Supporting Information

Park et al. 10.1073/pnas.1214400109



Fig. S1. Metabolic profiles of PyMT/PANIC mice compared with PyMT mice. (A) Schematic for the hyperglycemia induction in PANIC-ATTAC mice. Caspase8 cassettes are activated upon dimerize treatment (AP20187; Clontech). (*B*) Serum glucose levels (feeding condition) were determined for PyMT/PANIC compared with PyMT. Hyperglycemia in PyMT/PANIC was induced by AP20187 injection (0.5 μ g/g, i.p., 5 d) and maintained over the tumor progression (*n* = 9–16 per group). ****P* < 0.001 vs. PyMT by two-way ANOVA. (C) Serum insulin levels were significantly decreased in PyMT/PANIC compared with PyMT because of concomitant increase of serum glucose levels (*n* = 9–16 per group). ***P* = 0.0023 vs. PyMT by unpaired Student's t test. (*D*) Body weight was determined over the tumor progression (*n* = 9–16 per group). ***P* < 0.01 nd ****P* < 0.001 vs. PyMT by two-way ANOVA. (*C*) Serum insulin levels were significantly decreased in PyMT/PANIC compared with PyMT because of concomitant increase of serum glucose levels (*n* = 9–16 per group). ***P* = 0.0023 vs. PyMT by unpaired Student's t test. (*D*) Body weight was determined over the tumor progression (*n* = 9–16 per group). ***P* < 0.01 and ****P* < 0.01 vs. PyMT by two-way ANOVA. (*E*) Serum leptin reflecting body fat mass was determined (*n* = 9–16 per group). ***P* = 0.0013 vs. PyMT by unpaired Student's t test. (*F*) Early-stage of tumor progression was determined by whole mount staining of mammary gland tissues at 8-wk-old PyMT/PANIC compared with PyMT. (Scale bars: 50 mm.) Higher magnification for squared boxes is presented in *Lower*.

A Cancer-related signaling pathways (PyMT/PANIC vs. PyMT)



Ingenuity pathways

В

Pathway analysis	Gene (symbol)	Folds	p-value	Illumina ID
Increased	ATP2A3	1.770	1.87E-03	ILMN 2900462
EGF/ Ras pathway	BCL6	1.566	2.19E-03	ILMN_1230353
NF-kB pathway	GPR77	1.515	8.08E-04	ILMN_2671707
Bcl2L1 mediated-	ΝΓκΒ1Α	1.469	4.54E-04	ILMN_3001914
anti-apoptosis	GADD45G	1.424	3.82E-02	ILMN_2903945
Protein modification	DUSP1	1.421	7.21E-03	ILMN_2622983
 glycosylation 	EGF	1.407	2.70E-02	ILMN_2684104
- disulfide	RAB26	1.355	3.70E-02	ILMN_2722058
- Ubl conjugation	RABGEF1	1.292	9.16E-03	ILMN_1226143
	CCRL2	1.250	2.45E-02	ILMN_2752624
	BCL2L1	1.224	2.24E-02	ILMN_2647885
Decreased	DGAT2	-5.246	2.72E-03	ILMN_1219915
linid metabolism	LEP	-5.149	5.08E-03	ILMN_2695964
alucose metabolism	SCD	-2.968	1.29E-02	ILMN_1237375
FR function	COX8B	-2.942	1.43E-02	ILMN_2727520
Mitochondria function	CPT1B	-2.265	4.47E-02	ILMN_1227666
	HSD11B1	-2.211	2.54E-02	ILMN_2774160
	PFK4	-2.177	3.89E-02	ILMN_1259322
	ΡΡΑΒγ	-2.002	1.19E-02	ILMN_1221060
	AGPAT2	-1.965	4.95E-02	ILMN_1217252
	CAV1	-1.798	3.51E-02	ILMN_2632665
	LPL	-1.672	4.27E-02	ILMN_2692723
	HK2	-1.625	3.10E-03	ILMN_1239397
	CPT2	-1.429	1.48E-02	ILMN_2885532
	DNMT3B	-1.389	4.36E-02	ILMN_1252310
	BCL3	-1.374	9.47E-03	ILMN 2749717

Fig. S2. cDNA microarray for tumor tissues of PyMT/PANIC mice compared with PyMT. (*A*) Cancer-related pathway analysis (Ingenuity), revealing that EGFR, Ras, ERK1/2, NF-κB, and BCL2-L1 pathways are up-regulated in tumor tissues of PyMT mice upon hyperglycemia challenge. (*B*) Molecules in pathway analysis. Highly up- or down-regulated genes by hyperglycemia are listed in the table.

Α		ID	Symbol	Entrez Gene Name
18. <u>.</u>	1	II MN 3162594	ARHGAP15	Rho GTPase activating protein 15
	2	ILMN 2814927	CHGA	chromogranin A (parathyroid secretory protein 1)
	3	ILMN 2503552	CSMD1	CUB and Sushi multiple domains 1
	4	ILMN 1251139	DENND4C	DENN/MADD domain containing 4C
	5	ILMN 2653462	EFEMP1	EGE-containing fibulin-like extracellular matrix protein 1
	6	ILMN 2716212	EFNB2	ephrin-B2
	7	ILMN 2669766	FID1	EP300 interacting inhibitor of differentiation 1
	8	IL MN 3131522	FBX03	E-box protein 3
	9	ILMN 1239397	HK2	hexokinase 2
	10	IL MN 2572649	KDR	kinase insert domain recentor (a type III recentor tyrosine kinase)
	11	IL MN 1243671	MAST4	microtubule associated serine/threonine kinase family member 4
	12	ILMN_3162138	MIP	migration and invasion inhibitory protein
	13	ILMN 2605307	MTMR10	myotubularin related protein 10
	14	IL MN 1227012	NDUEB4	NADH debydrogenase (ubiguinone) 1 beta subcomplex 4 15kDa
	15	ILMN 2971688	NRG1	neuregulin 1
	16	ILMN 2860174	PEKEB3	6-phosphofructo-2-kinase/fructose-2.6-biphosphatase 3
	17	IL MN 2626470	PI B1	nhospholinase B1
	18	ILMN 1243525	PLDN	pallidin homolog (mouse)
	19	ILMN 1221060	PPARG	peroxisome proliferator-activated receptor gamma
	20	ILMN 1253819	PRKAR2B	protein kinase, cAMP-dependent, regulatory, type II, beta
	21	ILMN 2636859	RBBP8	retinoblastoma binding protein 8
	22	ILMN 2838501	SLC24A3	solute carrier family 24 (sodium/potassium/calcium exchanger), member 3
	23	ILMN 1222499	SLC33A1	solute carrier family 33 (acetyl-CoA transporter), member 1
	24	ILMN 1216265	TSHZ1	teashirt zinc finger homeobox 1
	25	ILMN 2497067	ZMYND11	zinc finger, MYND domain containing 11
		· · · · · · · · · · · · · · · · · · ·		
D				
В	(15 7		Ш РуМ Т
в	(B4)	15		PyM T PyM T/PANIC
B 	36B4)	15 ₋		PyMT PyMT/PANIC
B seis	vith 36B4)	15 10- 10-		PyMT PyMT/PANIC
B B	1 with 36B4)	15-		PyMT PyMT/PANIC
A Levels B	zed with 36B4)	15- 10-	. I	PyMT PyMT/PANIC
RNA Levels B	alized with 36B4)	15- 10- 5-	II .	PyM T PyM T/PANIC
mRNA Levels	rmalized with 36B4)	15- 10- 5-	l. i	
mrna Levels	Normalized with 36B4)	15 10- 5-		
mrna Levels B	(Normalized with 36B4)	15 10- 5- 0- <u>m nì nì nì nì</u>		
B mrna Levels	(Normalized with 36B4)	15 10- 5- 0-6 01 01 01		
B mRNA Levels	(Normalized with 36B4)	15 10- 5- 0		
B mRNA Levels	(Normalized with 36B4)	15 10- 5- 0- 0- 0- 0- 0- 0- 0- 0- 0- 0- 0- 0- 0-	A A M A A A A	PyMT PyMT/PANIC
B mRNA Levels C	(Normalized with 36B4)	15 10- 5- 10- 10- 10- 10- 10- 10- 10- 10- 10- 10		PyMT PyMT/PANIC
C mRNA Levels	4) (Normalized with 36B4)	15 10- 5- 0- 10- 10- 10- 10- 10- 10- 10- 10- 10-	N ST	PyMT PyMT/PANIC ** ** ** ** ** ** ** ** ** *
C mRNA Levels B	6B4) (Normalized with 36B4)	15 10- 5- 0- 10- 10- 10- 10- 10- 10- 10- 10- 10-	A A A A A A A A A A A A A A A A A A A	PyMT PyMT/PANIC ** ** ** ** ** ** ** ** ** *
els O mRNA Levels B	n 36B4) (Normalized with 36B4)	15 10- 5- 0- 10- 10- 10- 10- 10- 10- 10-	al a management	PyMT PyMT/PANIC ** ** ** ** ** ** ** ** ** *
vels O mRNA Levels B	vith 36B4) (Normalized with 36B4)	15 10- 5- 0- 10- 10- 10- 10- 10- 10- 10-		PyMT PyMT/PANIC ** ** ** ** ** ** ** ** ** ** ** ** **
Levels O mRNA Levels B	d with 36B4) (Normalized with 36B4)	15 10- 5- 0- 10- 10- 10- 10- 10- 10- 10-		PyMT PyMT/PANIC
NA Levels O mRNA Levels B	zed with 36B4) (Normalized with 36B4)	15 10- 5- 0- 10- 10- 10- 10- 10- 10- 10-		PyMT PyMT/PANIC
nRNALeveis O mRNALeveis B	nalized with 36B4) (Normalized with 36B4)	15 10- 5- 0- n n n n n n n n n n n n n n n n n n n	a a a a a a a a a a a a a a a a a a a	PyMT PyMT/PANIC
mRNALevels O mRNALevels B	ormalized with 36B4) (Normalized with 36B4)	15 10 5 0 10 10 10 10 10 10 10 10 10	A A A A A A A A A A A A A A A A A A A	PyMT PyMT/PANIC
mRNA Levels O mRNA Levels B	(Normalized with 36B4) (Normalized with 36B4)	15 10 5 0 10 10 10 10 10 10 10 10 10		
mRNA Levels O mRNA Levels B	(Normalized with 36B4) (Normalized with 36B4)	15 10 5 0 10 10 10 10 10 10 10 10 10		PyMT $PyMT/PANIC$ T $PyMT/PANIC$ T
mRNA Levels O mRNA Levels B	(Normalized with 36B4) (Normalized with 36B4)	15 10- 5- 10- 10- 10- 10- 10- 10- 10- 10		PyMT PyMT/PANIC PyMT/PANIC T PyMT/PANIC PyMT/PANIC T PyMT/
mRNA Levels O mRNA Levels B	(Normalized with 36B4) (Normalized with 36B4)	15 10 		= PyMT $= PyMT/PANIC$ T $= PyMT/PANIC$ T

Fig. S3. Comparison microarray data with FAIRE-seq data. (*A*) List of candidate molecules. (*B*) Quantitative (qRT-PCR), revealing increased Nrg1 mRNA levels in tumor tissues of PyMT mice upon hyperglycemia challenge (n = 8 per group). ***P < 0.001, **P < 0.01 vs. PyMT by two-way ANOVA. (*C*) qRT-PCR, showing that consistent increase of Nrg1 mRNA levels in HyG tumors than Ctrl tumors (n = 6 per group). Results were normalized to 36B4 and represented as mean \pm SEM ***P < 0.001 vs. Ctrl \rightarrow WT by two-way ANOVA.

Nrg1/NRG1 enhancer sequences

Mm_Nrg1_Enhancer MACTTATCTAPATTTCCCCCTCTCTTACTCAAAACATTTACCATCCT	CATCTTGCTTGTTTCTGCTGGAGAA
Hs_NRG1_Enhancer AACTTATTGCAATTTCCTCTTATCATAACATTTCCCATATT	CAACATCTTTCTTGTTTGTGGTAGAGAA
Mm_Nrg1_Enhancer <mark>CAGAAAACAG</mark> GTC <mark>TATGCAACTATCAAAACATTAATANT</mark> CAAATTTCA	CAATTA <mark>CT</mark> TTGTTGGCCTTGATATACTT
Hs_NRG1_Enhancer <mark>CAGAAAACAG</mark> ATA <u>TGCCACAGTATCAAAAACATTAATAAC</u> GAATTTCAA	CAATTA <mark>TA</mark> TTGTTGGCCTTGATATACTT
Mm_Nrg1_Enhancer <mark>TCTGAATTCCAACC</mark> CGATAGCTTGCTC <mark>TTCAACTTCCTTCCCATTT</mark> CT	CATGCCTTCGTTTTTTTATGAAGTGGGG
Hs_NRG1_Enhancer <mark>TCTGAATTCCAACG</mark> AGATAGCTATTC <mark>TACTTCCTTCCCATTT</mark> GA	CATATCTT-GCTTTTCTGAAACAGGG
Mm_Nrg1_Enhancer CCACGMTACHAACCCACAGCTAACGAAGAAAGAACTGGCAAAGCAACCCCCTA	GATCT <mark>AACATGCCTAA</mark> GGGCT <mark>A</mark> TGGAAT
Hs_NRG1_Enhancer CCTTTCAGTTACCGGTTCTAAGGAGCAATAACTGGGAAAGTACCAA	GATCT <mark>T</mark> ACATGCCTAA <mark>A</mark> GGCT <mark>C</mark> TGGAAT
Mm_Nrg1_Enhancer_GC <mark>G</mark> ACA <mark>GCTTTTGTTTTCTTAGC</mark> TAGA <mark>GAGATGCAAATATTTAAC</mark> C	ATGAATCATCTGAGC <mark>CA</mark> GGACTTCCT <mark>C</mark> A
Hs_NRG1_Enhancer_GT <mark>G</mark> GTG <mark>GCTTTTATTTTCTTAGC</mark> C <mark>AGAGGAGATGCAAATATTTAAC</mark> T	ATGAATCATCTGAGC <mark>TG</mark> GGACTTCCT <mark>T</mark> A
Mm_Nrg1_Enhancer	ATAATCAGGG <mark>G</mark> CCT <mark>TG</mark> GAGGATATTTAC ATAATCAGGG <mark>A</mark> CCT <mark>CA</mark> GAGGATATTTAC
Mm_Nrg1_Enhancer TCAACTTCTGATCAGGCTCTGGTCCCCCTTTTCCGGTGCAACTGGCC	TCTGGAAAGT <mark>C</mark> TCAG <mark>CTCTCG</mark> GATTA
Hs_NRG1_Enhancer TCAACTTCTGATCAGAGACTTTGCTCCCCCTTTCCCGGGGAAGTGGCC	TCTGGAAAGT <mark>G</mark> TCAG <mark>AG</mark> CTCTC <mark>A</mark> GATTA
Mm_Nrg1_Enhancer_TTTTCTCTAGGATTCCACTGAGCCTTTCCATATTCGCGATAAGTGGAA	AG <mark>CTTTCA<mark>CA</mark>ATTCCAATCCA<mark>AC</mark>ACATT</mark>
Hs_NRG1_Enhancer_TTTTTCTAA <mark>GATTACACTGAGCTTTT</mark> TAATATT <mark>A</mark> GAGATAAGTAGAC	AG <mark>TTTTCA<mark>TG</mark>ATTCCAATCCA<mark>GG</mark>ACATT</mark>
	AP1 binding motif

Fig. S4. Homology between mouse and human NRG1 enhancer sequences. Square indicates conserved AP-1 binding motif.

PNAS PNAS

A Mouse Nrg1 mRNA sequences amplified in HyG-tumors compared to Ctrl-tumors

>uc009ljn.1_mm9_1_1 240 0 0 chr8:32928500 - 32928739 -

B Homologous sequences in human NRG1 mRNA

>uc009ljn.1_hg18_1_1 240 0 0 chr8:32741223-32741462 +

C Human NRG1 isoforms

Accession	Description	E-value	Max identity
NP_001153473.1	pro-neuregulin-1, isoform HRG-beta1d	1e-30	100%
NP_001153476.1	pro-neuregulin-1, isoform HRG-beta1c	2e-30	100%
NP_001153471.1	pro-neuregulin-1, isoform HRG-beta1b	3e-30	100%
NP_039250.2	pro-neuregulin-1, isoform HRG-beta1	4e-30	100%
NP_039251.2	pro-neuregulin-1, isoform HRG-beta2	5e-30	100%
NP_039258.1	pro-neuregulin-1, isoform HRG-alpha	9e-30	100%

Fig. S5. Mouse Nrg1 mRNA sequences amplified under hyperglycemic conditions and analogous human isoforms. RNA-seq analysis was performed with RNA samples subjected from the mouse HyG versus Ctrl tumors by the sequencing facility in the McDermott Center for Human Growth and Development, University of Texas Southwestern Medical Center. (Illumina Hi-Seq2000, SOLiD 5000xl). The murine Nrg1 isoforms have not yet been well defined, so a series of comparative alignments with human homologous are shown. (A) Mouse mRNA sequence up-regulated in HyG-tumors compared with Ctrl-tumors. (B) Human homologous sequences corresponding to mouse Nrg1 mRNA. (C) Blastx analysis (http://blast.ncbi.nlm.nih.gov) with human NRG1 homologous sequences indicates that several isoforms of heregulin are up-regulated in response to hyperglycemia.



Fig. S6. Nrg1 acquisition in tumor tissues of PyMT/PANIC compared with PyMT. (A) Nrg1 immunostaining, showing higher Nrg1 signal in tumor tissues from PyMT/PANIC compared with those from PyMT/PANIC compared with PyMT. (A) Nrg1 immunostaining, showing higher Nrg1 signal in tumor tissues from PyMT/PANIC compared with those from PyMT/PANIC compared with PyMT. (B) Activation of HER receptors in tumors from PyMT/PANIC in comparison with those in PyMT. c-Jun levels were increased in PyMT/PANIC, whereas the effects on AKT and ERK activation was limited. β -actin was used as a loading control. (C) Quantified results for Western blots were normalized to β -actin and represented as mean \pm SEM. Two independent cohort experiments were performed. **P* < 0.05, ***P* < 0.01 vs. PyMT by two-way ANOVA. (*D*) Western blotting, showing an inefficient inhibition of AKT and ERK activation by a RTK inhibitor, lapatinib treatment (100 mg/kg per day by oral gavage) for both Ctrl and HyG tumor-bearing mice. c-Jun is consistently increased in HyG tumors compared with Ctrl tumors. β -actin was used as loading control. (*E*) Quantified results represent as mean \pm SEM, *n* = 5–6 per group. ***P* < 0.01, ****P* < 0.001 Ctrl-veh vs. HyG-veh; **P* < 0.05, HyG-veh vs. HyG-lap by unpaired Student's t test.



Fig. S7. NRG1 immunostaining for DM-diagnosed breast cancer patients compared with control patients. Paraffin-embedded tumor samples were immunostained with anti-NRG1 (H210; Santa Cruz Biotechnology). Hematoxylin was used for costaining. (Scale bars: 50 µm.)

Group	ID	Tumor grade	Glucose, mg/dL	NRG1	HER3	ER	PR	HER2	P53	Ki-67, %
Non-DM (<i>n</i> = 12)	C-1	1	87	0	0	+	_	_	_	12
	C-2	1	88	0	0	+	+	—	—	5
	C-3	1	89	0	0	+	+	—	—	7
	C-4	1	90	3+	0	+	+	—	—	9
	C-5	1	85	0	0	+	+	—	—	2
	C-6	1	88	0	2+	+	+	—	+	16
	C-7	1	84	0	0	+	+	—	—	47
	C-8	2	84	0	0	—	—	_	+	16
	C-9	2	84	0	0	+	+	—	—	18
	C-10	2	88	0	0	+	+	_	—	30
	C-11	3	85	0	0	_	_	_	+	54
	C-12	3	89	0	0	—	—	_	+	38
DM(<i>n</i> = 13)	D-1	1	145	2+	2+	+	_	_	—	10
	D-2	1	130	3+	0	+	+	_	+	9
	D-3	1	136	1+	0	+	_	_	—	14
	D-4	1	449	3+	1+	+	+	_	—	6
	D-5	1	163	0	0	+	+	_	—	20
	D-6	2	130	0	0	+	+	—	—	11
	D-7	2	201	1+	1+	+	+	_	—	5
	D-8	2	349	1+	0	+	+	_	_	14
	D-9	2	154	3+	2+	+	+	_	—	16
	D-10	2	184	3+	0	+	+	_	+	31
	D-11	2	194	2+	0	+	+	_	_	15
	D-12	2	370	2+	0	+	+	_	+	19
	D-13	2	135	3+	2+	+	+	—	—	28

Table S1	Fasting alucose	levels and cance	r marker status	for human nationts
Table 31.	rasting glucose	levels and cance	i marker status	ior numan patients

HER2 negative, control (n = 12), and DM-diagnosed (n = 13) human breast cancer patient samples were included in this analysis. NRG1 and HER3 status were determined by immunohistochemistry. Scoring for NRG1 and HER3 staining intensity represents as 0 (negative) to 3 (strong). Other marker proteins such as ER (estrogen receptor), PR (progesterone receptor), HER2, p53, and Ki-67 were also assessed by immunohistochemistry. Specimens highlighted in red are represented in Fig. 5A.

PNAS PNAS

Table 32. Fillier	sequences used in this study	
Gene ID	Sense (5' to 3')	Antisense (5' to 3')
Arhgap15	AATCCAAGTTGTGGGTCCCTGGAG	TCAGTGCTCCGGTGACGACGT
Chga	CCGCCACCATCACCGCTGTT	GGGCAAAAACTTGCCCGGCG
Csmd1	CCCACTCGCTGGCTCTCTCTCCAG	GGATGGCTCCTCTCGGATGCG
Dennd4c	TGGGGAGCACACAGTCTTCGTCAG	TGCTCTTCAGGGGCGTGACCA
Efemp1	GGGGGCTTCCGCTGTTACCC	GGCAGCTCCCGGCACATAGTG
Efnb2	ACAGCGTCTTCTGCCCGCAC	CCTACCGTGTCCTCCCCGGG
Eid1	AGGTGTTCCTGCGAACCGCG	GGCTCTTCTCACACCGCGCC
FBXO3	TAGAGGCCGAGACGGGGCTG	TCCGAGGACACCTGGGCTCC
HK2	TGCCAAGCGTCTCCATAAGG	GGAGGAAGCGGACATCACAA
KDR	CAGACTGTGTCCCGCAGCCG	CACAGAGGCGGTATGCGCCC
Mast4	GGTTTAGCGGGAGAAGCCGCAG	TTGGACCACAGGAGCGCGTC
Miip	TTTCGGGCGGAGAGCGTTGG	CTAGCGGGACGAAGGCACGC
Mtmr10	CCGCCTCAGCACCGACAG	TGCGGTGCTGCCTCAGCATC
Ndufb4	ACCAGAAGGGCGCAGGTCGA	CGACACGCGTTTGGGGTCGT
NRG1	TCCGGCAGAGCCTTCGGTCA	TCTCCCGTAGCCTCGGTGGC
Nrg1 (Type1)	GGGAATGAGCTGAACCGTAG	ACAATGGTGATGTTGGCAGA
Nrg1 (Type3)	TGCATTGCTGGCCTAAAGTG	GTTCTTCCGGGTGGGTACTG
Pfkfb3	ACCATTCAGACCGCGGAGGC	TCCGGGAGCTCTTCATGTTCTCTGA
Plb1	ATGGCATGGTGGGAACAGAAGCCAC	ACCGGGGACACACCTGGGTAC
Pldn	GCGCCCAGTGCGGGGATATG	TGTCGCTTAAGCCCGGCGTG
PPARg	CACAATGCCATCAGGTTTGG	GCTGGTCGATATCACTGGAGATC
Prkar2b	GCTGTGCTCGCTCTGCTGCTG	GTGCCTCAGCACCTCCACCG
Rbbp8	ACGGCAGCCTTACAACGCGG	ACCACGAGGGCTCTTGTGCAG
Slc24a3	TCCACCGCAAAGCGTCCGTG	TGTCGTCCCTCCGGGTTCCG
Slc33a1	AGCGACCAGTCTCTCCGCGT	GGGCCTGAGGCTGGCGTTTT
Tshz1	TTGTCAGGGTGACGTTGGCGAGA	CGGACATTCACTGGGTGGGCG
Zmynd11	AGCAAAGTGCCGAGCGGGTC	TGGAGGCGAGCACCGAGACA
EGFR	TGCCACCTATGCCACGCCAAC	ATCCCAAGGGCCACCACCACT
HER2	GCAACTGTGGTGGGCGTCCT	CAGCTCCACTGGGCGTCAGC
HER3	CACGGGGGCGTCAAACGTCA	GAGAGCGGGGTGACGGGAGT
CyclinD1	AGCCTCCAGAGGGCTGTCGG	GGCTGTGGTCTCGGTTGGGC
36B4	GGCATGCGGCCCGTCTCTC	CTTCCCTGGGCATCACGGCG
β-actin	CCACACCCGCCACCAGTTCG	TACAGCCCGGGGAGCATCGT

Table S2. Primer sequences used in this study

PNAS PNAS