

SUPPLEMENTAL MATERIAL

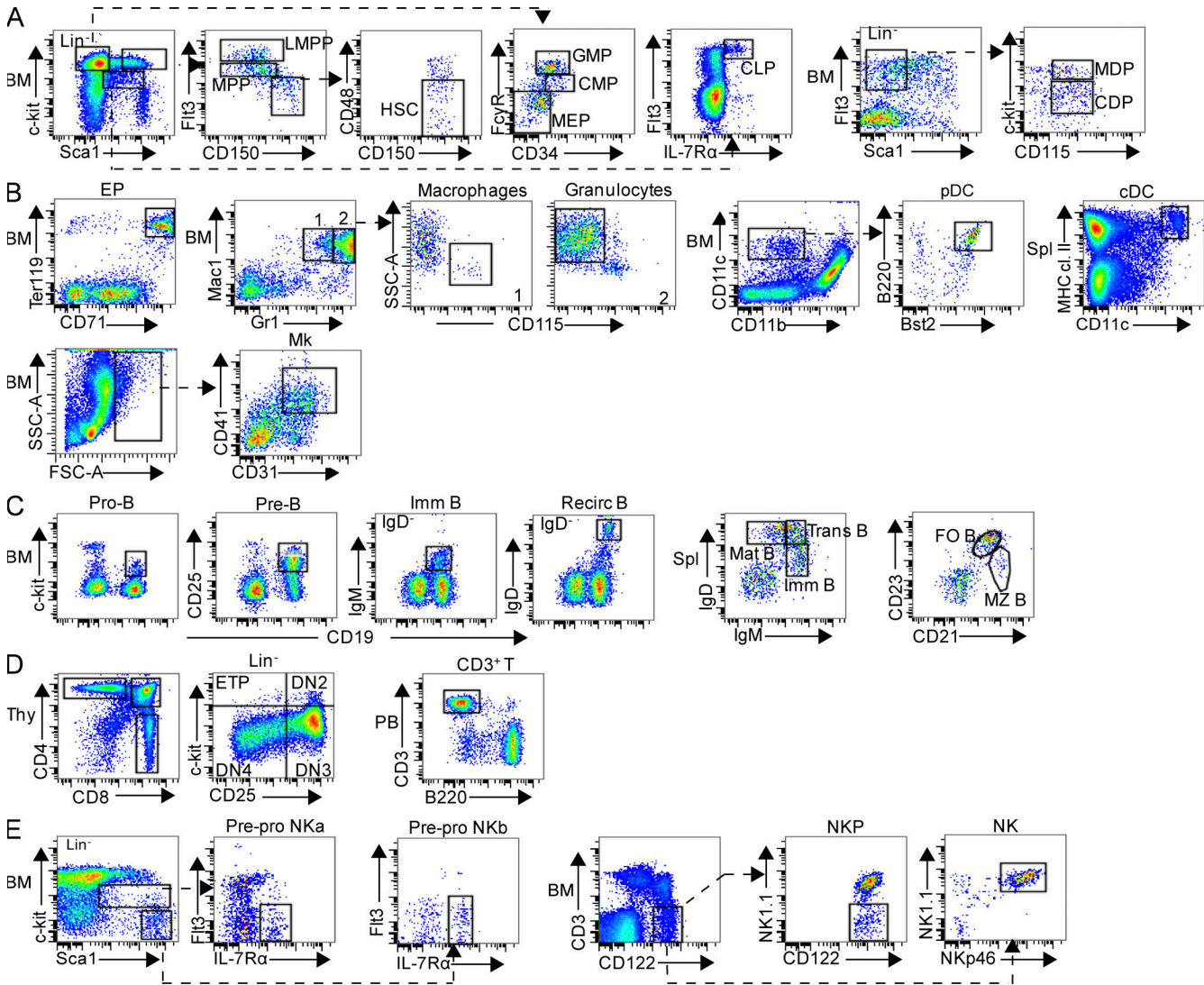
Yu et al., <http://www.jem.org/cgi/content/full/jem.20121846/DC1>

Figure S1. Gating strategies of hematopoietic populations. (A) HSC, MPP, LMPP, CMP, GMP, MEP, CLP, monocyte-dendritic precursor (MDP) and CDP. (B) EPs, macrophages, granulocytes, plasmacytoid dendritic cells (pDC), conventional dendritic cells (cDC), and megakaryocytes. (C) B cell compartment. (D) T cells. (E) NK cells in the BM. In all flow cytometry assays, at least 4 wild-type mice were analyzed for each cell type in independent experiments.

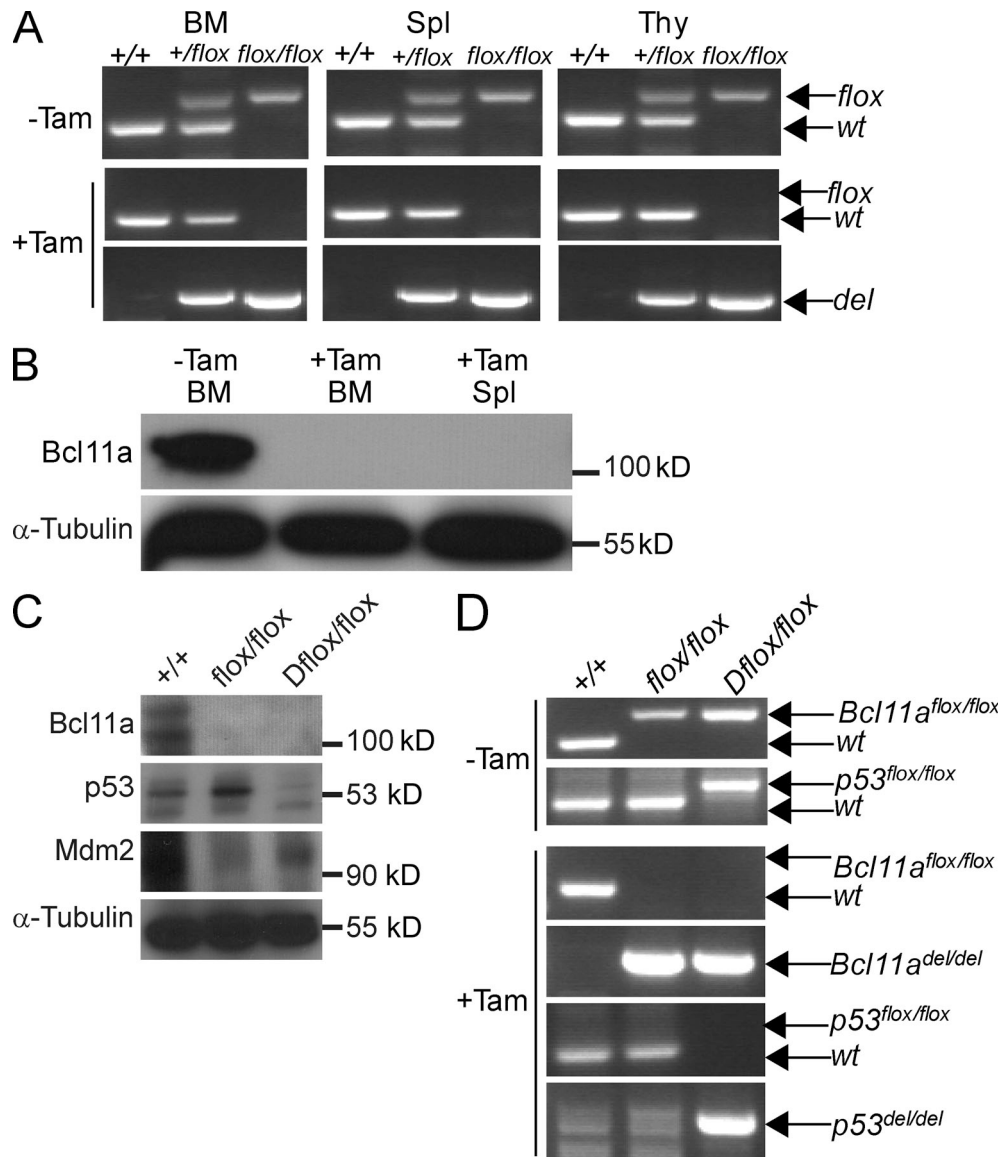


Figure S2. Genotyping strategies. (A) Genotyping cells of the BM, spleen (Spl), and thymus (Thy) for *Bcl11a* deletion (absence of the *flox* allele). *wt*, wild-type allele; *flox*, conditional knockout allele; *del*, deletion allele. (B) Western blotting analysis of BM and Spl cells for Bcl11a protein. α -Tubulin was the loading control. (C) Western blot analysis of Bcl11a, p53, and Mdm2 in cultured BM B cells of indicated genotypes. Loading control, α -tubulin. (D) Genotyping of cells for *Bcl11a* and p53 deletion. *Bcl11a^{flox/flox}*, *Bcl11a* conditional knockout allele; *p53^{flox/flox}*, p53 conditional knockout allele; *Bcl11a^{del/del}*: *Bcl11a* deletion allele. *p53^{del/del}*: p53 deletion allele.

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Table S1. Hematopoietic population definitions

Tissue	Population	Makers	Lineage
Bone marrow	HSC	Lin ⁻ Scal ⁺ c-kit ⁺ Flt3 ⁻ CD48 ⁻ CD150 ⁺	B220, CD3ε, CD4,
	MPP	Lin ⁻ Scal ⁺ c-kit ⁺ Flt3 ^{inter} CD150 ⁻	CD8α, NK1.1,
	LMPP	Lin ⁻ Scal ⁺ c-kit ⁺ Flt3 ⁺	CD11b, Ly6G,
	CLP	Lin ⁻ Flt3 ⁺ IL-7Rα ⁺ scal ^{low} c-kit ^{low}	Ter119
	Pre-pro NKa	Lin ⁻ Scal ⁺ c-kit ^{int} Flt3 ⁻ IL-7Rα ⁺	
	Pre-pro NKb	Lin ⁻ Scal ⁺ c-kit ⁻ Flt3 ⁻ IL-7Rα ⁺	
	CMP	Lin ⁻ Scal ⁻ c-kit ⁺ FcyR ^{int} CD34 ^{int}	B220, CD19
	GMP	Lin ⁻ Scal ⁻ c-kit ⁺ FcyR ^{hi} CD34 ^{hi}	CD3ε, CD4, CD8α,
	MEP	Lin ⁻ Scal ⁻ c-kit ⁺ FcyR ^{low} CD34 ^{low}	NK1.1, Ly6G,
	MDP	Lin ⁻ Scal ⁻ c-kit ^{hi} Flt3 ⁺ CD115 ⁺	IgM, IL-7Rα,
	CDP	Lin ⁻ Scal ⁻ c-kit ^{low} Flt3 ⁺ CD115 ⁺	Ter119
	Pro-B	B220 ⁺ CD19 ⁺ CD43 ⁺ c-Kit ⁺ IgM ⁻	
	Pre-B	B220 ⁺ CD19 ⁺ CD43 ⁻ CD25 ⁺ IgM ⁻	
	Imm B	B220 ⁺ CD19 ⁺ IgM ^{hi} IgD ⁻	
	Recirc B	B220 ⁺ CD19 ⁺ IgD ⁺	
	NKP	CD122 ⁺ NK1.1 ⁻ CD3 ⁻	
	NK	NK1.1 ⁺ CD122 ⁺ NKp46 ⁺ CD3 ⁻	
	Macrophages	Mac1 ⁺ Gr1 ^{low} CD115 ⁺	
	Granulocytes	Mac1 ⁺ Gr1 ^{hi} CD115 ⁻	
	EP	CD11b ⁻ Ly6G ⁻ CD3 ⁻ B220 ⁻ Ter119 ⁺ CD71 ⁺	
	pDC	CD11b ⁻ CD11c ^{low} B220 ⁻ Bst2 ⁺	
	cDC	CD11c ^{hi} MHC cI.II ⁺	
	Megakaryocytes	FSC ^{hi} CD41 ⁺ CD31 ⁺	
Thymus	ETP	Lin ⁻ c-kit ^{hi} CD25 ⁻	B220, CD19, CD3ε,
	DN2	Lin ⁻ c-kit ^{hi} CD25 ⁺	CD8α, TCRβ, TCRγδ,
	DN3	Lin ⁻ c-kit ⁻ CD25 ⁺	NK1.1, CD11c, CD11b
	DN4	Lin ⁻ c-kit ⁻ CD25 ⁻	, Ly-6G, Ter119
Spleen	Imm B	CD19 ⁺ IgM ^{hi} IgD ⁻	
	Trans B	CD19 ⁺ IgM ^{hi} IgD ^{hi}	
	Mat B	CD19 ⁺ IgM ^{low} IgD ⁻	
	MZ B	CD21 ^{hi} CD23 ^{low}	
	FO B	CD21 ^{int} CD23 ^{hi}	
	NK	NK1.1 ⁺ Ter119 ⁻ Gr1 ⁻ Mac1 ⁻ B220 ⁻ CD3 ⁻	
	CD3 ⁺ T	Ter119 ⁻ Gr1 ⁻ Mac1 ⁻ B220 ⁻ CD3 ⁺	
Peripheral blood	B	Ter119 ⁻ Gr1 ⁻ Mac1 ⁻ B220 ⁺ CD3 ⁻	
	T	Ter119 ⁻ Gr1 ⁻ Mac ⁻ B220 ⁻ CD3 ⁺	
	NK	NK1.1 ⁺ Ter119 ⁻ Gr1 ⁻ Mac ⁻ B220 ⁻ CD3 ⁻	
	GM	Ter119 ⁻ Mac1 ⁺ Gr1 ⁺ CD3 ⁻ B220 ⁻	

Table S2. Antibodies in this study

Epitope	Clone	Conjugate	Company
B220	RA3-6B2	Brilliant Violet 421	BioLegend
	RA3-6B2	FITC	eBioscience
	RA3-6B2	PE	eBioscience
	RA3-6B2	APC-Cy7	BioLegend
	RA3-6B2	PerCP-Cy5.5	BioLegend
CD19	RA3-6B2	PE-Cy5.5	BD
	6D5	Brilliant Violet 421	BioLegend
	1D3	FITC	eBioscience
	1D3	PE	BD
	1D3	APC	eBioscience
CD3ε	1D3	PE-Cy5.5	eBioscience
	145-2C11	FITC	eBioscience
	145-2C11	PE	BD
	145-2C11	Alexa Fluor 647	BioLegend
	145-2C11	PE-Cy5.5	BD
CD4	RM4-5	FITC	eBioscience
	RM4-5	PE	BD
	RM4-5	PE-Cy5.5	BD
CD8α	53-6.7	FITC	BioLegend
	53-6.7	PE	BD
	53-6.7	PE-Cy5.5	BD
TCRβ	B20.6	FITC	BD
	H57-597	PE-Cy5.5	BD
DX5	DX5	FITC	BD
	DX5	PE	BD
TCRγδ	GL3	FITC	BD
NK1.1	PK136	FITC	BD
	PK136	PE	eBioscience
	PK136	PerCP-Cy5.5	BD
	PK136	PE-Cy5.5	BioLegend
CD11b	M1/70	FITC	eBioscience
	M1/70	PE	BD
	M1/70	PE-Cy7	eBioscience
	M1/70	PE-Cy5.5	eBioscience
CD11c	N418	FITC	eBioscience
	HL3	PE	BD
	N418	PE-Cy5	eBioscience
	N418	Brilliant Violet 421	BioLegend
Gr1	RB6-8C5	FITC	BD
	RB6-8C5	PE	BioLegend
	RB6-8C5	PE-Cy5.5	eBioscience
Ter119	TER-119	FITC	eBioscience
	TER-119	PE	eBioscience
	TER-119	APC	BioLegend
	TER-119	PE-Cy5.5	BioLegend
IgM	RMM-1	FITC	BioLegend
	RMM-1	PE	BioLegend

	II/41	PE-Cy5	eBioscience
IgD	11-26C.2A	FITC	BD
	11-26C	APC	eBioscience
c-kit	2B8	PerCP-Cy5.5	BioLegend
	2B8	Brilliant Violet 421	BioLegend
Flt3	A2F10	PE	eBioscience
Sca1	D7	APC	eBioscience
	D7	PE-Cy7	BD
	D7	V450	BD
IL-7R α	SB/199	FITC	BioLegend
	SB/199	PE	BD
	A7R34	APC	BioLegend
	A7R34	Brilliant Violet 421	BioLegend
Fc γ R	2.4G2	PE-Cy7	BD
CD34	MEC 14.7	Brilliant Violet 421	BioLegend
	RAM34	Alexa Fluor 700	BD
CD150	TC15-12F 12.2	APC	BioLegend
CD48	HM48-1	Alexa Fluor 700	BioLegend
CD71	RI7217	PE	BioLegend
CD25	PC61	APC	BD
Ly6d	49-H4	Alexa Fluor 647	BD
CD115	AFS98	APC	BioLegend
CD43	S7	FITC	BD
	S7	PE	BD
CD317(Bst2)	927	Pacific Blue	BioLegend
MHC cl.II	M5/114.15.2	PerCP-Cy5.5	BioLegend
CD41	MWReg30	PE	BioLegend
	MWReg30	APC	eBioscience
CD31	MEC13.3	APC	BioLegend
CD21	eBio8D9	PE	eBioscience
	4E3	eFluor450	eBioscience
CD23	B3B4	PE	BD
CD45.1	A20	APC-eFluor 780	eBioscience
CD122	TM-B1	eFluor450	eBioscience

Table S3. Primers used in this study

Genotyping primers			
Genotyping PCR primers	Primer sequence (5'-3')		Size of PCR products (bp)
Bcl11a-cko-FW	TAGCTCCTGCTAGCCAGGTTTCTT		377(wild type)
Bcl11a-cko-RV	CGAGGCTTGCAGAAACAGAAAGAT		470(<i>flox</i>)
Bcl11a-cko-DEL	CTCGAAGGGAGGTTTCGGTATTGTG		700(<i>del</i>)
Bcl11a-eGFP-5'-FW	GAGACAAGAACAGGTGCAAGAGTGGATT		6746(targeted)
Bcl11a-eGFP-5'-RV	GGGTTATTGAATATGATCGGAATTGGGC		
Bcl11a-eGFP-3'-FW	GAAAGAACCAGCTGGGGCTCGACTAGAG		4507(targeted)
Bcl11a-eGFP-3'-RV	CAGCGAGGTCCCCTTTCTCACTAAAAAT		
Primers for ChIP assay qPCR			
PCR primers	Primer sequence (5'-3')		
Mdm2-Fwd	GCCAGCGTAGCCTAGGAG		
Mdm2-Rev	AAGAGGGAATGATCGGGACT		
Mdm4-Fwd	CAGGGTCCGCTCTATGGTT		
Mdm4-Rev	ACCCCAACACCACACCTTAC		
Cdkn1a-Fwd	GGGCCAGAGCTAAGAGCAC		
Cdkn1a-Rev	CACGCACGTACACAGACACA		
Bcl2-Fwd	GGGGACAAGCTGTTGTAGGA		
Bcl2-Rev	AGAGGGGACAGATAGCGATG		
Bcl2l1-Fwd	GCTGTGCAGGGAGGAGATAA		
Bcl2l1-Rev	CTCTGGGAGTAGGTGGTTGC		
Pax5-Fwd	GTGTGTGCATGAGCATGTGA		
Pax5-Rev	CCATTCCATGTGCAGAGATG		
Tcf3-Fwd	GGGACAGAGTGAGGACAGTCA		
Tcf3-Rev	CCCCATCTACTCTGTCACCA		
qRT-PCR primers			
Gene name	Forward primer (5'-3')	Reverse primer (5'-3')	Company
<i>Bcl11a</i>	AOD Probe	Mm01225235_m1	Applied Biosystems
<i>Sfp1</i>	AOD Probe	Mm00488142_m1	Applied Biosystems
<i>Ikzf1</i>	AOD Probe	Mm01187882_m1	Applied Biosystems
<i>Tcf3</i>	AOD Probe	Mm01175591_m1	Applied Biosystems
<i>Ebf1</i>	AOD Probe	Mm00395519_m1	Applied Biosystems
<i>Pax5</i>	AOD Probe	Mm00435501_m1	Applied Biosystems
<i>Cd79a</i>	AOD Probe	Mm00432423_m1	Applied Biosystems
<i>Foxo1</i>	AOD Probe	Mm00490672_m1	Applied Biosystems
<i>Il7r</i>	AOD Probe	Mm00434295_m1	Applied Biosystems
<i>Stat5a</i>	AOD Probe	Mm03053818_s1	Applied Biosystems
<i>Notch1</i>	AOD Probe	Mm00435249_m1	Applied Biosystems
<i>Bcl11b</i>	AOD Probe	Mm00480516_m1	Applied Biosystems
<i>Cebpa</i>	AOD Probe	Mm00514283_s1	Applied Biosystems
<i>Pbx1</i>	AOD Probe	Mm04207617_m1	Applied Biosystems
<i>Runx1</i>	AOD Probe	Mm01213405_m1	Applied Biosystems
<i>Mef2c</i>	AOD Probe	Mm01340842_m1	Applied Biosystems
<i>Ccr9</i>	AOD Probe	Mm01718955_g1	Applied Biosystems
<i>Flt3</i>	AOD Probe	Mm00439016_m1	Applied Biosystems
<i>Foxp1</i>	AOD Probe	Mm00474848_m1	Applied Biosystems
<i>Mpo</i>	AOD Probe	Mm01298424_m1	Applied Biosystems
<i>Csf3r</i>	AOD Probe	Mm00432735_m1	Applied Biosystems

Table S4. Lymphoid genes enriched for clustering from microarray data

<i>Sfpi1</i>	<i>Foxo3a</i>	<i>Tcf7</i>	<i>Scd1</i>
<i>Ikzf1</i>	<i>Pten</i>	<i>Mef2c</i>	<i>Car13</i>
<i>TCF3</i>	<i>Dntt</i>	<i>Gfi1</i>	<i>Cplx2</i>
<i>EBF1</i>	<i>Ets1</i>	<i>Evi1</i>	<i>E2f2</i>
<i>Pax5</i>	<i>Lef1</i>	<i>Bach2</i>	<i>Ikzf3</i>
<i>Foxo1</i>	<i>Syk</i>	<i>Hes1</i>	<i>Cbx2</i>
<i>Cd79a</i>	<i>Runx1</i>	<i>Xbp1</i>	<i>Hes5</i>
<i>Cd79b</i>	<i>Runx2</i>	<i>Igh-6</i>	<i>Pold4</i>
<i>Vpreb1</i>	<i>Runx3</i>	<i>Igll 1</i>	<i>Zfpm1</i>
<i>Vpreb2</i>	<i>Id2</i>	<i>Klf4</i>	<i>Dtx1</i>
<i>Vpreb3</i>	<i>Id3</i>	<i>C-kit</i>	<i>Bcar3</i>
<i>Cd19</i>	<i>Sox4</i>	<i>Cebpb</i>	<i>Nedd9</i>
<i>Erg</i>	<i>Stat3</i>	<i>Ccnd3</i>	<i>Sit1</i>
<i>Blk</i>	<i>IL-7Rα</i>	<i>Bcl7a</i>	<i>Btg2</i>
<i>Ccr7</i>	<i>Stat5a</i>	<i>Bcl7b</i>	<i>Tox</i>
<i>Ccr9</i>	<i>Stat5b</i>	<i>Cd93</i>	<i>Tmem66</i>
<i>Cxcr4</i>	<i>Sirt1</i>	<i>Flt3</i>	<i>CD22</i>
<i>Pou2f1</i>	<i>Sirt3</i>	<i>Foxo6</i>	<i>Evx1</i>
<i>Pou2af1</i>	<i>Gata3</i>	<i>Ccn3</i>	<i>Ncf4</i>
<i>Rag1</i>	<i>Notch1</i>	<i>Bank1</i>	<i>Tbp</i>
<i>Rag2</i>	<i>Notch2</i>	<i>Tcf4</i>	<i>Zap70</i>
<i>Irf4</i>	<i>Notch3</i>	<i>Bst1</i>	<i>Pde2</i>
<i>Irf8</i>	<i>Stat1</i>	<i>Klf7</i>	<i>Ccr2</i>
<i>Blnk</i>	<i>Foxp1</i>	<i>Clgn</i>	<i>Parp1</i>
<i>Ezh2</i>	<i>Bcl6</i>	<i>Car2</i>	<i>Hdac7</i>