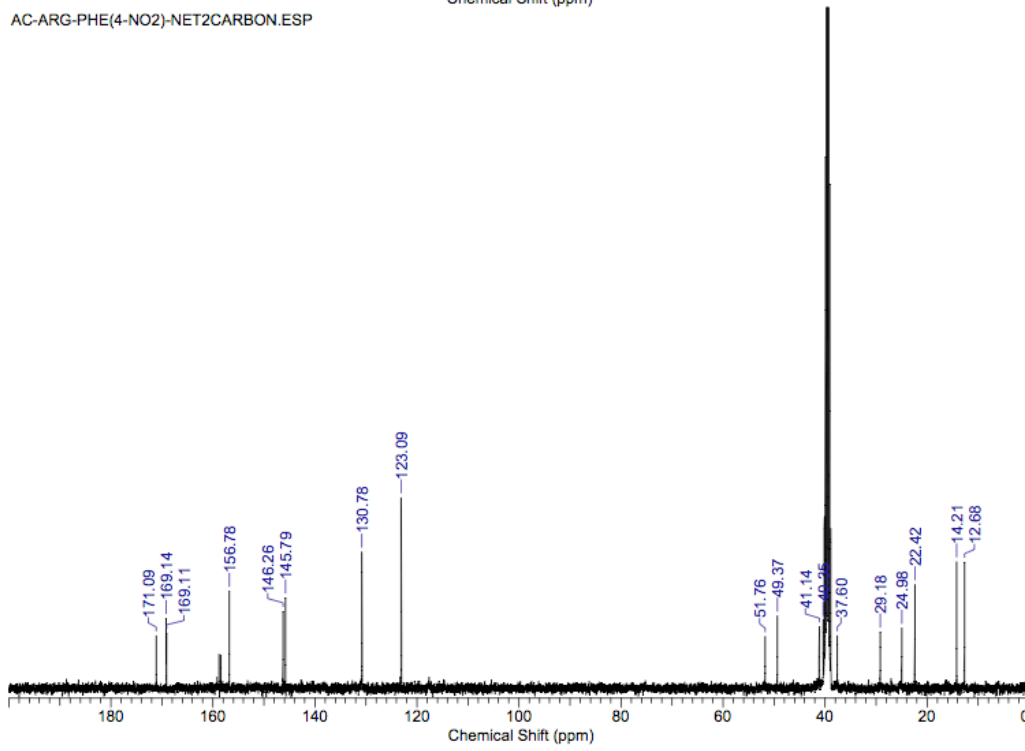
LRMS [M+H]⁺ 464 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 464.2616, [M+H]⁺ found = 464.2610.

Ac-Arg-Phe(4-NO2)-NEt2proton.esp

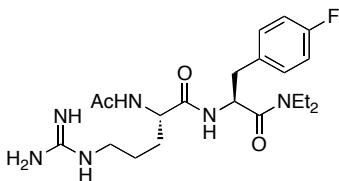


AC-ARG-PHE(4-NO2)-NET2CARBON.ESP



Ac-Arg-Phe(4-F)-NEt₂

The above protocol was followed, on a similar scale with the appropriate amino acids. The overall yield of synthesis was 19%.



Chemical Formula: C₂₁H₃₃FN₆O₃
Exact Mass: 436.2598
Molecular Weight: 436.5235

[α]_D²⁷ -29.1 (MeOH, c 0.54).

IR ν_{max} (neat) 3274 (m), 1618 (s), 1590 (s), 1509 (s) cm⁻¹.

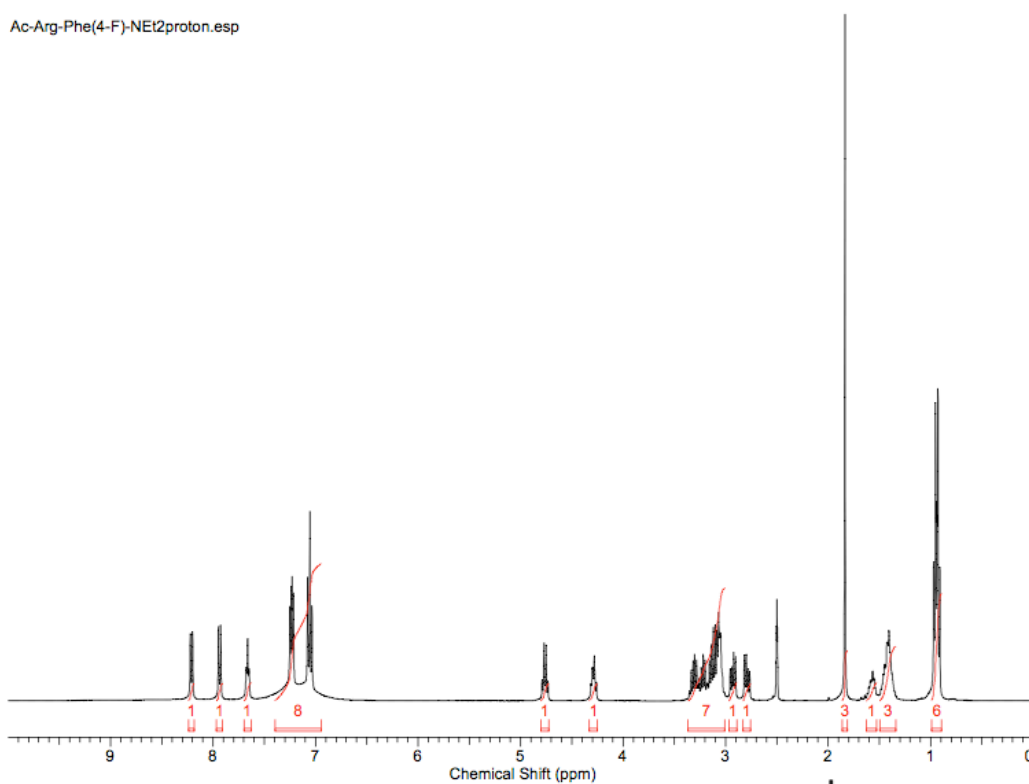
¹H NMR (400 MHz, DMSO-d₆) δ 8.21 (d, J = 8.1 Hz, 1H), 7.94 (d, J = 8.1 Hz, 1H), 7.66 (t, J = 5.6 Hz, 1H), 7.23 (dd, J = 6.1, 8.6 Hz, 2H), 7.06 (t, J = 9.1 Hz, 2H), 7.40 - 6.94 (br s, 3H), 4.76 (ddd, J = 7.1, 7.3, 8.1 Hz, 1H), 4.33 - 4.25 (m, 1H), 3.37 - 3.01 (m, 6H), 2.93 (dd, J = 7.1, 13.3 Hz, 1H), 2.79 (dd, J = 7.3, 13.3 Hz, 1H), 1.84 (s, 3H), 1.63 - 1.52 (m, 1H), 1.49 - 1.34 (m, 3H), 0.95 (t, J = 7.1 Hz, 3H), 0.92 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.0, 169.6, 169.2, 162.3, 159.9, 156.8, 133.4, 133.4, 131.2, 131.1, 114.9, 114.7, 51.7, 49.9, 41.1, 40.4, 39.8, 37.2, 29.3, 24.9, 22.4, 14.1, 12.7.

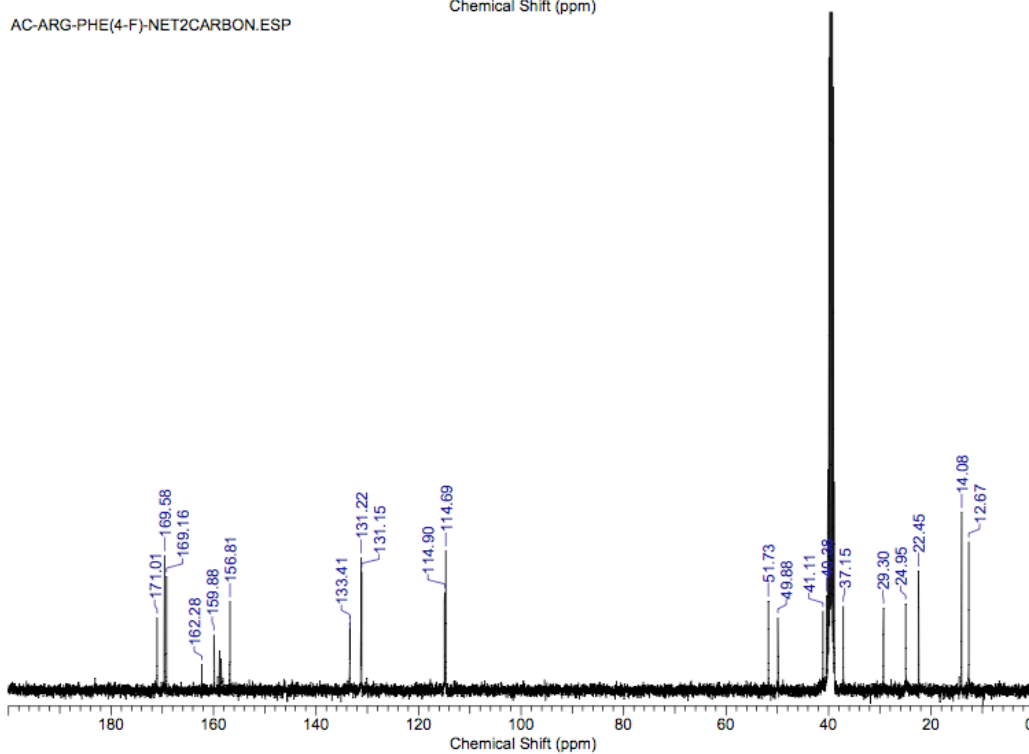
LRMS [M+H]⁺ 437 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 437.2671, [M+H]⁺ found = 437.2666.

Ac-Arg-Phe(4-F)-NET2proton.esp

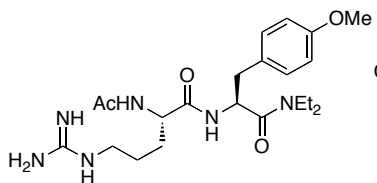


AC-ARG-PHE(4-F)-NET2CARBON.ESP



Ac-Arg-Tyr(Me)-NEt₂

The above procedure was repeated on a similar scale, with the appropriate amino acids, with a total yield of 22%.



Chemical Formula: C₂₂H₃₆N₆O₄
Exact Mass: 448.2798
Molecular Weight: 448.5590

[α]_D²⁷ -22.5 (MeOH, c 0.54).

IR ν_{max} (neat) 3273 (m), 1614 (s), 1539 (s), 1512 (s) cm⁻¹.

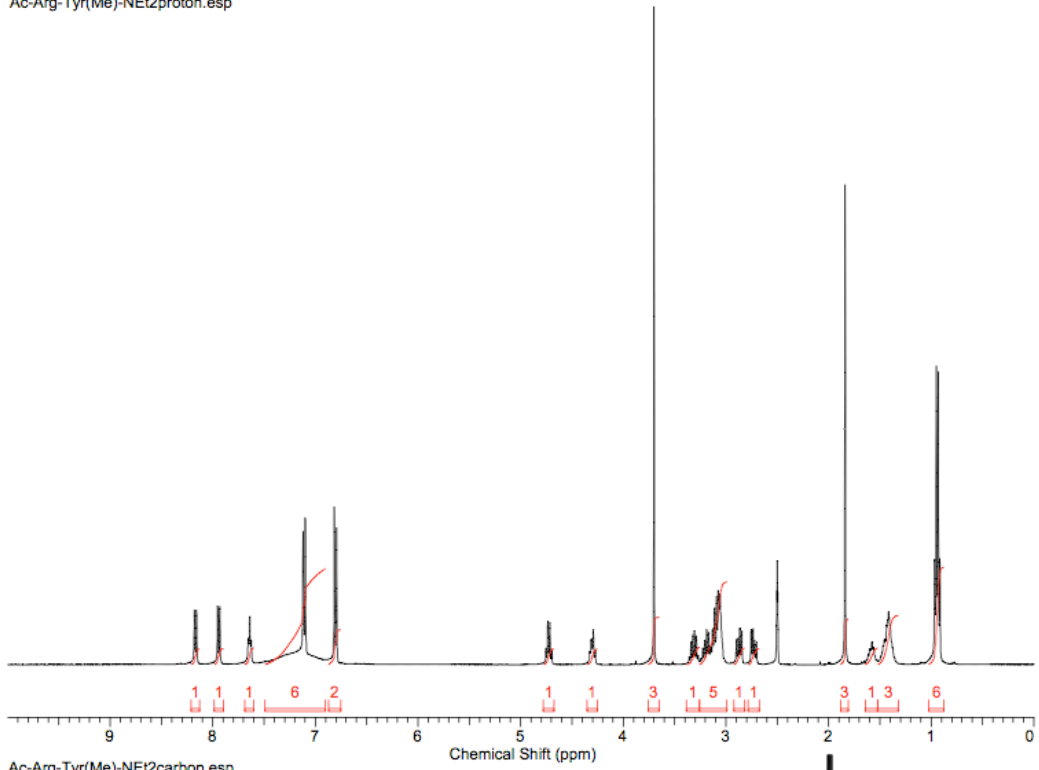
¹H NMR (400 MHz, DMSO-d₆) δ 8.17 (d, J = 8.1 Hz, 1H), 7.94 (d, J = 8.6 Hz, 1H), 7.64 (t, J = 5.6 Hz, 1H), 7.11 (d, J = 8.6 Hz, 2H), 7.49 - 6.93 (m, 3H), 6.80 (d, J = 8.6 Hz, 2H), 4.73 (ddd, J = 7.1, 7.3, 8.1 Hz, 1H), 4.35 - 4.25 (m, 1H), 3.70 (s, 3H), 3.39 - 3.26 (m, 1H), 3.26 - 2.99 (m, 5H), 2.87 (dd, J = 7.3, 13.2 Hz, 1H), 2.73 (dd, J = 7.1, 13.2 Hz, 1H), 1.84 (s, 3H), 1.64 - 1.53 (m, 1H), 1.51 - 1.34 (m, 3H), 0.99 - 0.89 (m, 6H).

¹³C NMR (100 MHz, DMSO-d₆) δ 170.9, 169.8, 169.1, 158.0, 156.8, 130.3, 129.1, 113.5, 55.0, 51.7, 50.1, 41.1, 40.4, 39.7, 37.3, 29.3, 24.9, 22.5, 14.0, 12.7.

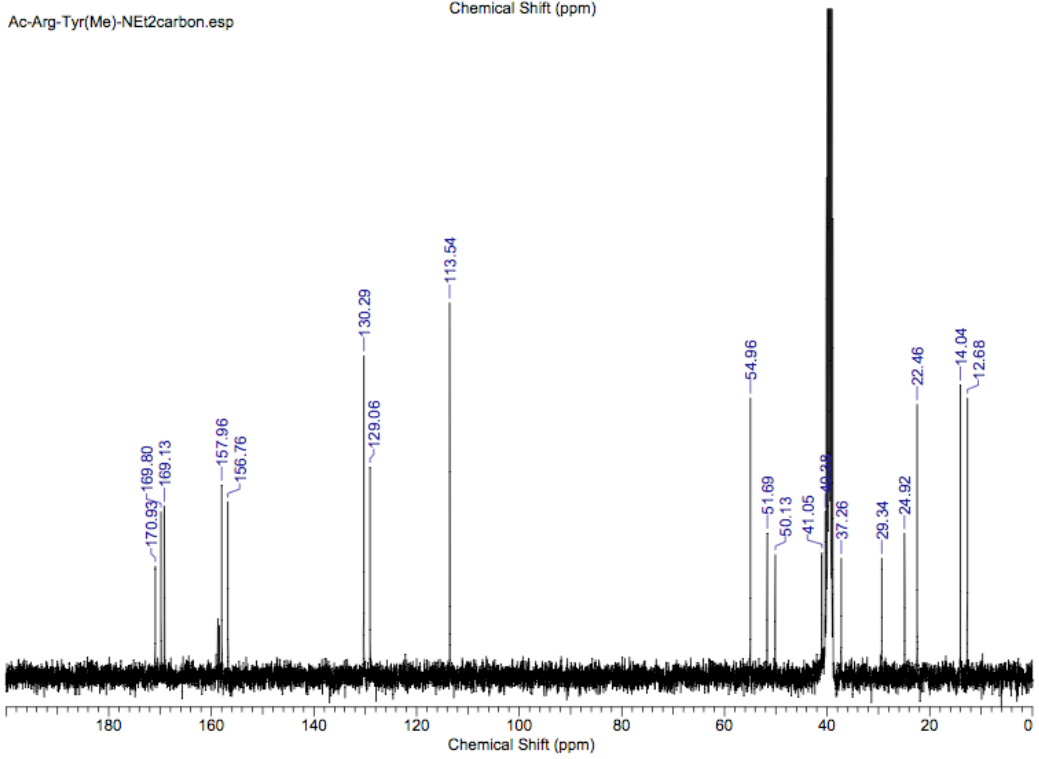
LRMS [M+H]⁺ 449 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 449.2871, [M+H]⁺ found = 449.2868.

Ac-Arg-Tyr(Me)-NEt2proton.esp

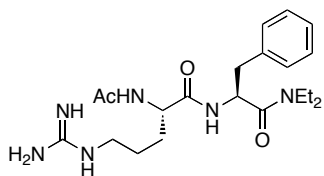


Ac-Arg-Tyr(Me)-NEt2carbon.esp



Ac-Arg-Phe-NEt₂

The above procedure was repeated on a similar scale, with the appropriate amino acids, with an overall yield of 19%.



Chemical Formula: C₂₁H₃₄N₆O₃
Exact Mass: 418.2692
Molecular Weight: 418.5331

[α]_D²⁵ -25.6 (MeOH, c 0.57).

IR ν_{max} (neat) 3273 (m), 1777 (w), 1618 (s), 1538 (s) cm⁻¹.

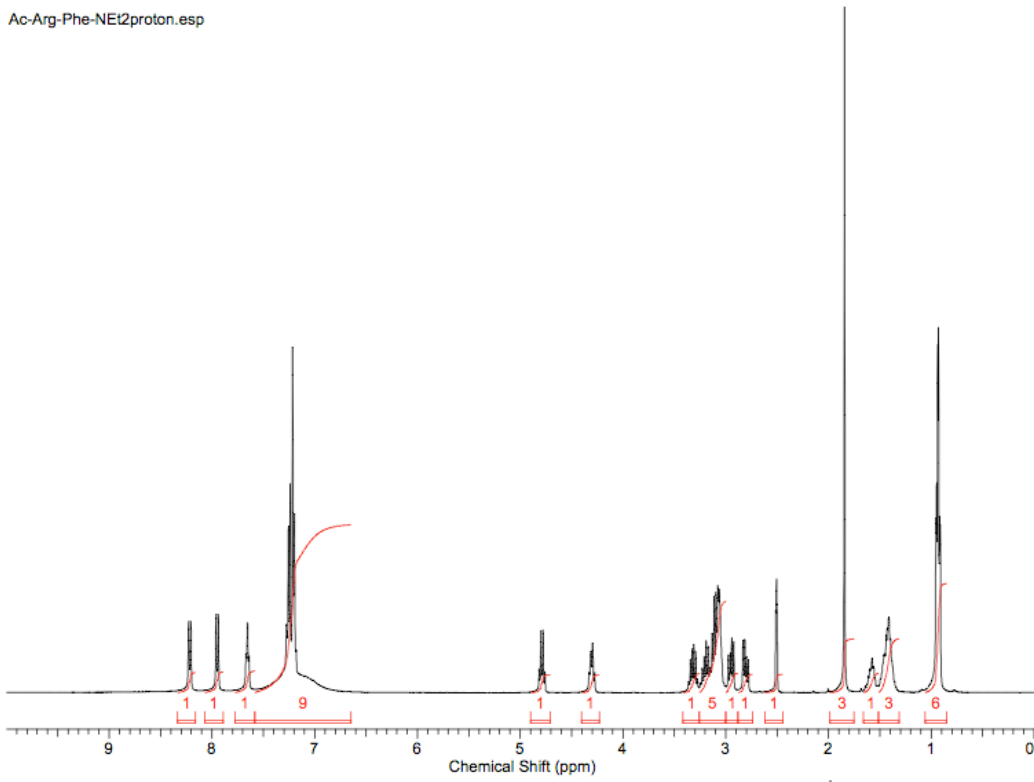
¹H NMR (400 MHz, DMSO-d₆) δ 8.22 (d, J = 8.1 Hz, 1H), 7.95 (d, J = 8.1 Hz, 1H), 7.65 (t, J = 5.3 Hz, 1H), 7.46 - 7.02 (m, 8H), 4.79 (ddd, J = 8.1, 7.3, 7.1 Hz, 1H), 4.35 - 4.26 (m, 1H), 3.42 - 3.26 (m, 1H), 3.26 - 3.00 (m, 5H), 2.95 (dd, J = 7.3, 13.5 Hz, 1H), 2.81 (dd, J = 7.1, 13.5 Hz, 1H), 1.84 (s, 3H), 1.66 - 1.51 (m, 1H), 1.51 - 1.33 (m, 3H), 0.94 (t, J = 7.1 Hz, 3H), 0.93 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.1, 169.1, 169.1, 156.8, 146.3, 145.8, 130.8, 123.1, 51.8, 49.4, 41.1, 40.4, 39.8, 37.6, 29.2, 25.0, 22.4, 14.2, 12.7.

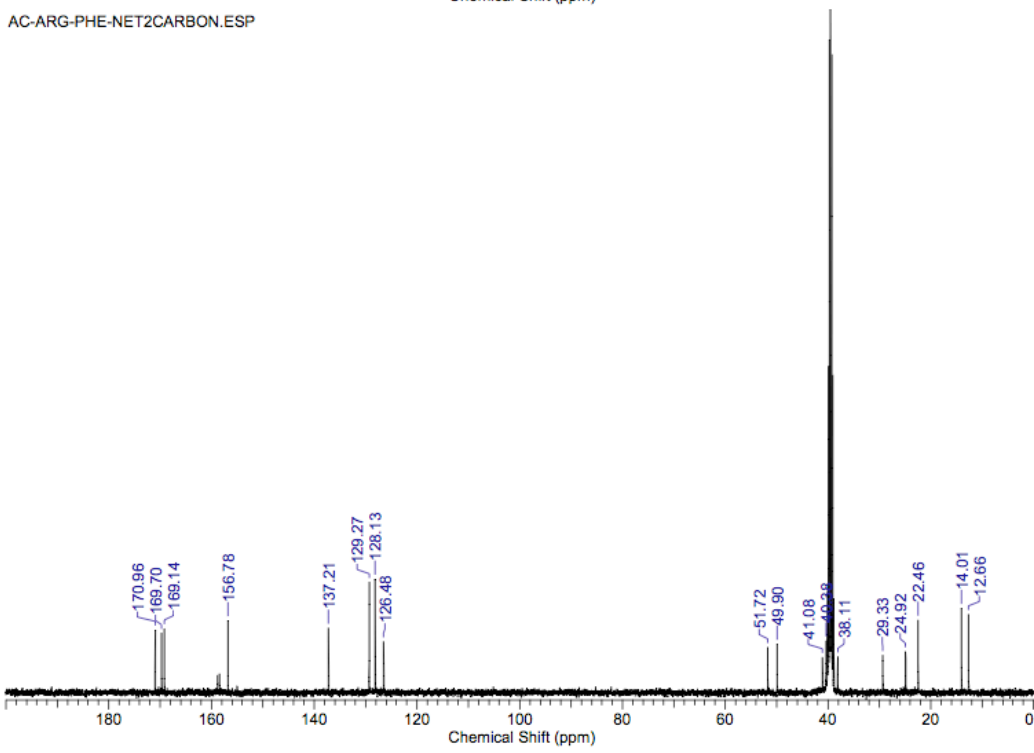
LRMS [M+H]⁺ 419 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 419.2765, [M+H]⁺ found = 419.2760.

Ac-Arg-Phe-NEt2proton.esp

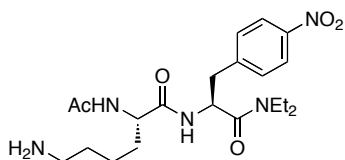


AC-ARG-PHE-NET2CARBON.ESP



Ac-Lys-Phe(4-NO₂)-NEt₂

The above protocol was repeated with the appropriate amino acids, on a similar scale, with an overall yield of 25%.



Chemical Formula: C₂₁H₃₃N₅O₅
Exact Mass: 435.2482
Molecular Weight: 435.5172

[α]_D²⁶ -46.9 (MeOH, c 0.48).

IR ν_{max} (neat) 3272 (m), 3077 (w), 2941 (m), 1776 (w), 1621 (s), 1519 (s) cm⁻¹.

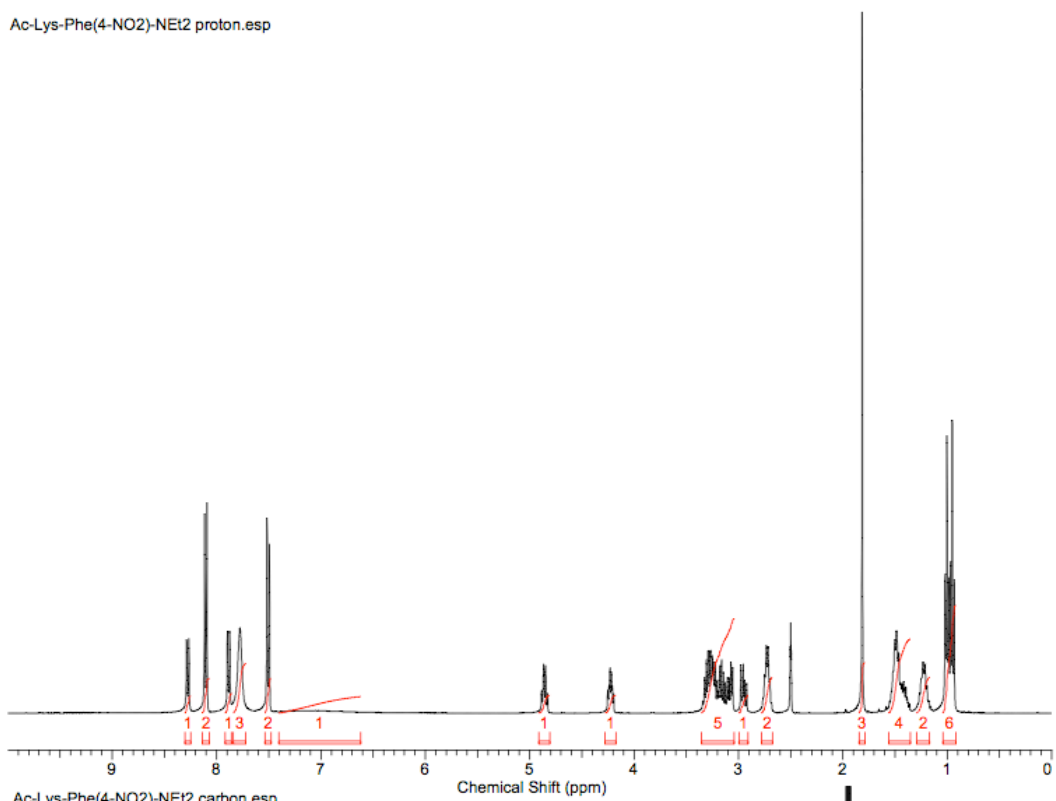
¹H NMR (400 MHz, DMSO-d₆) δ 8.28 (d, J = 8.6 Hz, 1H), 8.10 (d, J = 8.6 Hz, 2H), 7.88 (d, J = 8.6 Hz, 1H), 7.78 (br. s., 2H), 7.51 (d, J = 8.6 Hz, 2H), 7.01 (br. s., 1H), 4.86 (ddd, J = 6.3, 8.1, 8.6 Hz, 1H), 4.22 (dt, J = 5.1, 8.6 Hz, 1H), 3.35 - 3.12 (m, 4H), 3.08 (dd, J = 6.3, 13.4 Hz, 1H), 2.95 (dd, J = 8.1, 13.4 Hz, 1H), 2.78 - 2.67 (m, 2H), 1.81 (s, 3H), 1.56 - 1.35 (m, 4H), 1.29 - 1.17 (m, 2H), 1.00 (t, J = 7.1 Hz, 3H), 0.95 (t, J = 7.0 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.3, 169.2, 169.1, 146.3, 145.8, 130.8, 123.1, 52.0, 49.3, 41.1, 39.0, 38.7, 37.6, 31.4, 26.6, 22.5, 22.2, 14.2, 12.7.

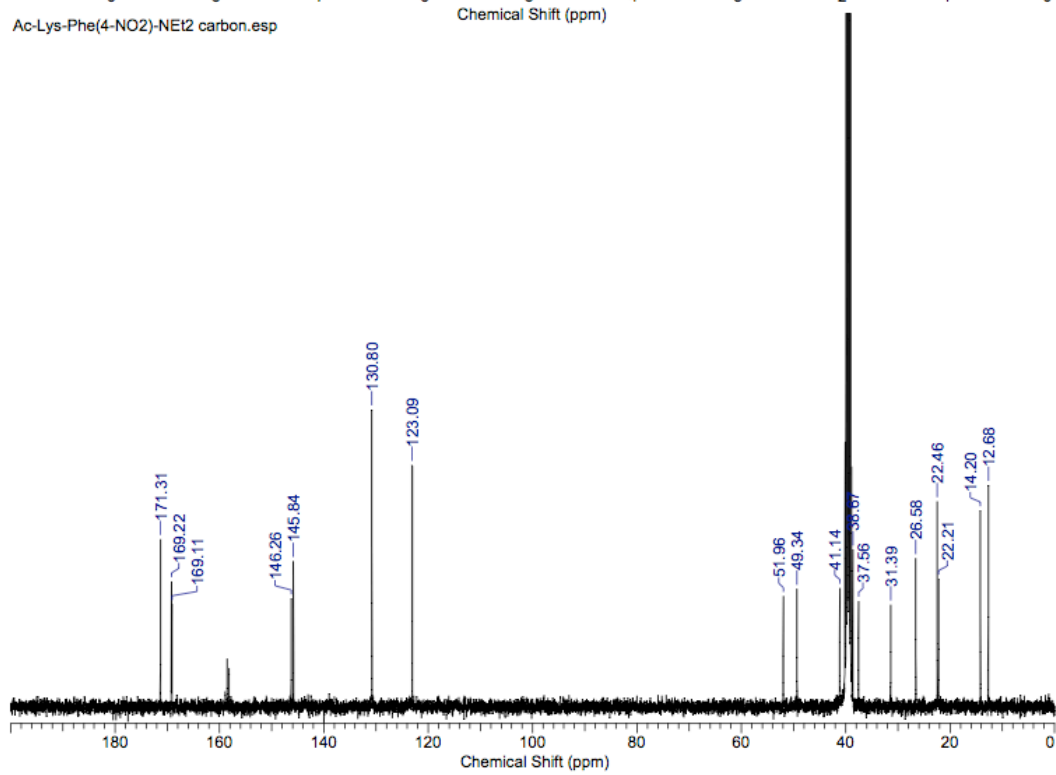
LRMS [M+H]⁺ 436 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 435.2482, [M+H]⁺ found = 436.2559.

Ac-Lys-Phe(4-NO2)-NEt2 proton.esp

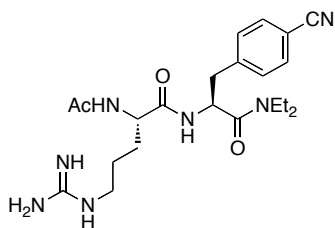


Ac-Lys-Phe(4-NO2)-NEt2 carbon.esp



Ac-Arg-Phe(4-CN)-NEt₂

The above procedure was followed at a similar concentration, with the appropriate amino acids,, with an overall yield of 22%.



Chemical Formula: C₂₂H₃₃N₇O₃
Exact Mass: 443.2645
Molecular Weight: 443.5425

[α]_D²⁶ -31.6 (MeOH, c 0.51).

IR ν_{max} (neat) 3278 (m), 2230 (w), 1777 (w), 1620 (s), 1538 (s) cm⁻¹.

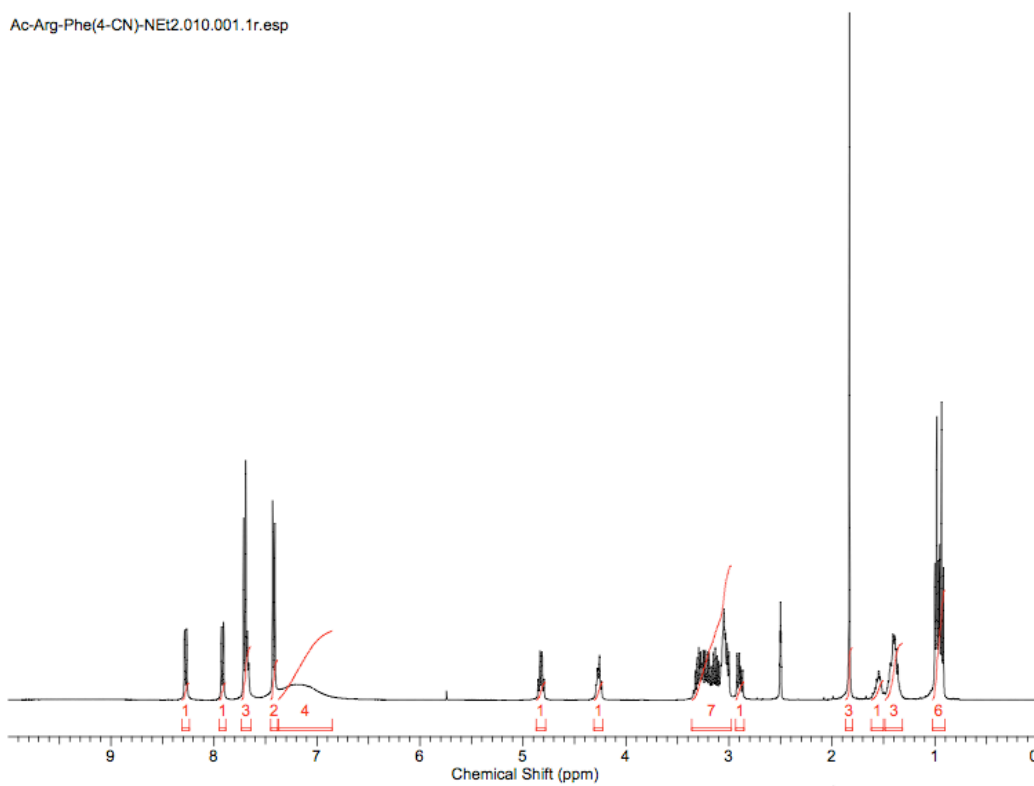
¹H NMR (400 MHz, DMSO-d₆) δ 8.28 (d, J = 8.6 Hz, 1H), 7.92 (d, J = 8.6 Hz, 1H), 7.74 - 7.64 (m, 3H), 7.42 (d, J = 8.1 Hz, 2H), 7.23 (br. s., 3H), 4.87 - 4.77 (m, 1H), 4.31 - 4.22 (m, 1H), 3.37 - 2.98 (m, 7H), 2.89 (dd, J = 7.8, 13.4 Hz, 1H), 1.83 (s, 3H), 1.63 - 1.50 (m, 1H), 1.49 - 1.31 (m, 3H), 0.98 (t, J = 7.1 Hz, 3H), 0.94 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.1, 169.2, 169.2, 156.8, 143.4, 131.9, 130.5, 118.9, 109.3, 51.7, 49.4, 41.1, 40.4, 39.8, 37.9, 29.2, 25.0, 22.4, 14.2, 12.7.

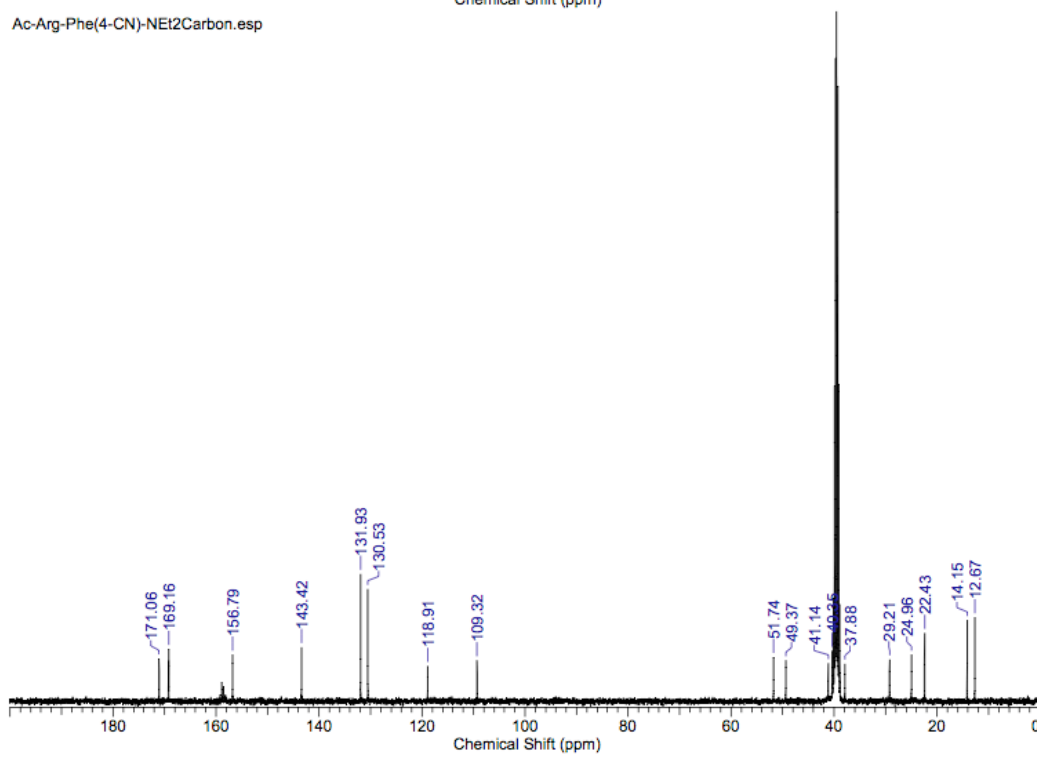
LRMS [M+H]⁺ 444 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 444.2718, [M+H]⁺ found = 444.2723.

Ac-Arg-Phe(4-CN)-NEt2.010.001.1r.esp

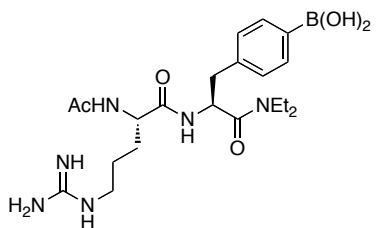


Ac-Arg-Phe(4-CN)-NEt2Carbon.esp



Ac-Arg-Phe(4-B(OH)₂)-NEt₂

The above procedure was repeated with the appropriate amino acids, on a similar scale, to give an overall yield of 13%.



Chemical Formula: C₂₁H₃₅BN₆O₅
Exact Mass: 462.2762
Molecular Weight: 462.3508

[α]_D²⁶ -21.8 (MeOH, c 0.50).

IR ν_{max} (neat) 3289 (m), 2981 (w), 1777 (w), 1614 (s), 1539 (s) cm⁻¹.

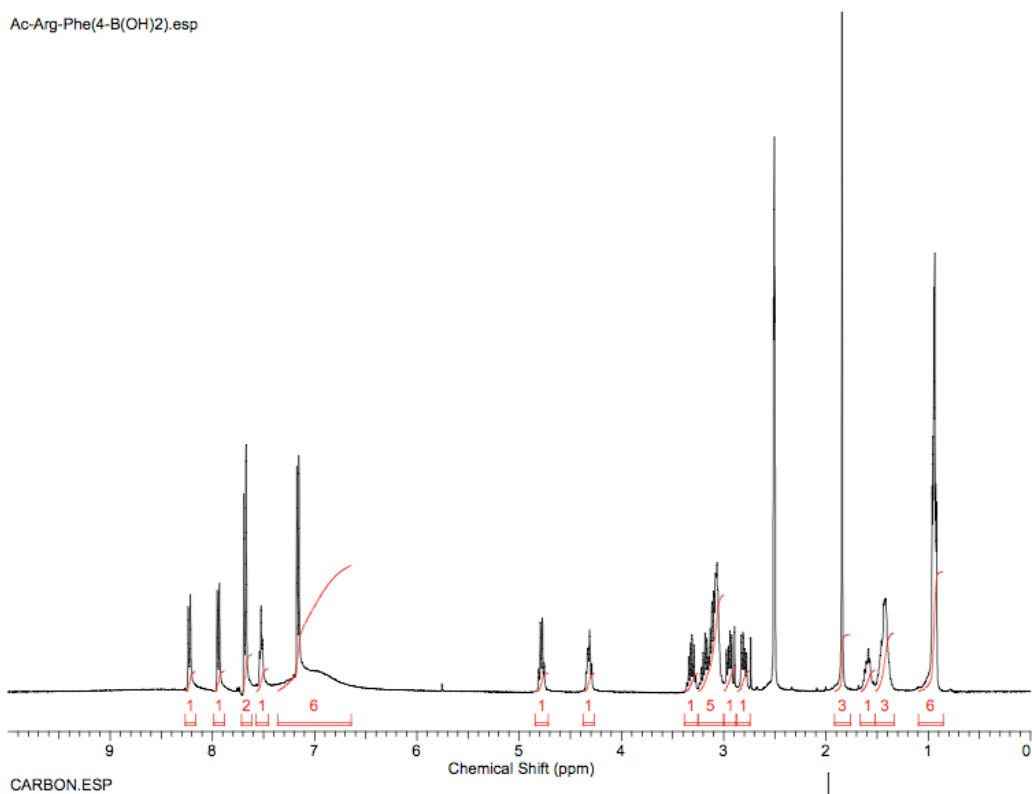
¹H NMR (400 MHz, DMSO-d₆) δ 8.23 (d, J = 8.1 Hz, 1H), 7.94 (d, J = 8.6 Hz, 1H), 7.68 (d, J = 8.1 Hz, 2H), 7.56 - 7.48 (m, 1H), 7.16 (d, J = 8.1 Hz, 2H), 4.78 (q, J = 7.2 Hz, 1H), 4.36 - 4.27 (m, 1H), 3.37 - 3.26 (m, ¹H), 3.25 - 3.01 (m, 5H), 2.95 (dd, J = 7.2, 13.2 Hz, 1H), 2.80 (dd, J = 7.2, 13.2 Hz, 1H), 1.84 (s, 3H), 1.66 - 1.52 (m, 1H), 1.52 - 1.34 (m, 3H), 0.96 - 0.90 (m, 6H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.0, 169.7, 169.2, 156.8, 139.1, 134.1, 128.3, 51.7, 49.9, 41.1, 40.4, 39.0, 38.2, 29.3, 24.9, 22.5, 14.0, 12.7.

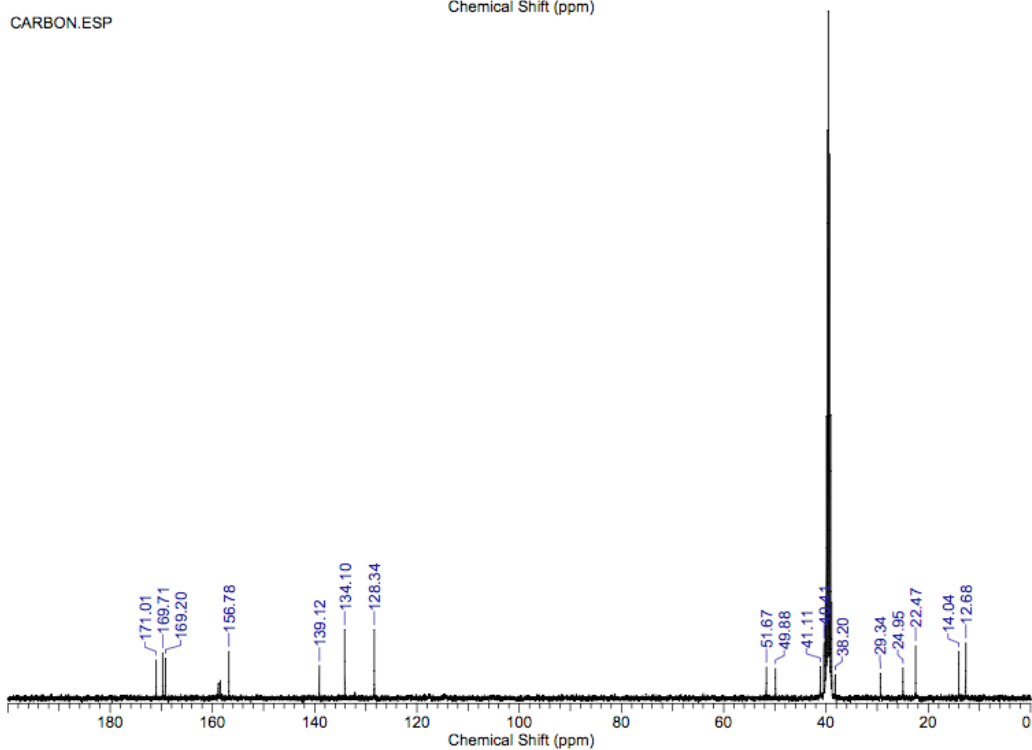
LRMS [M+H]⁺ 463 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 463.2835, [M+H]⁺ found = 463.2844.

Ac-Arg-Phe(4-B(OH)2).esp



CARBON.ESP



Synthesis of N-methylated dipeptides

The above procedure was followed, on a similar scale, starting with the appropriate amino acids, with the following deviation. After the first reaction with diethylamine, the boc-protected amino acid was N-methylated as outlined below. HATU/DIPEA were used in place of PyBOP for coupling to arginine, as outlined below.

Methylation of Boc-Protected Amino acids

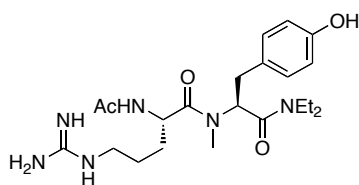
To a stirred solution of Boc-protected amino acid (1.54 mmol) dissolved in THF/DMF 20:1 (8ml) at 0°, 60% NaH mineral oil (136 mg, 3.39 mmol, 2.2 eq) was added. The reaction was stirred at 0°, and after 30 minutes, methyl iodide (0.714 ml, 7.70 mmol, 5 eq) was added. The reaction was stirred at 0° for a further 30 minutes, then at room temperature for 15 hours. The reaction was quenched with water (added slowly dropwise until no effervescence was observed). The organic layer was separated. The aqueous layer was re-extracted with EtOAc (3 x 10 ml). The combined organic phases were dried (MgSO₄) and concentrated in vacuo to give yellow/orange oil. Purification on SiO₂ eluting with EtOAc/hexane gave the methyl amine as a yellow oil (42% yield for tyrosine, 38% for p-nitrotyrosine).

Peptide coupling with N-Methyl amine

At room temperature, to a stirred solution of Fmoc-protected arginine (441 mg, 0.68 mmol) dissolved in CH₂Cl₂ (5 ml) was added HATU (260 mg, 0.68 mmol, 1 eq) and DIPEA (0.240 mg, 1.37 mmol, 2 eq). The N-methylated amino acid (0.68 mmol, 1 eq) in CH₂Cl₂ (5 ml) was added and the reaction was stirred for 15 hours. The reaction was diluted with 10% citric acid (10 ml). The organic layer was collected and the aqueous layer was re-extracted with CH₂Cl₂ (3 x 10 ml). The combined organic layers were washed with 10% citric acid (10 ml), saturated NaHCO₃ (10 ml) and brine (10 ml). The organic layer was dried (MgSO₄) and concentrated in vacuo to give a yellow oil (crude yield ~70%) that was used directly in the next step.

Ac-Arg-NMe-Tyr-NEt₂

The above procedure for the synthesis of N-methylated dipeptides was followed. The overall yield for the synthesis was 11%.



Chemical Formula: $C_{22}H_{36}N_6O_4$
Exact Mass: 448.2798
Molecular Weight: 448.5590

$[\alpha]_{26D} -139.6$ (MeOH, c 0.6).

IR ν_{max} (neat) 3273 (m), 3183 (m), 2978 (m), 1614 (s), 1515 (s) cm^{-1} .

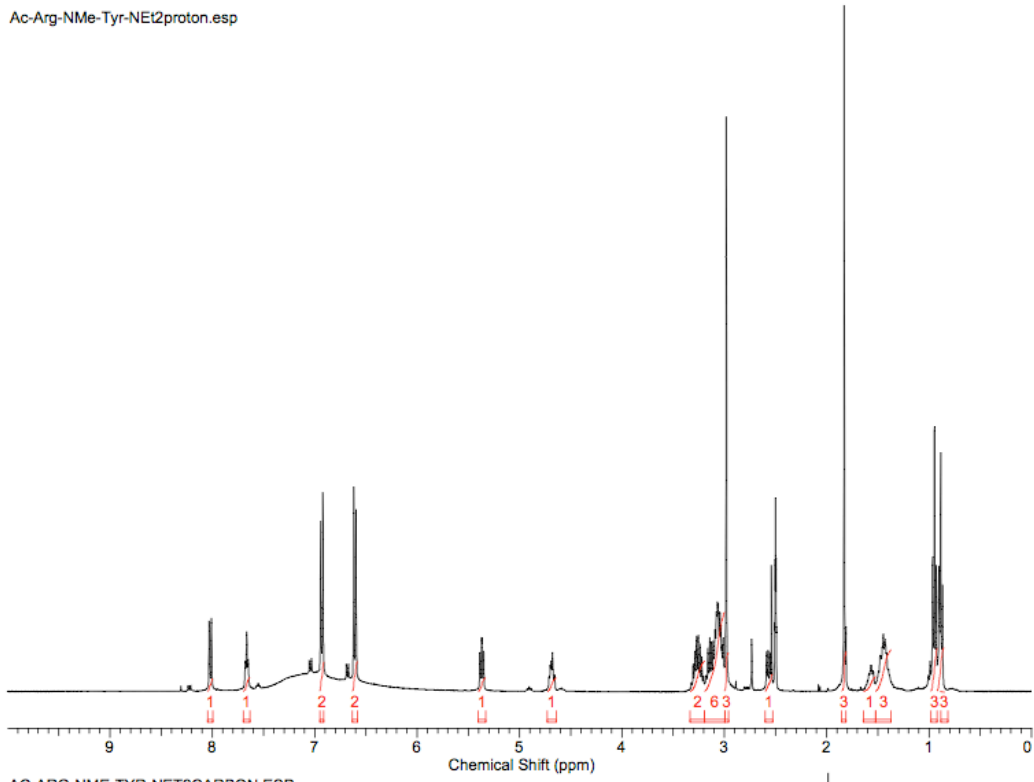
1H NMR (400 MHz, DMSO- d_6) δ 8.02 (d, $J = 8.6$ Hz, 1H), 7.67 (t, $J = 5.6$ Hz, 1H), 6.93 (d, $J = 8.6$ Hz, 2H), 6.61 (d, $J = 8.6$ Hz, 2H), 5.37 (dd, $J = 6.5, 8.1$ Hz, 1H), 4.72 - 4.65 (m, 1H), 3.33 - 3.19 (m, 2H), 3.19 - 3.00 (m, 6H), 2.98 (s, 3H), 2.56 (dd, $J = 6.5, 13.6$ Hz, 1H), 1.83 (s, 3H), 1.63 - 1.52 (m, 1H), 1.52 - 1.35 (m, 3H), 0.95 (t, $J = 6.8$ Hz, 3H), 0.89 (t, $J = 6.8$ Hz, 3H).

^{13}C NMR (100 MHz, DMSO- d_6) δ 171.3, 169.0, 168.1, 156.7, 155.7, 130.0, 127.6, 115.0, 53.7, 48.1, 40.7, 40.4, 39.6, 34.0, 30.2, 28.7, 24.8, 22.3, 14.2, 12.7.

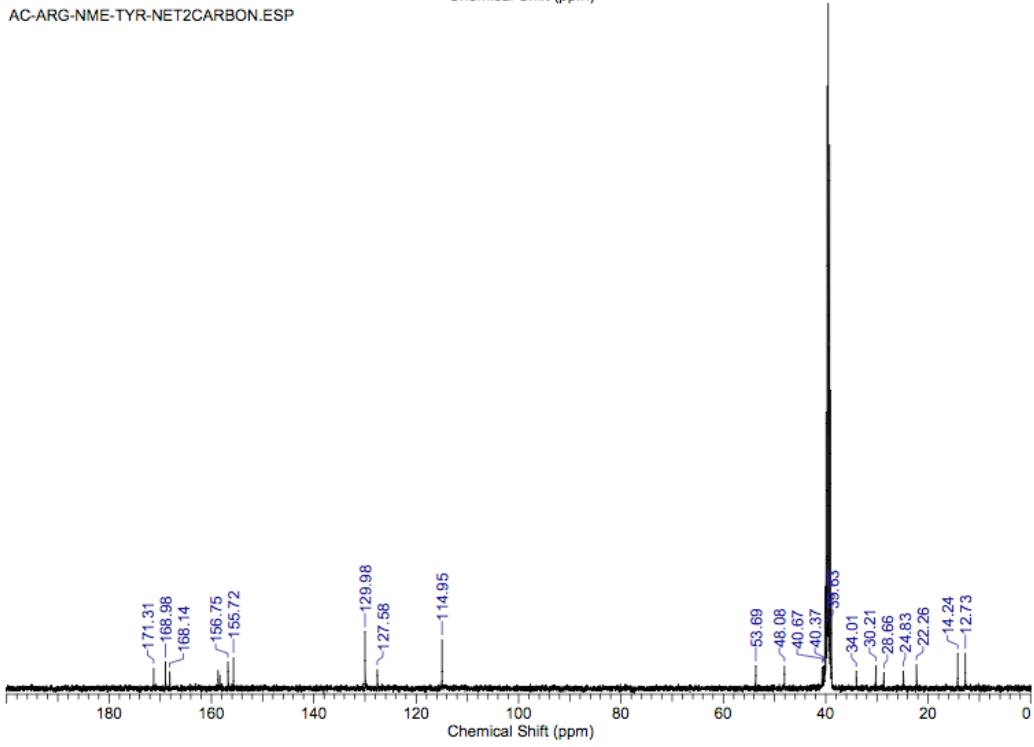
LRMS $[M+H]^+$ 449 (100%).

HRMS ESI+ $[M+H]^+$ calculated = 449.2871, $[M+H]^+$ found = 449.2866.

Ac-Arg-NMe-Tyr-NEt2proton.esp

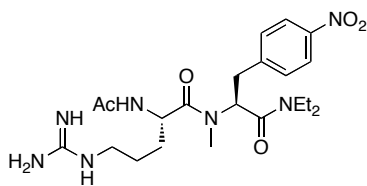


AC-ARG-NME-TYR-NET2CARBON.ESP



Ac-Arg-NMe-Phe(4-NO₂)-NEt₂

The above procedure for the synthesis of N-methylated dipeptides was followed. The overall yield for the synthesis was 15%.



Chemical Formula: C₂₂H₃₅N₇O₅
Exact Mass: 477.2700
Molecular Weight: 477.5572

[α]_D²⁶ -206.6 (MeOH, c 0.51).

IR ν_{max} (neat) 3294 (m), 3190 (w), 1776 (w), 1620 (s), 1518 (s) cm⁻¹.

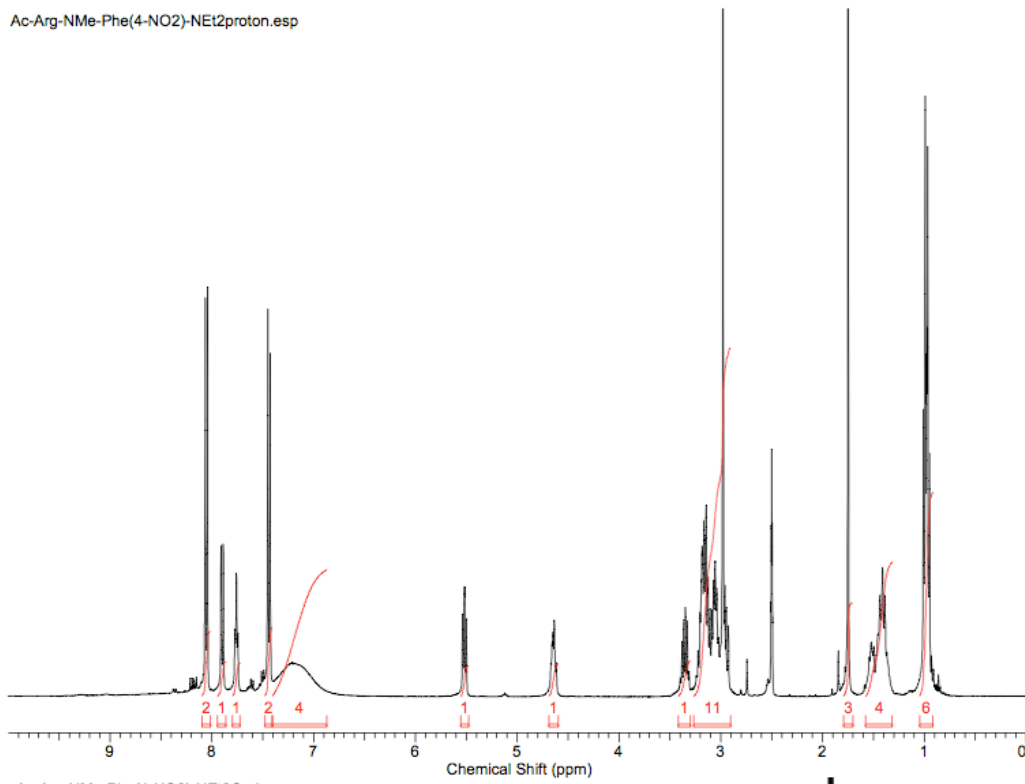
¹H NMR (400 MHz, DMSO-d₆) δ 8.05 (d, J = 8.6 Hz, 2H), 7.90 (d, J = 8.6 Hz, 1H), 7.76 (t, J = 5.6 Hz, 1H), 7.44 (d, J = 8.6 Hz, 2H), 7.19 (br. s., 3H), 5.56 - 5.46 (m, 1H), 4.69 - 4.60 (m, 1H), 3.36 (qd, J = 6.9, 13.7 Hz, 1H), 3.26 - 2.90 (m, 10H), 1.75 (s, 3H), 1.58 - 1.31 (m, 4H), 0.99 (t, J = 7.1 Hz, 3H), 0.98 (t, J = 7.1 Hz, 3H).

¹³C NMR (100 MHz, DMSO-d₆) δ 171.4, 168.8, 167.5, 156.8, 146.3, 146.1, 130.6, 123.0, 53.5, 47.8, 40.5, 40.3, 39.6, 34.3, 30.1, 28.4, 24.8, 22.1, 14.2, 12.7.

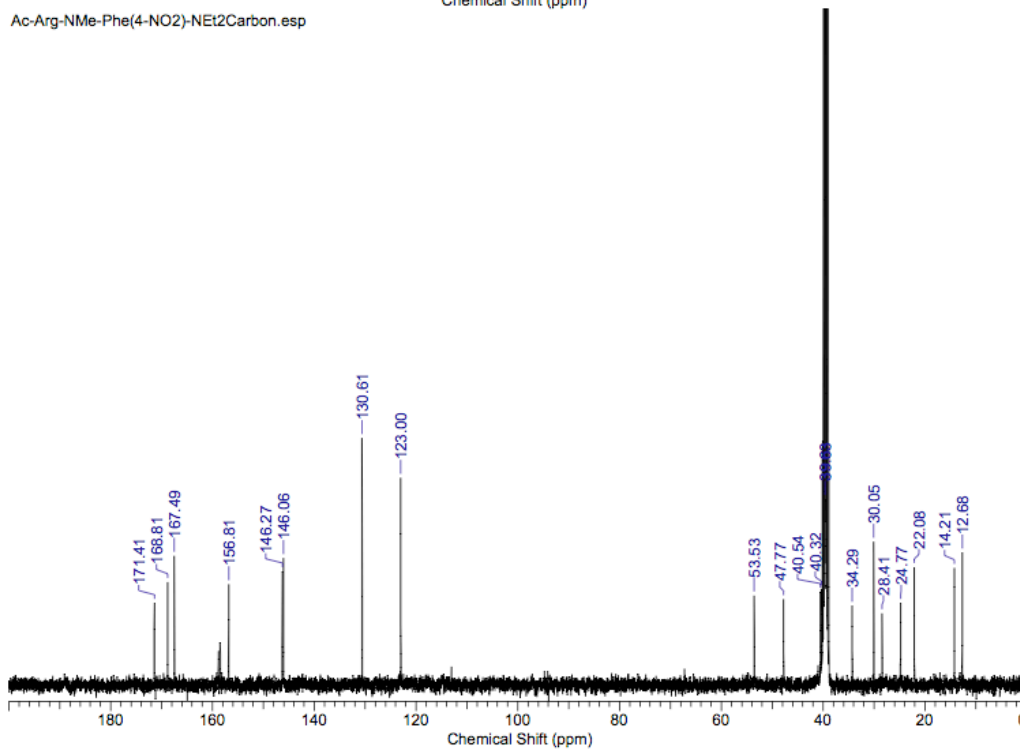
LRMS [M+H]⁺ 478 (100%).

HRMS ESI⁺ [M+H]⁺ calculated = 478.2772, [M+H]⁺ found = 478.2768.

Ac-Arg-NMe-Phe(4-NO2)-NEt2proton.esp

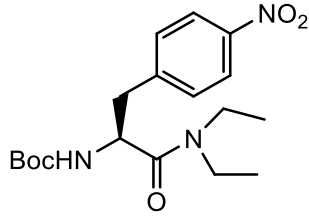


Ac-Arg-NMe-Phe(4-NO2)-NEt2Carbon.esp



Spectra for the intermediates in the synthesis of compound 14

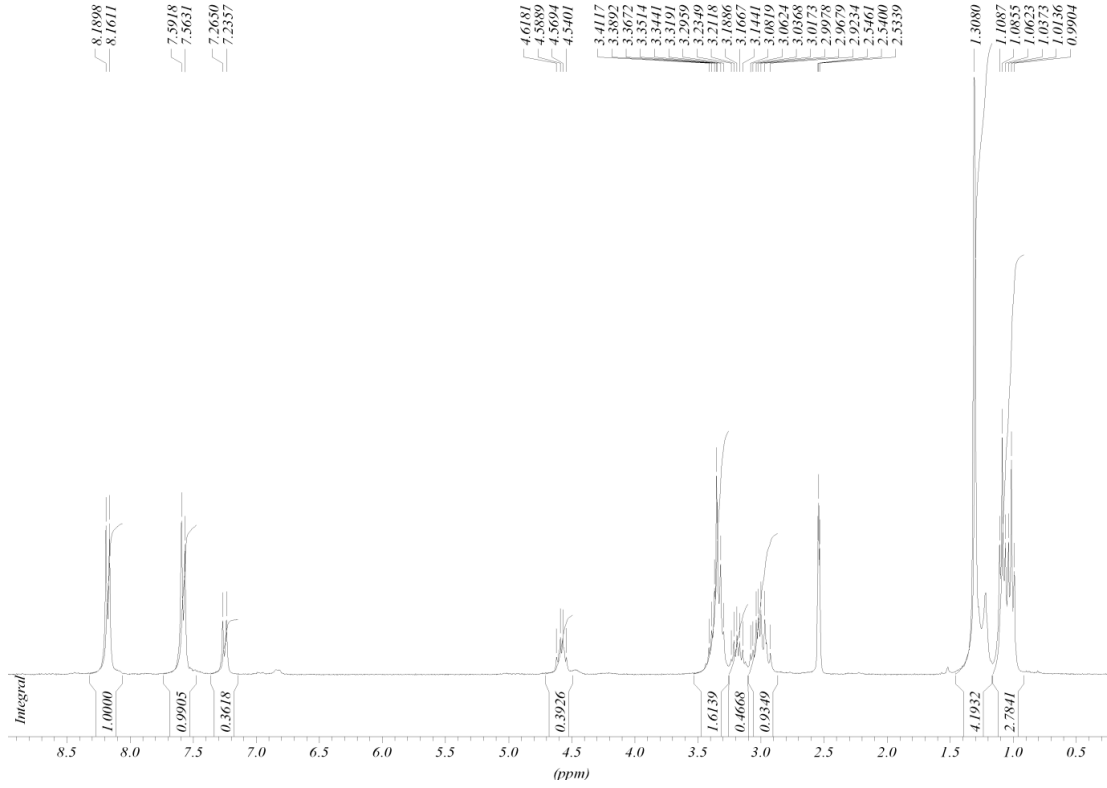
Boc-Phe(4-NO₂)-diethylamide

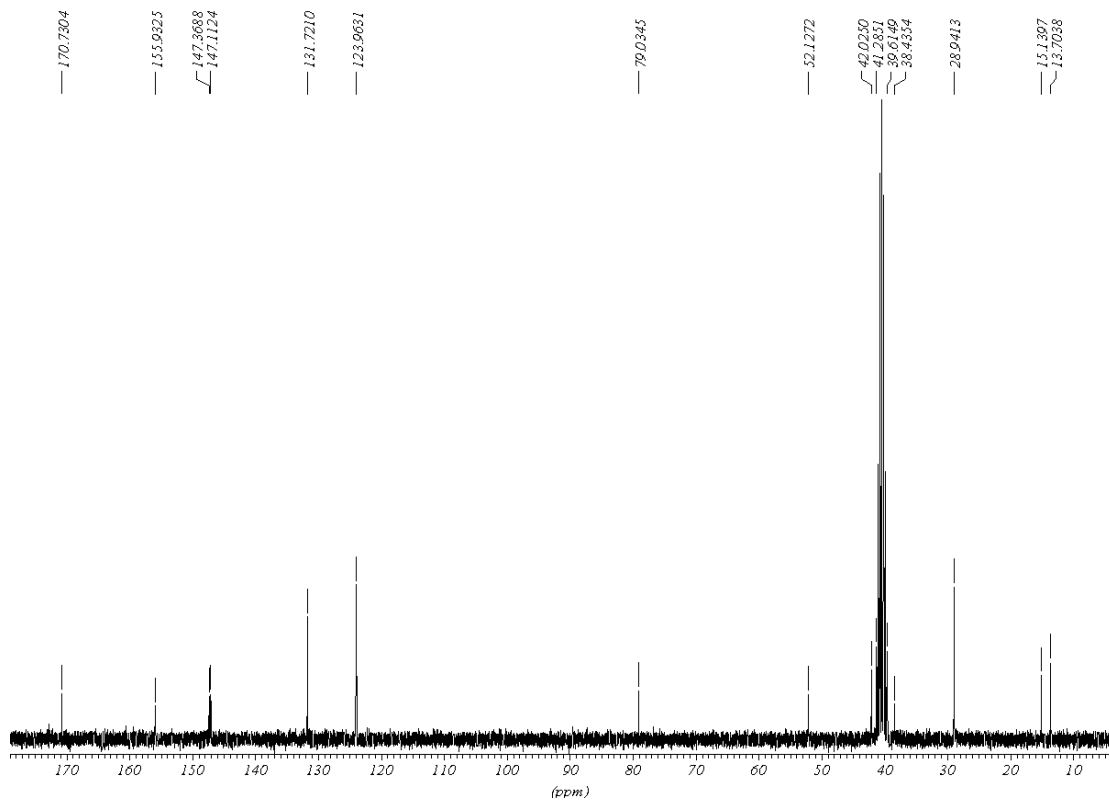


Chemical Formula: C₁₈H₂₇N₃O₅

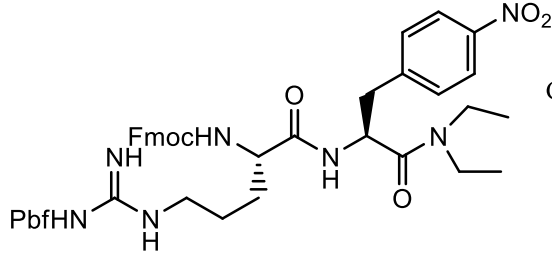
Exact Mass: 365.20

Molecular Weight: 365.42





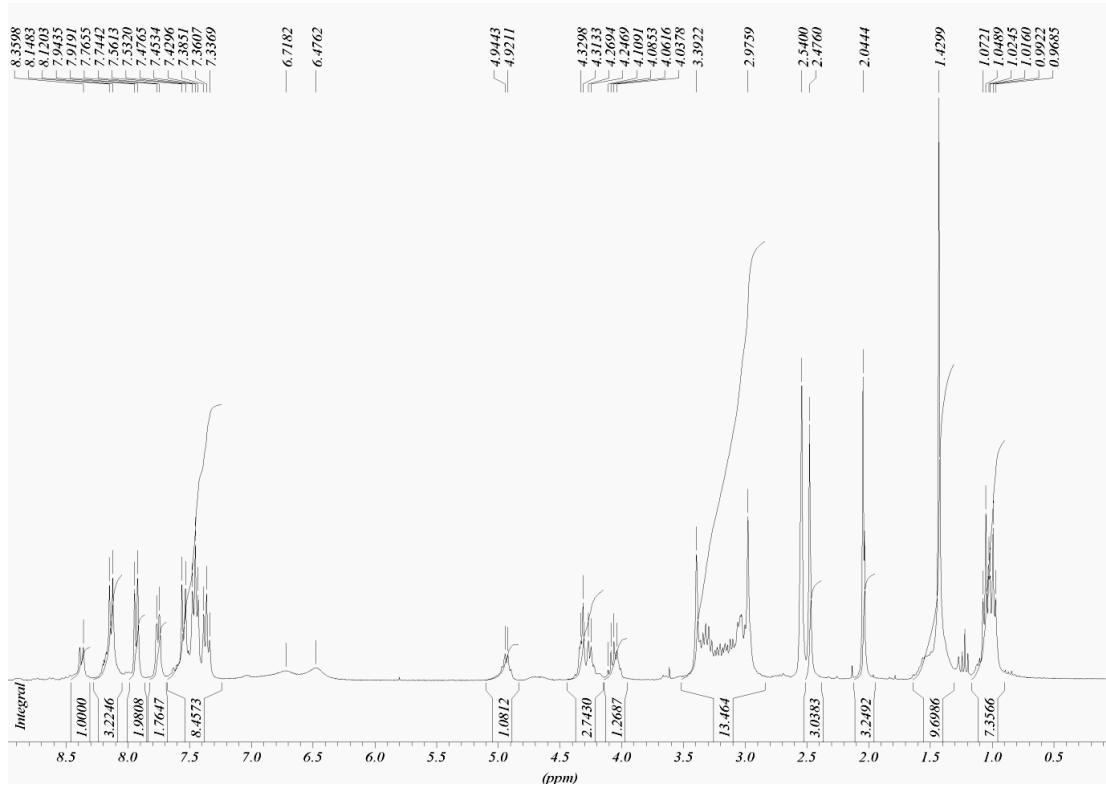
fmoc-Arg(Pbf)-CONH-Phe(4-NO₂)-L-Diethylamide

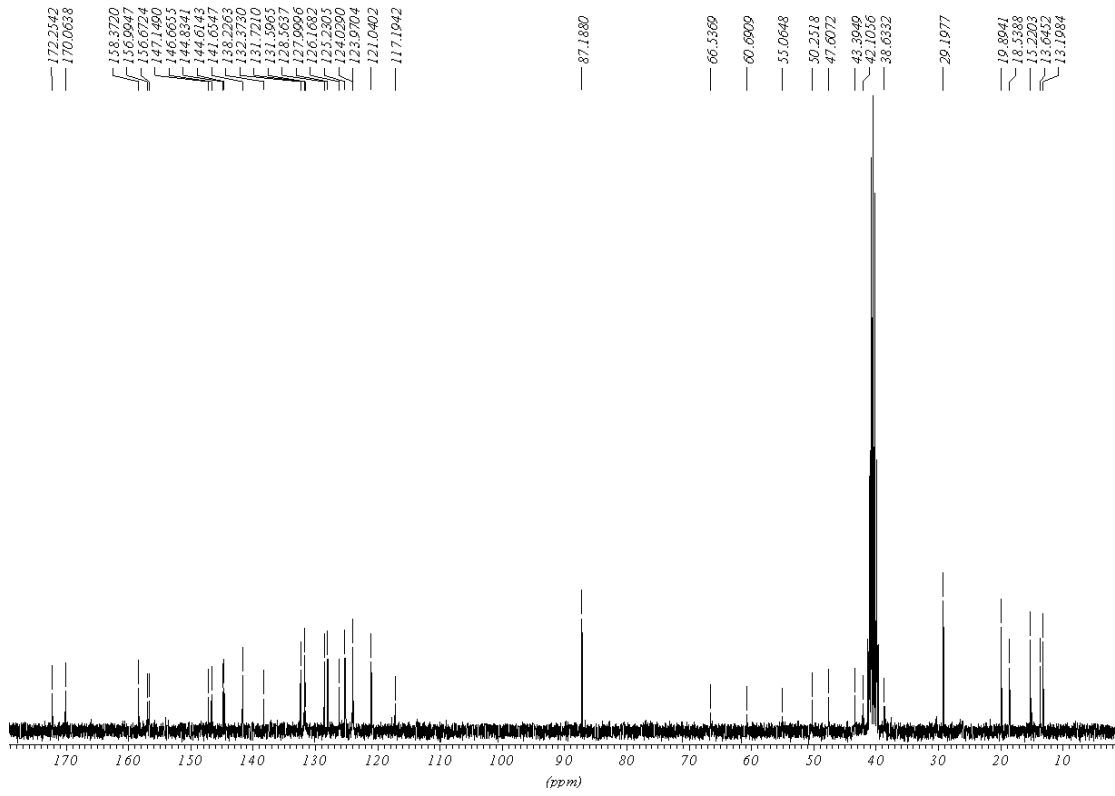


Chemical Formula: C₄₇H₅₇N₇O₉S

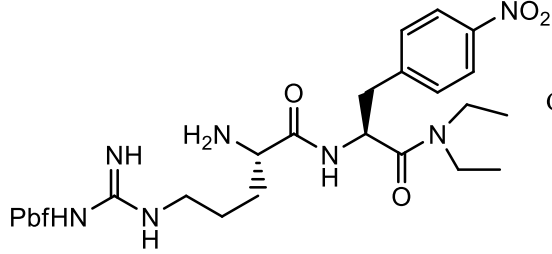
Exact Mass: 895.39

Molecular Weight: 896.06





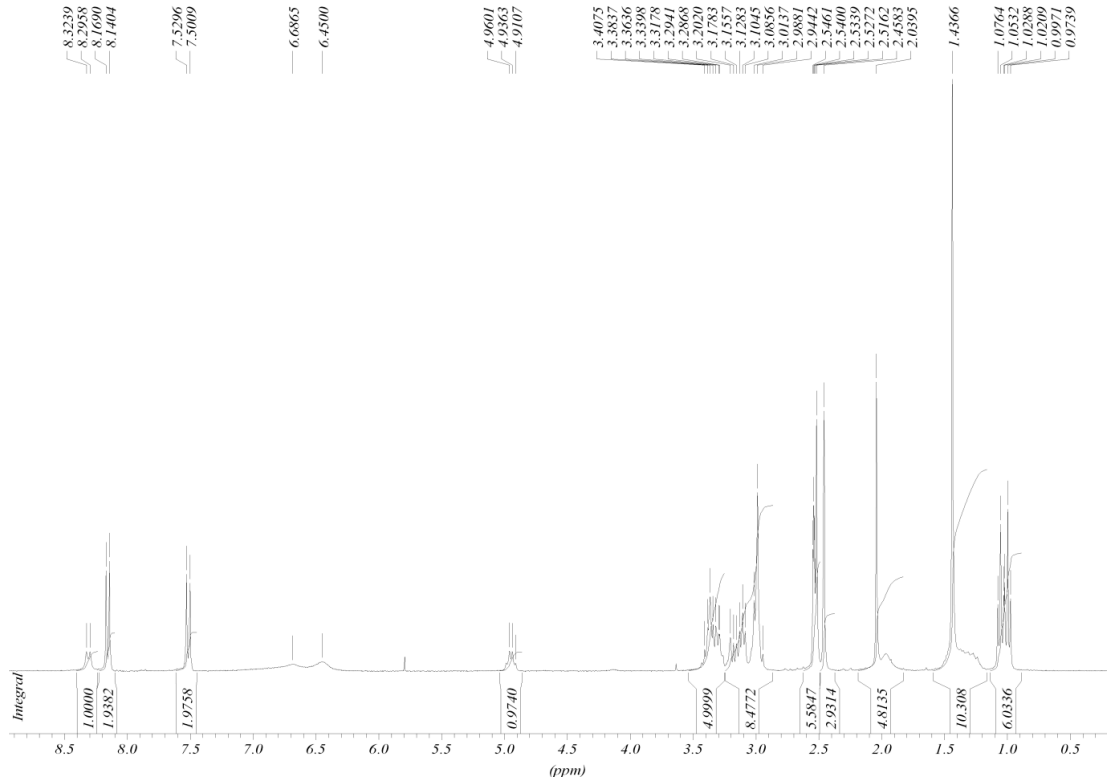
Amino-Arg(Pbf)-CONH-Phe(4-NO₂)-L-Diethylamide

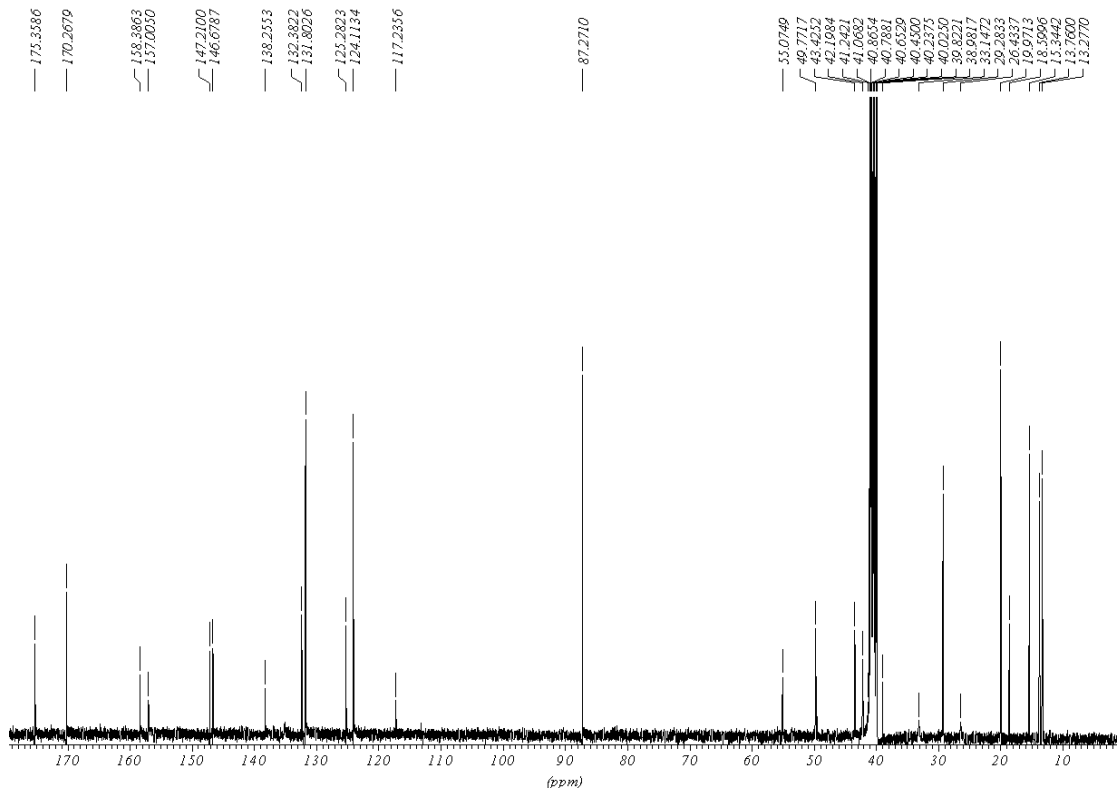


Chemical Formula: C₃₂H₄₇N₇O₇S

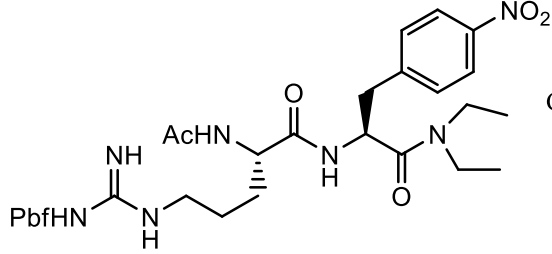
Exact Mass: 673.33

Molecular Weight: 673.82





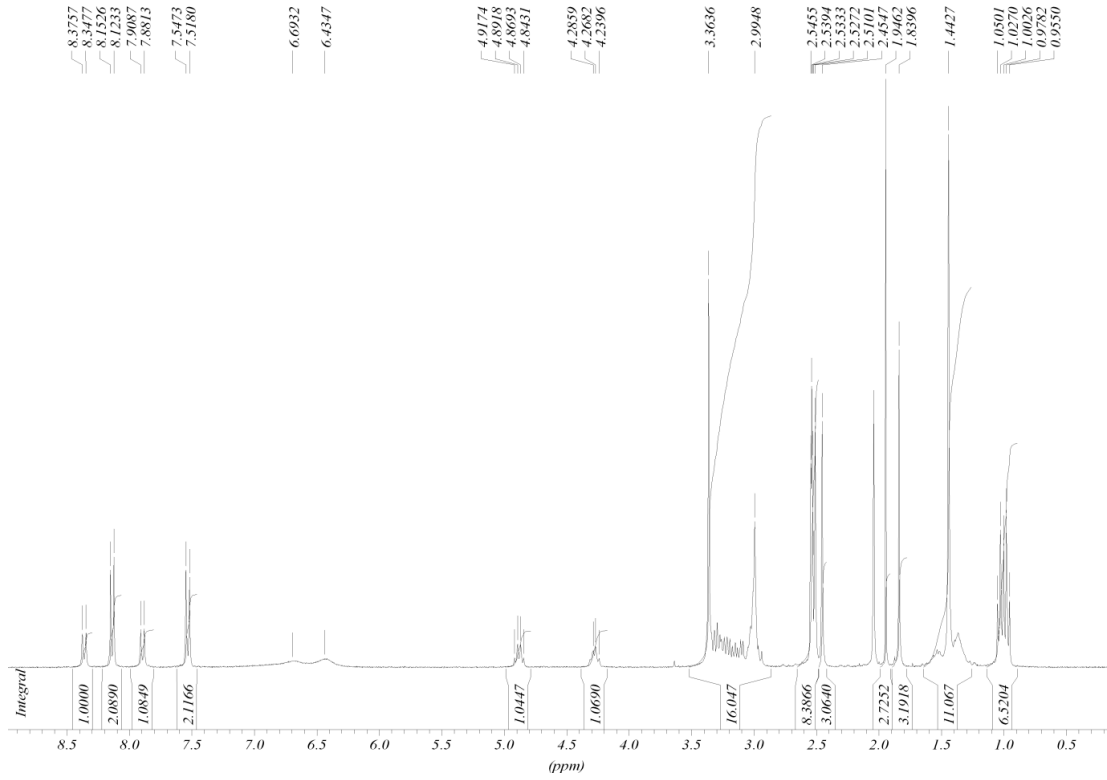
Acetyl-Arg(Pbf)-CONH-Phe(4-NO₂)-L-Diethylamide

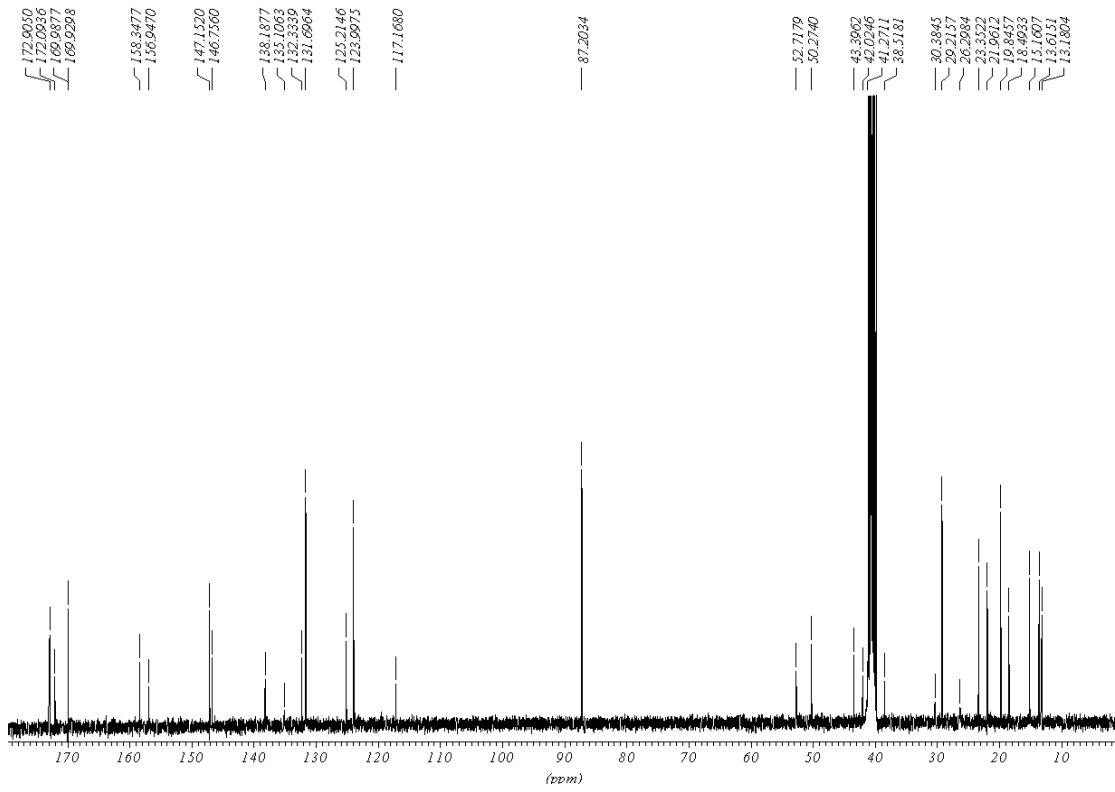


Chemical Formula: C₃₄H₄₉N₇O₈S

Exact Mass: 715.34

Molecular Weight: 715.86





Protein expression and enzyme kinetics

Avian and human ATIC fused to an *N*-terminal 6X histidine tag were expressed from pET28 vectors in BL21(DE3) strains of *E. coli*, and purified by standard protocols. The proteins were purified on an AKTAprime FPLC (GE Healthcare) by nickel affinity. AICAR Tfase assays were conducted with avian ATIC and carried out in 1 cm path length quartz cuvettes at 25 °C as previously described in "A. Tavassoli, S. J. Benkovic, *Angew Chem Int Ed Engl* (2005), 44, 2760-2763". Each inhibitor was dissolved in DMSO to give a 100 mM stock solution that was further diluted for use in each assay. Each data point was repeated in triplicate. The data was analyzed using KaleidaGraph (Synergy Software) and Microsoft Excel, by assuming competitive inhibition with 10-f-thf as previously described in "A. Tavassoli, S. J. Benkovic, *Angew Chem Int Ed Engl* (2005), 44, 2760-2763". Errors in the K_i were calculated by linear regression. Compounds 1, 8, and 14 were also assayed with human ATIC, with the same K_i (within error) being observed for each compound as for the avian enzyme.

Spectral data

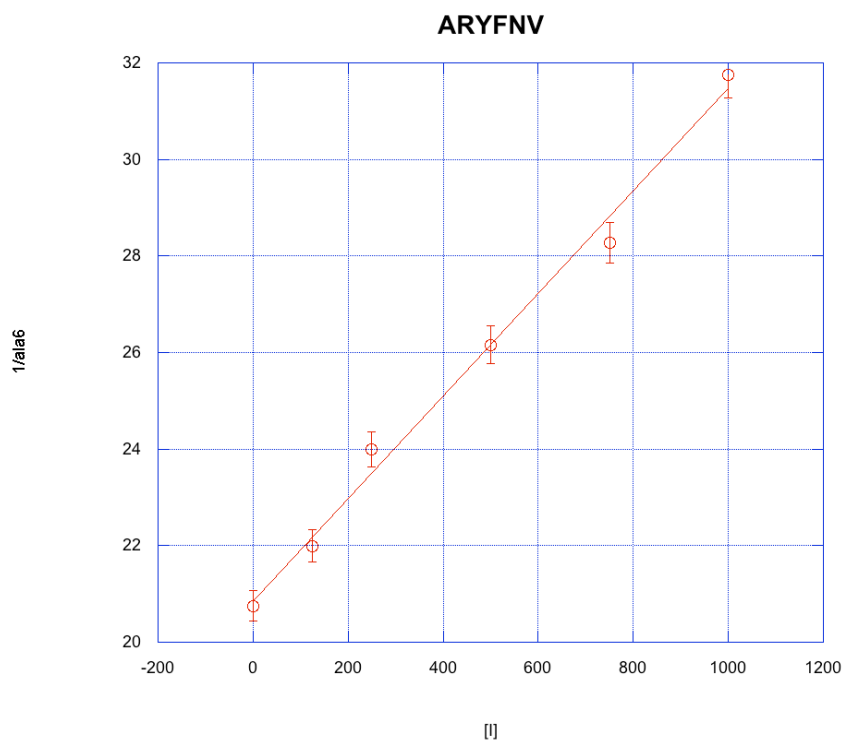


Figure 1. ARYFNV

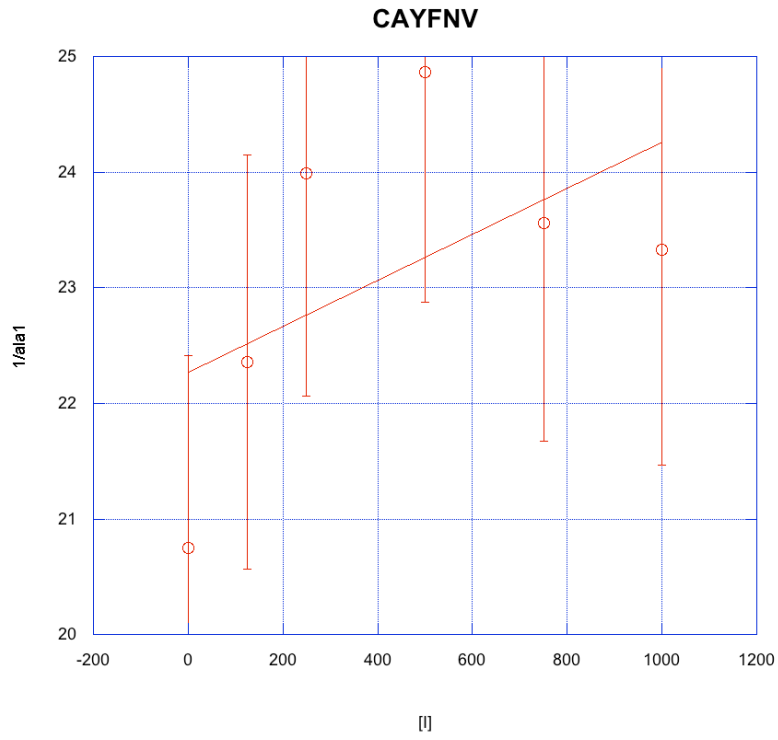


Figure 2. CAYFNV

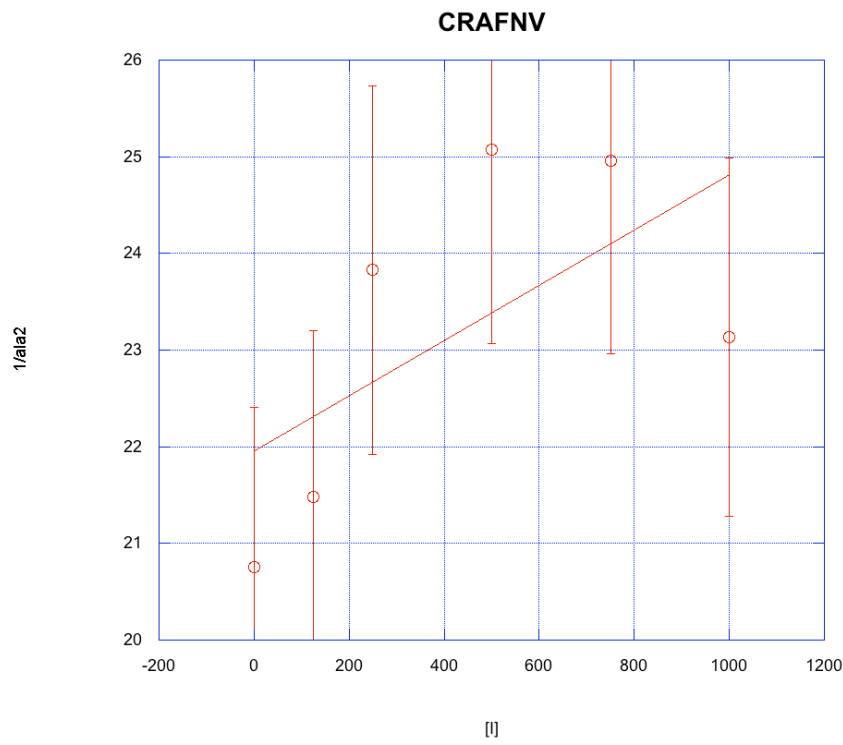


Figure 3. CRAFNV

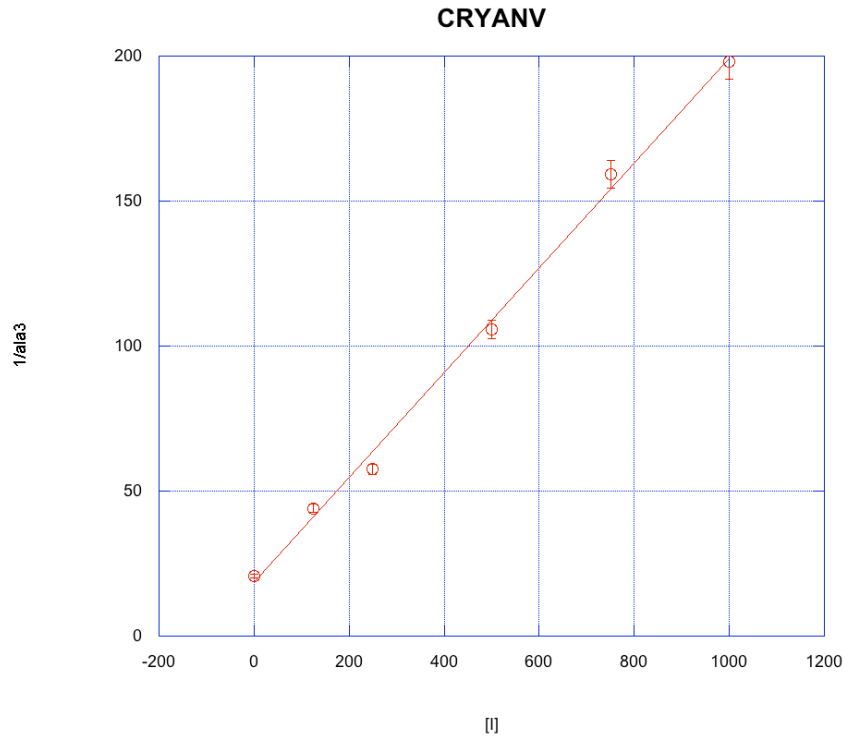


Figure 4. CRYANV

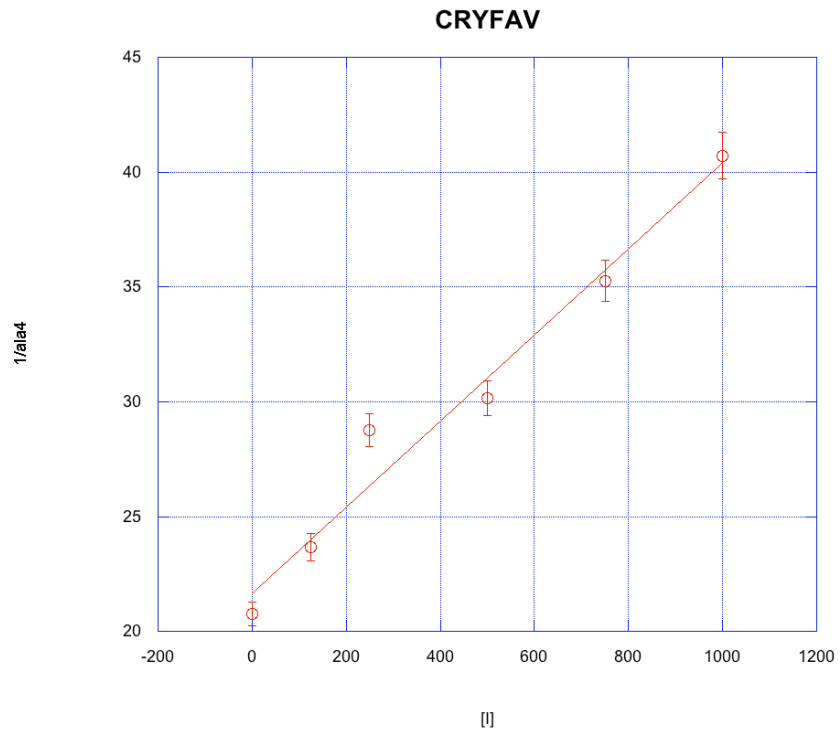


Figure 5. CRYFAV

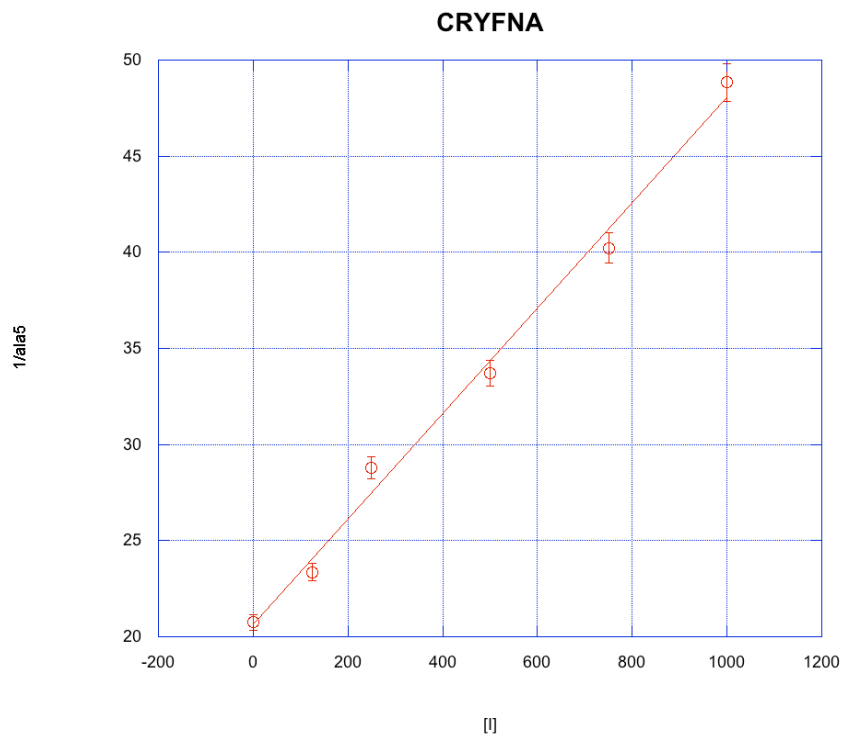


Figure 6. CRYFNA

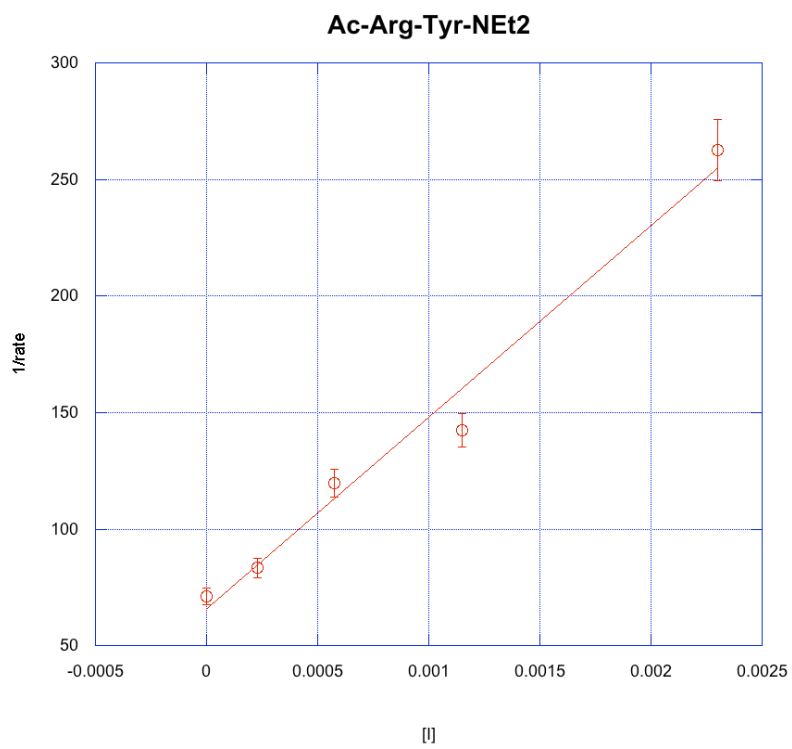


Figure 7. Ac-Arg-Tyr-NEt₂

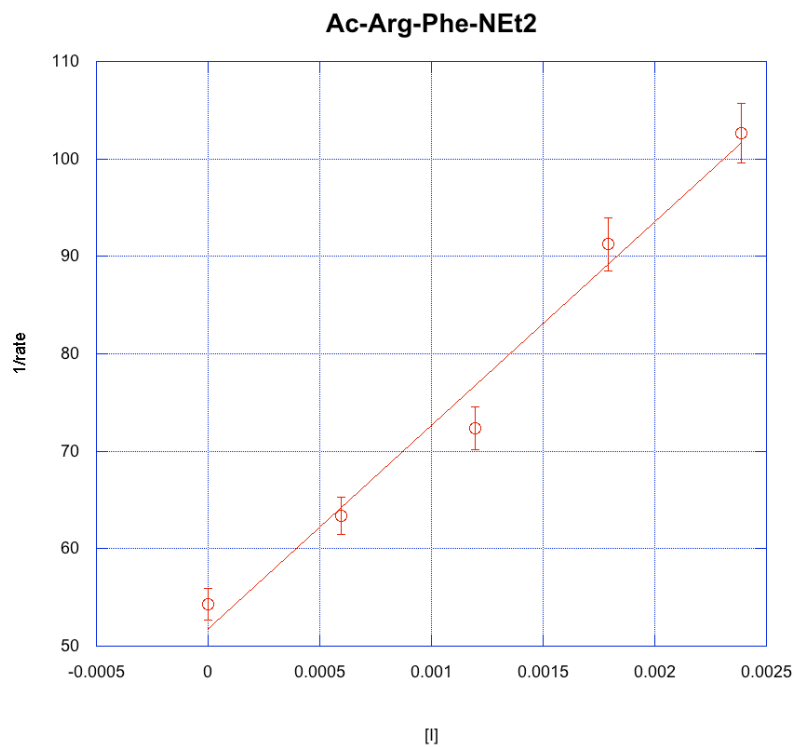


Figure 8. Ac-Arg-Phe-NEt₂

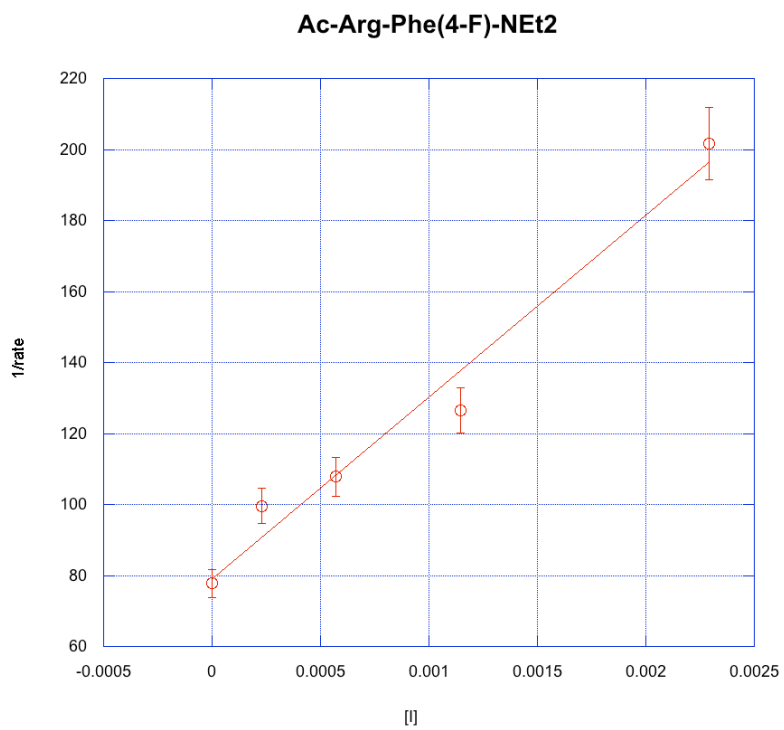


Figure 9. Ac-Arg-Phe(4-F)-NEt₂

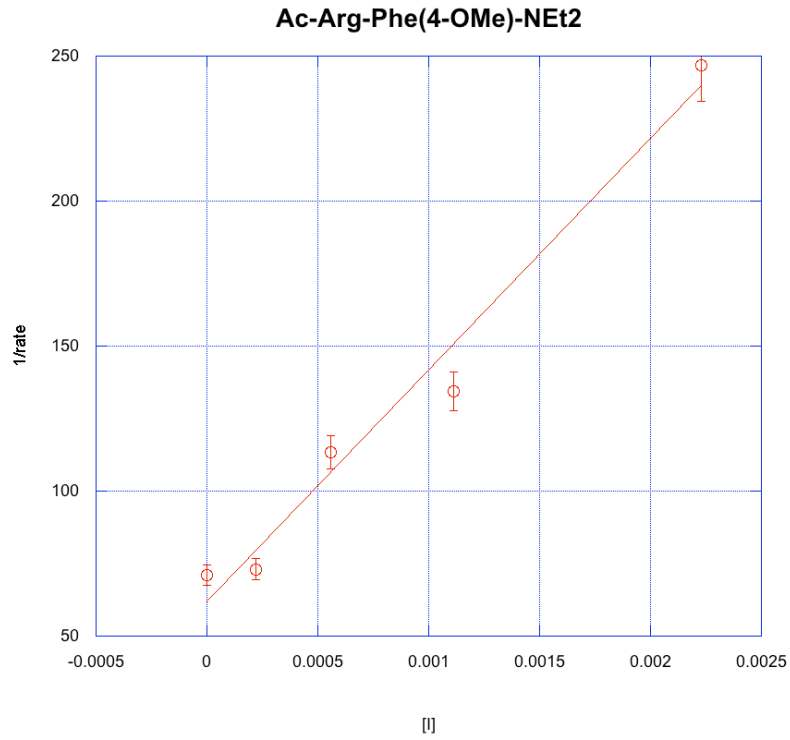


Figure 10. Ac-Arg-Phe (4-OMe)-NEt₂

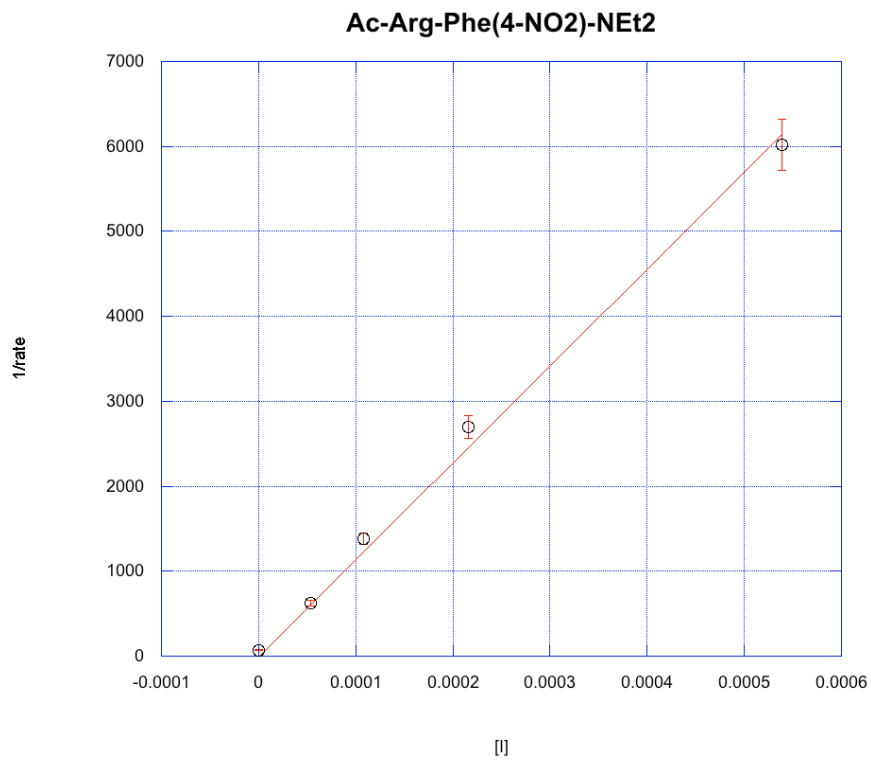


Figure 11. Ac-Arg-Phe(4-NO₂)-NEt₂

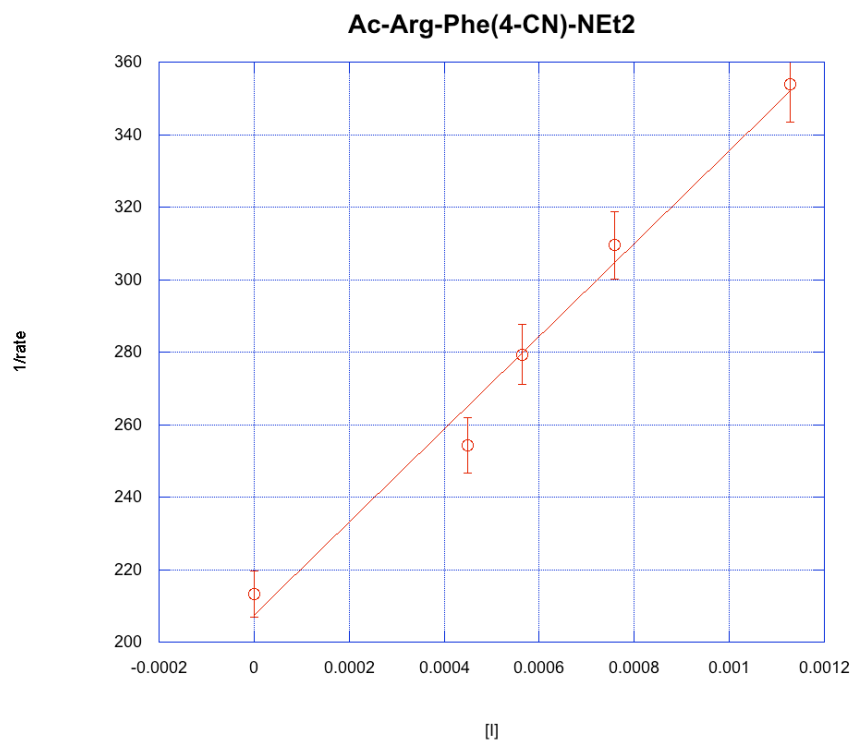


Figure 12. Ac-Arg-Phe(4-CN)-NEt₂

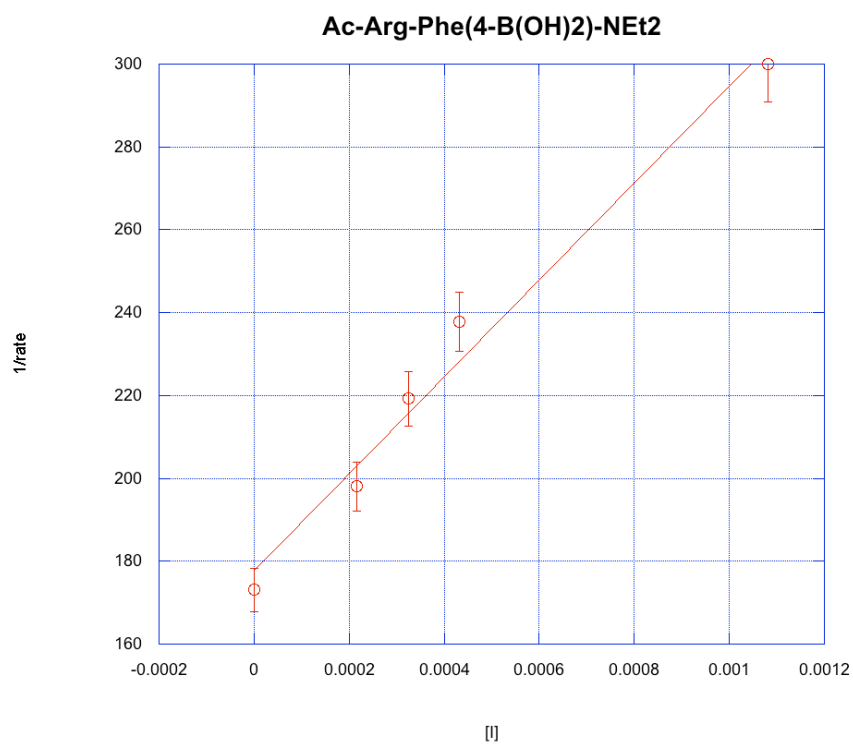


Figure 13. Ac-Arg-Phe(4-B(OH)₂)-NEt₂

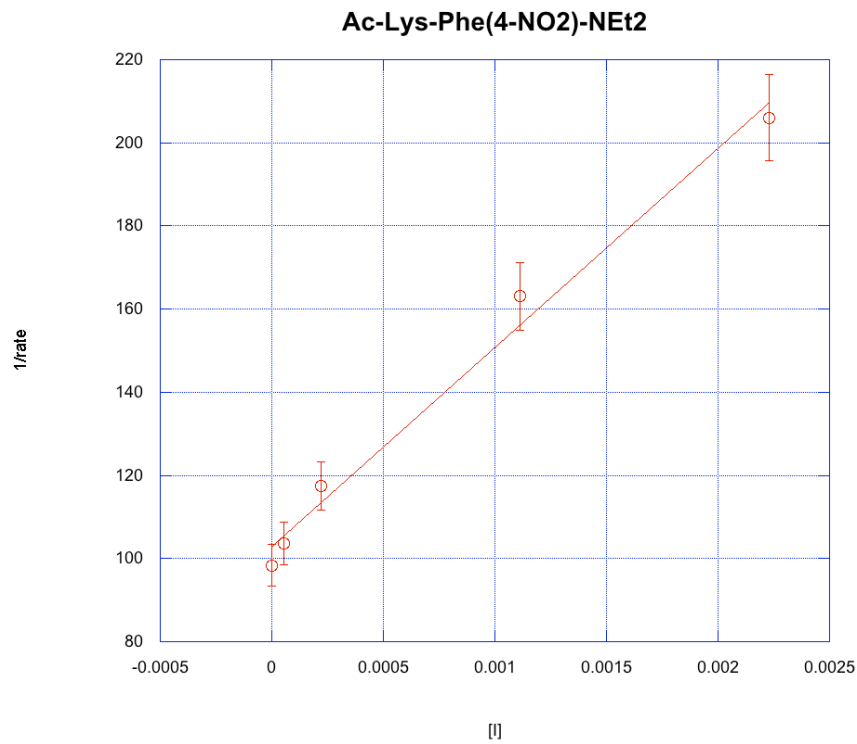


Figure 14. Ac-Lys-Phe(4-NO₂)-NEt₂

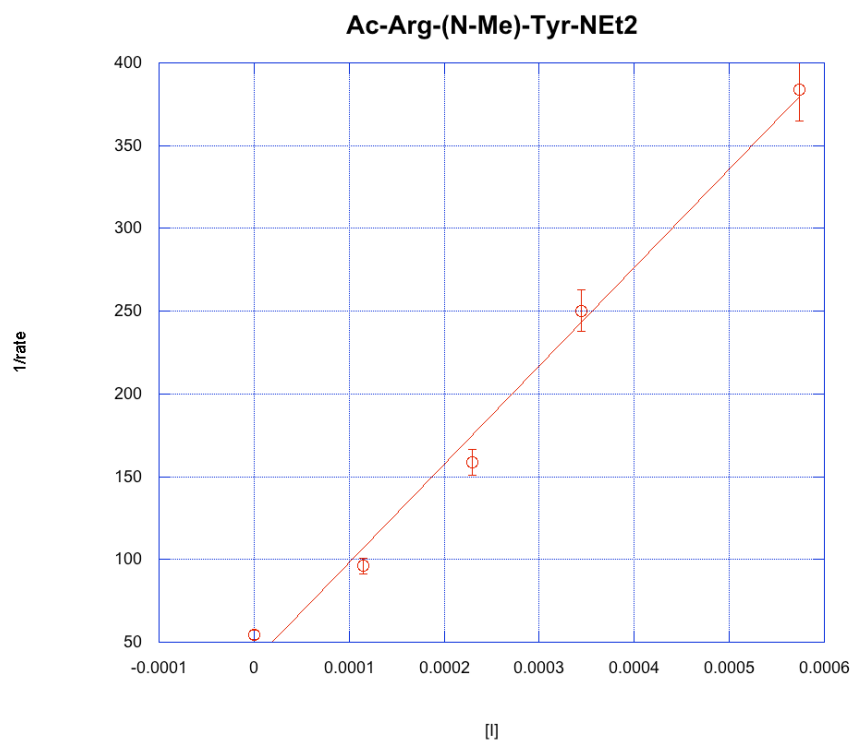


Figure 15. Ac-Arg-(N-Me)-Tyr-NEt₂

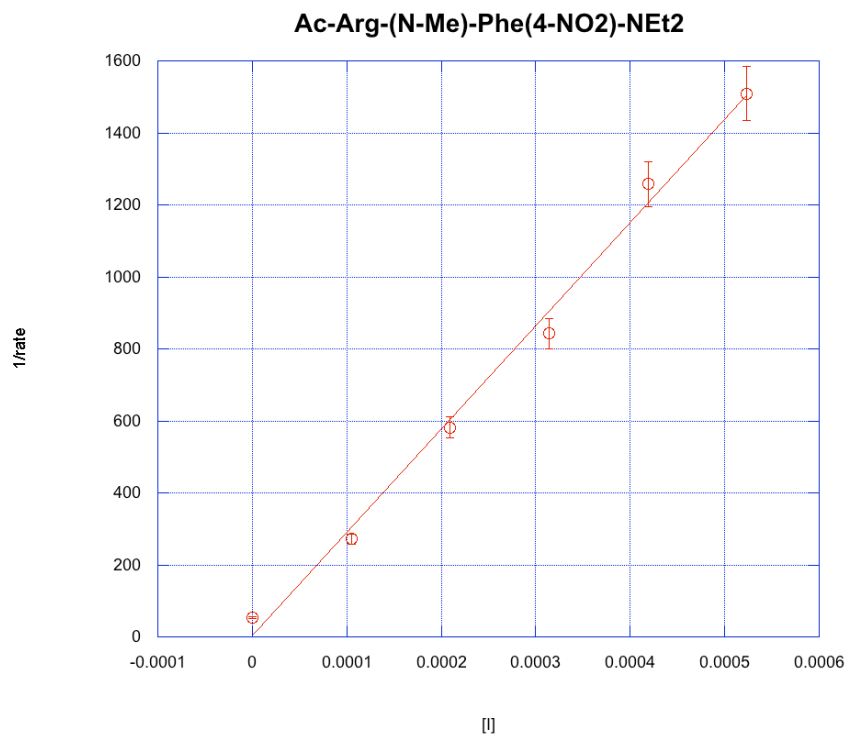


Figure 16. Ac-Arg-(N-Me)-Phe(4-NO₂)-NEt₂

Size exclusion chromatography

Human ATIC was prepared as detailed above, and divided into 100 nM solutions in 1 mL of buffer (20 mM Tris-Cl, pH 7.5, 150 mM NaCl, 50 mM KCl, 5 mM EDTA, 5 mM dithiothreitol). Inhibitor was added to each solution to give mixtures containing 0, 1, and 10 μ M of compound 14. Each solution was filtered through a Millex GP 0.22 μ m filter (Millipore), and loaded onto a HiLoad 16/60 Superdex 200 gel filtration column (GE Healthcare) pre-equilibrated with the above buffer. The flow rate was 1 mL/min and the flow monitored by UV on an AKTAprime FPLC (GE Healthcare). The column was calibrated using a HMW gel filtration calibration kit (GE Healthcare).

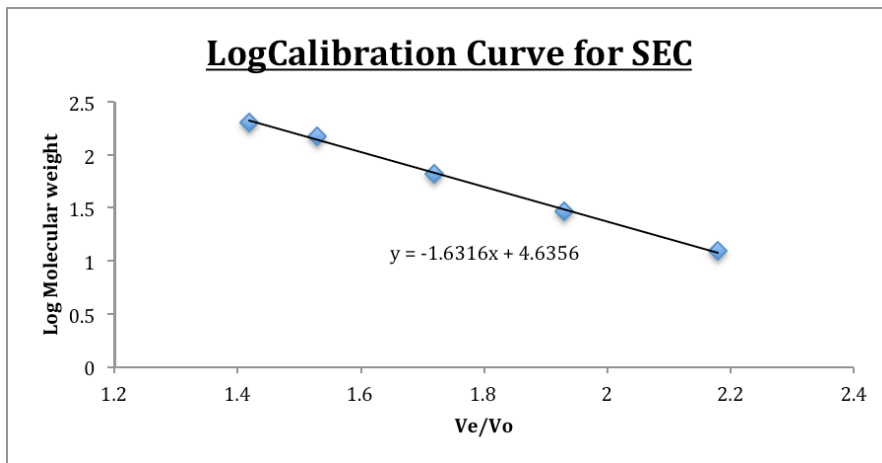


Figure 17: SEC calibration curves.

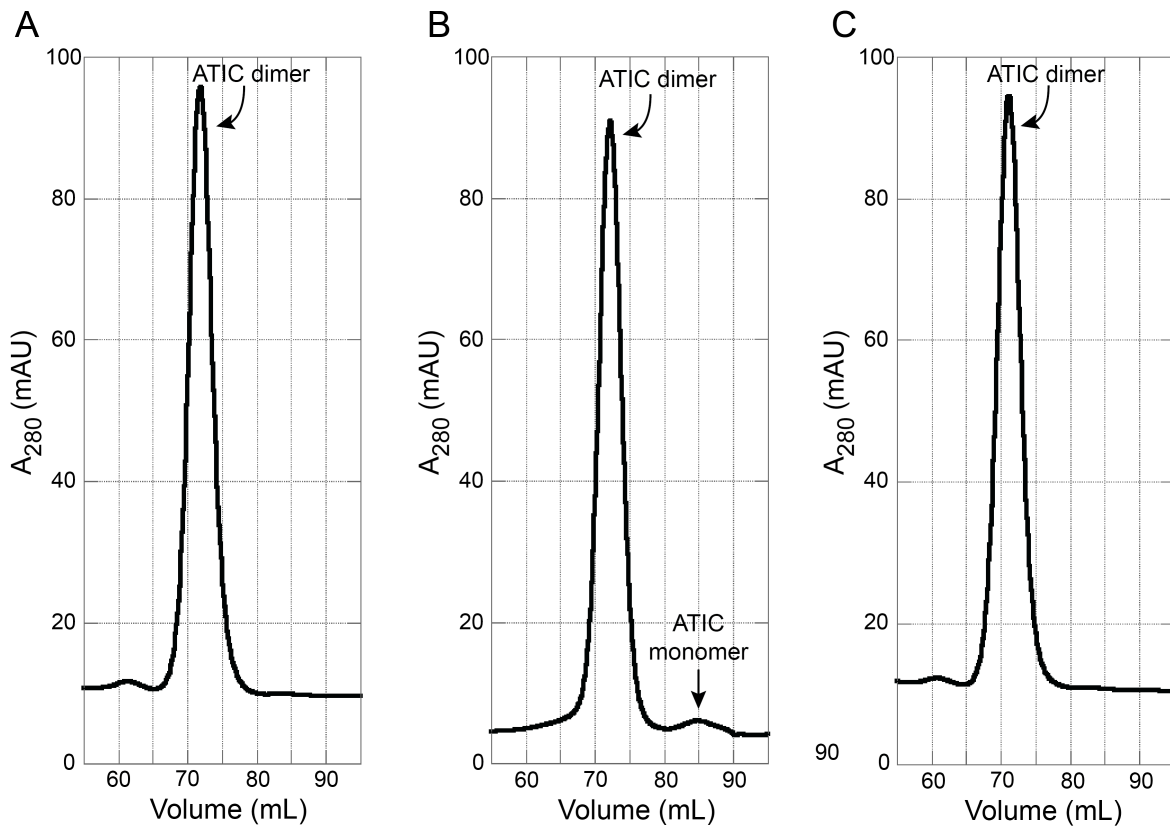


Figure 18: Size exclusion chromatography. (A) The ratio of monomer to dimer is shown in a 100 nM solution of ATIC (B) The same mixture with 10 μ M of compound 14 shows the presence of monomeric ATIC at 85 mL (C) ATIC incubated with compound 9 (negative control) shows no monomer species.