

SUPPORTING INFORMATION

Oxidation of Methyl and Ethyl Nitrosamines by Cytochromes P450 2E1 and P450 2B1

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Biochemistry **52**, xxx-xxx (2013)

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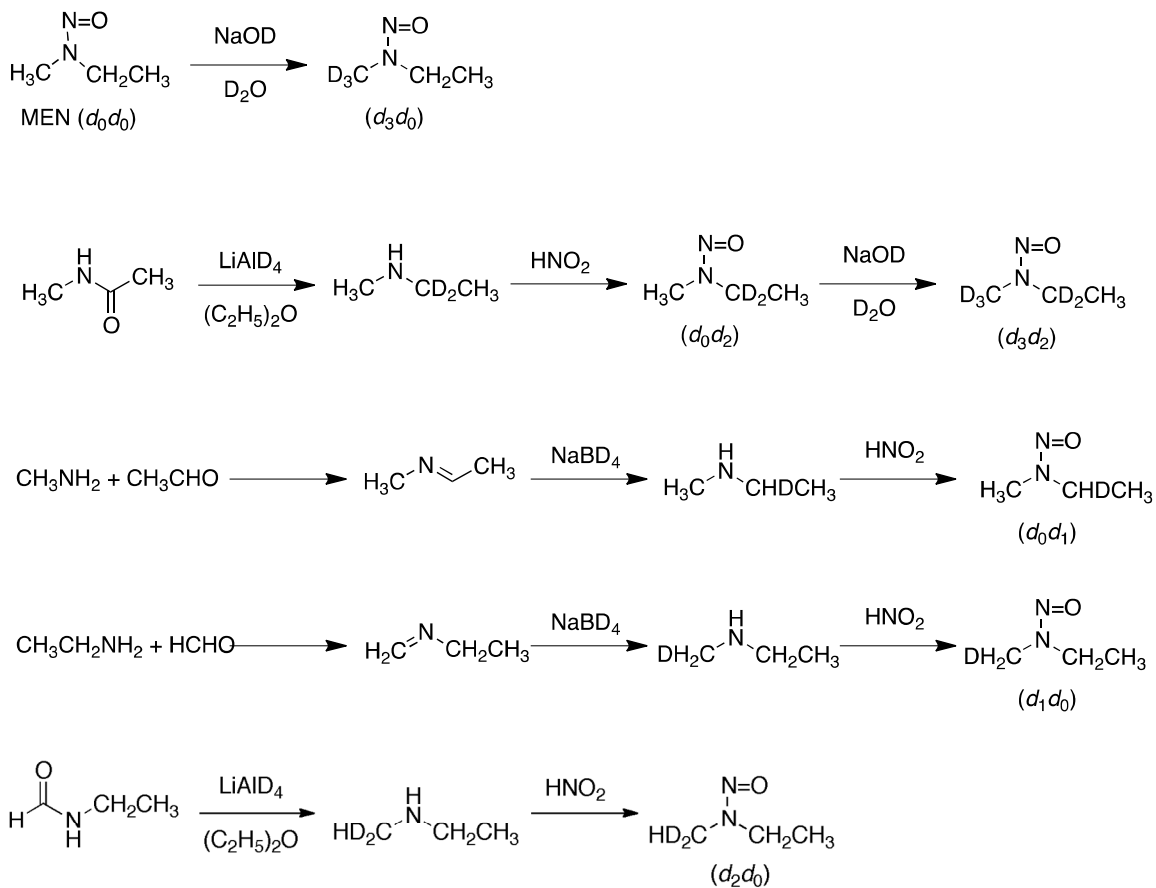
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Figure S1. (A) Synthesis of deuterated MEN substrates; (B) ^1H -NMR spectra. Note splitting of signals in nitrosamines, as discussed earlier.¹

A



[d_3d_0]-MEN. MEN (4.15 g, 47 mmol) was dissolved in 50 mL of 0.5 M NaOD in D_2O , which had been prepared by the addition of 0.57 g of Na° in 50 mL D_2O (at 0°C), with a CaSO_4 drying tube attached to the reaction flask. The solution was stirred at 70°C for 48 h. The mixture was cooled to room temperature and the pH (pD) was adjusted to ~ 4 (as judged by indicator paper) with D_3PO_4 (prepared by the careful addition of P_2O_5 to D_2O). NaCl (~ 12 g) was dissolved in the aqueous solution, and the solution was extracted three times with an equal volume of CHCl_3 . The combined organic layers were dried with MgSO_4 and, after removal of MgSO_4 by filtration, the CHCl_3 was evaporated carefully under a stream of N_2 in a water bath. Yield 77%. MS: m/z 92.3 (MH^+ for d_3). NMR (CDCl_3) δ 1.04 (t, $-\text{CH}_2\text{CH}_3$), 1.95 (t, $-\text{CH}_2\text{CH}_3$), 1.33 (t, $-\text{CH}_2\text{CH}_3$), 2.96 (s, residual $-\text{CH}_3$, $\leq 3\%$), 3.56 (q, $-\text{CH}_2\text{CH}_3$), 4.14 (q, $-\text{CH}_2\text{CH}_3$).

[d_0d_2]-MEN. LiAlD_4 (10 g, 0.24 mol) was stirred in 300 mL of dry $(\text{C}_2\text{H}_5)_2\text{O}$ (freshly opened can) in a 500 mL-round bottom flask, which was stirred and cooled to 0°C using an ice bath. (All glass had been heated to remove moisture, and a CaSO_4 drying tube was attached to the system.) *N*-Methylacetamide (17 g, 0.24 mol, dissolved in 60 mL of $(\text{C}_2\text{H}_5)_2\text{O}$), was added from an addition funnel over 1 h. The addition funnel was

replaced with a condenser and the solution was stirred and heated at reflux for 20 h. The reaction was cooled to 0 °C and quenched by the careful addition of 5 mL D₂O, followed by 5 mL of 15% NaOD in D₂O (*vide supra*) and then another 5 mL of D₂O. This mixture was stirred another 17 h. After cooling on ice, the solids were removed by filtration through Celite and washed with another cold 200 mL of (C₂H₅)₂O. The combined filtrate was chilled to 0 °C (ice) and sparged with HCl gas. The resulting CH₃NHCH₂CH₃-HCl was extracted from the (C₂H₅)₂O layer three times with 25 mL of aqueous 2.5 M HCl. The water was removed *in vacuo* (rotary evaporator) to yield a solid, which was dried *in vacuo* (vacuum pump) overnight.

The above compound (moist solid) was dissolved in 60 mL of cold glacial CH₃CO₂H and diluted with 30 g ice. The solution was stirred at 0 °C for 1 h, NaNO₂ (35 g, 0.51 mol) was added, and the reaction was stirred for 1.5 h at 0 °C and then overnight at 23 °C. Cold H₂O (25 mL) and NaCl (10 g) were added, and the product (*d*₀*d*₂-MEN) was extracted five times with 25 mL of CHCl₃. The combined CHCl₃ extracts were dried with MgSO₄ and filtered, and the CHCl₃ was carefully evaporated, under a stream of nitrogen at 23 °C, to give 9.7 g of [*d*₀*d*₂]-MEN (46% yield). NMR (CDCl₃) δ, 0.96 (s, -CD₂CH₃), 1.25 (s, -CD₂CH₃), 2.93 (s, CH₃-N), 3.62 (s, 3H, CH₃-N). The product was further purified by vacuum distillation (bp 50-55 °C, 12 Torr) (6.2 g).

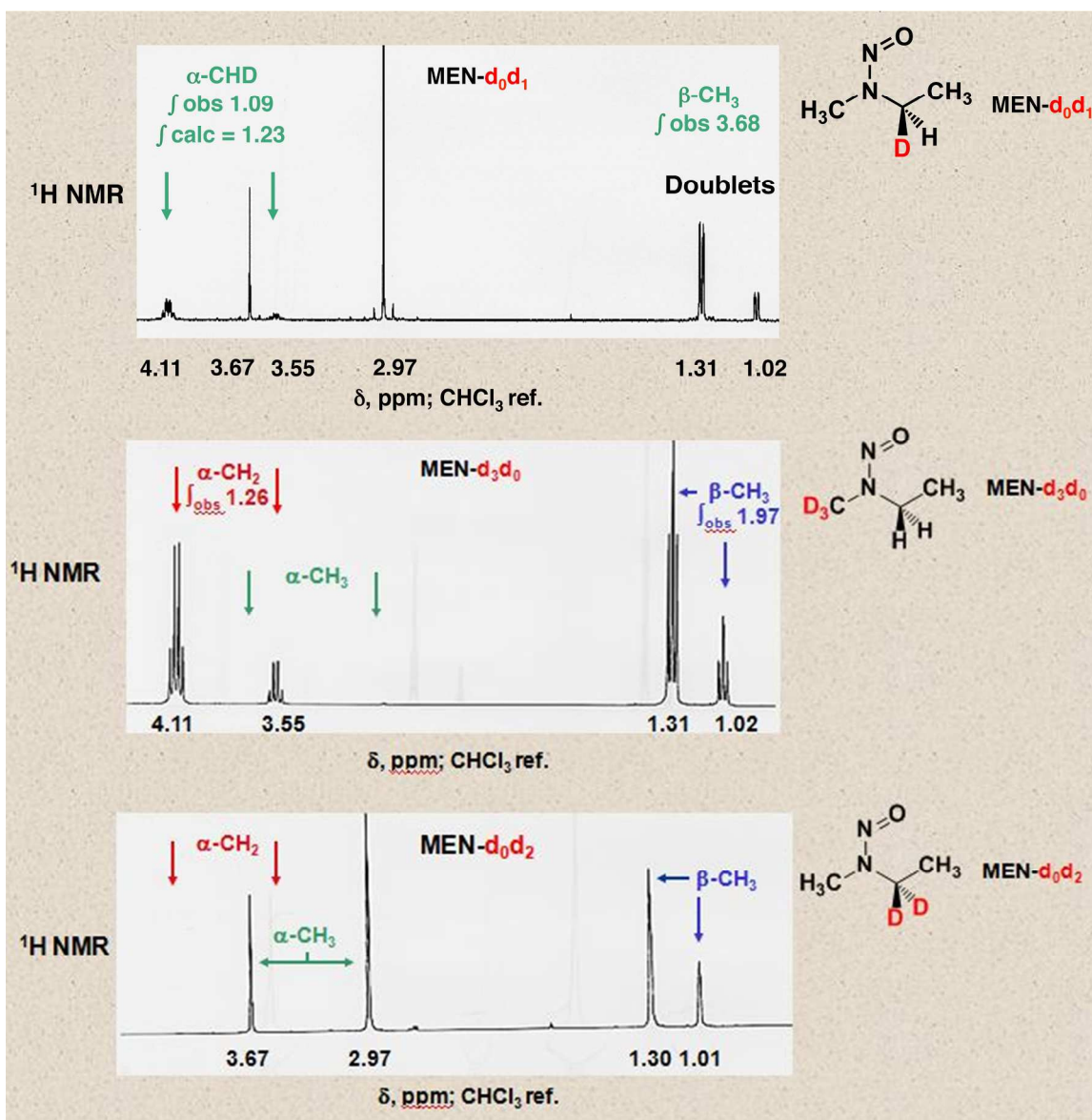
[*d*₃*d*₂]-MEN. The above product ([*d*₀*d*₂]-MEN, 5.7 g, distilled) was exchanged with 100 mL of 0.5 M NaOD in H₂O as described for [*d*₃*d*₀]-MEN (*vide supra*) for 130 h at 70 °C to yield [*d*₃*d*₂]-MEN with 98.5% excess atom D. MS: *m/z* 94.3 (MH⁺). NMR (CDCl₃) δ 1.01 (s, -CD₂CH₃), 1.30 (s, -CD₂CH₃).

[*d*₀*d*₁]-MEN. A cold aqueous solution of 40% CH₃CHO (w/v, 20 mL, 0.18 mol) was added dropwise to 24 mL of a 40% aqueous solution of CH₃NH₂ on ice, with vigorous stirring. After 2 h, NaOH pellets were added to the mixture until two layers formed, and then a few excess NaOH pellets were added. The mixture, containing the imine (Fig. 2), was transferred to a pre-chilled separatory funnel. The lower aqueous layer was discarded and the upper layer (imine, 16.9 g crude weight) was transferred to a clean round bottom flask and distilled (bp 26-32 °C, 760 Torr) to give 9.2 g of the imine: NMR (CDCl₃) δ 0.88 (s, 3H, CH₃-N), 1.81 (s, 3H, =CH-CH₃), 2.35 (s, residual CH₃NH₂), 3.12 (s, unknown). The imine was dissolved in 50 mL C₂H₅OH and reduced with 2.0 g of NaBD₄ (0.048 mol, in 100 C₂H₅OH), beginning at 0 °C (during addition), heating at 50 °C for 3 h, and then stirring at 23 °C overnight. The resulting *N*-methylethylamine was recovered and nitrosated as described above for [*d*₀*d*₂]-MEN. MS: *m/z* 90.2 (MH⁺). NMR (CDCl₃) δ 1.32 (d, 3H, -CHDC₂H₃), 2.98 (s, 3H, CH₃-N), 3.67 (s, 3H, CH₃-N), 4.10 (m, 1H, -CHDC₂H₃). See Part B for spectrum.

[*d*₁*d*₀]-MEN. The procedure was similar to that used above for [*d*₁*d*₀]-MEN, except that HCHO (20 mL of a 30% solution, w/v) and C₂H₅NH₂ (10.3 mL of a 70% aqueous solution) were used as starting materials (Fig. 2). The imine intermediate (crude yield 7.1 g, 45%) was not distilled but was reduced directly with NaBD₄ (4.0 g) in C₂H₅OH, with the workup of *N*-methylethylamine and nitrosation as before to yield the final product, *d*₁*d*₀-MEN. MS: *m/z* 90.2 (MH⁺).

[d_2d_0]-MEN. The procedure was similar to that used above for [d_0d_2]-MEN (*vide supra*). *N*-Ethylformamide was reduced with LiAlD_4 and the product was nitrosated in the usual manner. MS: m/z 91.2 (MH^+). NMR (CDCl_3) δ 1.05, 1.34 (t, 3H, $\text{N-CH}_2\text{CH}_3$); 2.97, 3.67 (m, 1H, $\text{CD}_2\text{H-N}$); 3.58, 4.14 (q, 2H, $\text{N-CH}_2\text{CH}_3$).

B



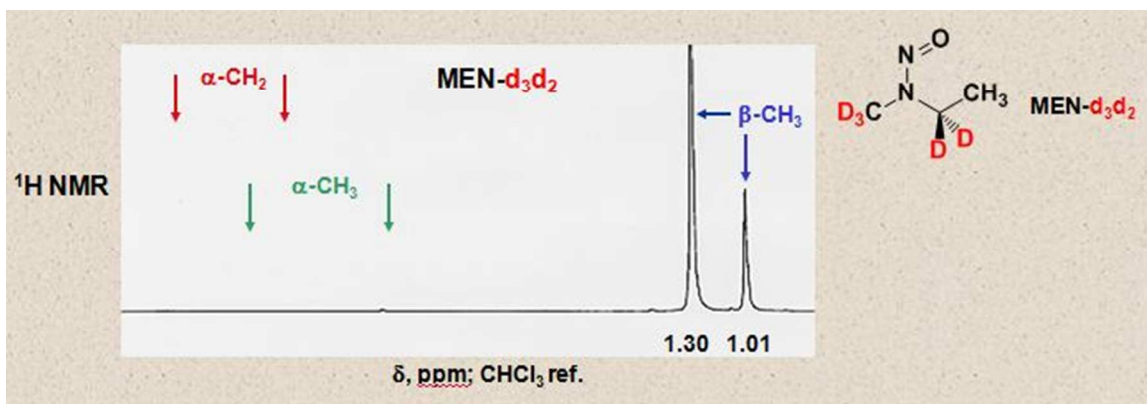
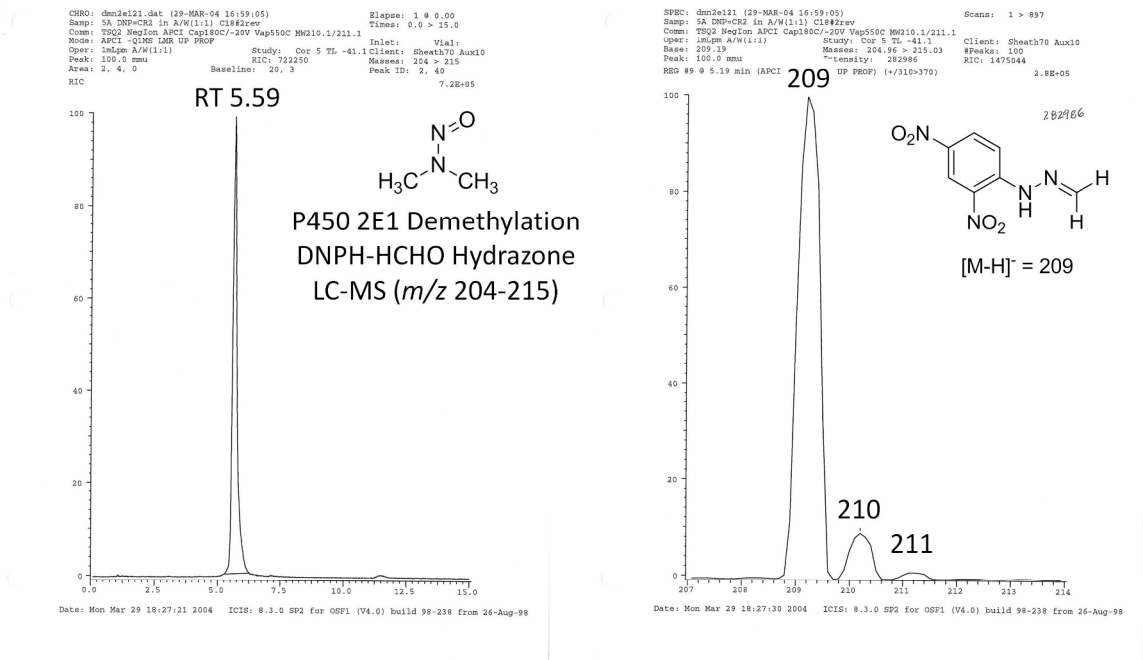


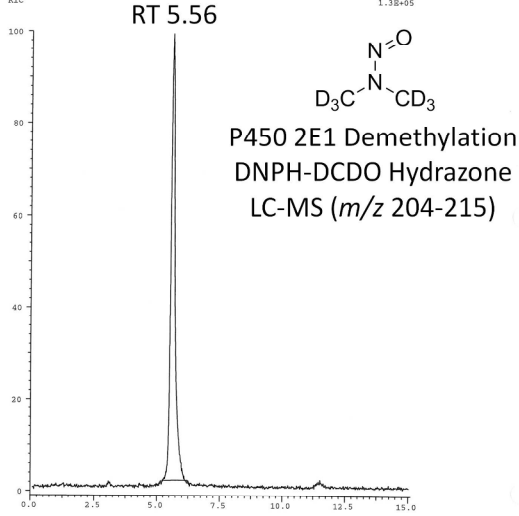
Figure S2. LC-MS of 2,4-dinitrophenyl hydrazone of CH₃CHO formed from DMN. (A) LC-MS chromatogram and mass spectrum of product obtained from an incubation with *d*₀ DMN. (B) LC-MS chromatogram and mass spectrum of product obtained from an incubation with *d*₆ DMN. (C) LC-MS chromatogram and mass spectrum of product obtained from an incubation with a 1/1 molar mixture of *d*₀ and *d*₆ DMN (competition experiment).

A

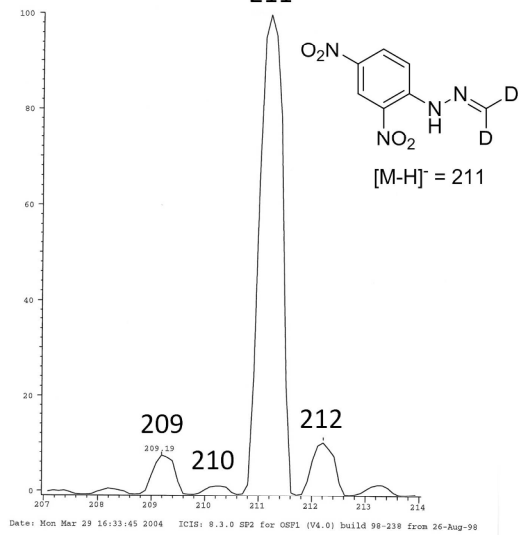


B

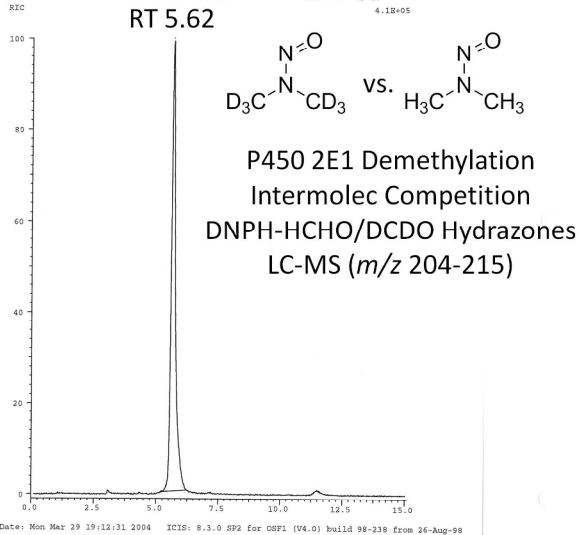
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 Mode: APCI -Q1MS LMS UP PROF Inlet: Vial:
 Oper: 10Lpm A/W(1:1) Study: Cor 5 TL -41.1 Client: Sheath70 Aux10
 Peak: 100.0 mmu RIC: 134530 Masses: 204 > 215
 Area: 2.4 C Baseline: 20.3 Peak ID: 2, 40
 RIC: 1.3E+05



SPRC: dm2e116 (29-MAR-04 15:40:54) Scans: 1 > 897
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 Mode: APCI -Q1MS LMS UP PROF Inlet: Vial:
 Oper: 10Lpm A/W(1:1) Study: Cor 5 TL -41.1 Client: Sheath70 Aux10
 Base: 211.21 Masses: 204.96 > 215.03 #Peaks: 100
 Peak: 100.0 mmu Intensity: 4.7E+04 RIC: 930749
 REG #9 @ 5.19 min (APCI -Q1MS LMR UP PROF) (-) 211

**C**

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 Mode: APCI -Q1MS LMS UP PROF Inlet: Vial:
 Oper: 10Lpm A/W(1:1) Study: Cor 5 TL -41.1 Client: Sheath70 Aux10
 Peak: 100.0 mmu RIC: 411247 Masses: 204 > 215
 Area: 2.4 C Baseline: 20.3 Peak ID: 2, 40
 RIC: 4.1E+05



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 Oper: 10Lpm A/W(1:1) Study: Cor 5 TL -41.1 Client: Sheath70 Aux10
 Base: 209.19 Masses: 204.96 > 215.03 #Peaks: 100
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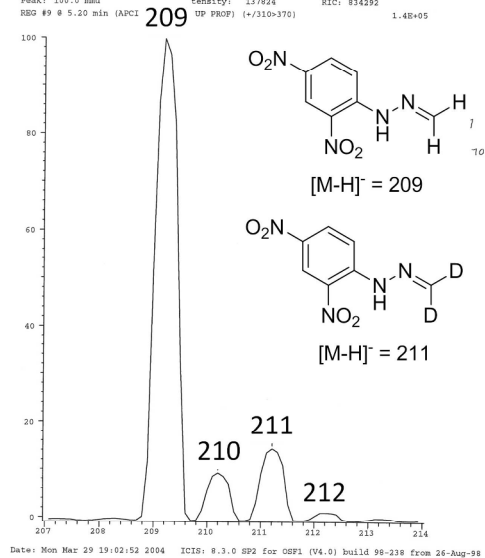
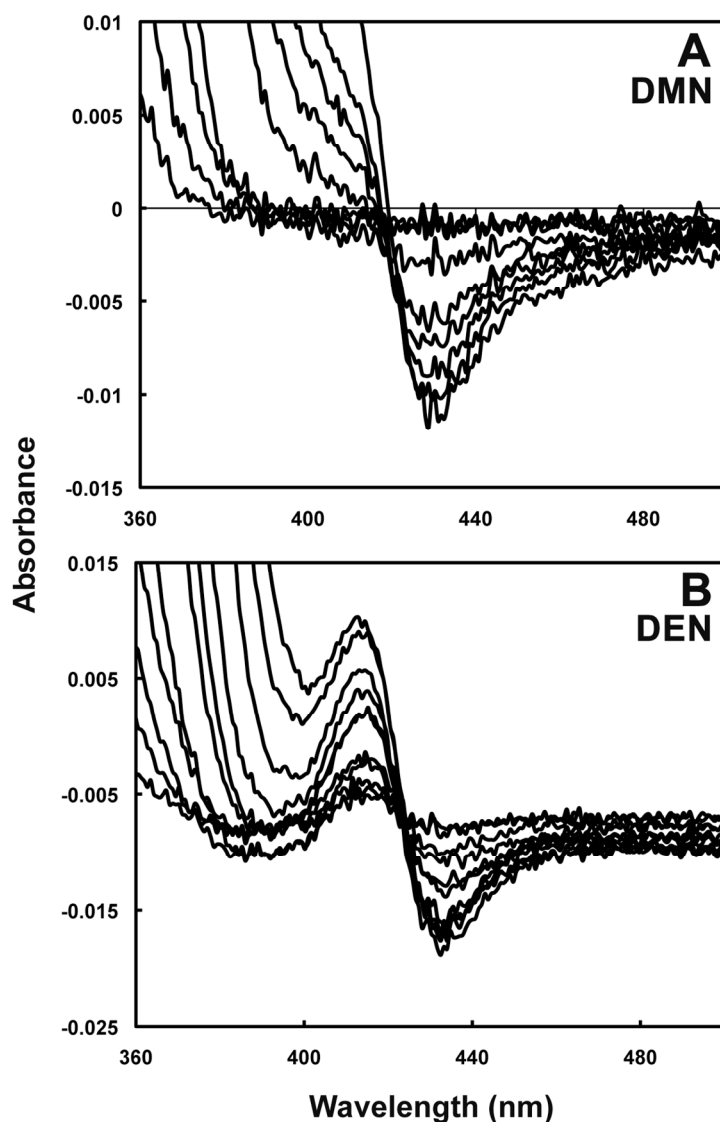
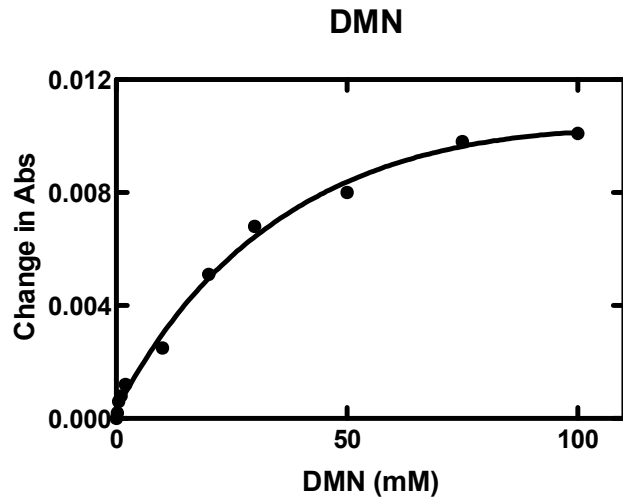


Figure S3. Binding spectra of P450 2E1 with DMN and DEN.⁴ Two cuvettes each contained 1.0 μM P450 in 0.10 M potassium phosphate buffer. A baseline was recorded and varying concentrations of the nitrosamine (in H_2O) were added to the sample cuvette, with the same amount of H_2O added to the reference cuvette. (A) DMN spectra; (B) DEN spectra; (C) plot of absorbance change (A_{430}) vs. [DMN]; (D) plot of absorbance change ($\Delta A_{413}-A_{431}$) vs. [DEN]. The K_d values for DMN and DEN were $69 (\pm 41)$ and $0.99 (\pm 0.20)$ mM, respectively.



C



D

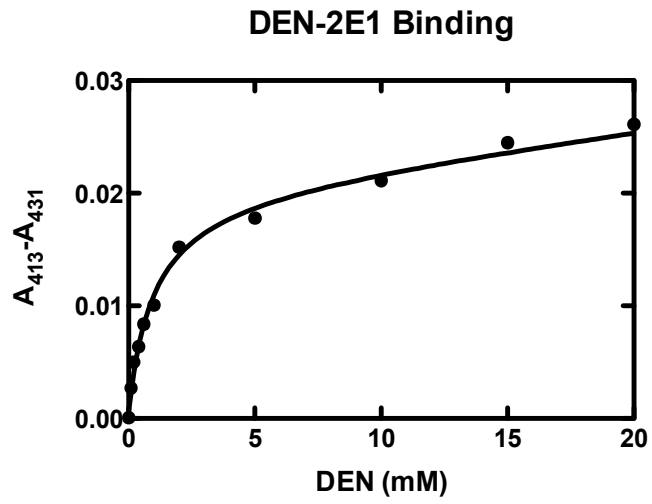


Figure S4. DynaFit modeling⁵ of theoretical time courses of formation of aldehydes and carboxylic acids based on k_{cat} and K_m values for separate reactions.

Script

```
;2E1DAN RUN 3  
;This is a situation in which Km & kcat results are available for several  
;pathways. The goal is to simulate the time course of different products as  
;a function of time.
```

```
; units are min & nM  
; E = P450  
; S = DMN  
; Q = aldehyde  
; P = carboxylic acid  
; Use k = Vmax = kcat  
; Use Km = Kdissoc
```

```
[task]
```

```
data = progress  
task = fit
```

```
[mechanism]
```

```
E + S <=> ES :      K1    dissoc  
ES --> EQ      :      k2  
E + Q <=> EQ :      K3    dissoc  
EQ ---> EP :      k4  
E + P <=> EP :      K5    dissoc
```

```
[constants]
```

```
K1 = 69000000  
k2 = 13.8  
K3 = 1300000  
k4 = 0.72  
K5 = 10000000000
```

```
[sweep]
```

```
[concentrations]
```

```
E = 2500  
S = 1000000
```

```
[progress]
```

mesh linear from 0 to 10 step 0.1

directory ./scripts

```
;variable S  
;file DMNHCO2Htime.txt  
;response S = 1
```

```
variable Q  
file DMNHCO2Htime.txt  
response Q = 1
```

```
variable P  
file DMNHCO2Htime.txt  
response P = 1
```

[output]
directory ./projects/P4502E1DAN/output/run3

[end]

Data file

DMNHCO2Htime.txt
(time, in min; concentration, in nM)

0	0
1	48
2	53
3	185
4	395
5	280
6	368
6	380

Differential equations

$$d[E]/dt = +K1[ES]-dissoc[E][S]+K3[EQ]-dissoc[E][Q]+K5[EP]-dissoc[E][P]$$

$$d[S]/dt = +K1[ES]-dissoc[E][S]$$

$$d[ES]/dt = -K1[ES]+dissoc[E][S]-k2[ES]$$

$$d[EQ]/dt = +k2[ES]-K3[EQ]+dissoc[E][Q]-k4[EQ]$$

$$d[Q]/dt = +K3[EQ]-dissoc[E][Q]$$

$$d[EP]/dt = +k4[EQ]-K5[EP]+dissoc[E][P]$$

$$d[P]/dt = +K5[EP]-dissoc[E][P]$$

Predicted traces (for HCHO (1) and HCO₂H (2, with data points shown)):

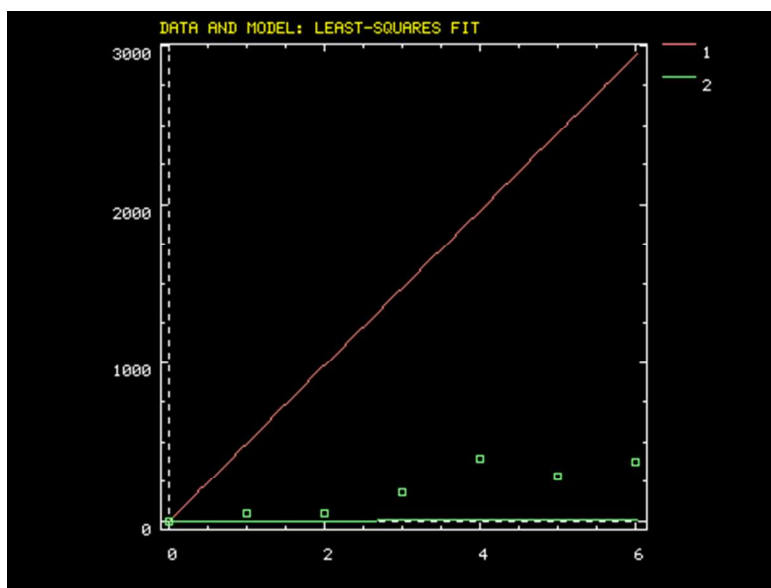
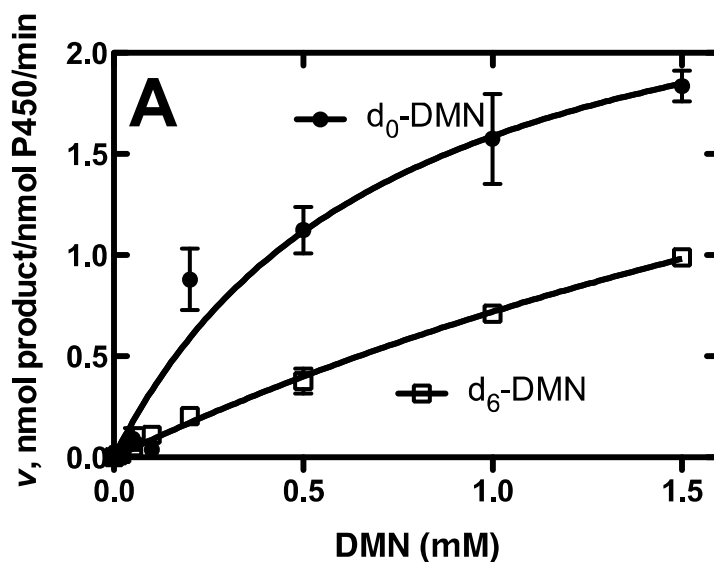
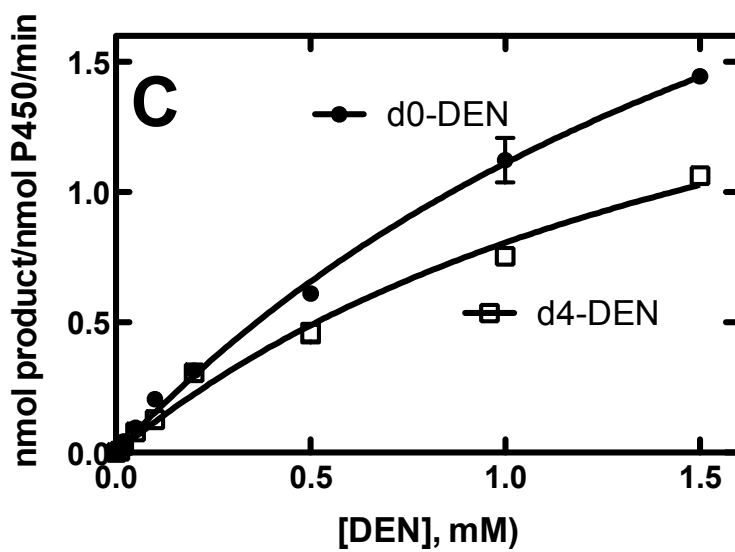
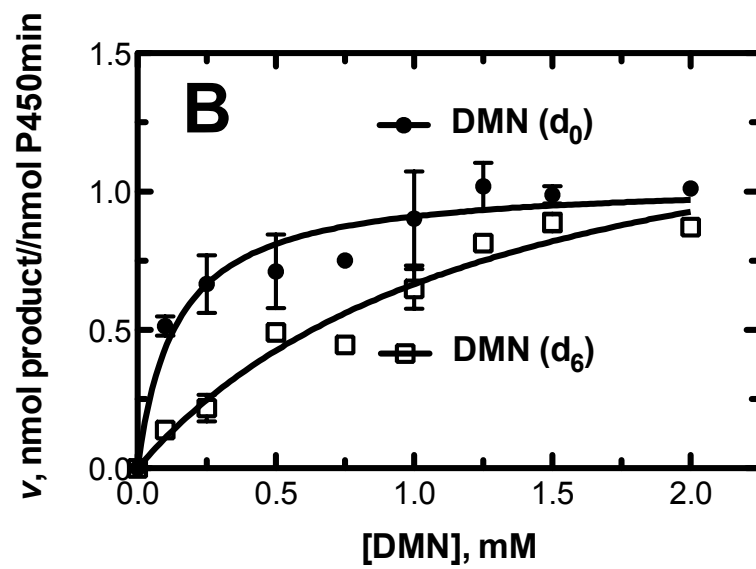


Figure S5. Steady-state kinetics of denitrosation products. Incubations were done for 15 min at 37 °C with 1 μ M human P450 2E1 in the typical reconstituted system containing NADPH-P450 reductase and b_5 . Analyses of alkylamines² and NO_2^- ³ are described in more detail under Experimental Procedures. (A) Formation of CH_3NH_2 from d_0 and d_6 DMN. (B) Formation of NO_2^- from d_0 and d_6 DMN. (C) Formation of $\text{CH}_3\text{CH}_2\text{NH}_2$ from d_0 and d_4 DEN ($\text{CH}_3\text{CD}_2\text{N}(\text{NO})\text{CD}_2\text{CH}_3$). (D) Formation of NO_2^- from d_0 and d_4 DEN ($\text{CH}_3\text{CD}_2\text{N}(\text{NO})\text{CD}_2\text{CH}_3$).





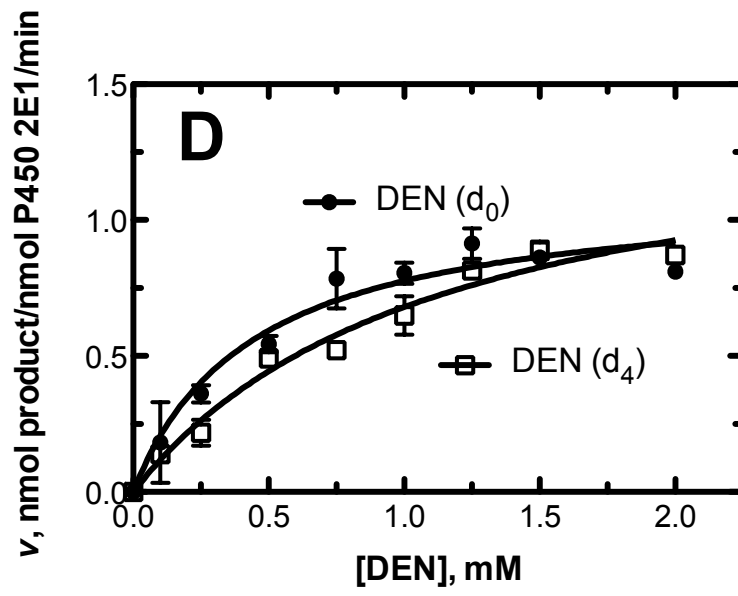


Figure S6. Steady-state kinetics of *N*-demethylation of [*d*₄]-DMN to formaldehyde by rat P450 2B1. Apparent k_{cat} $1.6 \pm 0.2 \text{ min}^{-1}$ (see text for correction for isotopes), K_m $2.5 \pm 0.5 \text{ mM}$. Dansyl hydrazone derivatives were analyzed by LC-MS, and the fit is to a hyperbolic (Michaelis-Menten) plot.

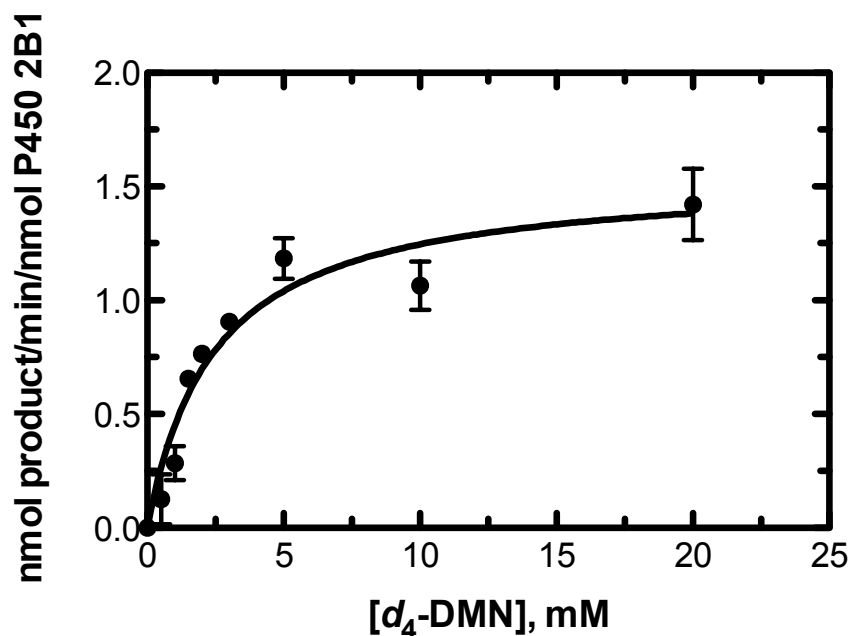
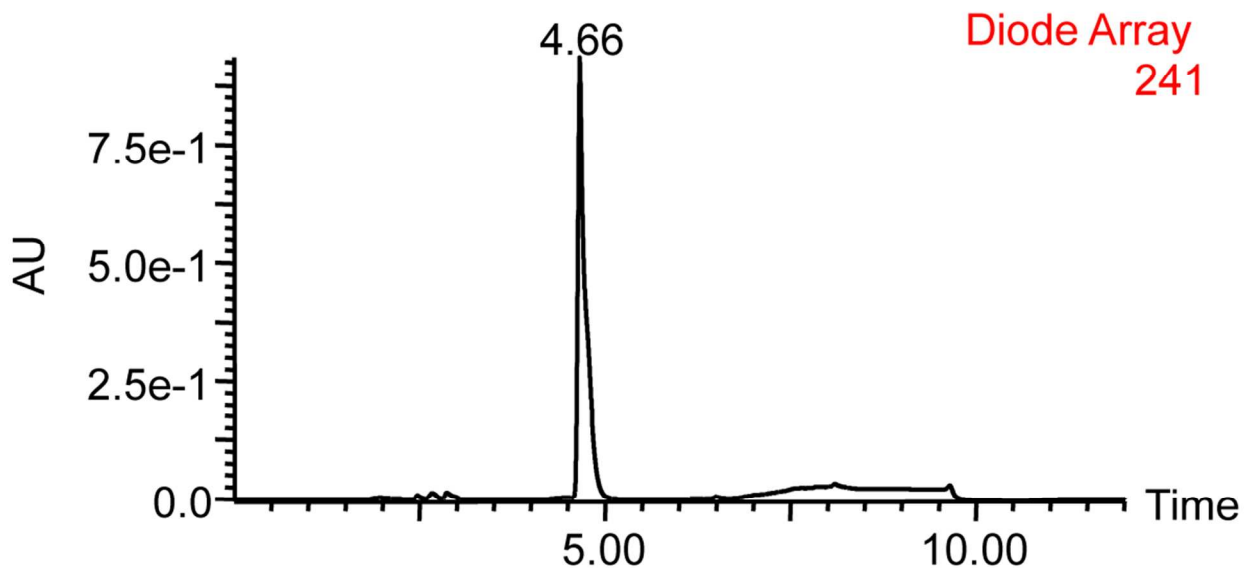


Figure S7. HPLC-UV of *N*-nitroso-*N*-ethylacetamide. UV absorbance at 241 nm was collected and is plotted on the y-axis.



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