



**Figure S1** Allele-specific Expression Assays to Measure the X Inactivation Ratio. **(A)** Location of genes on the X chromosome with assays for allele-specific expression. **(B)** *Pctk1* analysis using previously described Light Cycler Assay (PERCEC *et al.* 2002). The left panel shows the amplification curve of two control progeny samples: CastXm-3 is heterozygous for entire X chromosome; 246-1 is homozygous for 129S1 for the entire X chromosome except for the proximal end of the paternal X chromosome, which is Cast. The right panel

depicts the corresponding melting curves: peak at 60°C corresponds to the Cast allele product; peak at 65°C corresponds to 129S1 allele product. Peak heights were used to calculate the X inactivation ratio (129S1 /(Cast + 129S1)). The ratio for CastXm-3 = 0.23; the ratio for 246-1= 0.50. **(C)** *Mecp2* and *Xist* assays using RFLPs. Lanes shown are pBR322 DNA-*MspI* Digest (M), uncut PCR product (U) and cut PCR product (using *Tsp509I* for *Mecp2* and *SmlI* for *Xist*) for control and RX2 progeny samples. *Mecp2 Tsp509I* 129S1 digested fragment is 217 bp and Cast digested fragments are 155 bp and 62 bp. *Xist SmlI* 129S1 digested fragments are 279 bp, 82 bp and 24 bp, and the Cast digested fragments are 361 bp and 24 bp. Progeny tested in lanes 1-6 are CastXm-1, CastXm-7, 6443-1, 6443-3, 3695-1 and 3695-2, respectively. The ratio as measured by *Mecp2* for corresponding lanes are 0.31 (1), 0.22 (2), 0.40 (3), 0.28 (4), 0.58 (5) and 0.62 (6), and as measured by *Xist* for corresponding lanes are 0.31 (1), 0.21 (2), 0.46 (3) and 0.33 (4). Progeny tested in **(B)** and lanes 1-6 in **(C)** were from control or RX2-derived male mated with 129S1 female.

**Table S1 Genotype of X chromosome of progeny tested mice**

X Marker/Gene	Mb	Xce1 <sup>a</sup>	Xce2 <sup>b</sup>	Control Lines						RX1 Lines					RX2 Lines											
				CastXf	2175f	CastXm	199m	88m	246m	246f	109m	217m	78m	228m	218m	137m	183m	2173m	3695m	6443m	2181m	5005m	6570m	830m	800m	1114m
DXMit53	16.5																									
Pctk1	20.3																									
Hprt1	50.3																									
DXMit73	57																									
DXMit144	61.2																									
Mecp2	71.3																									
DXMit62	90.1																									
DXMit63	90.4																									
DXMit113	91.8/92.1																									
DXMit114	95.3																									
DXMit96	96.4																									
Eda Exon8	97.59																									
DXMit229	97.9																									
DXMit41	98.3																									
DXMit17	98.4																									
DXMit230	98.7																									
DXMit168	98.9																									
Snp 846	99.135															ND										
ss38408987	99.35																									
DXMit115	99.8																									
DXMit148	99.9																									
DXMit95	100.1/3																									
DXMit170	100.2																									
DXPas28	100.5																									
Tsx	100.61/62	ND																								
Xite	100.63	ND																								
DXPas29	100.63																									
Tsix	100.63/68																									
Xist	100.66/68																									
Xist exon3	100.66																									
Xisnpg Ex1	100.67																									
Xist Ex1	100.68																									
ss38407822	100.69																									
ss49779045	100.7																									
DXPas31	100.8																									
DXMit18	100.83																									
DXMit171	101.05																									
DXMit40	101.35																									
Abc7	101.5																									
DXMit64	103																									
DXMit97	116.47																									
DXMit234	138.5																									
DXMit152	144.1																									
Jarid1d1c	148.7																									
DXMit249/31	160.4																									

<sup>a</sup>Simmler et al. and see Figure 2A

<sup>b</sup>Chadwick et al. and see Figure 2A

Not Determined ND

Cast

129S1

Table S2 PCR primers and conditions

Gene or X Chr Location	Primer	Sequence 5' to 3'	NCBI SNP /Polymorphism	Product size /SNP location	Restriction site	Allele specific fragments (bp)	PCR conditions (Anneal Temp. /cycle number)
<b>Hprt1 cDNA</b>	Hprt F3	TGCTGACCTGCTGGATTACA	ss46946097	303bp	SfaNI	303-129S1	61°C
	Hprt R2	GGCCTGTATCCAACACTTCG	A-129/G-Cast Exon6	201bp		192,111-Cast	26-28 cycles
<b>Mecp2 cDNA</b>	Mecp2F3	CCAGTTCCTGCTTGATGTG	NA	217bp	Tsp509I	217-129S1	58°C
	Mecp2R3	TTGTAGTGGCTCATGCTTG	G-129/A-Cast	157bp		155,62-Cast	26-28 cycles
<b>Eda</b>	X97.59f	AGAGGCATTCTGCTGCATT	ss38410803	156bp	StyI	156-129S1	57°C
	X97.59r	TAGGCATGCATGTGGTCATT	G-129S1,C-Cast	120bp		120,36-Cast	35 cycles
<b>X99.35MB</b>	X99.35f	CGGTTGGCGAGTTAGAAAGA	ss38408987	250bp	Tsp45I	93,157-129S1	57°C
	X99.35r	CTGGCCGAGAGTTACCTGAG	G-129S1,T-Cast	96bp		250-Cast	35 cycles
<b>Tsx</b>	Tsx g1f	ATCATTTATGGCCCCCTGA	ss49779081	209pb	ApeKI	129,80-129S1	57°C
	Tsx g2r	AGCTTGGCAAGTGTCCCTCAT	T-129S1,C-Cast Exon4	131bp		209-Cast	35 cycles
<b>Xist cDNA</b>	Xist E2F1	TGGAGTCTGTTTGTGCTCCTGCC	ss38407831	385bp	SmlI	24,82,279-129S1	58°C
	Xist E4R1	CCTTGCTGGGTTCAAGGAAAGCGTC	G-129S1,A-Cast Exon3	106bp		24,361-Cast	26-28 cycles
<b>Xist</b>	Xist IN2F1	TCCGTTACTTGGTTGACTGAGA	ss38407831	245bp	SmlI	168,77-129S1	57°C
	Xist E3R3	TGTTTCAGAGTAGCGAGGACTTG	G-129S1,A-Cast Exon3	168bp		245-Cast	35 cycles
<b>Xist-LC Exon1</b>	XistF2	CTCGTTCCCGTGGATGTG	NA	489bp	No site	NA	57°C
	XistR2	CCGATGGGCTAACGGAGAACG	A-129S1,T-Cast Exon1	172bp			35 cycles
<b>XChr100.69MB</b>	X100.69f	ATATAGCGCCCGAGACTCAA	ss38407822	165bp	TaqαI	165-129S1	57°C
	X100.69r	TCTCGTTGGGACCACACATA	C-129, T-Cast	63bp		63,102-Cast	35 cycles
<b>XChr100.7MB</b>	X100.7f	TTTCTCTGTGTGATAGGGCTT	ss49779045	158bp	BsrI	60,98-129S1	57°C
	X100.7r	AGGAAGTACCCAGGCTCCTC	T-129, G-Cast	64bp		158-Cast	35 cycles
<b>Abcb7 cDNA</b>	Abc F4	TTCGAAAAGCACAAAGCATTC	NA	219bp	Hsp92II	51,158,10-129S1	58°C
	Abc R4	TATCAATGGCCATGTCTGGA	G 129S1,C Cast Exon1	51bp		209,10-Cast	26-28 cycles
<b>Jarid1c cDNA</b>	Jarid F5	TTCCCGAGGAGATGAAGATG	ss38488639	291bp	Hpy188I	292-129S1	58°C
	Jarid R2	CCGCCAAACTCCTCTCTA	C-129S1,T-Cast Exon 8	94bp		96,196-Cast	26-28 cycles

NA Not Applicable