



**Figure S1** Allele-specific Expression Assays to Measure the X Inactivation Ratio. **(A)** Location of genes on the X chromosome with assays for allele-specific expression. **(B)** *Pctk1* analysis using previously described Light Cycler Assay (PERCEC *et al.* 2002). The left panel shows the amplification curve of two control progeny samples: CastXm-3 is heterozygous for entire X chromosome; 246-1 is homozygous for 129S1 for the entire X chromosome except for the proximal end of the paternal X chromosome, which is Cast. The right panel

depicts the corresponding melting curves: peak at 60°C corresponds to the Cast allele product; peak at 65°C corresponds to 129S1 allele product. Peak heights were used to calculate the X inactivation ratio ( $129S1 / (Cast + 129S1)$ ). The ratio for CastXm-3 = 0.23; the ratio for 246-1 = 0.50. **(C)** *Mecp2* and *Xist* assays using RFLPs. Lanes shown are pBR322 DNA-*MspI* Digest (M), uncut PCR product (U) and cut PCR product (using *Tsp509I* for *Mecp2* and *SmlI* for *Xist*) for control and RX2 progeny samples. *Mecp2 Tsp509I* 129S1 digested fragment is 217 bp and Cast digested fragments are 155 bp and 62 bp. *Xist SmlI* 129S1 digested fragments are 279 bp, 82 bp and 24 bp, and the Cast digested fragments are 361 bp and 24 bp. Progeny tested in lanes 1-6 are CastXm-1, CastXm-7, 6443-1, 6443-3, 3695-1 and 3695-2, respectively. The ratio as measured by *Mecp2* for corresponding lanes are 0.31 (1), 0.22 (2), 0.40 (3), 0.28 (4), 0.58 (5) and 0.62 (6), and as measured by *Xist* for corresponding lanes are 0.31 (1), 0.21 (2), 0.46 (3) and 0.33 (4). Progeny tested in **(B)** and lanes 1-6 in **(C)** were from control or RX2-derived male mated with 129S1 female.



**Table S2 PCR primers and conditions**

Gene or X Chr Location	Primer	Sequence 5' to 3'	NCBI SNP /Polymorphism	Product size /SNP location	Restriction site	Allele specific fragments (bp)	PCR conditions (Anneal Temp. /cycle number)
<b>Hprt1 cDNA</b>	Hprt F3	TGCTGACCTGCTGGATTACA	ss46946097	303bp	SfaNI	303-129S1	61°C
	Hprt R2	GGCCTGTATCCAACACTTCG	A-129/G-Cast Exon6	201bp		192,111-Cast	26-28 cycles
<b>Mecp2 cDNA</b>	Mecp2F3	CCAGTTCCTGCTTTGATGTG	NA	217bp	Tsp509I	217-129S1	58°C
	Mecp2R3	TTGTAGTGGCTCATGCTTGC	G-129/A-Cast	157bp		155,62-Cast	26-28 cycles
<b>Eda</b>	X97.59f	AGAGGCATTCTTGCTGCATT	ss38410803	156bp	StyI	156-129S1	57°C
	X97.59r	TAGGCATGCATGTGGTCATT	G-129S1,C-Cast	120bp		120,36-Cast	35 cycles
<b>X99.35MB</b>	X99.35f	CGGTTGGCGAGTTAGAAAGA	ss38408987	250bp	Tsp45I	93,157-129S1	57°C
	X99.35r	CTGGCCGAGAGTTACCTGAG	G-129S1,T-Cast	96bp		250-Cast	35 cycles
<b>Tsx</b>	Tsx g1f	ATCATTTATTTGGCCCCTGA	ss49779081	209pb	ApeKI	129,80-129S1	57°C
	Tsx g2r	AGCTTGGCAAGTGCCTCAT	T-129S1,C-Cast Exon4	131bp		209-Cast	35 cycles
<b>Xist cDNA</b>	Xist E2F1	TGGAGTCTGTTTTGTGCTCCTGCC	ss38407831	385bp	SmlI	24,82,279-129S1	58°C
	Xist E4R1	CCTTGCTGGGTTTCAGGAAAGCGTC	G-129S1,A-Cast Exon3	106bp		24,361-Cast	26-28 cycles
<b>Xist</b>	Xist IN2F1	TCCGTTACTTGGTTGACTGAGA	ss38407831	245bp	SmlI	168,77-129S1	57°C
	Xist E3R3	TG TTCAGAGTAGCGAGGACTTG	G-129S1,A-Cast Exon3	168bp		245-Cast	35 cycles
<b>Xist-LC Exon1</b>	XistF2	CTCGTTTCCCGTGGATGTG	NA	489bp	No site	NA	57°C
	XistR2	CCGATGGGCTAAGGAGAAG	A-129S1,T-Cast Exon1	172bp			35 cycles
<b>XChr100.69MB</b>	X100.69f	ATATAGCGCCCGAGACTCAA	ss38407822	165bp	TaqαI	165-129S1	57°C
	X100.69r	TCTCGTTGGGACCACACATA	C-129, T-Cast	63bp		63,102-Cast	35 cycles
<b>XChr100.7MB</b>	X100.7f	TTTCTCCTGTGTGATAGGGTCTT	ss49779045	158bp	Bsrl	60,98-129S1	57°C
	X100.7r	AGGAAGTACCCAGGCTCCTC	T-129, G-Cast	64bp		158-Cast	35 cycles
<b>Abcb7 cDNA</b>	Abc F4	TTCGAAAAGCACAAAGCATTC	NA	219bp	Hsp92II	51,158,10-129S1	58°C
	Abc R4	TATCAATGGCCATGTCTGGA	G 129S1,C Cast Exon1	51bp		209,10-Cast	26-28 cycles
<b>Jarid1c cDNA</b>	Jarid F5	TTCCCGAGGAGATGAAGATG	ss38488639	291bp	Hpy188I	292-129S1	58°C
	Jarid R2	CCGCCAAAACCTCTTCTCTA	C-129S1,T-Cast Exon 8	94bp		96,196-Cast	26-28 cycles
NA Not Applicable							