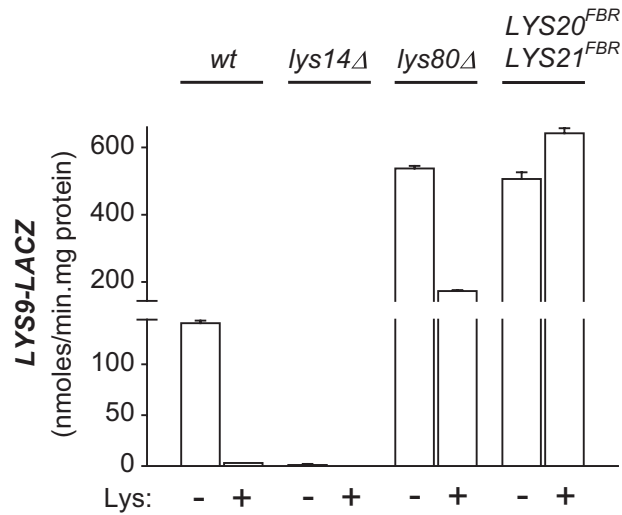
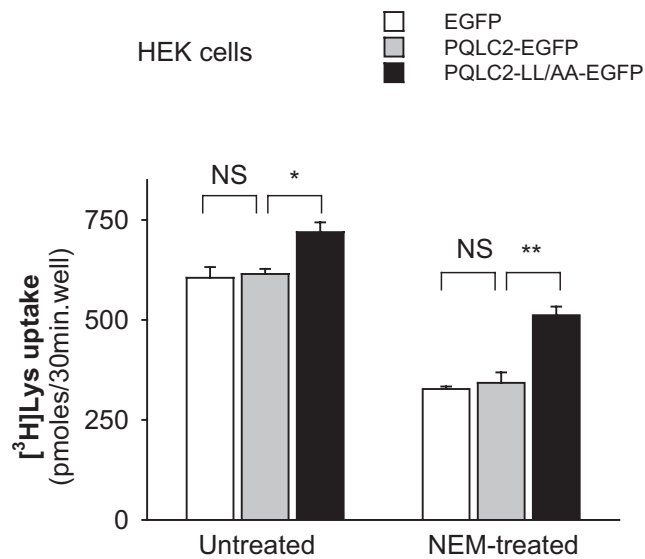


# Supporting Information

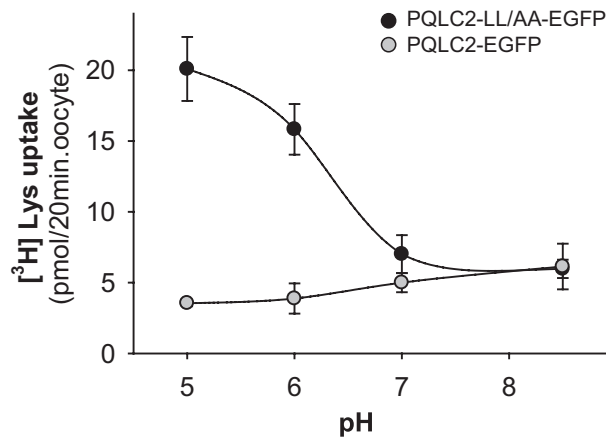
Jézégou et al. 10.1073/pnas.1211198109



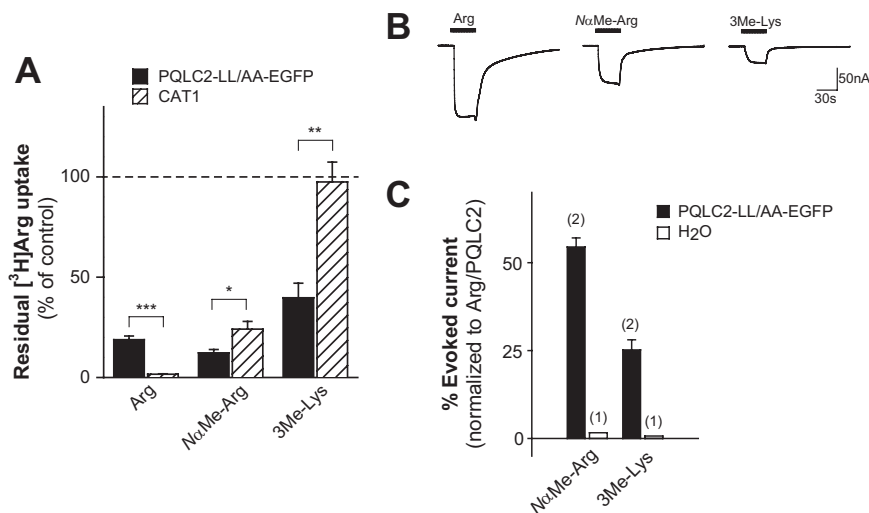
**Fig. S1.** Transcriptional regulation of the *LYS9* gene, a known member of the lysine-repressible *LYS* regulon. Strains of the indicated genotypes were transformed with a centromere-based plasmid expressing the *lacZ* reporter gene under the control of the *LYS9* gene promoter. Cells were grown on a minimal glucose/ammonium medium with (+) or without (-) lysine (1 mM).  $\beta$ -Galactosidase activities are the means of at least two independent experiments.



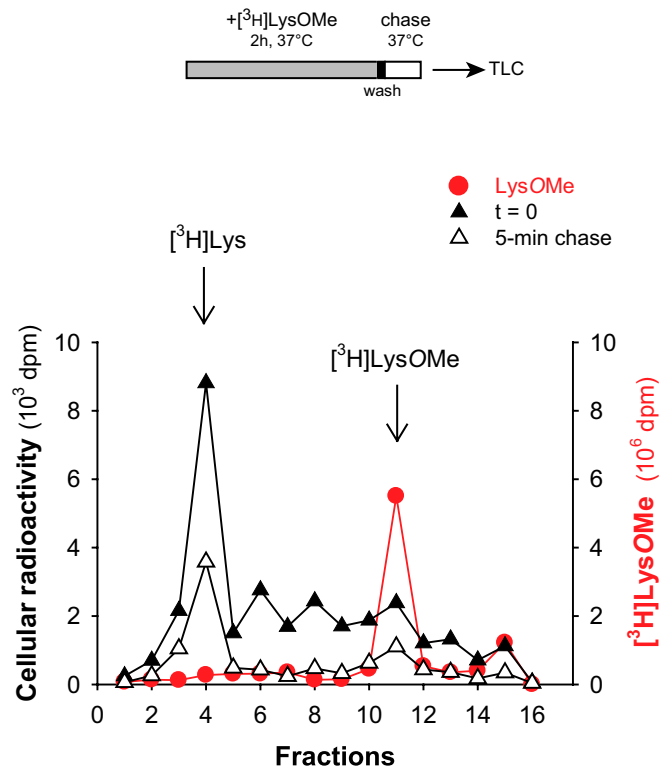
**Fig. S2.** Expression of PQLC2 in mammalian cells induces lysine uptake. HEK-293 cells (300,000 cells per well) were transfected with an empty pEGFP-N1 vector or with plasmids encoding WT PQLC2-EGFP or PQLC2-LL290/291AA mutation (LL/AA)-EGFP. Two days later, cells were washed and pretreated for 30 min at room temperature with 1 mM *N*-ethylmaleimide (NEM) to decrease the endogenous transport of cationic amino acid. Untreated and NEM-treated cells were then washed, incubated for 30 min with  $[^3\text{H}]\text{lysine}$  (0.1 mM) at pH 5.0, washed twice in ice-cold buffer, and subjected to scintillation counting. \* $P < 0.02$ ; \*\* $P < 0.01$ ; NS, not significant by the Student *t* test ( $n = 3$ ).



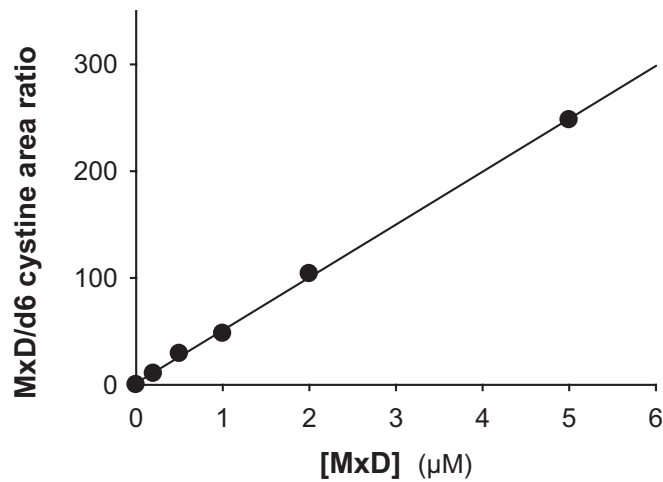
**Fig. 53.** Extracellular pH dependence of PQLC2-mediated lysine uptake. [ $^3\text{H}$ ]Lysine (0.1 mM) uptake into oocytes expressing WT PQLC2 or the LL290/291AA (LL/AA) mutant was measured at distinct pH values. Uptake is activated in conditions mimicking the lysosomal environment of PQLC2. Means  $\pm$  SEMs of four to five oocytes are shown.



**Fig. 54.** Selectivity of PQLC2 toward methylated cationic amino acids resembles that of lysosomal system c. (A) Arginine uptake into paired sets of oocytes expressing either PQLC2-LL290/291AA mutation (LL/AA)-EGFP or the plasma membrane transporter CAT-1 was analyzed at pH 5.0, in the absence or presence of 10 mM unlabeled L-arginine, *N*- $\alpha$ -methyl-L-arginine (NaMe-Arg), or  $\epsilon$ -*N*-trimethyl-L-lysine (3Me-Lys) added simultaneously to [ $^3\text{H}$ ]L-arginine (40 nM). NaMe-Arg and 3Me-Lys preferentially inhibit PQLC2 rather than CAT-1, in agreement with earlier biochemical studies on lysosome fractions. Means  $\pm$  SEMs of 8–14 oocytes from two batches are shown. \*\*\* $P < 10^{-9}$ ; \*\* $P < 0.01$ ; \* $P < 0.02$  by the Student *t* test. (B and C) Two-electrode voltage clamp recording of PQLC2-LL/AA oocytes at  $-40$  mV and pH 5.0 shows that the two methylated compounds (10 mM) are translocated by PQLC2. Raw traces from a PQLC2-LL/AA-EGFP oocyte and mean current responses normalized to the L-arginine current from two oocytes are shown in B and C, respectively.



**Fig. 55.** L- $^3\text{H}$ lysine methyl ester ( $^3\text{H}$ LysOMe) applied to human fibroblasts is predominantly converted to lysine. Fibroblasts were incubated for 2 h at 37 °C in 5%  $\text{CO}_2$  with 0.2 mM  $^3\text{H}$ LysOMe, washed, and further incubated at 37 °C (chase period). Water-soluble cell extracts (triangles) were then analyzed by TLC and compared with an equivalent amount of  $^3\text{H}$ LysOMe solution (red circles). Most of the water-soluble radioactivity was recovered as lysine. This intracellular  $^3\text{H}$ lysine pool decays with a time scale of minutes.



**Fig. 56.** Calibration curve of the mixed disulfide (MxD) liquid chromatography tandem MS assay. Increasing dilutions from a known MxD solution were tested. The assay is linear up to 5  $\mu\text{M}$  ( $R^2 = 0.9992$ ).

