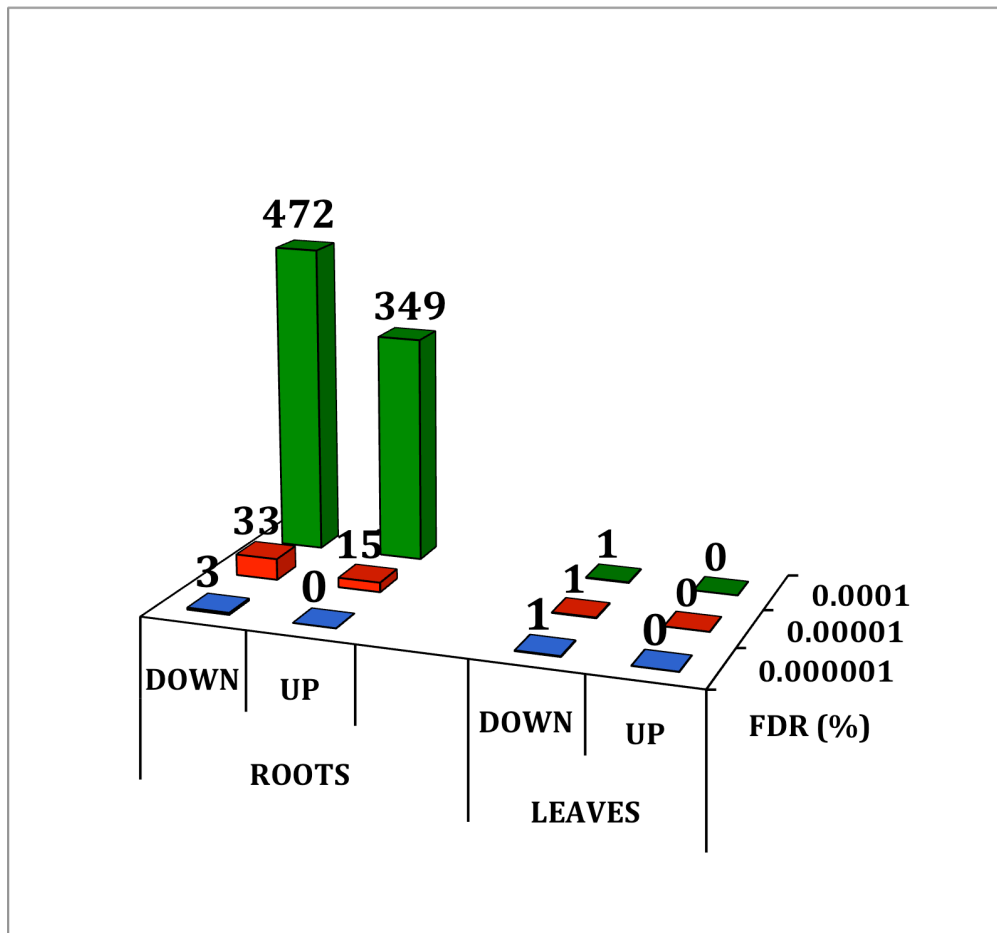
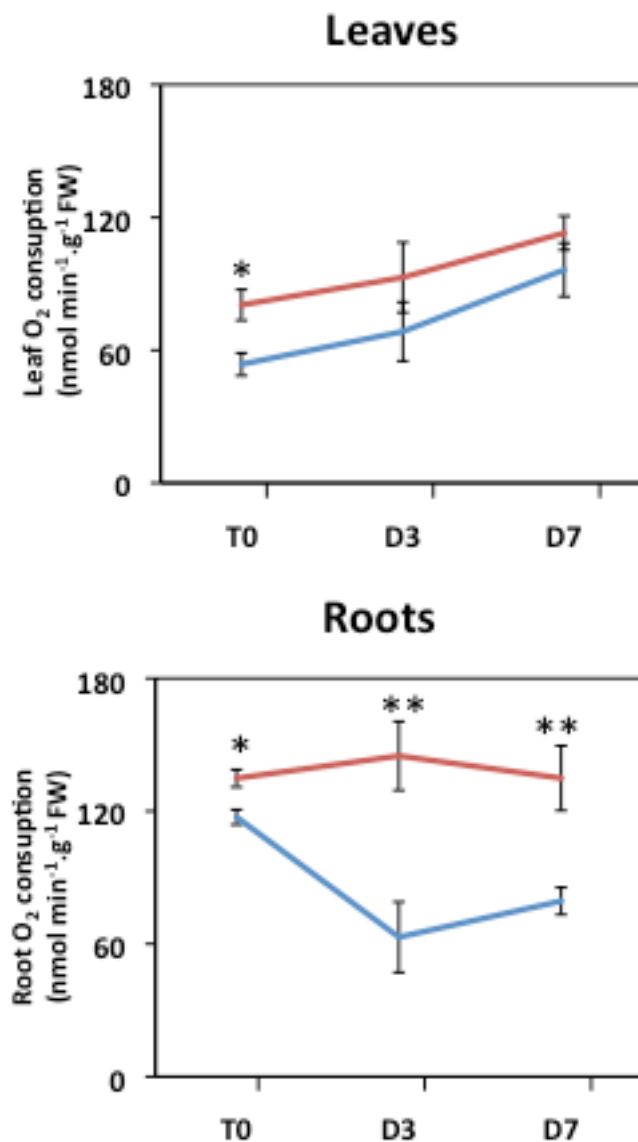


Supplemental Figure 2: Phenotype of the WT and of the *gdh1-2-3* Mutant
(**A**) Phenotype of the WT and of the *gdh1-2-3* mutant at the rosette stage grown for 28 days under hydroponic conditions and corresponding to T0 in the dark-induced experiment.
(**B**) Phenotype of the WT and of the *gdh1-2-3* mutant after floral induction.
(**C**) Phenotype of the WT and of the *gdh1-2-3* mutant after 7 days in the dark
Bars = 1cm



Supplemental Figure 3. Overall Changes in Transcript Accumulation in Roots and Leaves of *gdh1-2-3* Mutants. The rank product method was used to detect differentially expressed genes according to different levels of False Discovery Rate (FDR). Numbers on top of the columns indicate the number of differentially expressed genes.



Supplemental Figure 4. Respiration in Leaves and Roots of the WT and *gdh1-2-3* Mutant Plants Following Prolonged Darkness. T0 = plants grown under short day conditions and harvested 2 h after the beginning of the light period, D3 = after 3 continuous days in the dark, D7 = after 7 continuous days in the dark. Respiration was measured by monitoring O₂ consumption of fresh roots and leaves for 30 to 45 min. The red line corresponds to the *gdh1-2-3* mutant and the blue line to the WT. Results are presented as mean values for five plants with standard errors. Asterisks indicate a bilateral t-test P-value <0.01(**) and <0.05(*) for statistically significant differences.

Supplemental Table 1. Quantification of GDH Protein in Different Tissue Sections of NADH-GDH Mutant Plants. Immunolocalization of the enzyme was performed using transmission electron microscopy on roots and leaves of the Wild Type (WT), and of the *gdh1-2* double mutant and *gdh1-2-3* triple mutant.

	Number of gold particles/ $\mu\text{m}^2 \pm \text{sd}$ in mitochondria of phloem companion cells			
	WT	<i>gdh1-2</i>	<i>gdh1-2-3</i>	Preimmune serum
Leaf	100 \pm 35	10 \pm 6 ^a	7 \pm 4 ^a	6 \pm 3 ^a
Root	310 \pm 40	100 \pm 30	6 \pm 3 ^a	6 \pm 4 ^a

^a statistically not different

Supplemental Table 2. Comparison of the Results Obtained with the CATMA Microarray and a qRT-PCR Experiment for Transcripts Abundance in the *Arabidopsis gdh1-2-3* Mutant Compared to the Wild Type (WT).

AGI ^d	Target transcript	qRT-PCR Average FC ^e <i>gdh1-2-3</i> /WT	SD ^f	Microarray FC ^e <i>gdh1-2-3</i> /WT
AT3G0247C	S-ADENOSYLMETHIONINE DECARBOXYLASE ^a	3.14	0.90	2.34
AT1G2084C	TONOPLAST MONOSACCHARIDE TRANSPORTER1	1.76	0.51	1.93
AT1G6596C	GLUTAMATE DECARBOXYLASE 2	1.78	0.29	1.76
AT3G4845C	^b nitrate-responsive NOI protein, putative	0.68	0.16	0.29
AT1G7206C	Serine-type endopeptidase inhibitor	0.75	0.12	0.30
AT4G3173C	GLUTAMINE DUMPER 1	0.73	0.05	0.59
AT5G0829C	YLS8 ^c	1.16	0.07	1.00
AT4G1342C	K TRANSPORTER	1.14	0.07	1.00
AT1G6180C	Gluc6PT = GPT2	1.19	0.22	1.00

^{a,b,c} Three transcripts exhibiting an increase^a, a decrease^b or no changes^c in the *gdh1-2-3* mutant in comparison to the WT were selected from those listed in Supplementary Dataset 3 and 4.

^d Arabidopsis gene identification

^e Average fold change (FC) between the two replicated experiments

^f Standard deviation (SD)

Supplemental Table 3: Oligonucleotides Used for Screening *Arabidopsis GDH* Mutant Plants. *GDH1*, *GDH2* and *GDH3* indicate the three primers pairs used in the tested gene. "+ ins" indicates the primer pair used to test the presence of the T-DNA in the tested gene with primers located at the intersection between the insertion of the T-DNA and the gene encoding GDH.

Gene	Primer	Primer sequence	Fragment size (bp)
<i>GDH1</i>	Forward	TTCTGATCAAACTCCAGTGAAA	1559
	Reverse	TGATGACACCACCAAGTGCT	
<i>GDH1</i> + ins	Forward	CGGCTACTGGAAGAGGAGTG	1381
	Reverse	TCGCAAGACCCTTCCTCTA	
<i>GDH2</i>	Forward	ACAAGGACGCAACTGGAAGT	736
	Reverse	TCCATTTGGAAGCTCAAT	
<i>GDH2</i> + ins	Forward	ACAAGGACGCAACTGGAAGT	1227
	Reverse	TCGCAAGACCCTTCCTCTA	
<i>GDH3</i>	Forward	GACTCTCTGATCAAATAAACCCAAA	2103
	Reverse	AAGGTTAACTGCAATTAGCACA	
<i>GDH3</i> + ins	Forward	GACTCTCTGATCAAATAAACCCAAA	1256
	Reverse	TCGCAAGACCCTTCCTCTA	

Supplemental Table 4: Oligonucleotides Used for Amplification of RNA to Obtain DNA Fragments Specific for the Three Genes Encoding NADH-GDH and for the β -actin Genes Used as Controls.

Gene	Primer	Primer sequence	Fragment size (bp)
<i>GDH1</i>	Forward	TGGCTCAAGCTACCATTCTCAGA	141
	Reverse	TCGCTCAAGCTACCACTATCAG	
<i>GDH2</i>	Forward	GTGGTTGGGAAGCTTAATTCAGTT	122
	Reverse	TCGCTCAAGCTACCACTATCAG	
<i>GDH3</i>	Forward	TCGCTCAAGCTACCACTATCAG	151
	Reverse	CTGCAATTAGCACATAGTTTTTACTC	
β -actin-2	Forward	AGTGGTCGTACAACCGGTATTGT	93
	Reverse	GATGGCATGGAGGAAGAGAGAAAC	
β -actin-7	Forward	AGTGGTCGTACAACCGGTATTGT	95
	Reverse	GAGGAAGAGCATTCCCCTCGTA	
β -actin-8	Forward	AGTGGTCGTACAACCGGTATTGT	96
	Reverse	GAGGATAGCATGTGGAAGTACTGAGAA	

Supplemental Table 5: Oligonucleotides Used for Amplification of RNA to Obtain DNA Fragments for the qRT-PCR Used to Validate the Microarray Experiment.

Gene	Primer	Primer sequence
<i>PP2A3</i>	Forward	GCAATCTCTCATTCCGATAGTC
	Reverse	ATACCGAACATCAACATCTGG
<i>Q-TIP41</i>	Forward	GCTCATCGGTACGCTCTTTT
	Reverse	TCCATCAGTCAGAGGCTTCC
<i>UBI-10</i>	Forward	GGCCTTGATAATCCCTGATGAATAAG
	Reverse	AAAGAGATAACAGGAACGGAAACATAGT
<i>SAMDC</i>	Forward	GCAATCGGTTTCGAAGGTTA
	Reverse	TCCTTTAGCTCCACCTCGAA
<i>TMT</i>	Forward	ACCCCTATTTCAACGCCTCT
	Reverse	GAGAGTCGCTCCCTTCATTG
<i>GAD2</i>	Forward	GAGAGTCGCTCCCTTCATTG
	Reverse	TCCTTTAGCTCCACCTCGAA
<i>YLS8</i>	Forward	ACTGGGATGAGACCTGTATG
	Reverse	TGTTGTTGTTACCAGTTCCA
<i>HAK5</i>	Forward	TGTTTTCGGACATTTTCGTG
	Reverse	ACCGGCTTTCGAAGTCATAC
<i>G6T</i>	Forward	GACTTTCCTCTTCCTGTCT
	Reverse	AATGGAGTGAGGATCACAAG
<i>NOI</i>	Forward	CAATAAAGCTCGCGATGACA
	Reverse	TCTTTGATCTTGGGGTTTGG
<i>ENDOP</i>	Forward	AAGCTTCCTCCAAGATGCAA
	Reverse	GAAATCGATGAGTGGGAGGA
<i>GDU1</i>	Forward	AATCGCCTTTGCTCTTCTCA
	Reverse	CAAATCTTCTCCGGCCATTA

