

SUPPLEMENTAL MATERIAL:

Figure S1. Normal CTL effector function in *Def6*^{-/-} OT-I CD8⁺ T cells.

A, Naïve WT B6 (CD45.2⁺) mice received 5000 OT-I WT (CD45.1⁺) or *Def6*^{-/-} OT-I (CD45.1⁺) CD8⁺ T cells. One day later, mice were challenged with 5 x 10⁶ Act-mOVA/K^{b/-} splenocytes. **A**, Data are presented as the frequency of expanded OT-I cells (CD45.1⁺ H-2K^b-OVA tetramer⁺) generated in WT recipient mice (first and third panels from left) and % of specific killing of specific target cells pulsed with OVA₂₅₇₋₂₆₄ and control, irrelevant target cells (Ski9) (second and fourth panels from left). **B**, *In vitro* killing of target cells by WT and KO OT-I CD8⁺ T cells at the indicated effector:target (E:T) ratios was evaluated by measuring ³H retention in target cells, as described in Materials and Methods. Data are presented as percent ± SD of specific cytotoxicity of quadruplicate samples and are representative of three independent experiments. **C**, Cell surface expression of CD107a was analyzed in CD8⁺ OT-I recovered from immunized mice and restimulated *in vitro* with OVA peptide in the presence of Brefeldin A for 5 h.

Figure S2. Impaired secondary expansion of *Def6*^{-/-} CD8⁺ T cells in response to LM-OVA infection is rescued by constitutive *Cdc42* or *NFAT1* activation.

Naive WT B6 (CD45.1/2⁺) mice (n=5) received 50 WT OT-I (CD45.1⁺) and 50 *Def6*^{-/-} OT-I (CD45.2⁺) CD8⁺ T cells. One day later, the mice were challenged with 5x10⁶ Act-mOVA/K^{b/-} splenocytes. On day 7, the mice were rechallenged with 1 x 10⁷ cfu of Act-A⁻ Lm-OVA followed by analysis on day 12. **A**, Representative FACS plots of WT and KO OT-I CD8⁺ T cell frequency in peripheral blood. **B**, Average ratio of WT to KO OT-I CD8⁺ T cells per mouse. The WT/KO ratio in non-immunized animals is set at 1. **C**, Average absolute numbers ± SEM of WT and KO OT-I CD8⁺ T cells in spleens from recipient mice on day 12. **D**, Average expression ±

SEM of T-Bet and Eomes by WT and KO OT-I CD8⁺ T cells 7 days after immunization. Data are representative of at least three independent experiments. **E**, Average frequency \pm SEM of OT-I cells that express KLRG-1 and CD127 in the peripheral blood. **F**, Average frequency \pm SEM of WT and KO OT-I CD8⁺ T cells producing IFN- γ alone (IFN- γ ⁺TNF- α ⁻) or both IFN- γ and TNF- α (IFN- γ ⁺TNF- α ⁺) among total splenocytes determined by ICS 12 days post-immunization. **G**, The NFAT1^{CA} Δ dimer mutant protein can constitutively transactivate an NFAT-responsive reporter luciferase construct. Jurkat-TAg cells were cotransfected with an empty control vector (empty vector; 10 μ g) or with an NFAT^{CA} Δ dimer-expressing NFAT^{CA} Δ dimer, together with NFAT-luciferase reporter (10 μ g) plus a β -Gal (5 μ g) reporter gene. Cells were left unstimulated for 16 hr and normalized luciferase activity (mean \pm SD) was measured in triplicates. NFAT expression was detected using an anti-HA Ab, and actin immunoblotting served as a loading control. **H-I**, Constitutive Cdc42 or NFAT1 activation rescue secondary expansion of *Def6*^{-/-} CD8⁺ T cells. Retrogenic mice generation, adoptive transfer, priming with Act-mOVA/K^{b/-} splenocytes and challenge with LM-OVA were performed as in **Fig. 5**. Analysis was performed 5 days after LM-OVA challenge. **H**, Average frequency (mean \pm SEM, n=4-5) of WT and KO OT-I CD8⁺ T cells among peripheral blood CD8⁺ cells. **I**, Average ratio of WT to KO OT-I CD8⁺ T cells. The ratio in non-immunized animals is set at 1.

Figure S3. *Def6* deficiency results in defective CD8⁺ T cell response to *Listeria monocytogenes* infection.

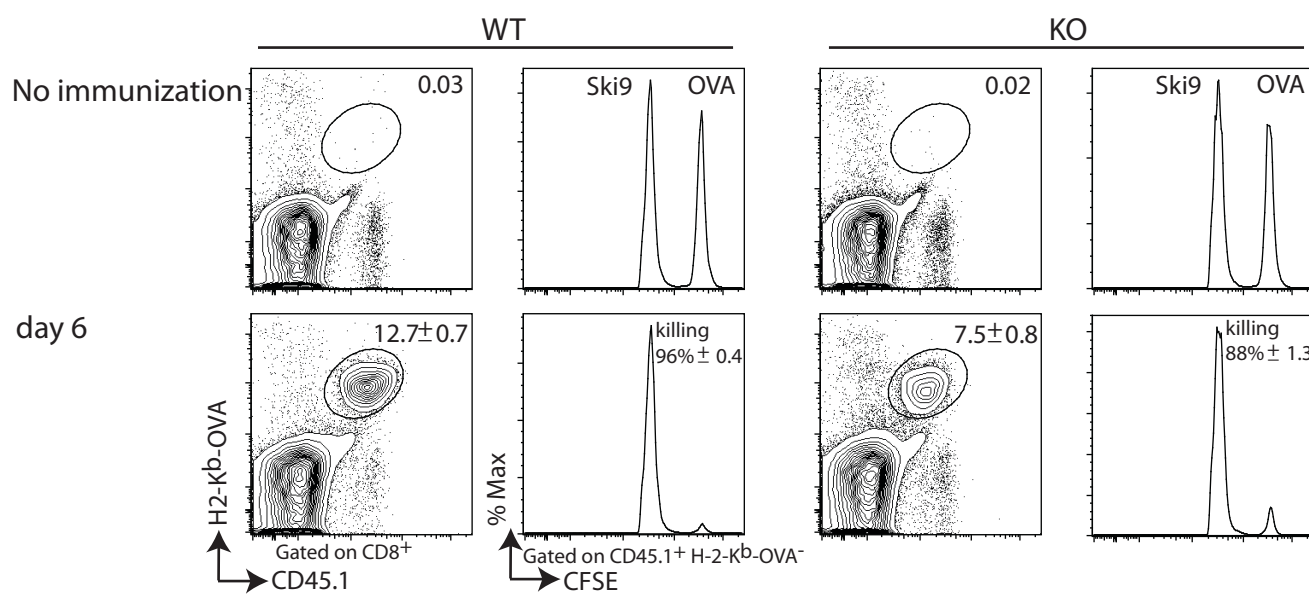
Fifty naïve WT (CD45.1⁺) and *Def6*^{-/-} (KO; CD45.2⁺) OT-I CD8⁺ T cells were adoptively co-transferred into CD45.1/2⁺ B6 recipient that were immunized one day later with 3000 cfu OVA-expressing *Listeria monocytogenes* (LM-OVA) followed by analysis at day 7 (primary), at day 49 (memory), and at day 54 (secondary) in mice that had received 1x10⁷ cfu OVA-expressing *Listeria monocytogenes* Act A deficient rechallenge 5 days earlier. Frequency of WT and KO

OT-I CD8⁺ T cells in the blood from recipient mice (n=6) is shown. *P<0.05, ***P<0.0005 (two-tailed unpaired *t*-test).

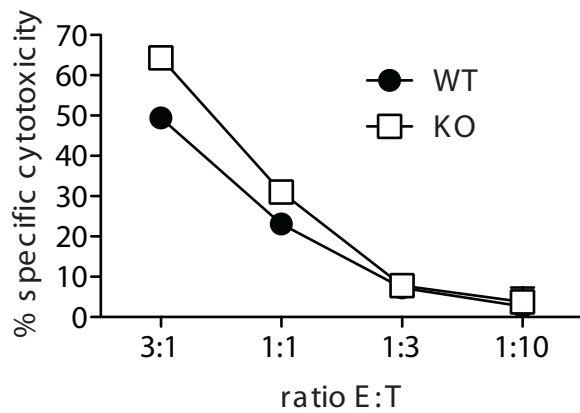
Figure S4. Defective IL-2 production of Def6^{-/-} CD8⁺ T cells upon CD3/CD28 stimulation *in vitro*.

A, Purified peripheral CD8⁺ T cells from WT and Def6^{-/-} (KO) mice were stimulated with the indicated concentrations of plate-coated anti-CD3 plus soluble CD28 mAbs for 40 hours. **A**, [³H]thymidine was added for the final 16 hours, and uptake was measured. **B**, CD8⁺ T cells were stimulated for 24 hours as in **A**, and IL-2 production was measured by an ELISA. **C**, CD8⁺ T cells from WT and KO mice were stimulated with the indicated concentrations of anti-CD3 plus 2.5µg/ml anti-CD28 mAbs for 16 hours, and analyzed by flow cytometry for CD25 and CD69 expression. Results are expressed as mean ± SD. Statistical differences were determined as in **Figure 1**. ***P<0.0005 (two-tailed unpaired *t*-test). Data are representative of three independent experiments.

A



B



C

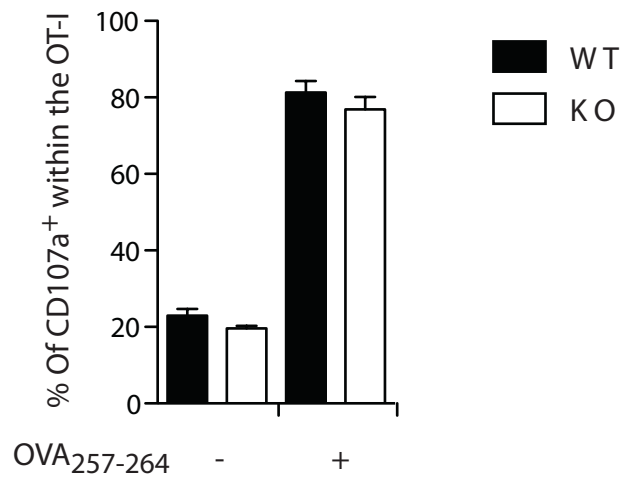


Figure S2, Feau et al.

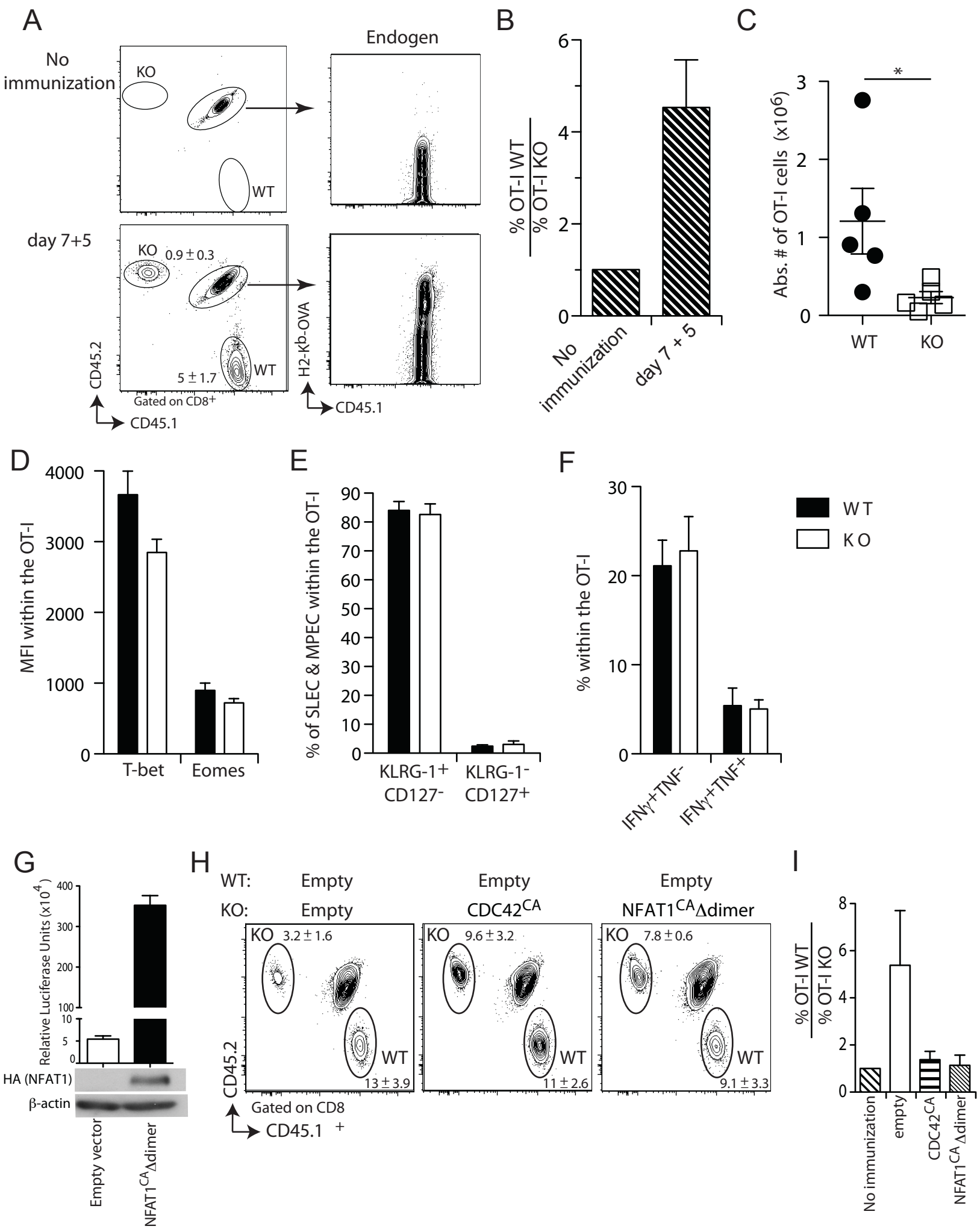
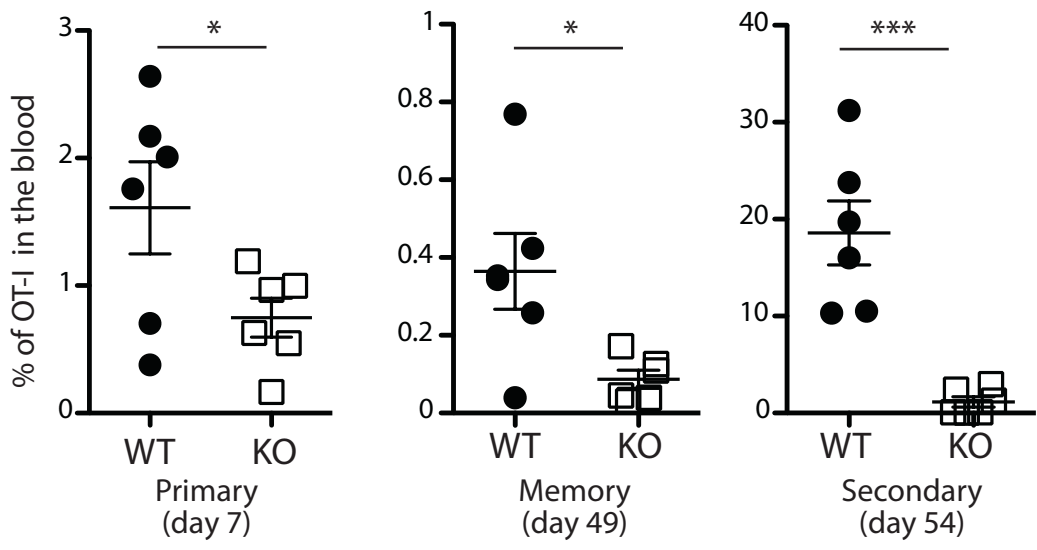
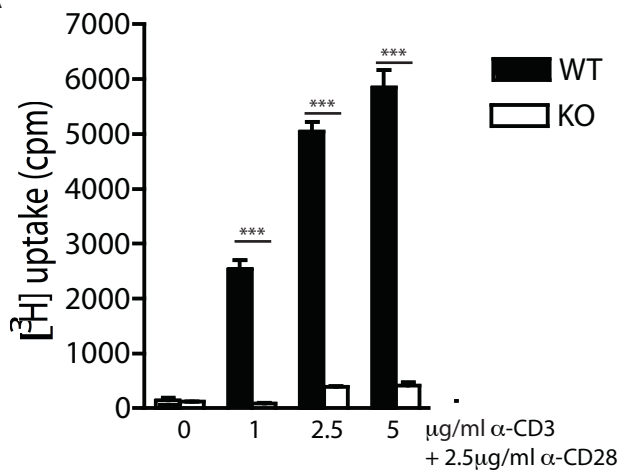


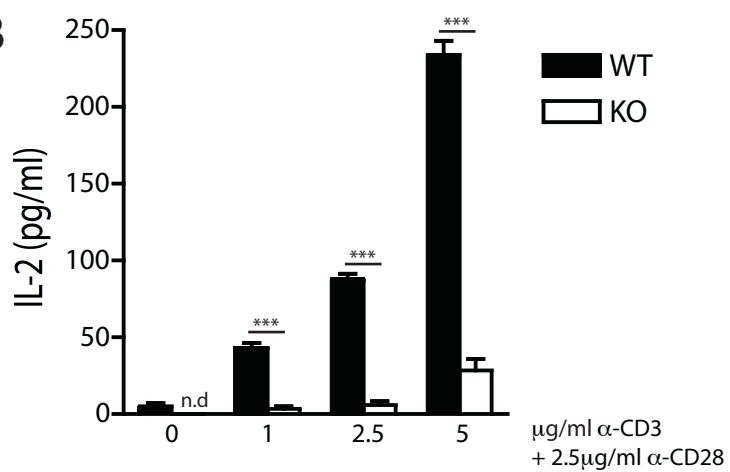
Figure S3, Feau et al.



A



B



C

