

# **Supplementary Material**

## **The Reactivity of Human Serum Albumin towards *trans*-4-Hydroxy-2-nonenal**

Qingyuan Liu, David C. Simpson, Scott Gronert\*

Department of Chemistry, Virginia Commonwealth University, Richmond, VA, USA

### **Data Analysis Expanded Details**

Sequest (version 28, revision 13; Thermo, San Jose, CA, USA) [1], controlled through BioWorks Browser (version 3.3.1, SP1; Thermo), was used to obtain peptide identifications from recorded tandem mass spectra (MS/MS). Scan grouping, when extracting peak lists, was disabled in all cases. For searching linear ion trap MS/MS from the LTQ XL (Thermo) instrument, average masses were used for parent ions while monoisotopic masses were used for fragment ions. For LTQ Orbitrap Velos (Thermo) data, monoisotopic masses were used for both parent and fragment ions. This approach was taken because the LTQ Orbitrap Velos system can, while the LTQ XL linear ion trap system cannot, reliably identify the monoisotopic peak. Only peptides fully conforming to digestion by trypsin were considered. Trypsin was defined as cutting to the C-terminal side of lysine and arginine except when the residue to the C-terminal side of the proposed break was proline. A maximum of two missed cleavages were allowed. For searching linear ion trap MS/MS from the LTQ XL instrument, the parent ion tolerance was set to

$\pm 2$  AMU while the fragment ion tolerance for all dissociation methods was set to  $\pm 1$  AMU. For Orbitrap data, the parent ion tolerance was set to  $\pm 15$  ppm while the fragment ion tolerance for collision-induced dissociation (CID) fragmentation was set to  $\pm 1$  AMU and for higher-energy C-trap dissociation (HCD) fragmentation was set to  $\pm 0.1$  AMU (HCD MS/MS were recorded using the Orbitrap while CID MS/MS were recorded in the linear ion trap region of the LTQ Orbitrap Velos instrument). A maximum of three differential modifications were permitted for each peptide. Precise modification mass shifts are listed in Table S1 for experiments without iTRAQ labels (Applied Biosystems, Foster City, CA, USA) and in Table S2 for experiment with iTRAQ labels.

Modification Description	Modification Composition	Monoisotopic Mass Shift	Type
Oxidation at Met	+O	+15.994915	Differential
Iodoacetamide Alkylation at Cys	+C <sub>2</sub> H <sub>3</sub> NO	+57.021464	Fixed
Michael Addition of HNE at Cys, Reduced	+C <sub>9</sub> H <sub>18</sub> O <sub>2</sub> -[acetamide cap]	+101.109216	Differential
Michael Addition of HNE at His, Reduced	+C <sub>9</sub> H <sub>18</sub> O <sub>2</sub>	+158.130680	Differential
Michael Addition of HNE at Lys, Reduced	+C <sub>9</sub> H <sub>18</sub> O <sub>2</sub>	+158.130680	Differential
Schiff Base Formation with HNE at Lys, Reduced	+C <sub>9</sub> H <sub>16</sub> O	+140.120115	Differential

Table S1: Modification mass shifts considered in non-iTRAQ experiments

Modification Description	Modification Composition	Monoisotopic Mass Shift	Type
Oxidation at Met	+O	+15.994915	Differential
Iodoacetamide Alkylation at Cys	+C <sub>2</sub> H <sub>3</sub> NO	+57.021464	Fixed
iTRAQ Label at Lys	+ <sup>12</sup> C <sub>5</sub> <sup>13</sup> C <sub>2</sub> <sup>1</sup> H <sub>12</sub> <sup>14</sup> N <sub>2</sub> <sup>18</sup> O/ + <sup>12</sup> C <sub>4</sub> <sup>13</sup> C <sub>3</sub> <sup>1</sup> H <sub>12</sub> <sup>14</sup> N <sup>15</sup> N <sup>16</sup> O	+144.103991 <sup>†</sup>	Fixed
iTRAQ Label at Peptide N-terminus	+ <sup>12</sup> C <sub>5</sub> <sup>13</sup> C <sub>2</sub> <sup>1</sup> H <sub>12</sub> <sup>14</sup> N <sub>2</sub> <sup>18</sup> O/ + <sup>12</sup> C <sub>4</sub> <sup>13</sup> C <sub>3</sub> <sup>1</sup> H <sub>12</sub> <sup>14</sup> N <sup>15</sup> N <sup>16</sup> O	+144.103991 <sup>†</sup>	Fixed
iTRAQ Label at Tyr <sup>‡</sup>	+ <sup>12</sup> C <sub>5</sub> <sup>13</sup> C <sub>2</sub> <sup>1</sup> H <sub>12</sub> <sup>14</sup> N <sub>2</sub> <sup>18</sup> O/ + <sup>12</sup> C <sub>4</sub> <sup>13</sup> C <sub>3</sub> <sup>1</sup> H <sub>12</sub> <sup>14</sup> N <sup>15</sup> N <sup>16</sup> O	+144.103991 <sup>†</sup>	Differential
Michael Addition of HNE at Cys, Reduced	+C <sub>9</sub> H <sub>18</sub> O <sub>2</sub> -[acetamide cap]	+101.109216	Differential
Michael Addition of HNE at His, Reduced	+C <sub>9</sub> H <sub>18</sub> O <sub>2</sub>	+158.130680	Differential
Michael Addition of HNE at Lys, Reduced	+C <sub>9</sub> H <sub>18</sub> O <sub>2</sub> -[iTRAQ label]	+14.026689	Differential
Schiff Base Formation with HNE at Lys, Reduced	+C <sub>9</sub> H <sub>16</sub> O -[iTRAQ label]	-3.983876	Differential

Table S2: Modification mass shifts considered in iTRAQ experiments; <sup>†</sup> iTRAQ label mass shift value was the average for the four tags (separation between heavy pair and light pair is circa 0.004 Da, too small to resolve with available instrumentation); <sup>‡</sup> iTRAQ label addition at tyrosine only considered with Orbitrap data and found to be uncommon

Sequest used a protein sequence database that was composed of entries for normal and reversed versions of NCBI RefSeq human sequences and the UniProt porcine trypsin sequence. NCBI RefSeq human sequences were downloaded on November 20, 2010, from [ftp://ftp.ncbi.nlm.nih.gov/refseq/H\\_sapiens/mRNA\\_Prot/human.protein.faa.gz](ftp://ftp.ncbi.nlm.nih.gov/refseq/H_sapiens/mRNA_Prot/human.protein.faa.gz), while the UniProt porcine trypsin sequence (TRYP\_PIG) was obtained on the same day from <http://www.uniprot.org/uniprot/P00761>. The final database contained 68,040 sequences (normal and reversed versions of 34,019 human and one pig protein).

Initial Sequest output was refined using the Trans-Proteomic Pipeline software package (version 4.4, VUVUZELA, revision 1, build 201010121551, MinGW; Institute for Systems Biology, Seattle, WA, USA). PeptideProphet [2] was operated in semi-

supervised mode [3] so that reversed (decoy) protein sequence entries could be used to improve discrimination between correct and incorrect identifications. At the PeptideProphet stage, repeat runs were combined and iTRAQ reporter ion intensities were extracted and corrected (using the Libra module; isotope corrections were those provided by Applied Biosystems, the iTRAQ kit manufacturer). PeptideProphet output was exported and then manipulated using Microsoft Access database software. Instances where the same MS/MS produces two identifications capable of passing the scoring threshold were removed (can occur in linear ion trap data when MS/MS are searched as potentially resulting from doubly- and triply-charged precursors).

## Results Expanded Details

MODIFICATIONS: We report modifications resulting from the exposure of human serum albumin (HSA) to *trans*-4-hydroxy-2-nonenal (HNE) at HNE:HSA ratios of 1:1 and 10:1. For both treatment ratios, five complete repeats were performed. Four were analyzed using the linear ion trap system while one was analyzed using the Orbitrap system. For each preparation, three replicate LC–MS/MS runs were collected. Therefore, a total of twelve runs were performed at each ratio using the LTQ XL and three runs were performed at each ratio using the LTQ Orbitrap Velos. In the main manuscript, Table 1 gives the modifications discovered in terms of a count of runs in which the modification in question was detected by (i) CID MS/MS using the LTQ XL, (ii) electron-transfer dissociation (ETD) MS/MS using the LTQ XL and (iii) CID MS/MS using the LTQ Orbitrap Velos. In this document, Tables S3 and S4 give the peptides that support the modification identifications given in Table 1. Results are presented as counts of runs in

which the peptide in question was detected in Table S3 and as simple detection counts in Table S4 (counts for the fragmentation methods and instrumental platforms are given in the same way as found in Table 1). Using a PeptideProphet score threshold of 0.9, 38 modified peptides were initially proposed from the LTQ XL CID scans while 31 were proposed from the LTQ XL ETD scans. After manual scrutiny, 35 peptides from the LTQ XL CID scans and 27 peptides from the LTQ XL ETD scans were accepted. Naturally, there was considerable overlap, resulting in a total of 39 accepted peptides from the LTQ XL. Using the same PeptideProphet scoring threshold, 18 modified peptides were initially identified using the LTQ Orbitrap Velos. After inspection, 17 were accepted. Example MS/MS for all accepted peptides, from all fragmentation methods and instrumental platforms, are provided at the end of this document.

Modified Peptide (with flanking residues)	Modification Site or Sites (MA indicates Michael addition; SB indicates Schiff base formation)	Counts of LC–MS/MS runs in which the modified peptide was identified using LTQ XL CID scans (maximum = 12)/LTQ XL ETD scans (maximum = 12)/LTQ Orbitrap Velos CID scans associated with high mass accuracy Orbitrap precursor ion mass measurements (maximum = 3)	
		1:1 HNE to HSA Ratio	10:1 HNE to HSA Ratio
K.DLGEENFKALVIAFAQYLQQC#PFEDHVK.L	Cys-034 (MA)	not detected	0/0/1
K.ALVIAFAQYLQQC#PFEDHVK.L	Cys-034 (MA)	9/1/0	9/3/3
K.SLH@TLFGDK.L	His-067 (MA)	12/0/3	12/5/3
K.SLHTLFGDK\LCTVATLR.E	Lys-073 (MA)	not detected	3/0/0
R.NECFLQHK-DDNPNLPR.L	Lys-106 (SB)	not detected	0/1/0
R.LVRPEVDVMCTAFH@DNEETFLK.K	His-128 (MA)	0/2/0	2/3/0
K.K^YLYEiar.R	Lys-137 (MA)	not detected	1/0/0
R.RH@PYFYAPELLFFAK.R	His-146 (MA)	12/11/0	12/12/3
R.RH@PYFYAPELLFFAKR.Y	His-146 (MA)	not detected	1/0/0
R.RH@PYFYAPELLFFAK^R.Y	His-146 (MA) & Lys-159 (MA)	not detected	0/2/1
R.YK^AAFTECCQAADK.A	Lys-162 (MA)	not detected	12/2/2
R.YK-AAFTECCQAADK.A	Lys-162 (SB)	not detected	10/1/0
R.LK^CASLQK.F	Lys-199 (MA)	6/0/2	11/0/3
R.AFK^AWAVAR.L	Lys-212 (MA)	not detected	12/0/1
K.AEFAEVSK\LVTDLTK.V	Lys-233 (MA)	not detected	9/4/0
K.LVTDLTK^VH@TECCHGDLLECADDR.A	Lys-240 (MA) & His-242 (MA)	8/0/0	3/0/0
K.VH@TECCHGDLLECADDR.A	His-242 (MA)	0/7/0	0/7/0
K.VH@TECCH@GDLLECADDR.A	His-242 (MA) & His-247 (MA)	not detected	9/4/0
K.VH@TECCH@GDLLECADDRADLAK.Y	His-242 (MA) & His-247 (MA)	not detected	3/7/0
R.ADLAG^YICENQDSISSK.L	Lys-262 (MA)	not detected	5/3/0
K.SH@CIAEVENDEMPADLPSLAADFVESK.D	His-288 (MA)	10/0/0	12/1/3
K.SH@CIAEVENDEM^PADLPSLAADFVESK.D	His-288 (MA)	1/0/0	11/1/1
R.RH@PDYSVVLRL.L	His-338 (MA)	4/0/0	12/3/3
R.H@PDYSVVLRL.L	His-338 (MA)	not detected	2/0/0
R.H@PDYSVVLRLRAKTYETTLEK.C	His-338 (MA)	0/6/0	1/9/0
R.LAK^TYETTLEK.C	Lys-351 (MA)	10/0/0	9/0/0
R.LAK-TYETTLEK.C	Lys-351 (SB)	not detected	3/1/0
K.TYETTLEK^CCAAADPHECYAK.V	Lys-359 (MA)	not detected	1/3/0
K.CCAAADPH@ECYAK.V	His-367 (MA)	not detected	9/0/0
K.VFDEFK^PLVEEPQNLIK.Q	Lys-378 (MA)	not detected	5/1/0
K.QNCELFEQLGEYK^FQNALLVR.Y	Lys-402 (MA)	not detected	4/0/2
K.K^VPQVSTPTLVEVSR.N	Lys-414 (MA)	not detected	11/1/3
K.K-VPQVSTPTLVEVSR.N	Lys-414 (SB)	8/0/0	12/1/3
R.VTK^CCTESLVNR.R	Lys-475 (MA)	not detected	1/0/0
K.EFNAETFTFH@ADICTLSEK.E	His-510 (MA)	1/8/0	12/11/0
K.EFNAETFTFHADICTLSEK^ER.Q	Lys-519 (MA)	not detected	1/0/0
K.K^QTALVELVK.H	Lys-525 (MA)	not detected	0/1/3
K.K-QTALVELVK.H	Lys-525 (SB)	3/0/3	8/3/3
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (MA)	4/7/0	11/11/3
K.EQLK-AVMDDFAAFVEK.C	Lys-545 (SB)	not detected	4/7/0

Table S3: Counts of LC–MS/MS runs in which the indicated modified peptide was identified. The PeptideProphet score threshold was 0.9. Five independent preparations were made for each HNE:HSA ratio. For each ratio, four preparations were analyzed using a Thermo LTQ XL linear ion trap mass spectrometer and one preparation was analyzed using a Thermo LTQ Orbitrap Velos instrument. In all cases, three replicate LC–MS/MS runs were recorded for each preparation. For the LTQ XL runs, one collision-induced dissociation (CID) and one electron-transfer dissociation (ETD) MS/MS were recorded for each precursor ion selected for fragmentation; resulting CID and ETD spectrum counts are presented separately. For the LTQ Orbitrap Velos runs, only CID MS/MS were recorded. C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K- indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.

Modified Peptide (with flanking residues)	Modification Site or Sites (MA indicates Michael addition; SB indicates Schiff base formation)	Counts of spectra that passed the PeptideProphet score threshold obtained using LTQ XL CID scans (from 12 LC-MS/MS runs)/LTQ XL ETD scans (from 12 LC-MS/MS runs)/LTQ Orbitrap Velos CID scans associated with high mass accuracy Orbitrap precursor ion mass measurements (from 3 LC-MS/MS runs)	
		1:1 HNE to HSA Ratio	10:1 HNE to HSA Ratio
K.DLGEENFKALVLIAFAQYLQQC#PFEDHVK.L	Cys-034 (MA)	not detected	0/0/1
K.ALVLIAFAQYLQQC#PFEDHVK.L	Cys-034 (MA)	13/1/0	16/3/10
K.SLH@TLFGDK.L	His-067 (MA)	16/0/8	68/23/30
K.SLHTLFGDK^LCTVATLR.E	Lys-073 (MA)	not detected	3/0/0
R.NECFLQHK~DDNPNLPR.L	Lys-106 (SB)	not detected	0/1/0
R.LVRPEVDVMCTAFH@DNEETFLK.K	His-128 (MA)	0/2/0	2/4/0
K.K^YLYEiar.R	Lys-137 (MA)	not detected	1/0/0
R.RH@PYFYAPELFFAK.R	His-146 (MA)	27/24/0	157/130/26
R.RH@PYFYAPELFFAK.R.Y	His-146 (MA)	not detected	1/0/0
R.RH@PYFYAPELFFAK^R.Y	His-146 (MA) & Lys-159 (MA)	not detected	0/2/1
R.YK^AAFTECCQAADK.A	Lys-162 (MA)	not detected	20/2/4
R.YK~AAFTECCQAADK.A	Lys-162 (SB)	not detected	10/1/0
R.LK^CASLQK.F	Lys-199 (MA)	6/0/4	11/0/6
R.AFK^AWAVAR.L	Lys-212 (MA)	not detected	12/0/2
K.AEFAEVSK^LVTDLTK.V	Lys-233 (MA)	not detected	9/4/0
K.LVTDLK^VH@TECCHGDLLEADDR.A	Lys-240 (MA) & His-242 (MA)	8/0/0	3/0/0
K.VH@TECCHGDLLEADDR.A	His-242 (MA)	0/7/0	0/10/0
K.VH@TECCH@GDLLEADDR.A	His-242 (MA) & His-247 (MA)	not detected	11/4/0
K.VH@TECCH@GDLLEADDRADLAK.Y	His-242 (MA) & His-247 (MA)	not detected	3/7/0
R.ADLAG^YICENQDSISSK.L	Lys-262 (MA)	not detected	6/3/0
K.SH@CIAEVENDEMPADLPSLAADFVESK.D	His-288 (MA)	10/0/0	86/8/6
K.SH@CIAEVENDEM*PADLPSLAADFVESK.D	His-288 (MA)	1/0/0	16/2/1
R.RH@PDYSVVLLR.L	His-338 (MA)	4/0/0	46/3/21
R.H@PDYSVVLLR.L	His-338 (MA)	not detected	2/0/0
R.H@PDYSVVLLRLAKTYETTLEK.C	His-338 (MA)	0/6/0	1/9/0
R.LAK^TYETTLEK.C	Lys-351 (MA)	10/0/0	9/0/0
R.LAK-TYETTLEK.C	Lys-351 (SB)	not detected	3/1/0
K.TYETTLEK^CCAAADPHECYAK.V	Lys-359 (MA)	not detected	1/3/0
K.CCAAADPH@ECYAK.V	His-367 (MA)	not detected	9/0/0
K.VFDEFK^PLVEEPQNLIK.Q	Lys-378 (MA)	not detected	5/1/0
K.QNCELFEQLGEYK^FQNALLVR.Y	Lys-402 (MA)	not detected	4/0/4
K.K^VPQVSTPTLVEVSR.N	Lys-414 (MA)	not detected	25/4/5
K.K~VPQVSTPTLVEVSR.N	Lys-414 (SB)	8/0/0	36/4/17
R.VTK^CCTESLVNR.R	Lys-475 (MA)	not detected	1/0/0
K.EFNAETFTFH@ADICTLSEK.E	His-510 (MA)	1/8/0	20/21/0
K.EFNAETFTFHADICTLSEK^ER.Q	Lys-519 (MA)	not detected	1/0/0
K.K^QTALVELVK.H	Lys-525 (MA)	not detected	0/1/7
K.K~QTALVELVK.H	Lys-525 (SB)	3/0/8	8/5/17
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (MA)	4/7/0	19/21/6
K.EQLK~AVMDDFAAFVEK.C	Lys-545 (SB)	not detected	4/7/0

Table S4: Counts of MS/MS supporting the indicated modified peptides. The PeptideProphet score threshold was 0.9. Five independent preparations were made for each HNE:HSA ratio. For each ratio, four preparations were analyzed using a Thermo LTQ XL linear ion trap mass spectrometer and one preparation was analyzed using a Thermo LTQ Orbitrap Velos instrument. In all cases, three replicate LC-MS/MS runs were recorded for each preparation. For the LTQ XL runs, one collision-induced dissociation (CID) and one electron-transfer dissociation (ETD) MS/MS were recorded for each precursor ion selected for fragmentation; resulting CID and ETD spectrum counts are presented separately. For the LTQ Orbitrap Velos runs, only CID MS/MS were recorded. C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K- indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.

**MASS LIST:** Measurements of iTRAQ reporter ion intensities obtained using pulsed-Q dissociation (PQD) in a linear ion trap system are not very precise, so a targeted mass list was used to increase the number of measurements obtained. The targeted mass list was also used for iTRAQ experiments performed using the LTQ Orbitrap Velos instrument. Those peptides that appeared to be most important from the initial survey runs and from previously published studies [4,5] were included in the mass list. The mass list contains *m/z* ratios for singly-, doubly- and triply-charged ions falling within the analyzed *m/z* window (300–2000). Both modified peptides and unmodified versions of the same peptide were included. In cases where HNE addition was indicated at lysine, the full length peptide, which now contains a missed cleavage, and the two shorter peptides covering the same sequence that would result from cleavage with trypsin were included (provided they were long enough to be efficiently detected). The mass list was constructed with the aim of covering eighteen modification sites and contained 138 entries. The *m/z* ratio window around the listed masses that was accepted was  $\pm 2$ . Approximately 27% of the *m/z* ratio window from which precursor ions were selected was covered. The complete targeted mass list is presented as Table S5 while a shorter list of targeted modified peptides is provided as Table 2 in the main manuscript.

Modification Site or Modification Site(s) Associated with Unmodified Peptide (MA indicates Michael addition; SB indicates Schiff base formation)	Peptide	Mass List Constituents		
		[M+H] <sup>+</sup>	[M+2H] <sup>2+</sup>	[M+3H] <sup>3+</sup>
Cys-034 (MA)	ALVLIAFAQYLQQC#PFEDHVK	2879.60	1440.31	960.54
Cys-034-associated	ALVLIAFAQYLQQCPFEDHVK	2778.49	1389.75	926.84
Lys-051 (MA)	LVNEVTEFAK^TCVAD	1998.06	999.53	666.69
Lys-051-associated	LVNEVTEFAKTCVAD	1984.03	992.52	662.02
Lys-051-associated	LVNEVTEFAK	1437.82	719.42	479.95
Lys-051-associated	TCVAD	709.33	355.17	237.12
His-067 (MA)	SLH@TLFGDK	1463.88	732.44	488.63
His-067-associated	SLHTLFGDK	1305.74	653.38	435.92
His-105 (MA)	NECFLQH@K	1521.84	761.42	507.95
His-105-associated	NECFLQHK	1363.71	682.36	455.24
Lys-162 (MA)	YK^AAFT ECCQAADK	2109.06	1055.04	703.69
Lys-162-associated	YKAAFT ECCQAADK	2095.04	1048.02	699.02
Lys-162-associated	AAFT ECCQAADK	1659.78	830.39	553.93
Lys-199 (MA)	LK^CASLQK	1393.87	697.44	465.30
Lys-199-associated	LKCASLQK	1379.85	690.43	460.62
Lys-199-associated	CASLQK	994.56	497.79	332.19
Lys-212 (MA)	AFK^AWAVAR	1321.81	661.41	441.28
Lys-212-associated	AFKAWAVAR	1307.79	654.40	436.60
Lys-212-associated	AWAVAR	817.48	409.25	273.17
Lys-233 (MA)	AEFAEVSK^LVTDLTK	2097.23	1049.12	699.75
Lys-233-associated	AEFAEVSKLVTDLTK	2083.21	1042.11	695.07
Lys-233-associated	AEFAEVSK	1168.65	584.83	390.22
Lys-233-associated	LVTDLTK	1077.68	539.34	359.90
His-242 (MA)	VH@TECCHGDLLECADDR	2389.07	1195.04	797.03
His-247 (MA)	VHTECCH@GDLLECADDR	2389.07	1195.04	797.03
His-242-associated & His-247-associated	VHTECCHGDLLECADDR	2230.94	1115.97	744.32
Lys-262 (MA)	ADLAK^YICENQDSISSLK	2388.26	1194.63	796.76
Lys-262-associated	ADLAKYICENQDSISSLK	2374.23	1187.62	792.08
Lys-262-associated	ADLAK	805.51	403.26	269.17
Lys-262-associated	YICENQDSISSLK	1731.85	866.43	577.96
His-288 (MA)	SH@CIAEVENDEM*PADLPSLAADFVESK	3436.68	1718.84	1146.23
His-288 (MA)	SH@CIAEVENDEMPADLPSLAADFVESK	3420.68	1710.85	1140.90
His-288-associated	SHCIAEVENDEM*PADLPSLAADFVESK	3278.55	1639.78	1093.52
His-288-associated	SHCIAEVENDEMPADLPSLAADFVESK	3262.55	1631.78	1088.19
Lys-351 (MA)	LAK^TYETTLEK	1743.04	872.03	581.69
Lys-351-associated	LAKTYETTLEK	1729.02	865.01	577.01
Lys-351-associated	TYETTLEK	1272.70	636.85	424.90
His-367 (MA)	CCAAADPH@ECYAK	1998.94	999.97	666.98
His-367-associated	CCAAADPHECYAK	1840.81	920.91	614.27
Lys-378 (MA)	VFDEFK^PLVEEPQNLIK	2491.43	1246.22	831.15
Lys-378-associated	VFDEFKPLVEEPQNLIK	2477.41	1239.21	826.47
Lys-378-associated	VFDEFK	1072.60	536.80	358.20
Lys-378-associated	PLVEEPQNLIK	1567.93	784.47	523.32
Residue 403-410 peptide	FQNALLVR	1104.67	552.84	368.89
Lys-414 (MA)	K^VPQVSTPTLVEVSR	1942.17	971.59	648.06
Lys-414 (SB)	K-VPQVSTPTLVEVSR	1924.16	962.59	642.06
Lys-414-associated	KVPQVSTPTLVEVSR	1928.15	964.58	643.39
Lys-414-associated	VPQVSTPTLVEVSR	1655.95	828.48	552.65
His-510 (MA)	EFNAETFTFH@ADICTLSEK	2706.36	1353.68	902.79
His-510-associated	EFNAETFTFHADICTLSEK	2548.23	1274.62	850.08
Lys-525 (MA)	K^QTALVELVK	1575.04	788.02	525.68
Lys-525 (SB)	K-QTALVELVK	1557.03	779.02	519.68
Lys-525-associated	KQTALVELVK	1561.01	781.01	521.01
Lys-525-associated	QTALVELVK	1288.81	644.91	430.28

Table S5: List of targeted peptides and associated monoisotopic mass-to-charge ratios that were used as a targeted mass list for choosing parent ions for fragmentation. Mass-to-charge ratios in gray are outside the analyzed range (300-2000). Targeted peptides are grouped by modification site. C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K- indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met. Eighteen sites were targeted and both Schiff base formation and Michael addition were considered at two of these sites (Lys-414 and Lys-525). The peptide, FQNALLVR, was included in error and is listed here for completeness. In total, the mass list contained 138 entries.

**RELATIVE ABUNDANCE MEASUREMENTS:** Changes in relative abundance for HNE-modified and corresponding unmodified HSA peptides were measured using the iTRAQ approach. The targeted mass list was used in all cases. Table S6 gives relative abundances for a comparison of no added HNE with applied HNE:HSA ratios of 50:1 and 100:1. A single repeat experiment was performed and three replicate LC–MS/MS runs were collected using PQD fragmentation on the LTQ XL system. Only mass list peptides are reported in the table. Bolded peptides were used in constructing Figure 1 in the main document. Tables S7 and S8 give the results of an experiment in which incubation time was varied at the 100:1 HNE:HSA ratio. Incubation times of 0, 1, 3, and 24 h were investigated. Three complete repeats were performed. For each repeat, three replicate LC–MS/MS runs were collected using PQD fragmentation on the LTQ XL system and three further replicates were recorded using HCD fragmentation on the LTQ Orbitrap Velos system. Linear ion trap/PQD results are given as Table S7 and Orbitrap/HCD results are given as Table S8. Together, these results were used to construct Figures 2 and 3 in the main document (Figure 2 displays results for the histidines while Figure 3 does the same for the lysines). Bolded entries in the tables indicate those peptides used in constructing the main document figures. Where more than one peptide was available to represent a site (most common for the unmodified peptides corresponding to the lysine modification sites), the most frequently-detected peptide was used. As can be seen from Tables S6–S8, the same peptides were chosen for use in constructing the main document figures in all cases. For peptide identification acceptance, a PeptideProphet score of 0.9 or greater was required and MS/MS for which all four iTRAQ reporter ion intensities were zero were discarded.

Modification	Peptide	Detection Count	No HNE Added	50:1 HNE:HSA	100:1 HNE:HSA
Cys-034-associated	ALVLIAFAQYLQQCPFEDHVK	412	78.9%	11.0%	10.2%
Cys-034 (MA)	ALVLIAFAQYLQQC#PFEDHVK	2	15.0%	0.0%	85.0%
His-067-associated	<b>SLHTLFGDK</b>	795	<b>70.8%</b>	<b>18.6%</b>	<b>10.7%</b>
His-105-associated	NECFLQHK	345	50.1%	27.8%	22.0%
His-242- & His-247-associated	VHTECCHGDLLECADDR	239	65.3%	19.0%	15.7%
His-288-associated	SHCIAEVENDEMPADLPSLAADFVESK	433	84.0%	10.9%	5.0%
His-367-associated	<b>CCAAADPHECYAK</b>	642	<b>54.4%</b>	<b>28.4%</b>	<b>17.1%</b>
His-510-associated	<b>EFNAETFTFHADICTLSEK</b>	748	<b>75.5%</b>	<b>17.6%</b>	<b>6.9%</b>
His-067 (MA)	<b>SLH@TLFGDK</b>	916	1.3%	51.2%	47.5%
His-105 (MA)	NECFLQH@K	89	1.2%	42.7%	56.1%
His-367 (MA)	CCAAADPH@ECYAK	326	2.1%	42.6%	55.2%
His-510 (MA)	EFNAETFTFH@ADICTLSEK	126	2.7%	34.4%	62.8%
Lys-051-associated	LVNEVTEFAK	1356	46.6%	31.2%	22.3%
Lys-162-associated	<b>AAFTECCQAADK</b>	527	<b>49.7%</b>	<b>30.4%</b>	<b>19.9%</b>
Lys-199-associated	CASLQK	21	82.2%	14.2%	3.6%
Lys-233-associated	<b>AEFAEVSK</b>	1018	<b>44.2%</b>	<b>29.7%</b>	<b>26.2%</b>
Lys-233-associated	<b>LVTDLTK</b>	313	42.4%	30.6%	27.1%
Lys-262-associated	ADLAK	6	47.2%	25.5%	27.4%
Lys-262-associated	<b>YICENQDSISSLK</b>	1195	<b>47.6%</b>	<b>29.6%</b>	<b>22.8%</b>
Lys-351-associated	<b>TYETTLEK</b>	212	<b>59.2%</b>	<b>26.7%</b>	<b>14.1%</b>
Lys-378-associated	<b>VFDEFKPLVEEPQNLIK</b>	895	<b>46.5%</b>	<b>29.4%</b>	<b>24.1%</b>
Lys-414-associated	<b>KVPQVSTPTLVEVSR</b>	778	<b>46.5%</b>	<b>32.0%</b>	<b>21.5%</b>
Lys-414-associated	VPQVSTPTLVEVSR	575	37.4%	36.5%	26.1%
Lys-525-associated	KQTALVELVK	174	79.4%	15.9%	4.7%
Lys-525-associated	<b>QTALVELVK</b>	794	<b>70.1%</b>	<b>18.4%</b>	<b>11.5%</b>
Lys-162 (MA)	<b>YK^AAFTECCQAADK</b>	44	1.6%	44.0%	54.4%
Lys-199 (MA)	<b>LK^CASLQK</b>	1	0.0%	41.2%	58.8%
Lys-233 (MA)	<b>AEFAEVSK^LVTDLTK</b>	79	1.7%	28.6%	69.6%
Lys-262 (MA)	<b>ADLAK^YICENQDSISSLK</b>	2	0.0%	13.0%	87.0%
Lys-351 (MA)	<b>LAK^TYETTLEK</b>	43	1.2%	44.8%	54.1%
Lys-378 (MA)	<b>VFDEFK^PLVEEPQNLIK</b>	43	21.3%	27.6%	51.0%
Lys-414 (MA)	<b>K^VPQVSTPTLVEVSR</b>	248	10.5%	45.9%	43.6%
Lys-414 (SB)	<b>K~VPQVSTPTLVEVSR</b>	118	12.5%	43.9%	43.7%
Lys-525 (MA)	<b>K^QTALVELVK</b>	135	1.0%	54.1%	44.9%
Lys-525 (SB)	<b>K~QTALVELVK</b>	12	2.2%	70.3%	27.5%

Table S6: Change in relative abundance for HNE-modified and corresponding unmodified HSA peptides in response to HNE-exposure at stated HNE:HSA molar ratios. Raw iTRAQ reporter ion intensities for each MS/MS are converted to relative percentages and then averaged for each peptide; since the control was duplicated, raw intensity for the two control signals was averaged before relative percentages were calculated. iTRAQ reporter ion intensities were obtained using pulsed-Q dissociation (PQD) in a linear ion trap mass spectrometer. This dataset is the result of a single repeat preparation subjected to three replicate LC–MS/MS runs. Incubation time with HNE was 3 h. The mass list described in Table S5 was used and only mass list peptides are included in this table. Bolded entries were used in constructing Figure 1 while italicized entries were not used. For a peptide identification to be accepted, a PeptideProphet score of 0.9 or greater was required. MS/MS where all four iTRAQ reporter ion intensities were zero were discarded. C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K- indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.

Modification	Peptide	Detection Count	Repeat Count	100:1 HNE:HSA for 0 h	100:1 HNE:HSA for 1 h	100:1 HNE:HSA for 3 h	100:1 HNE:HSA for 24 h
Cys-034-associated	ALVLIAFAAQYLQQCPFEDHVK	342	3	31.6%	20.2%	19.1%	29.1%
Cys-034 (MA)	ALVLIAFAAQYLQQC#PFEDHVK	70	1	3.2%	32.4%	24.2%	40.3%
His-067-associated	SLHTLFGDK	953	3	52.4%	28.4%	13.6%	5.5%
His-105-associated	NECFLQHK	437	3	33.2%	28.1%	24.5%	14.1%
His-242- & His-247-associated	VHTECCHGDLLECADDR	353	3	51.4%	15.6%	21.0%	12.0%
His-288-associated	SHCIAEVENDEM^PADLPSLAADFVESK	857	2	14.4%	57.8%	15.1%	12.7%
His-288-associated	SHCIAEVENDEMPADLPSLAADFVESK	866	3	60.2%	17.7%	13.3%	8.8%
His-367-associated	CCAAADPHECYAK	966	3	35.5%	30.2%	22.1%	12.2%
His-510-associated	EFGNAETFTFHADICTLSEK	2204	3	48.0%	27.8%	13.9%	10.4%
His-067 (MA)	SLH@TLFGDK	3622	3	0.9%	28.2%	37.6%	33.3%
His-105 (MA)	NECFLQH@K	260	3	0.9%	24.9%	27.5%	46.7%
His-242 (MA)	VH@TECCHGDLLECADDR	8	3	0.4%	26.4%	43.5%	29.7%
His-288 (MA)	SH@CIAEVENDEM^PADLPSLAADFVESK	834	2	1.2%	13.8%	45.7%	39.4%
His-288 (MA)	SH@CIAEVENDEMPADLPSLAADFVESK	2751	2	2.2%	13.6%	45.1%	39.1%
His-367 (MA)	CCAAADPH@ECYAK	1401	3	1.2%	14.8%	23.8%	60.2%
His-510 (MA)	EFGNAETFTFH@ADICTLSEK	1722	3	1.2%	23.8%	36.9%	38.2%
Lys-051-associated	LVNEVTEFAK	2442	3	26.4%	27.4%	23.9%	22.3%
Lys-162-associated	AAFFTECCQAADK	1052	3	32.2%	29.4%	23.3%	15.1%
Lys-199-associated	CASLQK	29	2	79.8%	6.0%	6.6%	7.6%
Lys-233-associated	AEFAEVSK	1404	3	26.5%	27.7%	24.1%	21.6%
Lys-233-associated	AEFAEVSKLVTDLK	1	1	6.3%	39.7%	24.5%	29.5%
Lys-233-associated	LVTDLK	499	3	27.1%	26.7%	25.2%	21.1%
Lys-262-associated	ADLAK	5	2	35.0%	17.7%	19.6%	27.7%
Lys-262-associated	YICENQDSISSK	1803	3	26.2%	26.4%	23.8%	23.6%
Lys-351-associated	TYETTLEK	344	3	39.4%	26.8%	19.1%	14.7%
Lys-378-associated	VFDDEFKPLVEEPQNLIK	6292	3	24.2%	28.2%	25.0%	22.6%
Lys-414-associated	KVPQVSTPTLVEVSR	1293	3	32.1%	27.0%	22.9%	18.0%
Lys-414-associated	VPOQVSTPTLVEVSR	1052	3	19.8%	36.5%	24.8%	18.9%
Lys-525-associated	KQTALVELVK	270	3	62.9%	13.9%	12.5%	10.7%
Lys-525-associated	QTALVELVK	1020	3	48.5%	17.5%	16.6%	17.3%
Lys-162 (MA)	YK^AAFTECCQAADK	210	3	1.3%	32.1%	38.5%	28.1%
Lys-199 (MA)	LK^CASLQK	33	3	1.1%	26.7%	38.3%	33.9%
Lys-212 (MA)	AFK^AWAVAR	48	2	9.2%	14.4%	21.2%	55.2%
Lys-233 (MA)	AEFAEVSK^LVTDLK	355	3	0.9%	11.2%	21.2%	66.7%
Lys-262 (MA)	ADLAK^YICENQDSISSK	120	3	0.9%	18.0%	26.8%	54.4%
Lys-351 (MA)	LAK^TYETTLEK	222	3	2.2%	20.7%	38.4%	38.7%
Lys-378 (MA)	VFDDEFK^PLVEEPQNLIK	357	3	11.6%	22.0%	24.8%	41.6%
Lys-414 (MA)	K^VPQVSTPTLVEVSR	501	3	6.4%	33.4%	26.5%	33.7%
Lys-414 (SB)	K-VPQVSTPTLVEVSR	145	3	11.7%	44.6%	28.2%	15.5%
Lys-525 (MA)	K^QTALVELVK	345	3	1.1%	20.2%	30.8%	47.8%
Lys-525 (SB)	K-QTALVELVK	1	1	0.0%	0.0%	20.2%	79.8%

Table S7: Change in relative abundance for HNE-modified and corresponding unmodified HSA peptides in response to HNE-exposure time at a 100:1 HNE:HSA molar ratio. Raw iTRAQ reporter ion intensities for each MS/MS are converted to relative percentages and then averaged for each peptide. iTRAQ reporter ion intensities were obtained using pulsed-Q dissociation (PQD) in a linear ion trap mass spectrometer. This dataset is the result of a three repeat preparations subjected to three replicate LC-MS/MS runs each. The Repeat Count column gives a count of repeats in which the peptide in question was identified. The same set of three repeat preparations was analyzed using the LTQ Orbitrap Velos system (see Table S8). The mass list described in Table S5 was used and only mass list peptides are included in this table. Bolded entries were used in constructing Figures 2 and 3 while italicized entries were not used. For a peptide identification to be accepted, a PeptideProphet score of 0.9 or greater was required. MS/MS where all four iTRAQ reporter ion intensities were zero were discarded. C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K~ indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.

Modification	Peptide	Detection Count	Repeat Count	100:1 HNE:HSA for 0 h	100:1 HNE:HSA for 1 h	100:1 HNE:HSA for 3 h	100:1 HNE:HSA for 24 h
Cys-034-associated	ALVLIAFAQYLQQCPFEDHVK	587	3	36.2%	19.3%	20.0%	24.6%
Cys-034 (MA)	ALVLIAFAQYLQQC#PFEDHVK	42	3	0.1%	28.0%	29.4%	42.5%
His-067-associated	SLHTLFGDK	415	3	52.7%	26.6%	14.4%	6.2%
His-105-associated	NECFLQHK	136	3	32.6%	26.6%	24.7%	16.1%
His-242- & His-247-associated	VHTECCHGDLLEADDR	165	3	67.1%	6.5%	15.4%	11.0%
His-288-associated	SHCIAEVENDEM^PADLPSLAADFVESK	427	3	58.2%	23.1%	8.1%	10.5%
His-288-associated	SHCIAEVENDEMPADLPSLAADFVESK	447	3	63.8%	8.5%	9.2%	18.5%
His-367-associated	CCAAADPHCYAK	242	3	35.5%	29.2%	22.9%	12.4%
His-510-associated	EFNAETFTFHADICTLSEK	3968	3	53.1%	25.3%	12.5%	9.2%
His-067 (MA)	SLH@TLFGDK	1726	3	0.7%	27.6%	37.6%	34.0%
His-105 (MA)	NECFLQH@K	144	3	0.3%	17.3%	24.1%	58.4%
His-242 (MA)	VH@TECCHGDLLEADDR	1	1	0.0%	5.8%	26.3%	67.9%
His-288 (MA)	SH@CIAEVENDEM^PADLPSLAADFVESK	432	3	0.7%	18.2%	44.0%	37.2%
His-288 (MA)	SH@CIAEVENDEMPADLPSLAADFVESK	517	3	0.9%	17.0%	45.8%	36.3%
His-367 (MA)	CCAAADPH@ECYAK	300	3	1.6%	14.6%	23.8%	60.0%
His-510 (MA)	EFNAETFTFH@ADICTLSEK	1709	3	0.3%	24.6%	37.3%	37.8%
Lys-051-associated	LVNEVTEFAK	1711	3	26.3%	25.8%	24.0%	23.9%
Lys-162-associated	AAFTECCQAADK	326	3	32.9%	27.2%	23.4%	16.5%
Lys-199-associated	CASLQK	3	1	84.3%	0.9%	5.8%	9.0%
Lys-212-associated	AWAVAR	18	2	25.8%	26.4%	27.0%	20.8%
Lys-233-associated	AEFAEVSK	1405	3	25.5%	25.9%	25.2%	23.4%
Lys-233-associated	LVTDLTK	83	2	21.4%	25.2%	28.1%	25.3%
Lys-262-associated	ADLAK	33	2	21.1%	24.3%	26.7%	27.9%
Lys-262-associated	YICENQDSISSK	433	3	25.7%	26.5%	24.3%	23.6%
Lys-351-associated	LAKTYETTLEK	3	1	40.2%	23.5%	23.3%	13.0%
Lys-351-associated	TYETTLEK	318	3	38.4%	25.2%	20.3%	16.1%
Lys-378-associated	VFDEFKPLVEEPQNLIK	3490	3	25.0%	26.8%	24.8%	23.4%
Lys-414-associated	KVPQVSTPTLVEVSR	2689	3	34.2%	23.8%	21.9%	20.0%
Lys-414-associated	VPOVSTPTLVEVSR	335	3	24.3%	20.2%	24.5%	30.9%
Lys-525-associated	KQTALVELVK	105	3	64.5%	11.2%	13.0%	11.3%
Lys-525-associated	QTALVELVK	337	3	49.7%	14.0%	17.4%	18.9%
Lys-162 (MA)	YK^AAFTECCQAADK	109	3	1.2%	30.3%	38.9%	29.5%
Lys-199 (MA)	LK^CASLQK	82	3	1.3%	27.5%	37.7%	33.5%
Lys-212 (MA)	AFK^AWAVAR	183	3	1.3%	10.7%	21.2%	66.8%
Lys-233 (MA)	AEFAEVSK^LVTDLTK	259	3	0.0%	7.5%	18.8%	73.6%
Lys-262 (MA)	ADLAK^YICENQDSISSK	221	3	0.8%	14.1%	24.8%	60.2%
Lys-351 (MA)	LAK^TYETTLEK	358	3	1.3%	23.9%	34.4%	40.3%
Lys-378 (MA)	VFDEFK^PLVEEPQNLIK	222	3	11.0%	18.4%	23.1%	47.5%
Lys-414 (MA)	K^VPQVSTPTLVEVSR	168	3	8.5%	30.2%	26.9%	34.4%
Lys-414 (SB)	K-VPQVSTPTLVEVSR	139	3	13.0%	40.3%	29.7%	17.0%
Lys-525 (MA)	K^QTALVELVK	269	3	0.6%	23.9%	33.3%	42.3%
Lys-525 (SB)	K-QTALVELVK	150	3	0.7%	54.6%	33.8%	10.8%

Table S8: Change in relative abundance for HNE-modified and corresponding unmodified HSA peptides in response to HNE-exposure time at a 100:1 HNE:HSA molar ratio. Raw iTRAQ reporter ion intensities for each MS/MS are converted to relative percentages and then averaged for each peptide. iTRAQ reporter ion intensities were obtained using higher-energy C-trap dissociation (HCD) in an Orbitrap mass spectrometer. This dataset is the result of a three repeat preparations subjected to three replicate LC-MS/MS runs each. The Repeat Count column gives a count of repeats in which the peptide in question was identified. The same set of three repeat preparations was analyzed using the LTQ XL system (see Table S7). The mass list described in Table S5 was used and only mass list peptides are included in this table. Bolded entries were used in constructing Figures 2 and 3 while italicized entries were not used. For a peptide identification to be accepted, a PeptideProphet score of 0.9 or greater was required. MS/MS where all four iTRAQ reporter ion intensities were zero were discarded. C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K~ indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.

## References

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## Index of Example LTQ XL Linear Ion Trap Tandem Mass Spectra

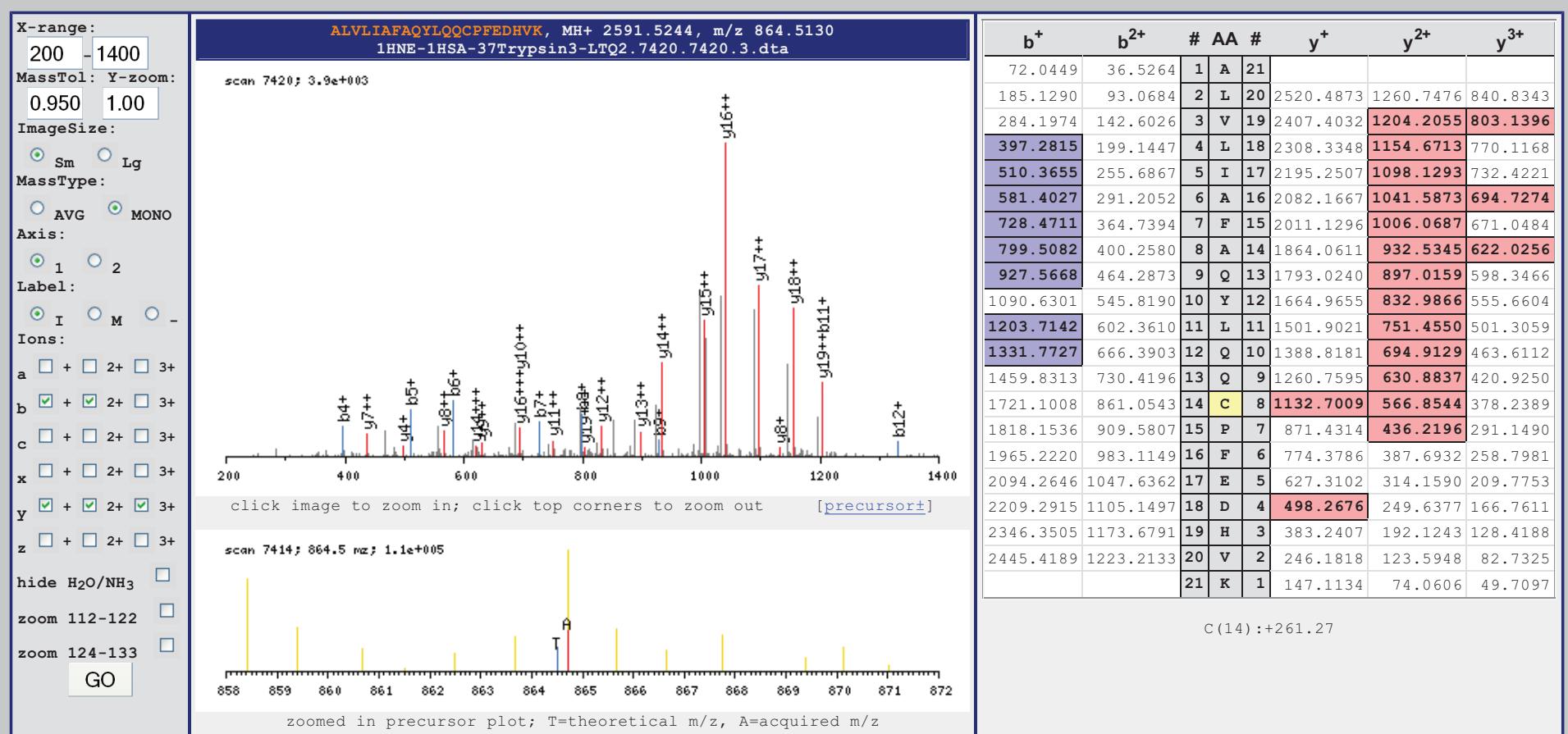
Modified Peptide (with flanking residues)	Modification Site or Sites (MA indicates Michael addition; SB indicates Schiff base formation)	Type	Sequest Xcorr	Sequest DeltaCn	Sequest RankSp	Peptide Prophet Score	Page Number
K.ALVLIAFAQYLQQC#PFEDHVK.L	Cys-034 (MA)	CID	4.873	0.477	1	1.0000	17
K.ALVLIAFAQYLQQC#PFEDHVK.L	Cys-034 (MA)	ETD	5.488	0.458	1	1.0000	18
K.SLH@TLFGDK.L	His-067 (MA)	CID	3.949	0.302	1	0.9999	19
K.SLH@TLFGDK.L	His-067 (MA)	ETD	3.687	0.287	1	0.9998	20
K.SLHTLFGDK\LCVTATL.R.E	Lys-073 (MA)	CID	2.956	0.391	1	0.9770	21
R.NECFLQHK\DDNPNLPR.L	Lys-106 (SB)	ETD	4.241	0.372	1	0.9982	22
R.LVRPEVDVMCTAFH@DNEETFLK.K	His-128 (MA)	CID	5.787	0.436	1	1.0000	23
R.LVRPEVDVMCTAFH@DNEETFLK.K	His-128 (MA)	ETD	5.225	0.457	1	1.0000	24
K.K^LYLEIAR.R	Lys-137 (MA)	CID	3.041	0.312	1	0.9919	25
R.RH@PYFYAPELLFFAK.R	His-146 (MA)	CID	4.582	0.501	1	1.0000	26
R.RH@PYFYAPELLFFAK.R	His-146 (MA)	ETD	5.467	0.433	1	1.0000	27
R.RH@PYFYAPELLFFAK.R.Y	His-146 (MA)	CID	6.144	0.446	1	1.0000	28
R.RH@PYFYAPELLFFAK.R.Y	His-146 (MA) & Lys-159 (MA)	ETD	5.282	0.353	1	0.9999	29
R.YK^AAFTECQAADK.A	Lys-162 (MA)	CID	5.758	0.560	1	1.0000	30
R.YK^AAFTECQAADK.A	Lys-162 (MA)	ETD	5.303	0.489	1	1.0000	31
R.YK~AAFTECQAADK.A	Lys-162 (SB)	CID	4.249	0.331	1	0.9970	32
R.YK~AAFTECQAADK.A	Lys-162 (SB)	ETD	4.803	0.354	1	0.9980	33
R.LK^CASLQK.F	Lys-199 (MA)	CID	4.196	0.261	1	0.9961	34
R.AFK^AWAVAR.L	Lys-212 (MA)	CID	3.815	0.257	2	0.9782	35
K.AEFAEVSK\LVTDLTK.V	Lys-233 (MA)	CID	3.979	0.313	1	0.9931	36
K.AEFAEVSK\LVTDLTK.V	Lys-233 (MA)	ETD	3.436	0.371	1	1.0000	37
K.LVTDLTK\vh@TECCHGDLLECADDR.A	Lys-240 (MA) & His-242 (MA)	CID	4.775	0.319	1	0.9843	38
K.VH@TECCHGDLLECADDR.A	His-242 (MA)	ETD	4.505	0.274	1	0.9997	39
K.VH@TECCH@GDLLECADDR.A	His-242 (MA) & His-247 (MA)	CID	4.561	0.419	1	1.0000	40
K.VH@TECCH@GDLLECADDR.A	His-242 (MA) & His-247 (MA)	ETD	3.321	0.329	1	0.9995	41
K.VH@TECCH@GDLLECADDRADLAK.Y	His-242 (MA) & His-247 (MA)	CID	4.297	0.361	1	0.9923	42
K.VH@TECCH@GDLLECADDRADLAK.Y	His-242 (MA) & His-247 (MA)	ETD	5.437	0.432	1	0.9996	43
R.ADLAK^YICENQDSISSK.L	Lys-262 (MA)	CID	4.947	0.447	1	0.9999	44
R.ADLAK^YICENQDSISSK.L	Lys-262 (MA)	ETD	4.260	0.427	1	0.9978	45
K.SH@CIAEVENDEMPADLPSLAADFVESK.D	His-288 (MA)	CID	6.548	0.471	1	1.0000	46
K.SH@CIAEVENDEMPADLPSLAADFVESK.D	His-288 (MA)	ETD	5.269	0.555	1	1.0000	47
K.SH@CIAEVENDEM*PADLPSLAADFVESK.D	His-288 (MA)	CID	7.038	0.576	1	1.0000	48
K.SH@CIAEVENDEM*PADLPSLAADFVESK.D	His-288 (MA)	ETD	3.552	0.224	1	0.9868	49
R.RH@PDYSVVLRL.I	His-338 (MA)	CID	4.528	0.420	1	1.0000	50
R.RH@PDYSVVLRL.I	His-338 (MA)	ETD	4.562	0.432	1	1.0000	51
R.H@PDYSVVLRL.I	His-338 (MA)	CID	4.439	0.250	1	0.9997	52
R.H@PDYSVVLRL.RAKTYETTLEK.C	His-338 (MA)	CID	3.784	0.360	1	0.9821	53
R.H@PDYSVVLRL.RAKTYETTLEK.C	His-338 (MA)	ETD	6.466	0.463	1	0.9962	54
R.LAK^TYETTLEK.C	Lys-351 (MA)	CID	4.137	0.420	1	0.9998	55
R.LAK-TYETTLEK.C	Lys-351 (SB)	CID	3.265	0.311	2	0.9704	56
R.LAK-TYETTLEK.C	Lys-351 (SB)	ETD	3.874	0.412	1	0.9988	57
K.TYETTLEK^CCAAADPHCYAK.V	Lys-359 (MA)	CID	5.601	0.527	1	1.0000	58
K.TYETTLEK^CCAAADPHCYAK.V	Lys-359 (MA)	ETD	4.199	0.381	1	0.9957	59
K.CCAAADPH@ECYAK.V	His-367 (MA)	CID	4.350	0.460	1	0.9987	60
K.VFDEFK^PLVEEPQNLIK.Q	Lys-378 (MA)	CID	4.943	0.479	1	1.0000	61
K.VFDEFK^PLVEEPQNLIK.Q	Lys-378 (MA)	ETD	2.804	0.178	2	0.9295	62
K.QNCELFEQLGEYK^FQNALLVR.Y	Lys-402 (MA)	CID	3.700	0.442	1	0.9991	63
K.K^VPQVSTPTLVEVSR.N	Lys-414 (MA)	CID	4.007	0.435	1	1.0000	64
K.K^VPQVSTPTLVEVSR.N	Lys-414 (MA)	ETD	4.331	0.346	1	0.9999	65
K.K~VPQVSTPTLVEVSR.N	Lys-414 (SB)	CID	4.161	0.488	1	1.0000	66
K.K~VPQVSTPTLVEVSR.N	Lys-414 (SB)	ETD	4.454	0.297	1	0.9998	67
R.VTK^CCTESLVNR.R	Lys-475 (MA)	CID	3.635	0.402	1	0.9991	68
K.EFNAETFTFH@ADICLSEK.E	His-510 (MA)	CID	5.059	0.588	1	1.0000	69
K.EFNAETFTFH@ADICLSEK.E	His-510 (MA)	ETD	3.799	0.404	1	0.9997	70
K.EFNAETFTFHADICLSEK^ER.Q	Lys-519 (MA)	CID	4.686	0.393	1	0.9992	71
K.K^QTALVELVK.H	Lys-525 (MA)	ETD	2.547	0.293	5	0.9961	72
K.K-QTALVELVK.H	Lys-525 (SB)	CID	4.406	0.189	1	0.9991	73
K.K-QTALVELVK.H	Lys-525 (SB)	ETD	4.142	0.197	2	0.9988	74
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (MA)	CID	5.270	0.512	1	1.0000	75
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (MA)	ETD	6.854	0.527	1	1.0000	76
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (SB)	CID	3.975	0.414	1	0.9993	77
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (SB)	ETD	5.433	0.552	1	1.0000	78

Iodoacetamide addition to cysteine is highlighted in the following Trans-Proteomic Pipeline screenshots, but not in this table (all cysteines were assumed to have been alkylated with iodoacetamide unless HNE addition occurred). C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K- indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.

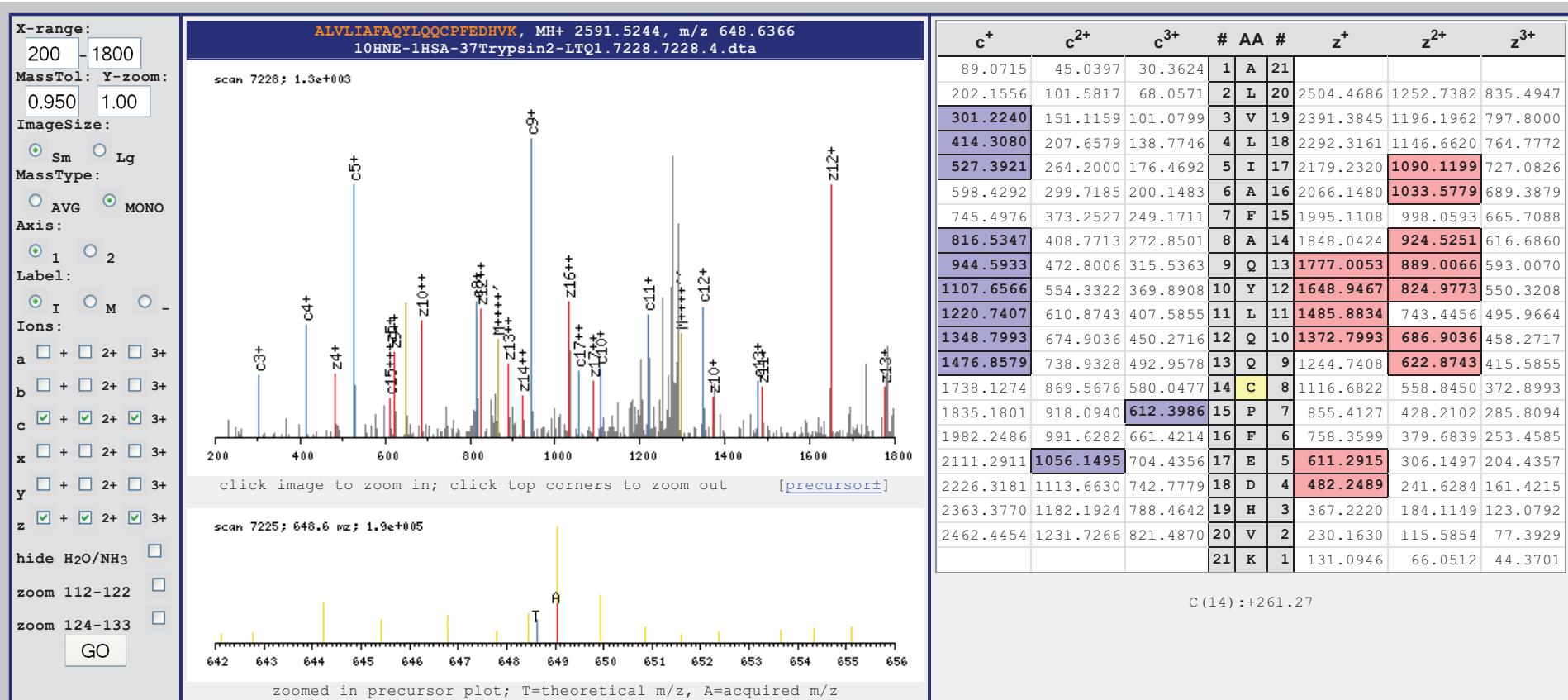
## Index of Example LTQ Orbitrap Velos Linear Ion Trap Tandem Mass Spectra

Modified Peptide (with flanking residues)	Modification Site or Sites (MA indicates Michael addition; SB indicates Schiff base formation)	Type	Sequest Xcorr	Sequest DeltaCn	Sequest RankSp	Peptide Prophet Score	Page Number
K.DLGEENFKALVIAFAQYQLQQC#PFEDHVK.L	Cys-034 (MA)	CID	4.225	0.329	1	0.9248	79
K.ALVLIAFAQYQLQQC#PFEDHVK.L	Cys-034 (MA)	CID	5.397	0.355	1	0.9953	80
K.SLH@TLFGDK.L	His-067 (MA)	CID	3.033	0.459	1	0.9988	81
R.RH@PYFYAPELLFFAK.R	His-146 (MA)	CID	6.258	0.597	1	1.0000	82
R.RH@PYFYAPELLFFAK'R.Y	His-146 (MA) & Lys-159 (MA)	CID	4.641	0.460	1	0.9989	83
R.YK^AAFTTECCQAADK.A	Lys-162 (MA)	CID	4.453	0.480	1	0.9919	84
R.LK^CASLQK.F	Lys-199 (MA)	CID	4.123	0.387	1	0.9845	85
R.AFK^AWAVAR.L	Lys-212 (MA)	CID	3.733	0.488	1	0.9926	86
K.SH@CIAEVENDEMPAIDLPSLAADFVESK.D	His-288 (MA)	CID	6.444	0.592	1	1.0000	87
K.SH@CIAEVENDEM*PADLPSLAADFVESK.D	His-288 (MA)	CID	6.357	0.610	1	1.0000	88
R.RH@PDYSVVLLR.I	His-338 (MA)	CID	5.221	0.449	1	0.9997	89
K.QNCELFEQLGEYK^FQNALLVR.Y	Lys-402 (MA)	CID	4.428	0.489	1	0.9881	90
K.K^PQVSTPTLVESR.N	Lys-414 (MA)	CID	3.622	0.470	1	0.9990	91
K.K~PQVSTPTLVESR.N	Lys-414 (SB)	CID	4.254	0.580	1	0.9998	92
K.K^QTALVELVK.H	Lys-525 (MA)	CID	3.197	0.298	1	0.9854	93
K.K~QTALVELVK.H	Lys-525 (SB)	CID	5.027	0.366	1	0.9993	94
K.EQLK^AVMDDFAAFVEK.C	Lys-545 (MA)	CID	4.783	0.438	1	0.9868	95

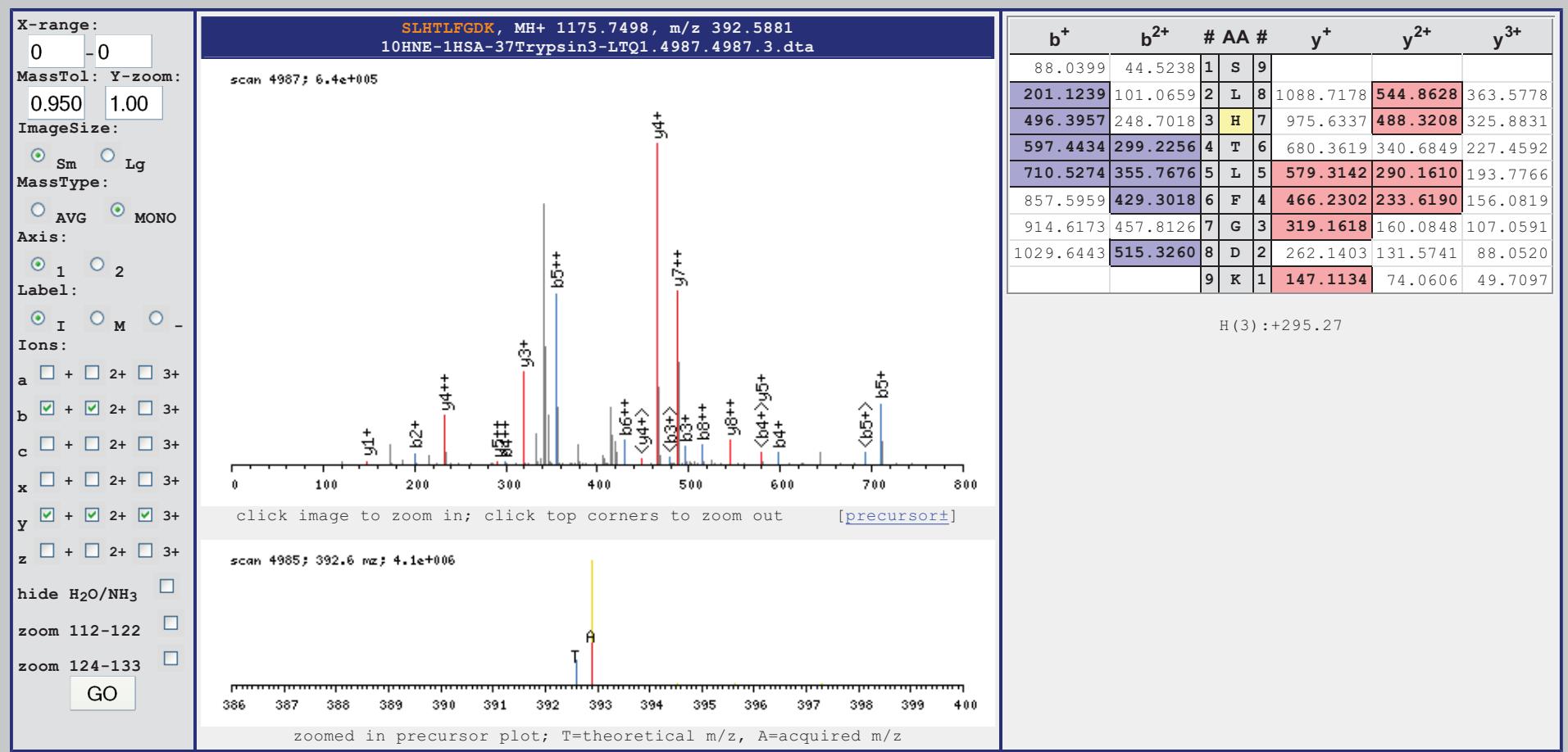
Iodoacetamide addition to cysteine is highlighted in the following Trans-Proteomic Pipeline screenshots, but not in this table (all cysteines were assumed to have been alkylated with iodoacetamide unless HNE addition occurred). C# indicates HNE Michael addition at Cys followed by reduction; H@ indicates HNE Michael addition at His followed by reduction; K^ indicates HNE Michael addition at Lys followed by reduction; K~ indicates Schiff base formation with HNE at Lys followed by reduction; M\* indicates oxidation at Met.



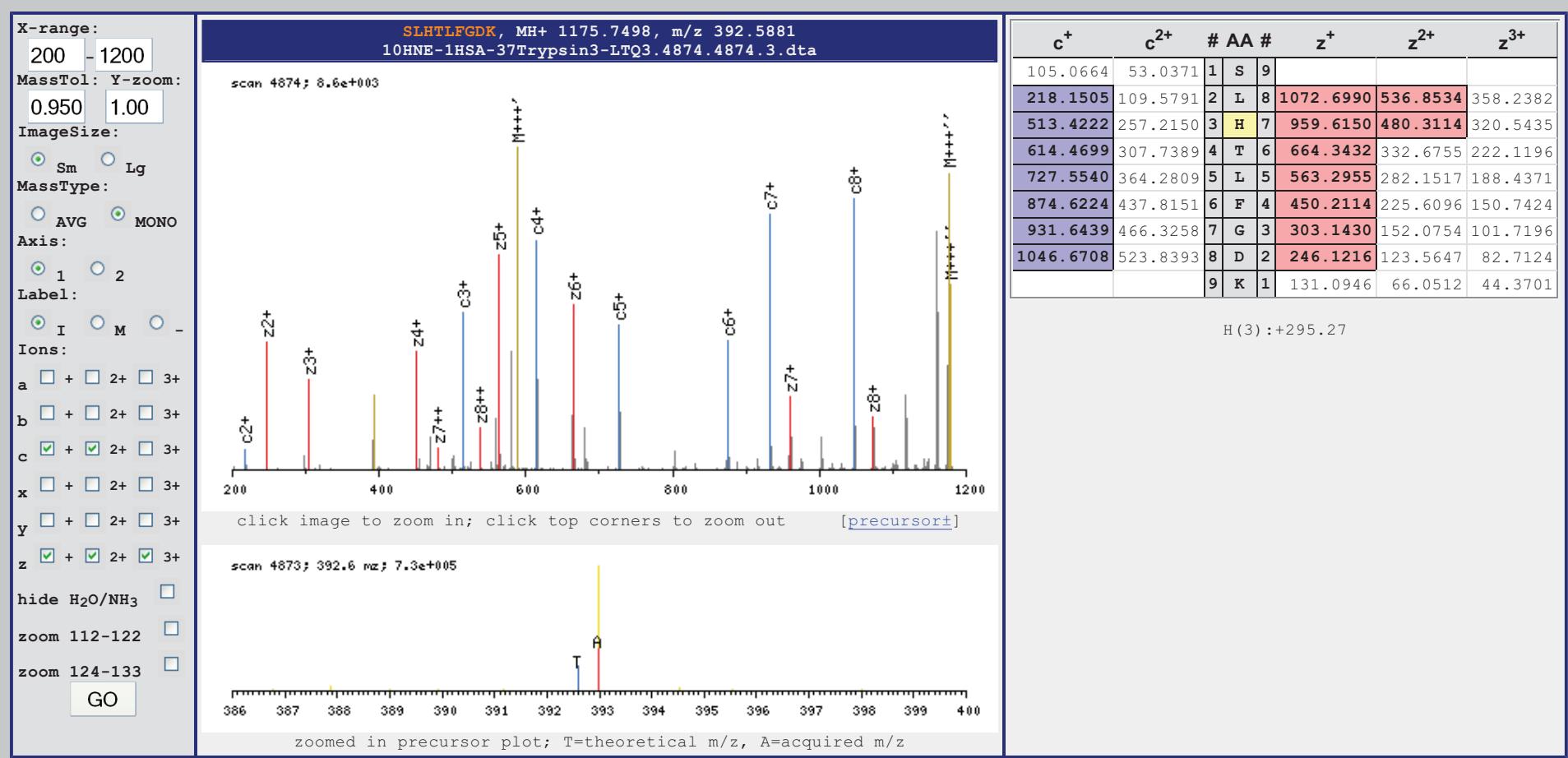
COMET Spectrum View by J.Eng (c) ISB 2001  
 (TPP v4.4 VUVUZELA rev 1, Build 201010121551 (MinGW))



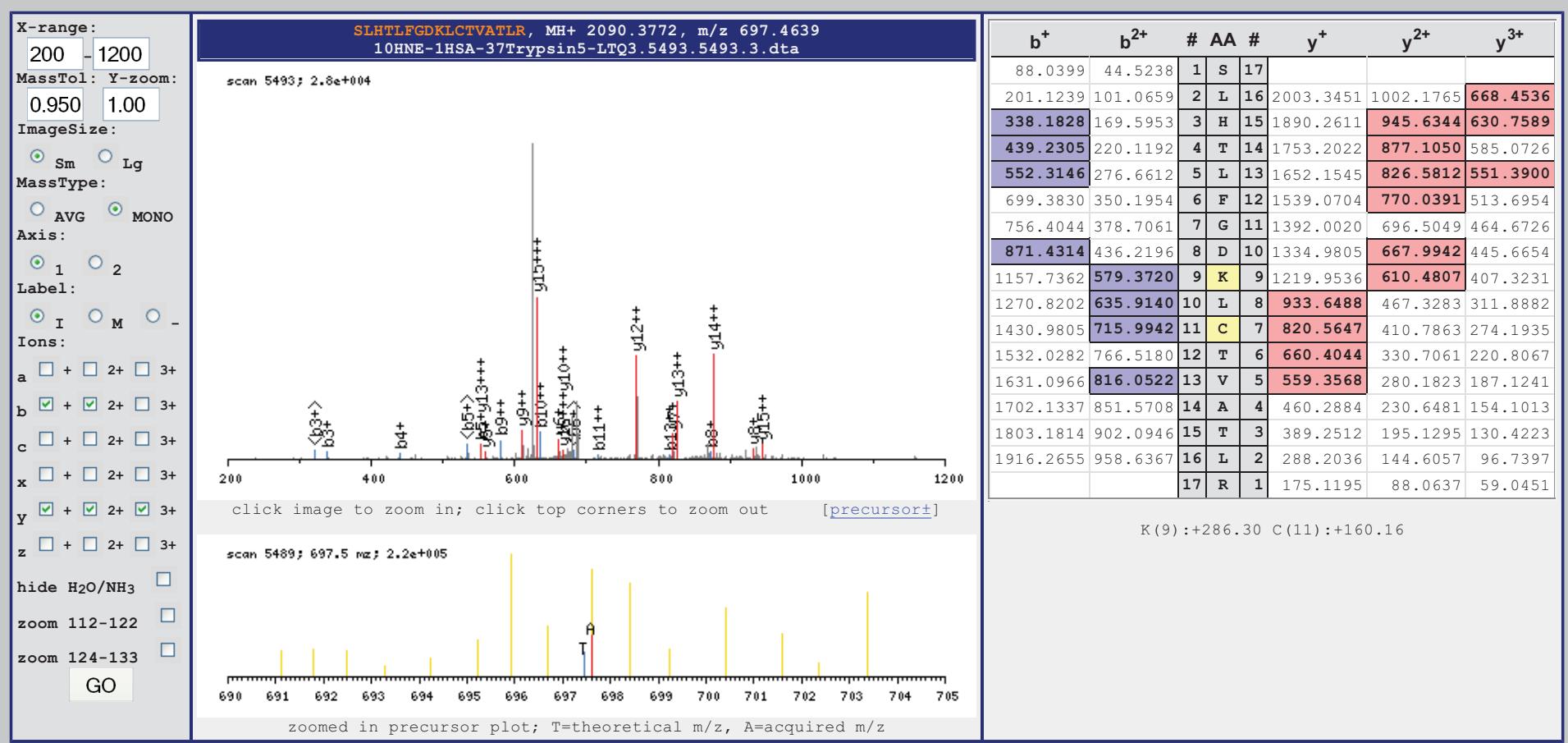
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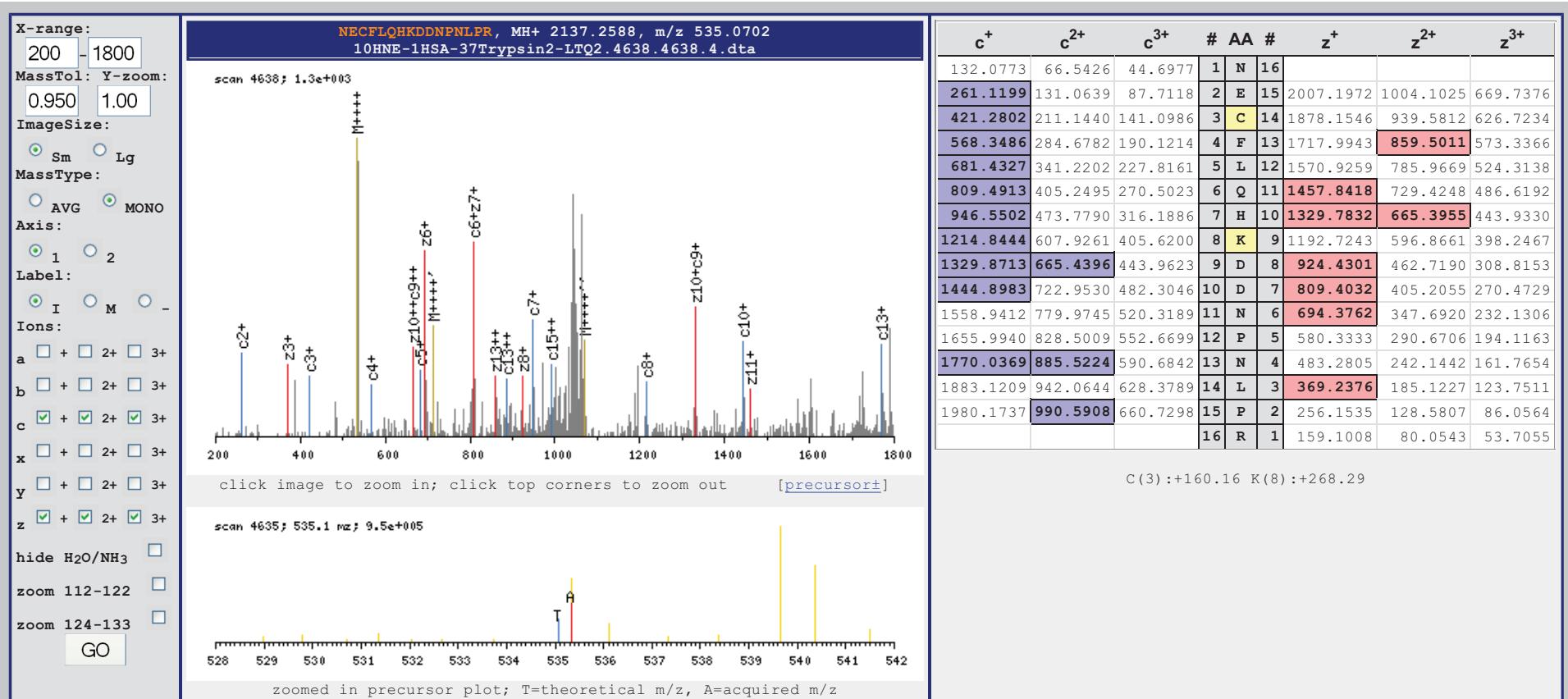
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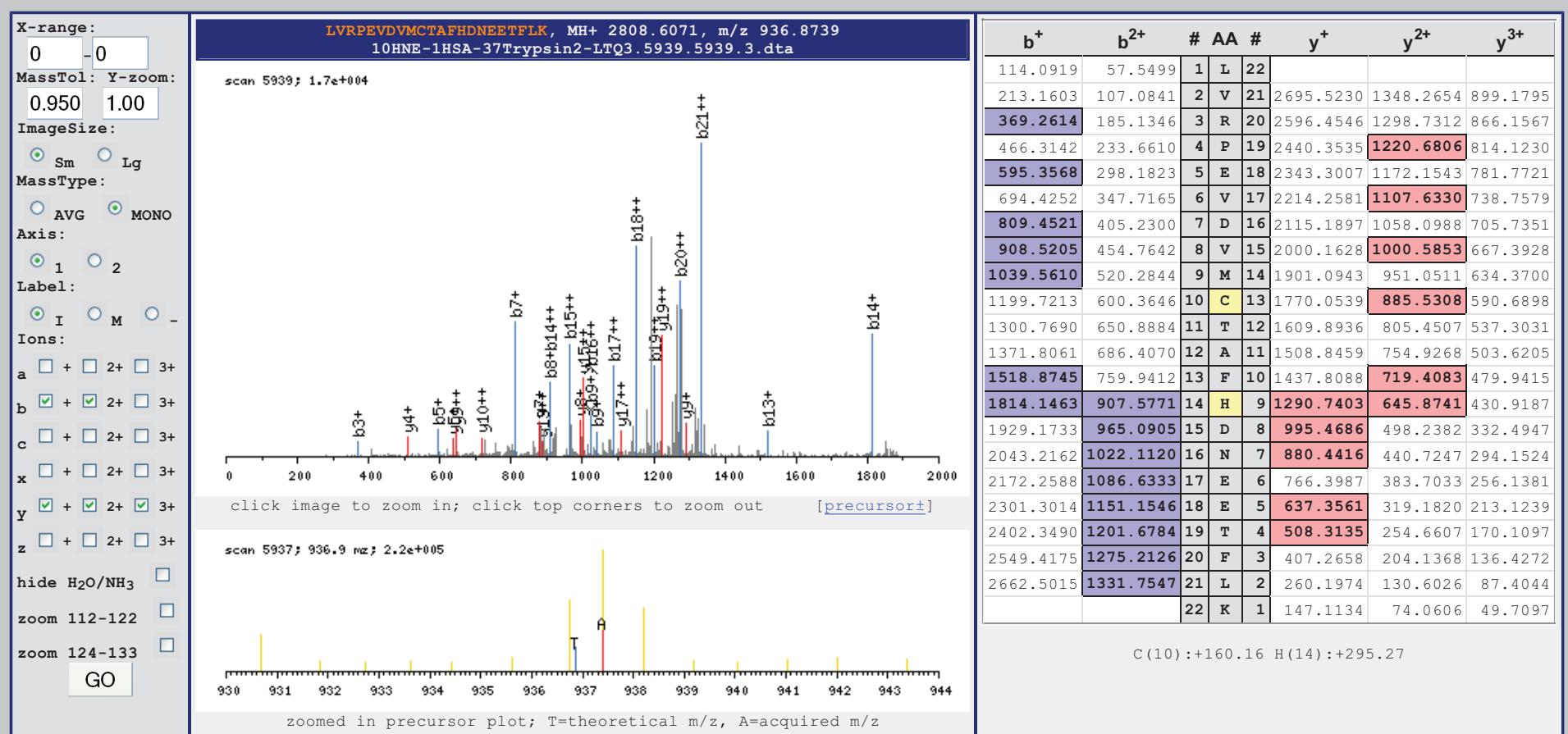
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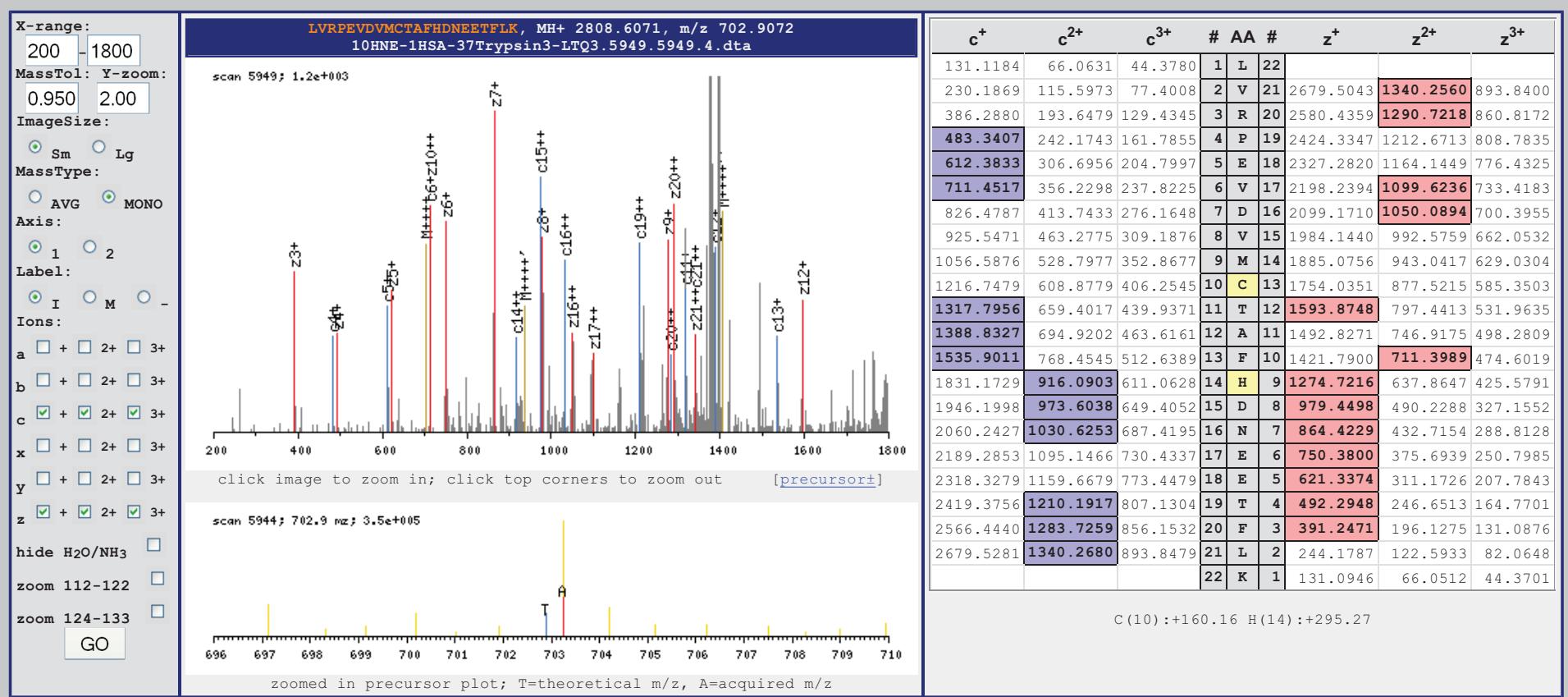
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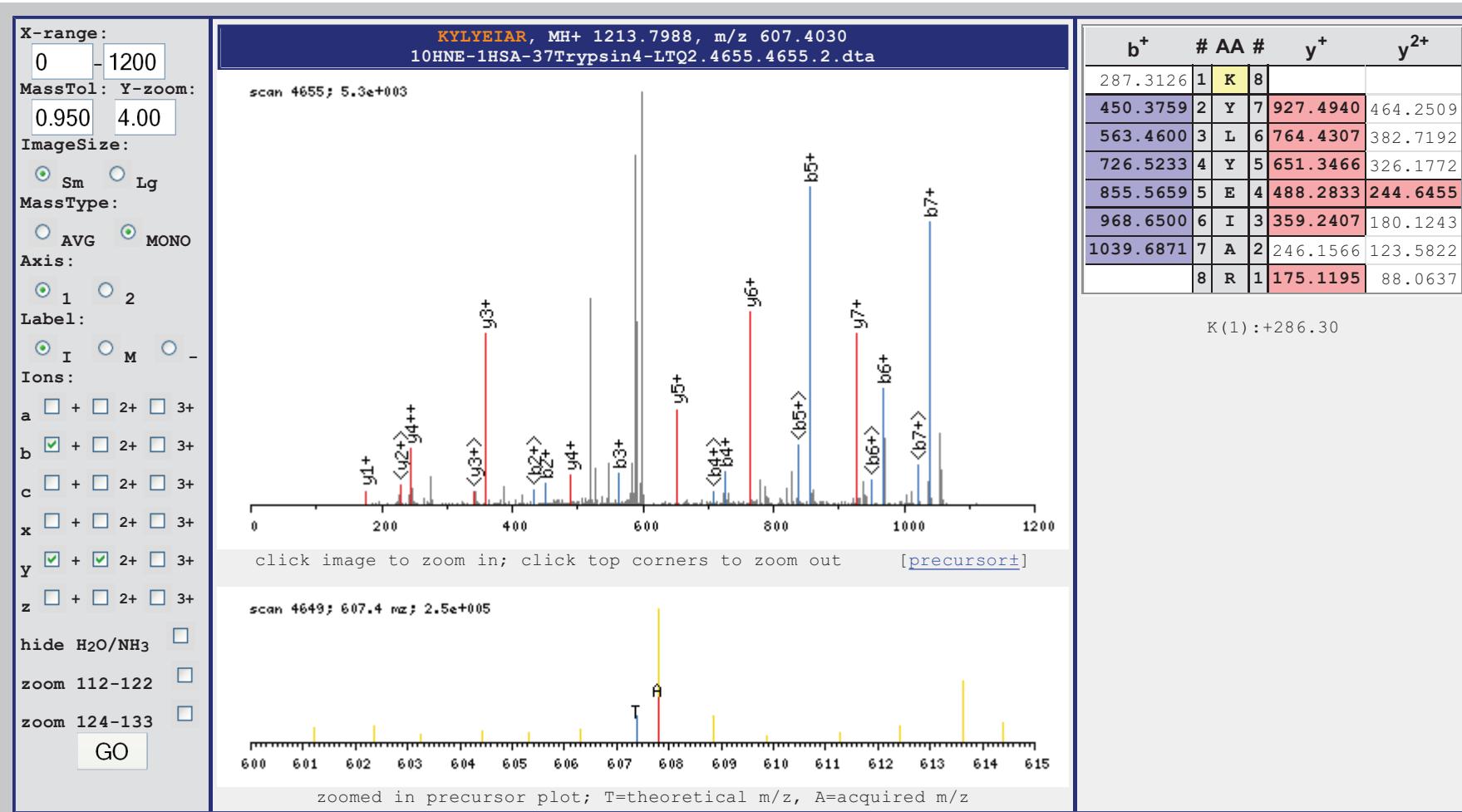
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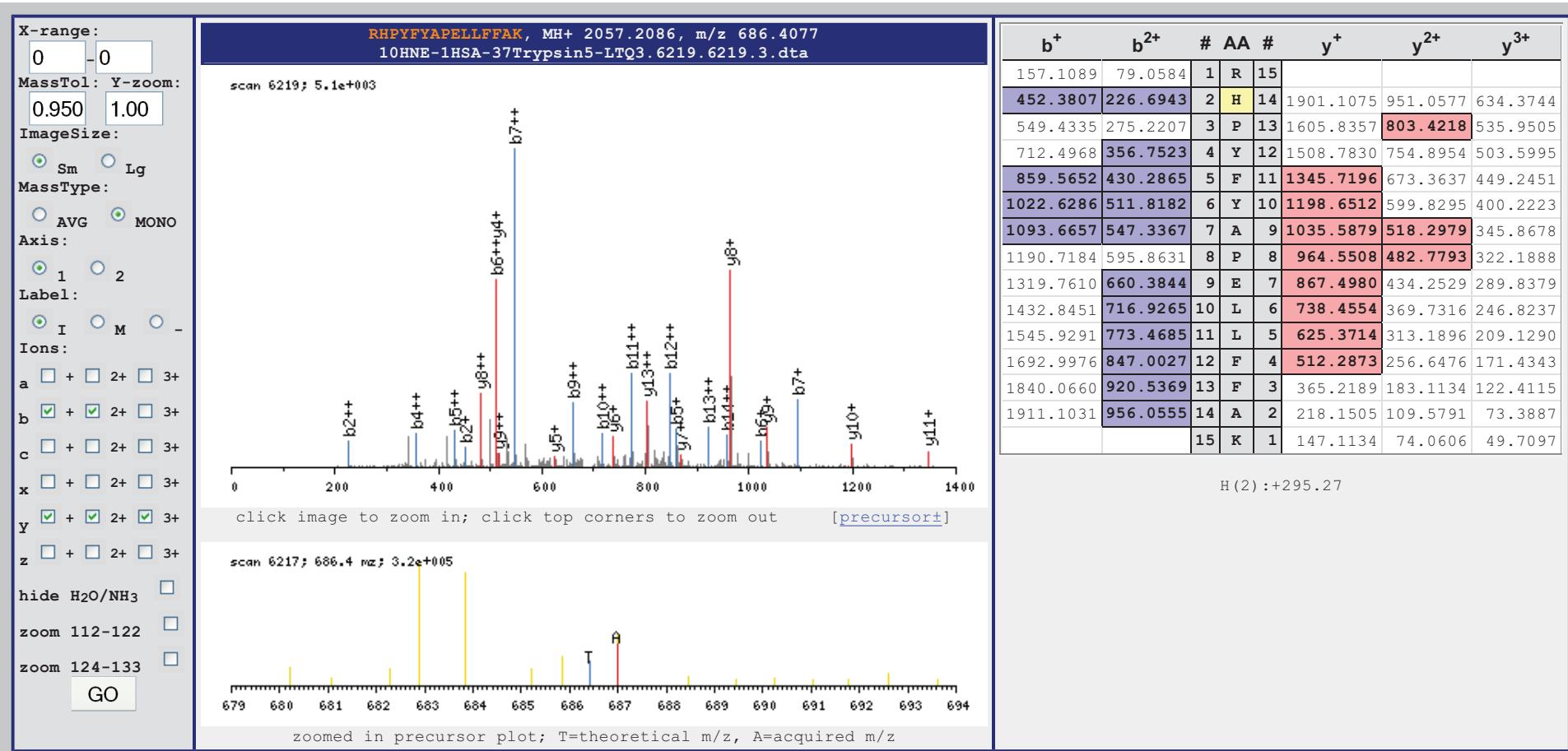
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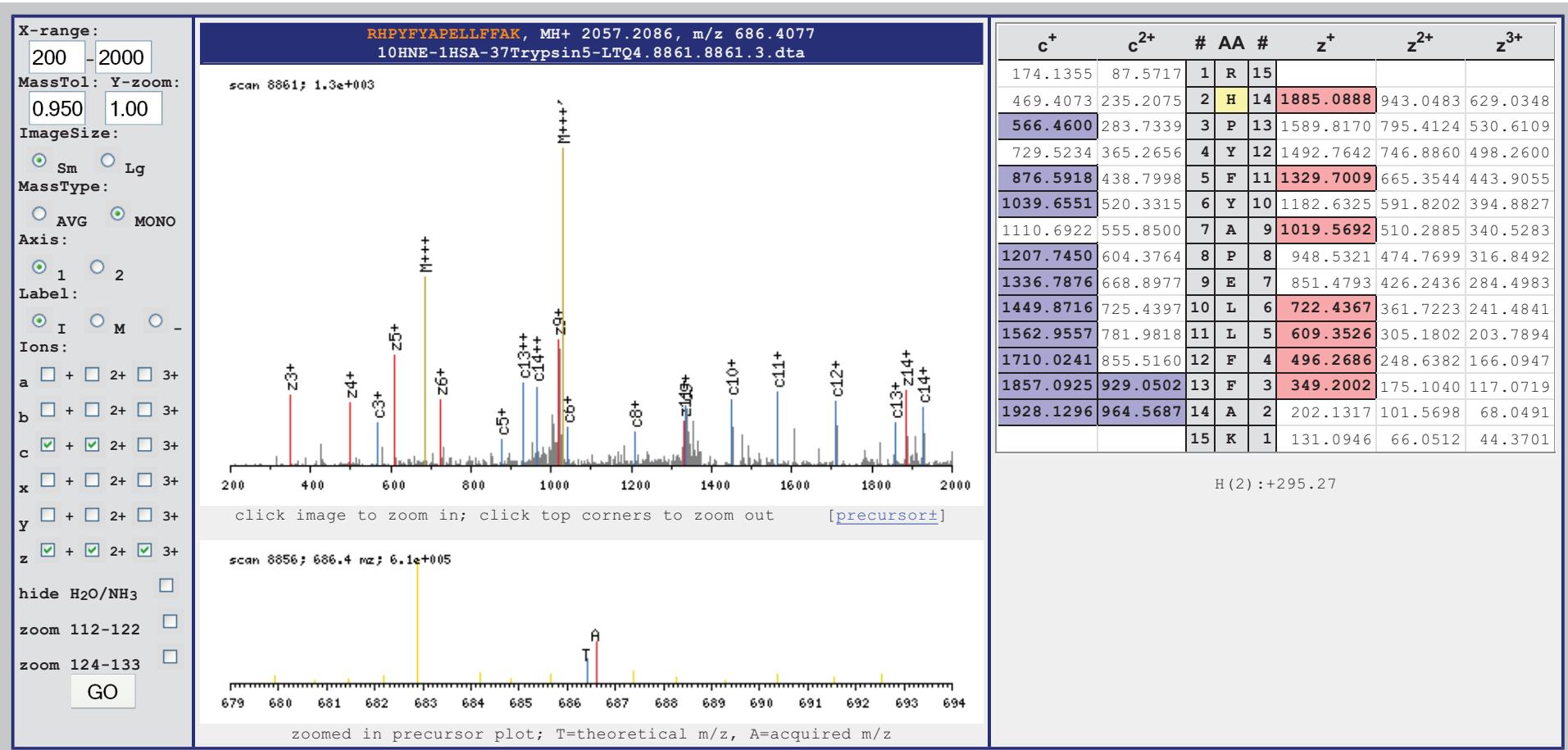
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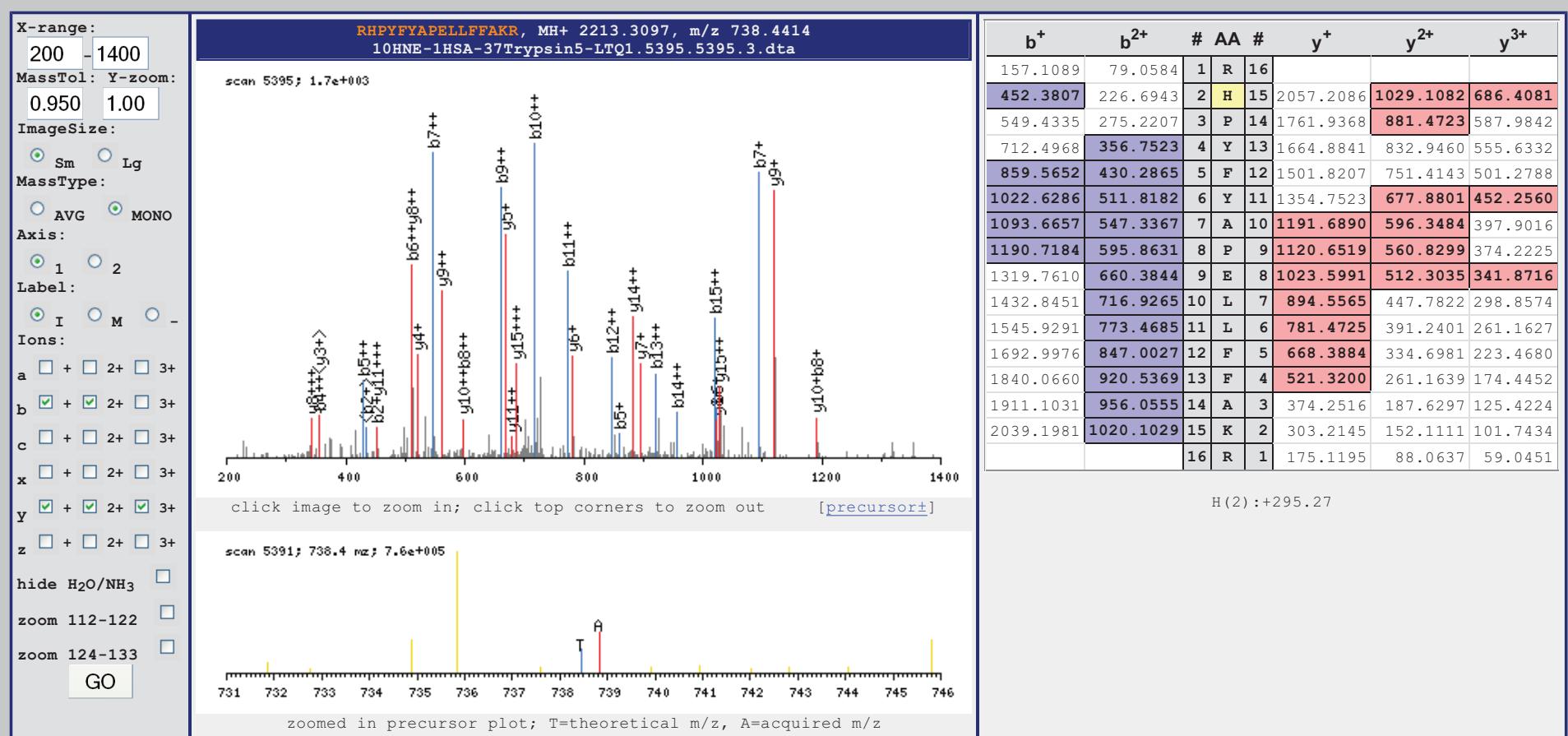
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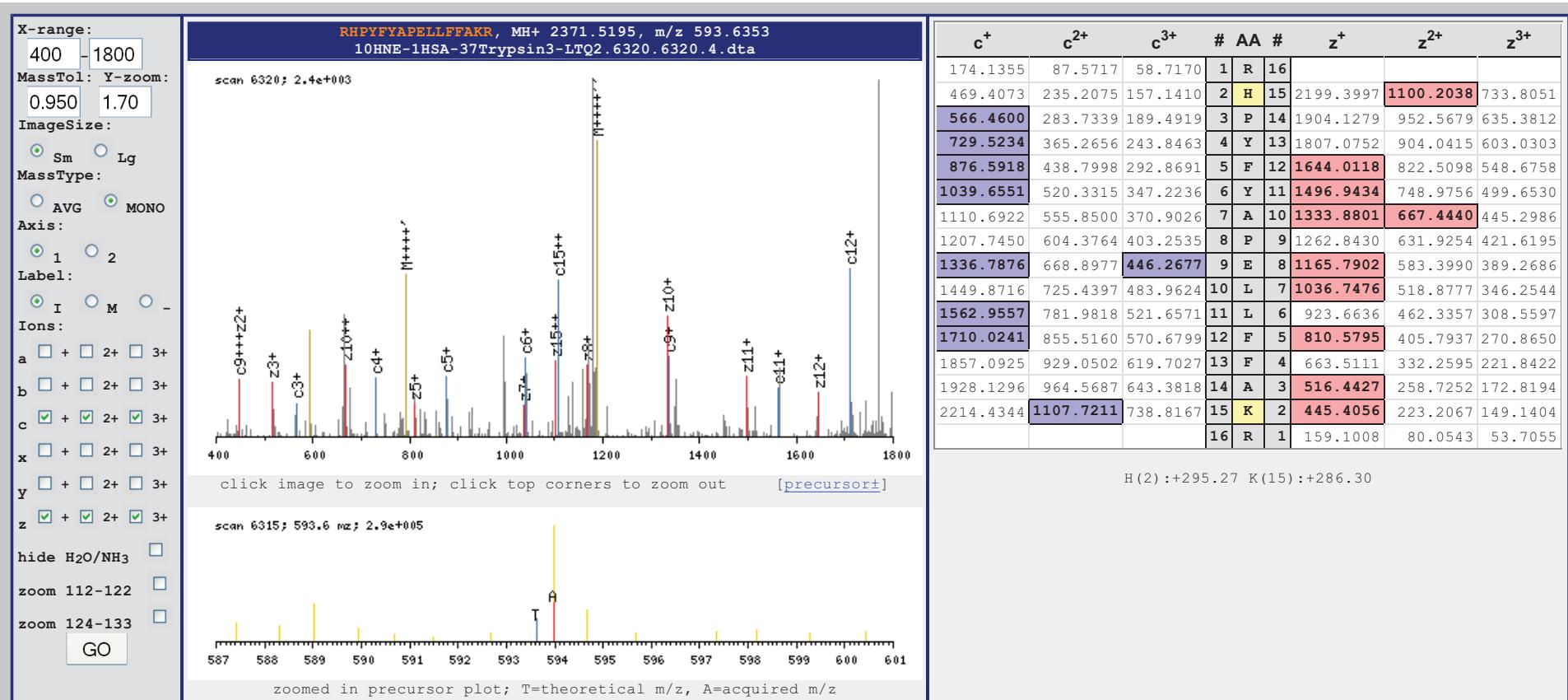
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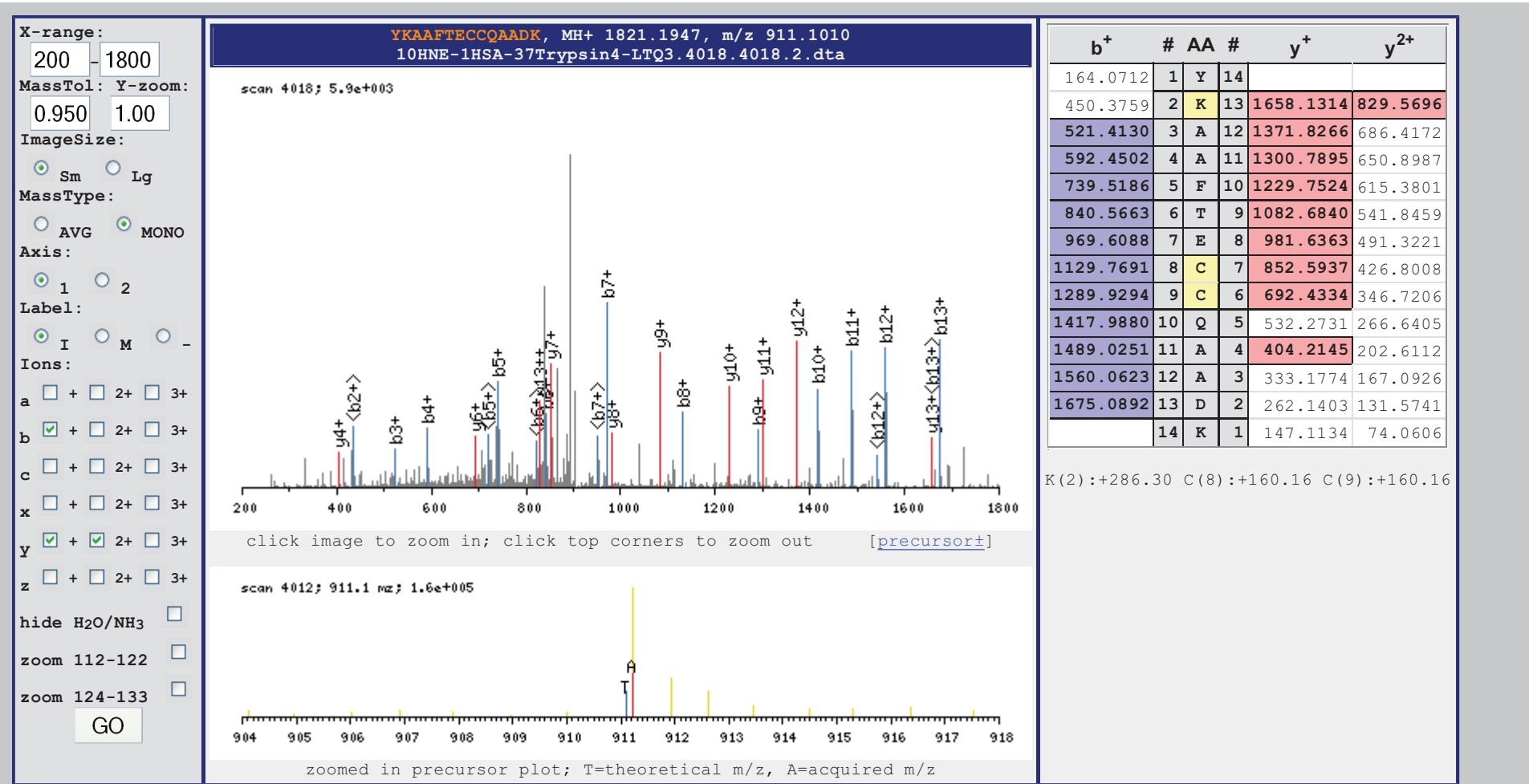
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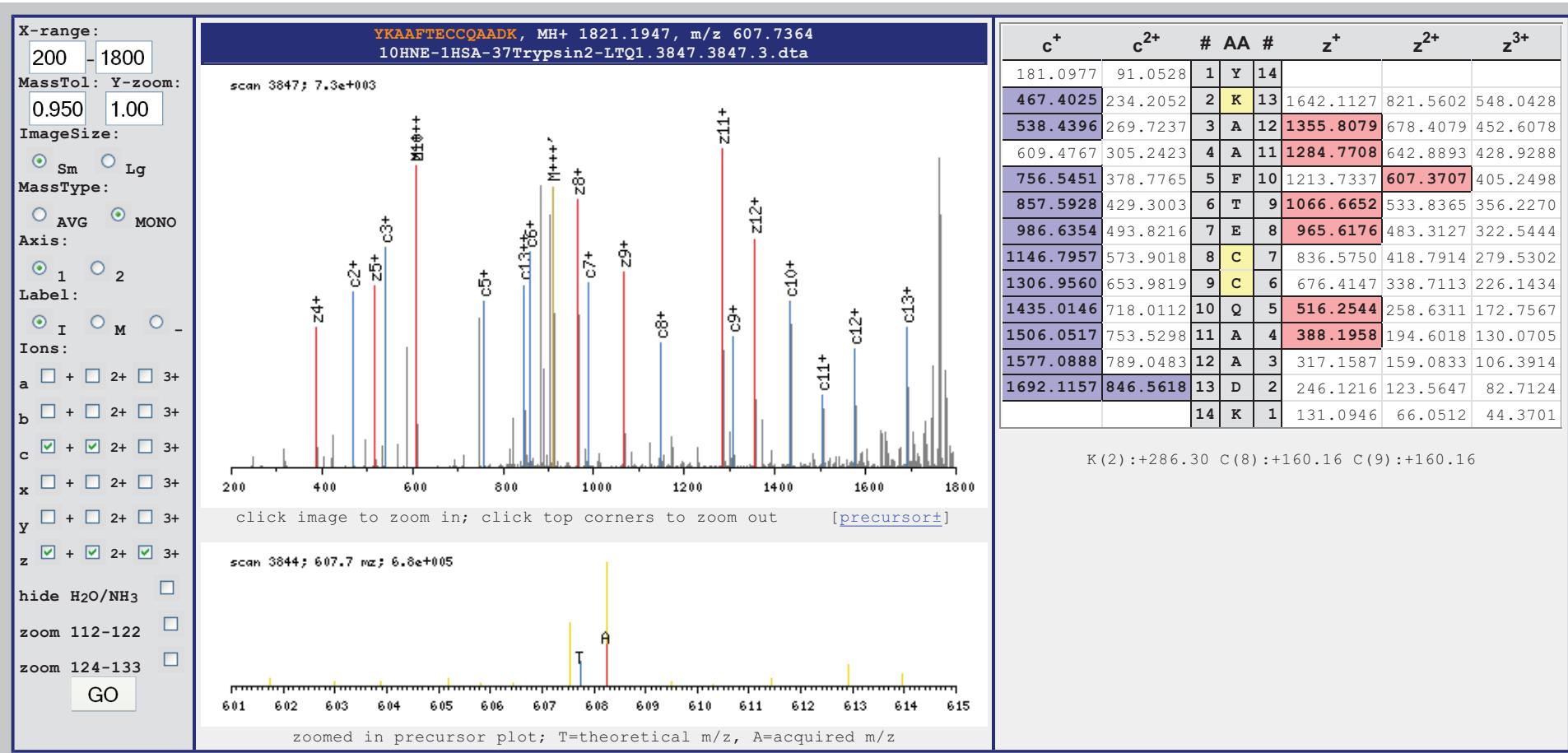
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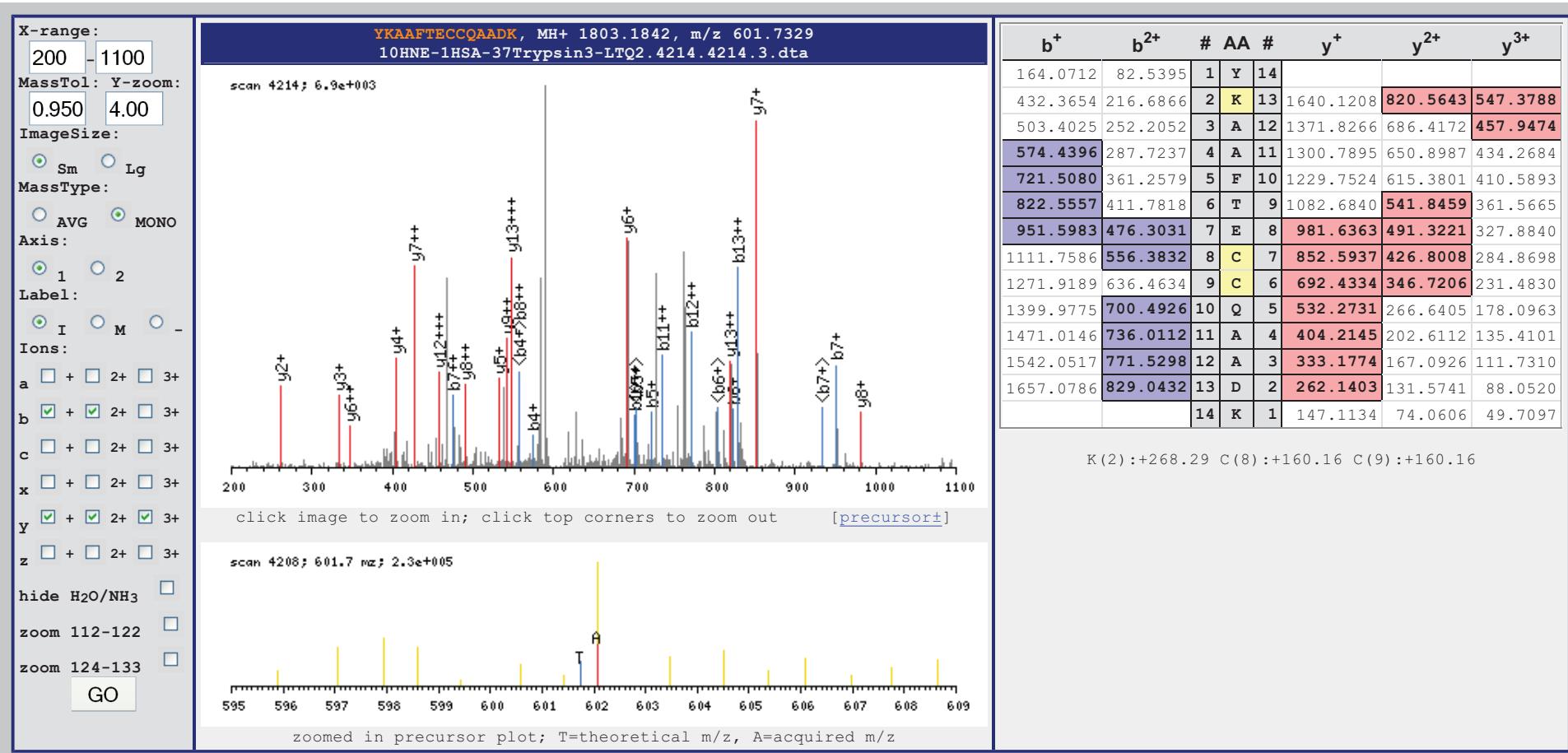
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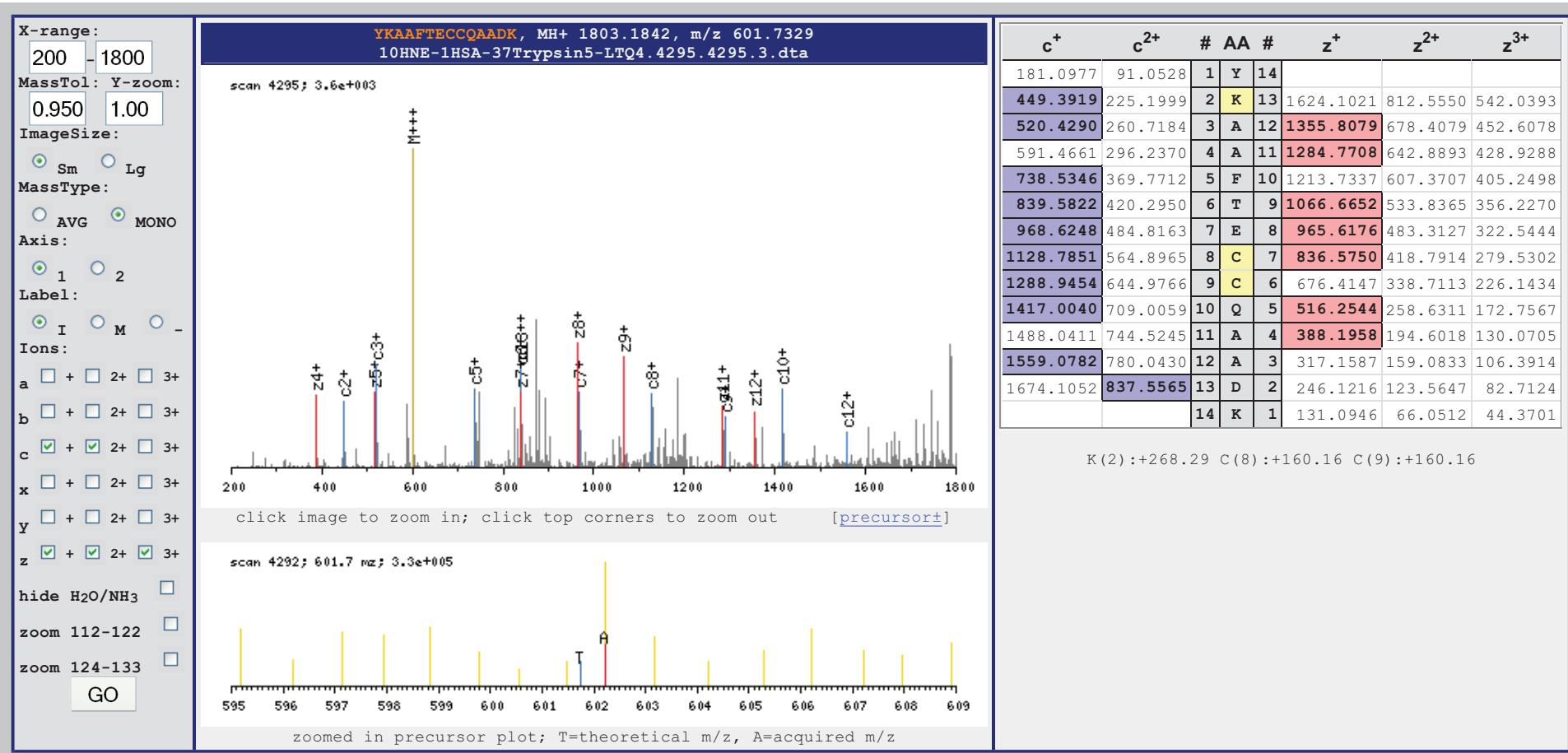
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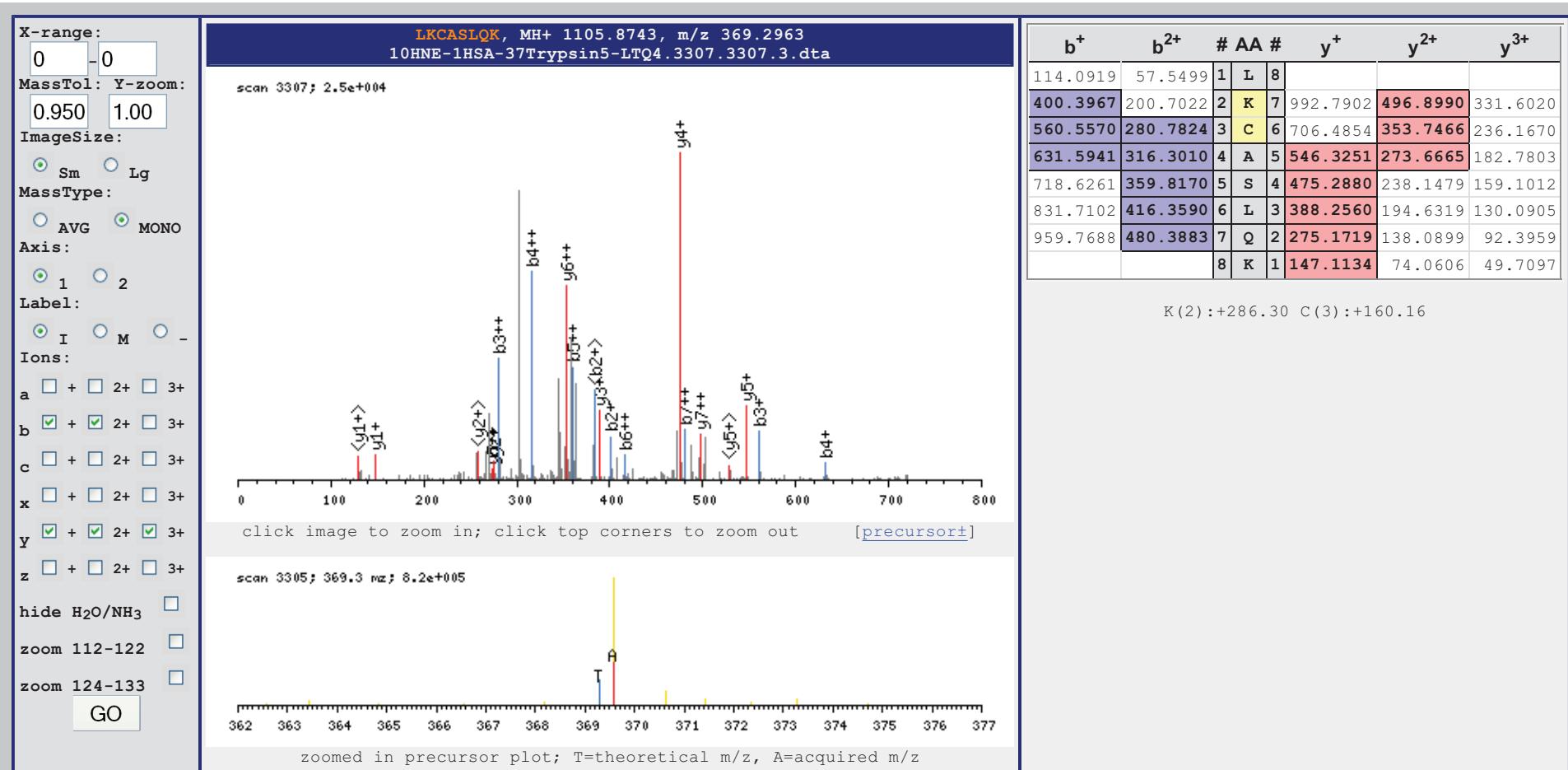
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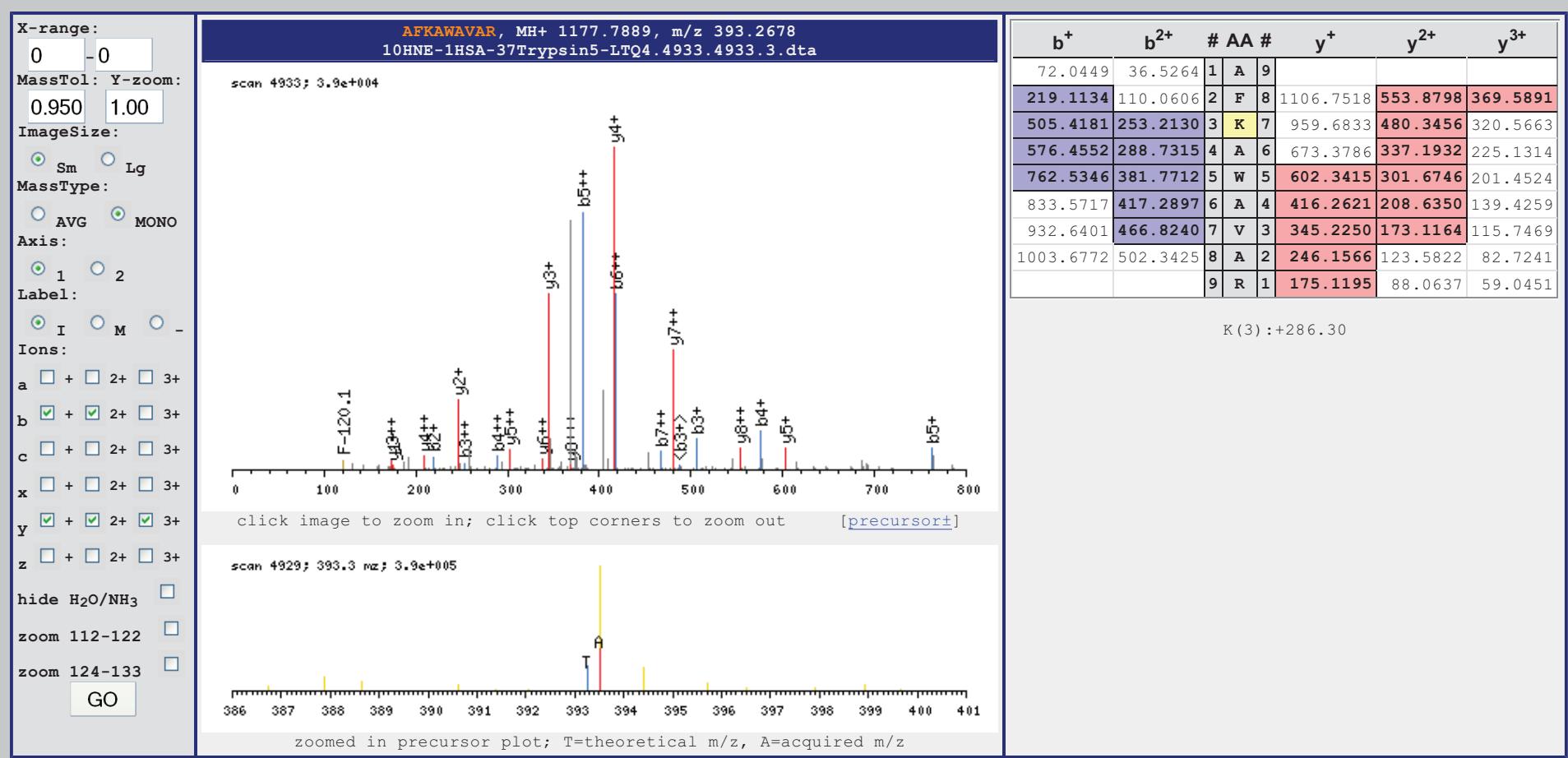
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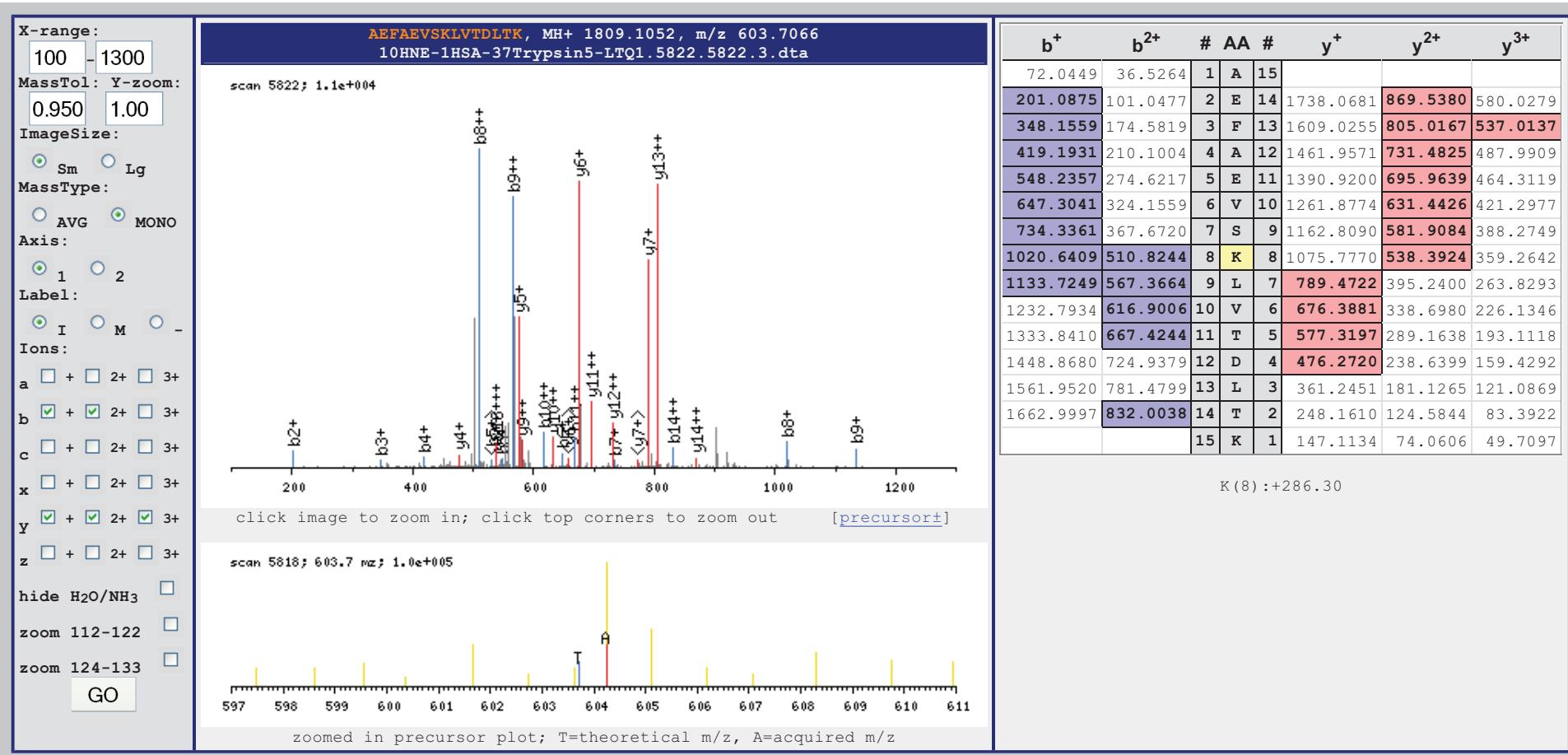
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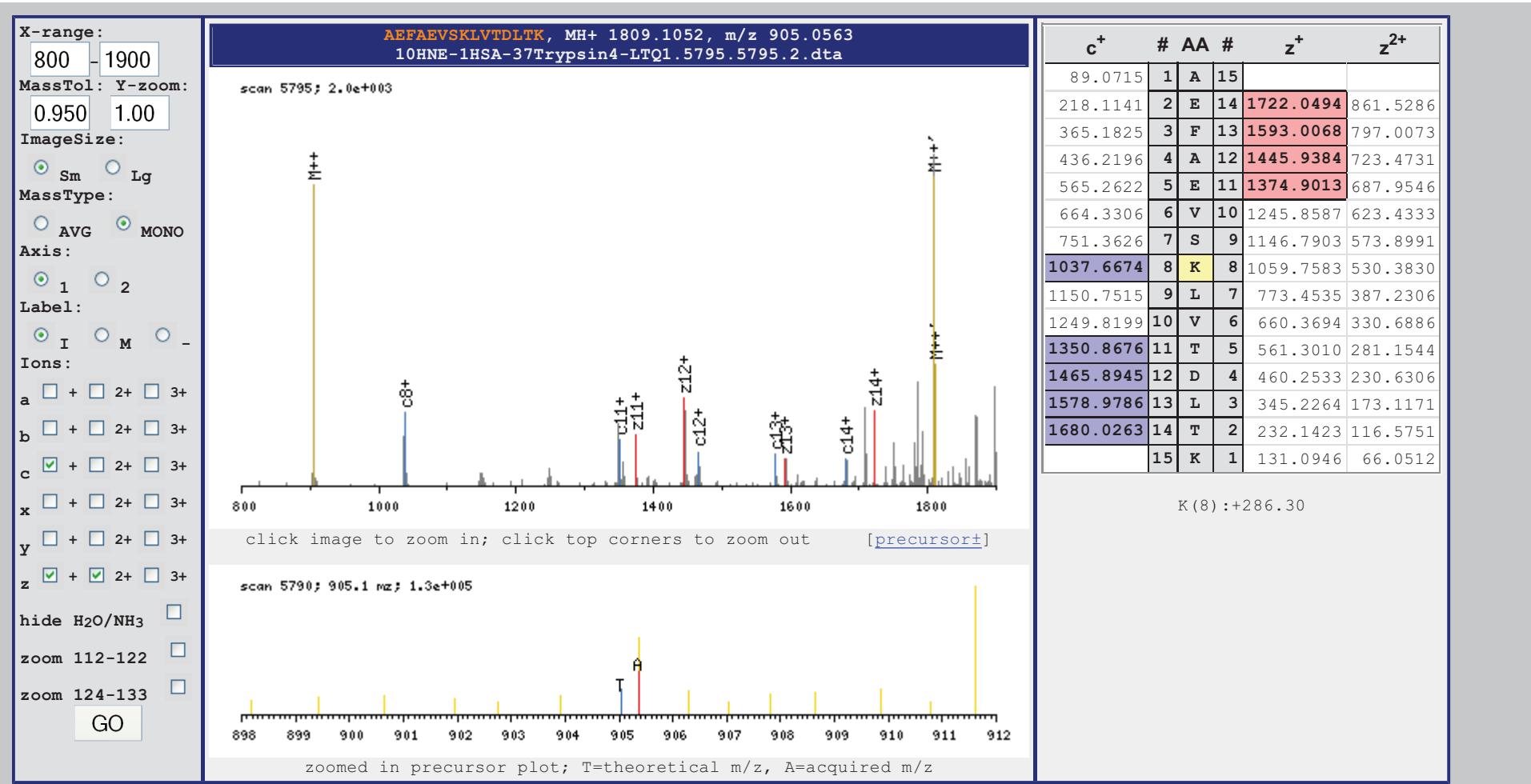
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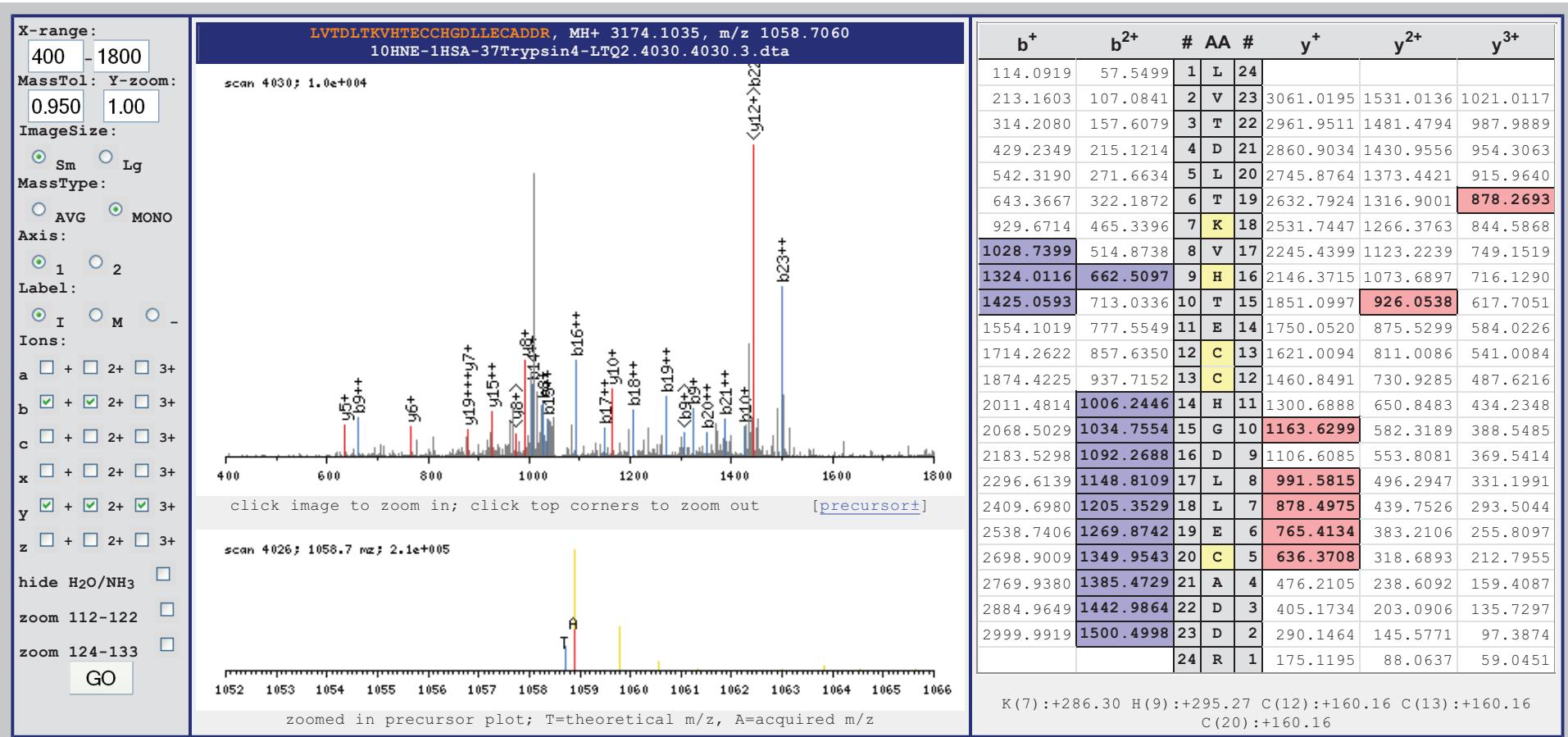
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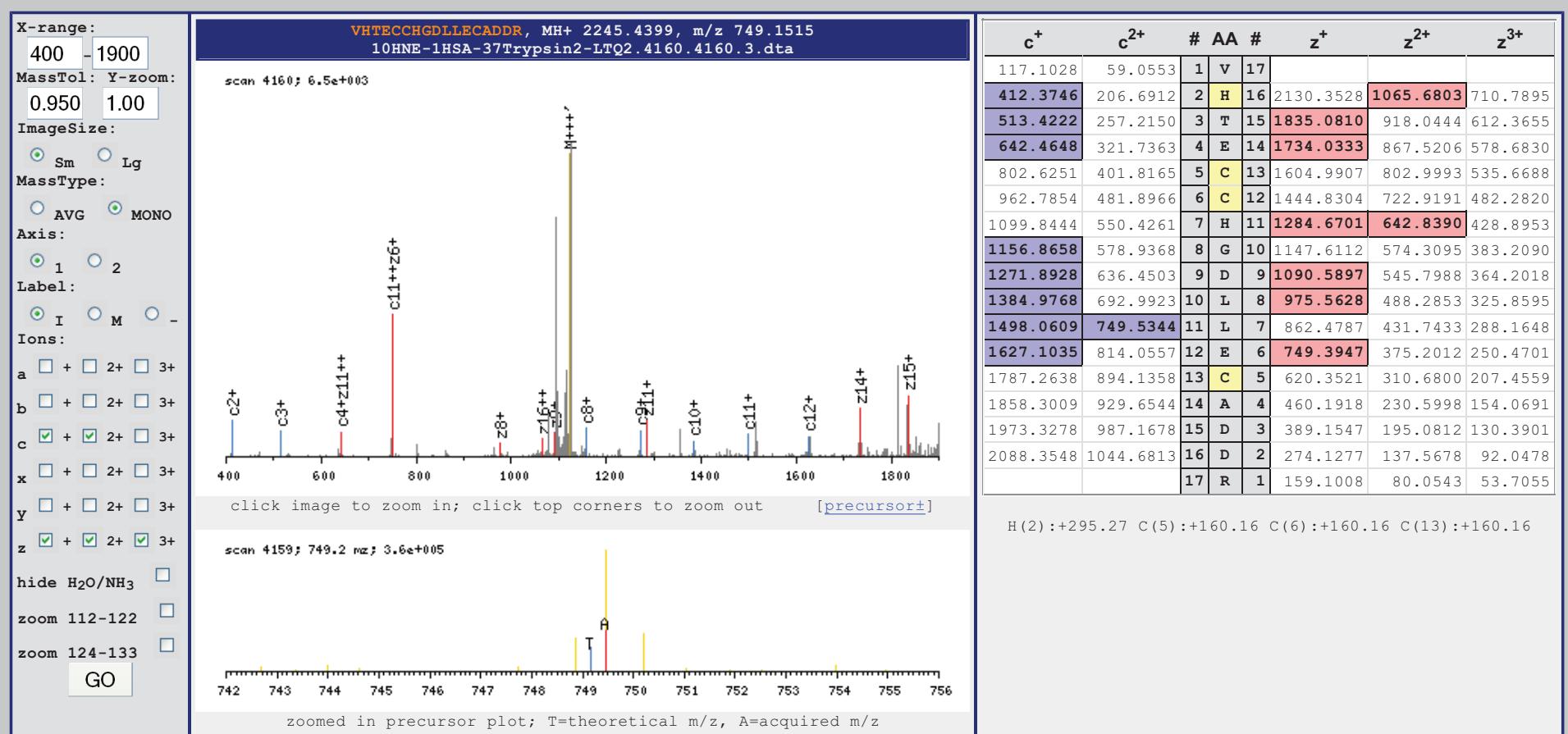
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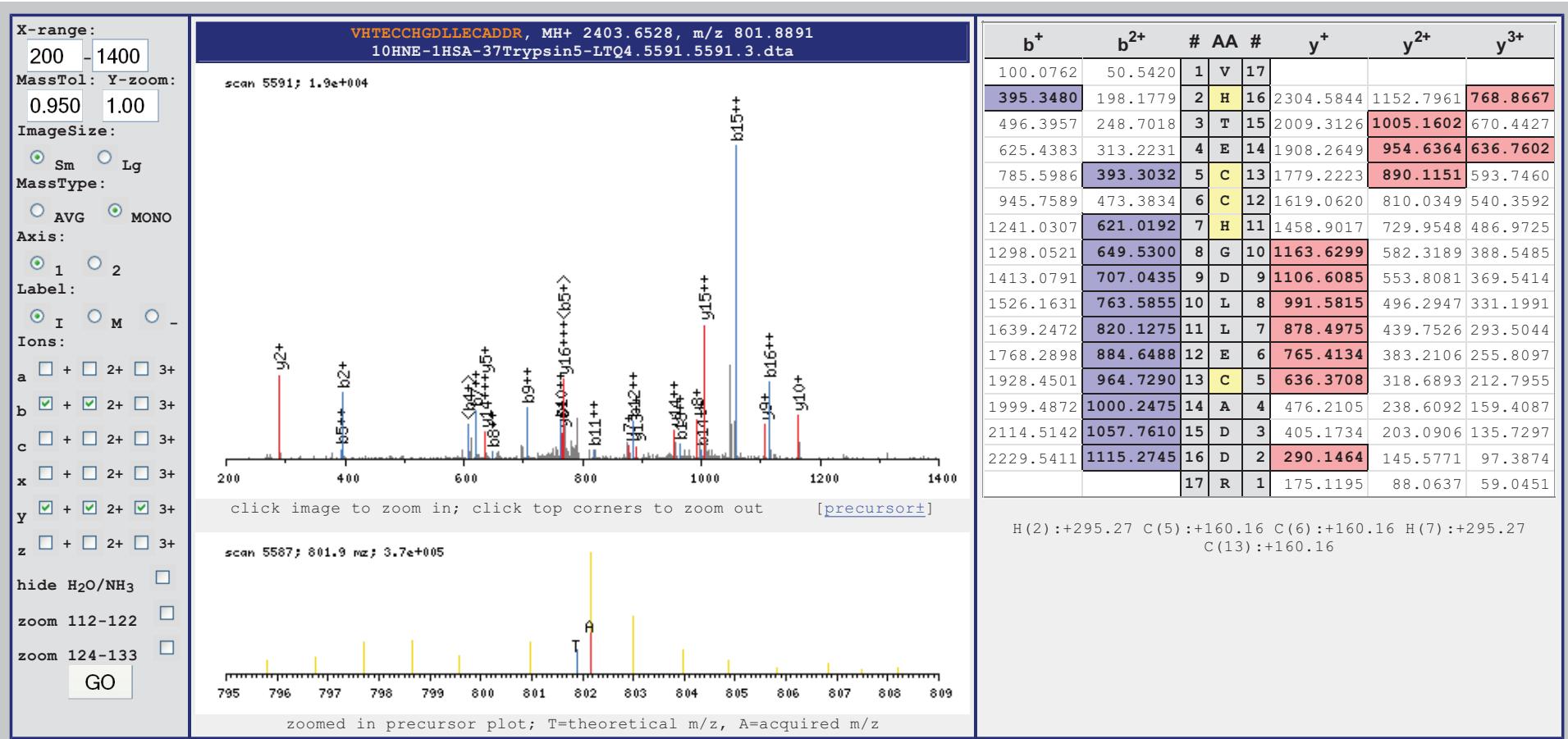
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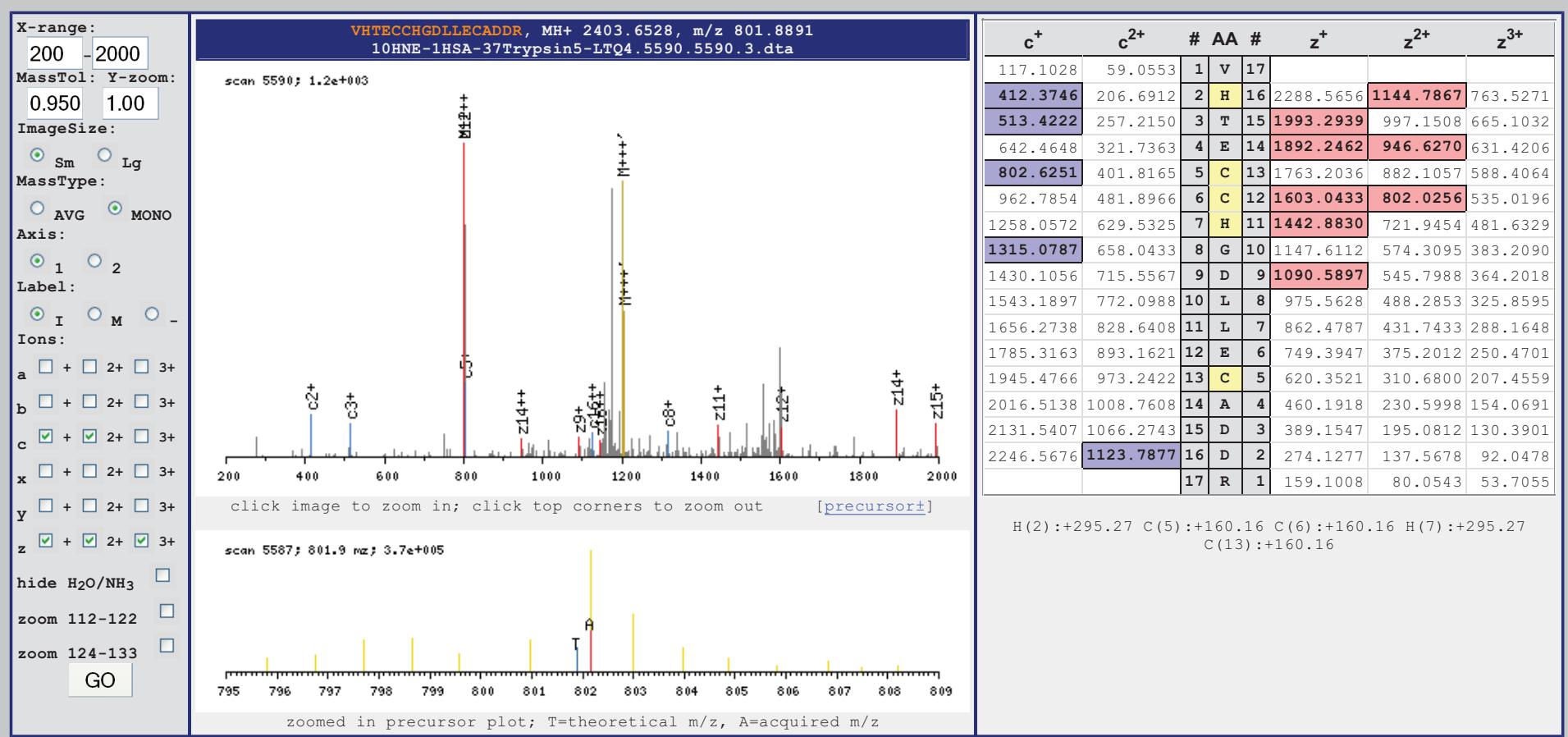
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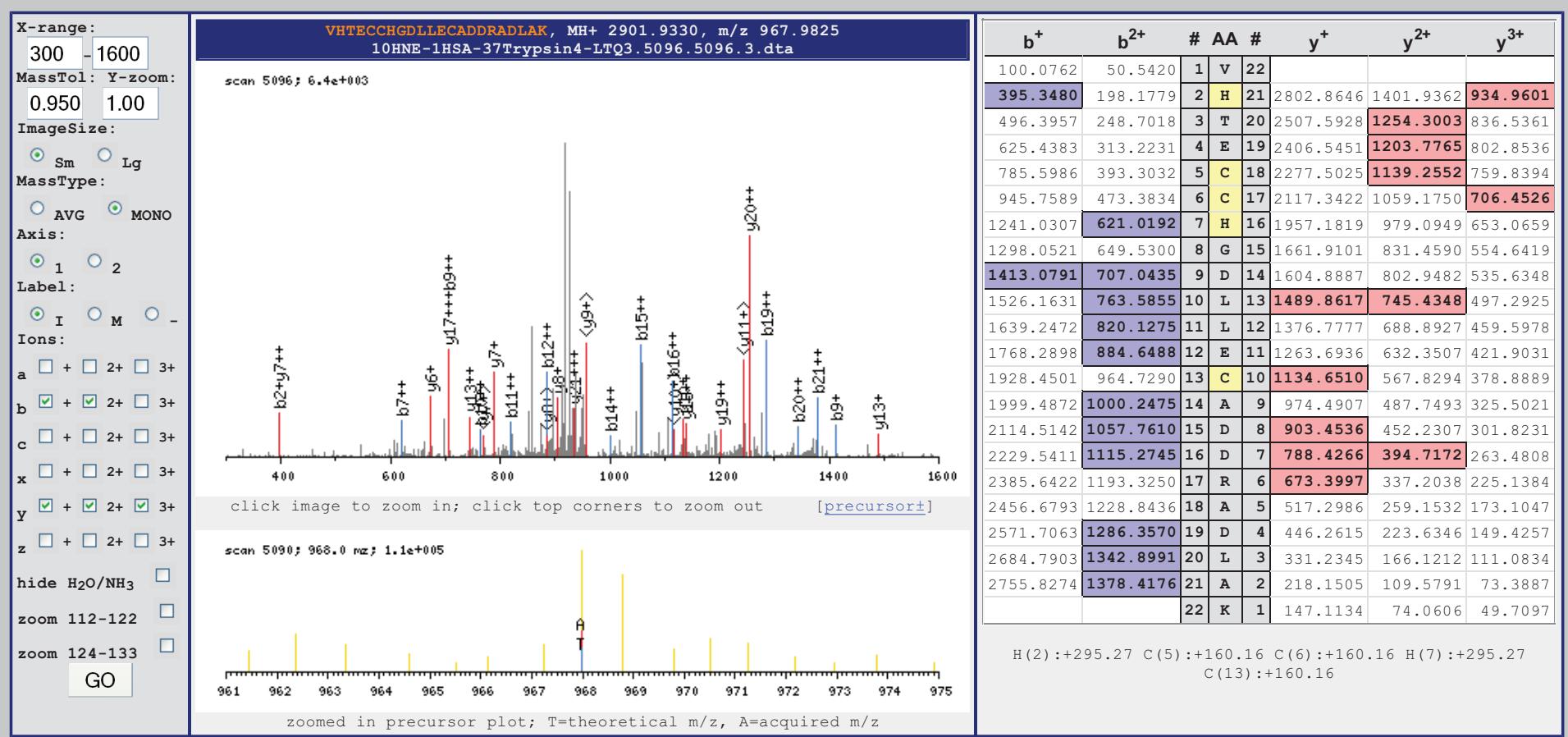
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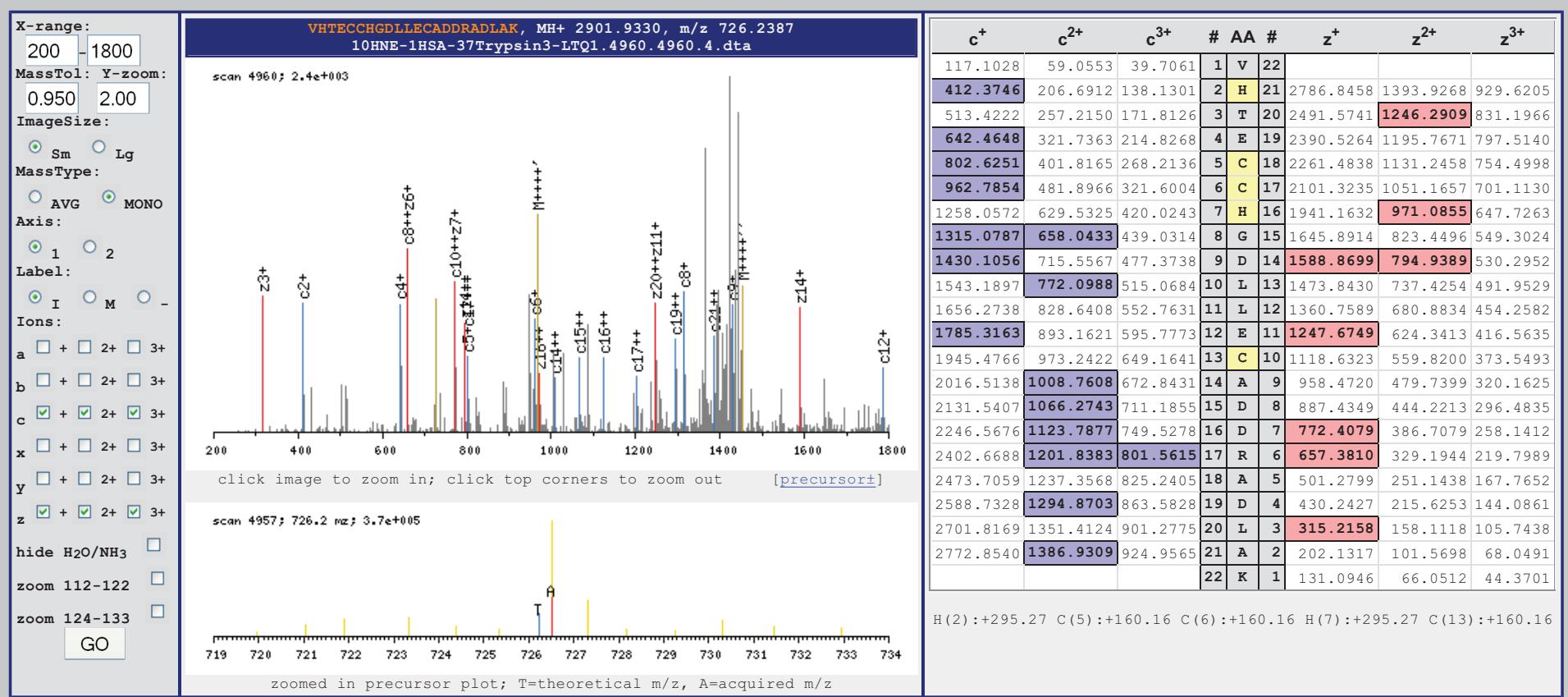
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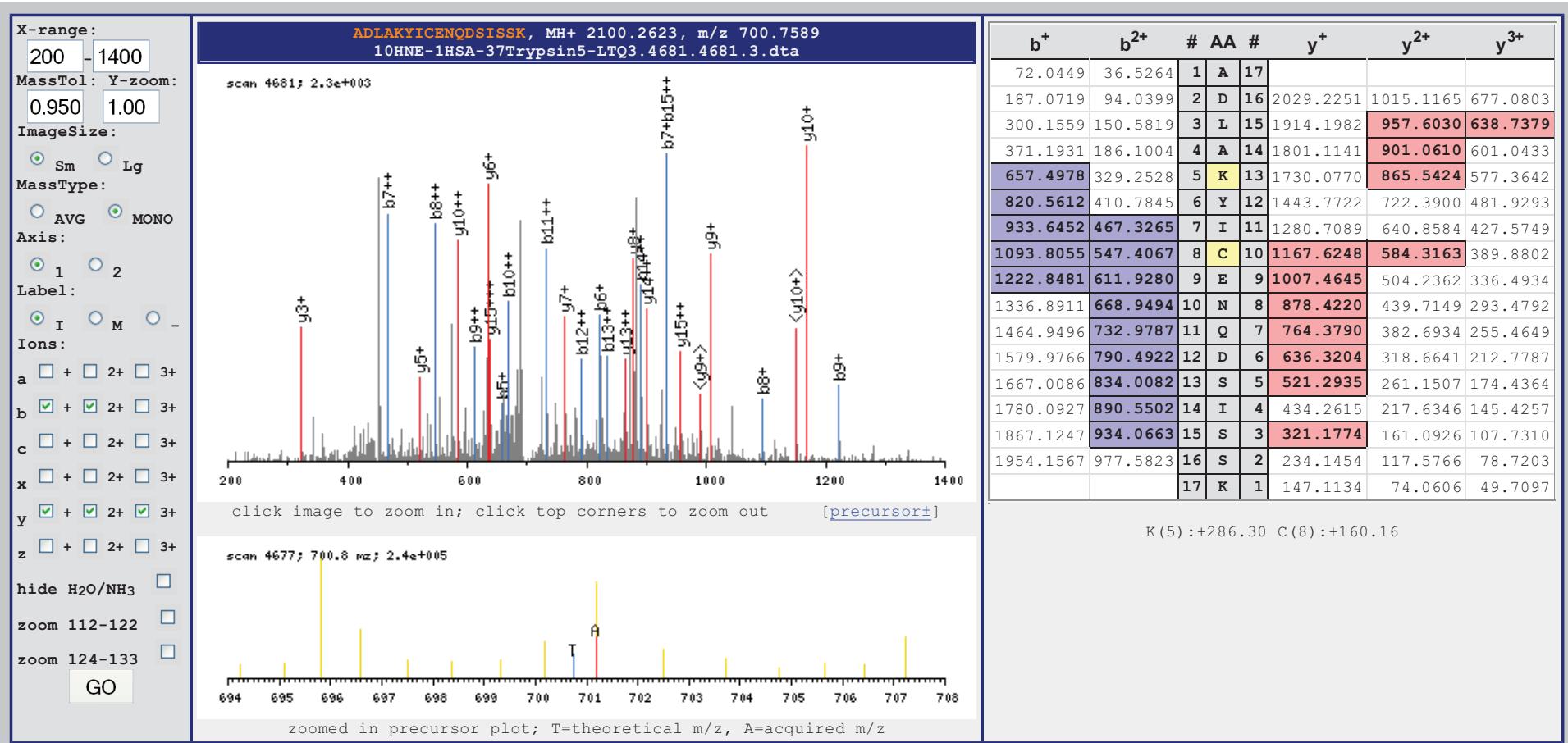
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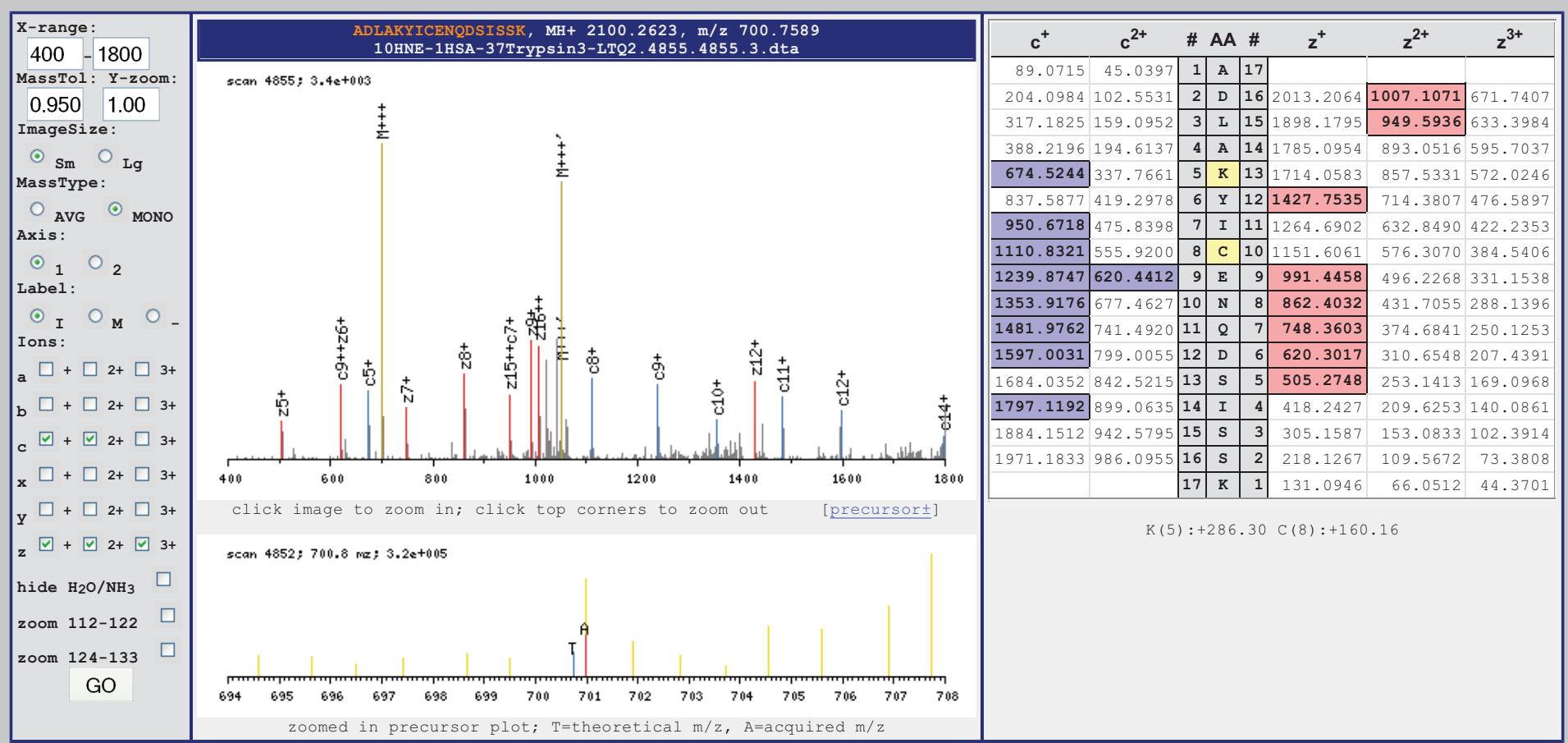
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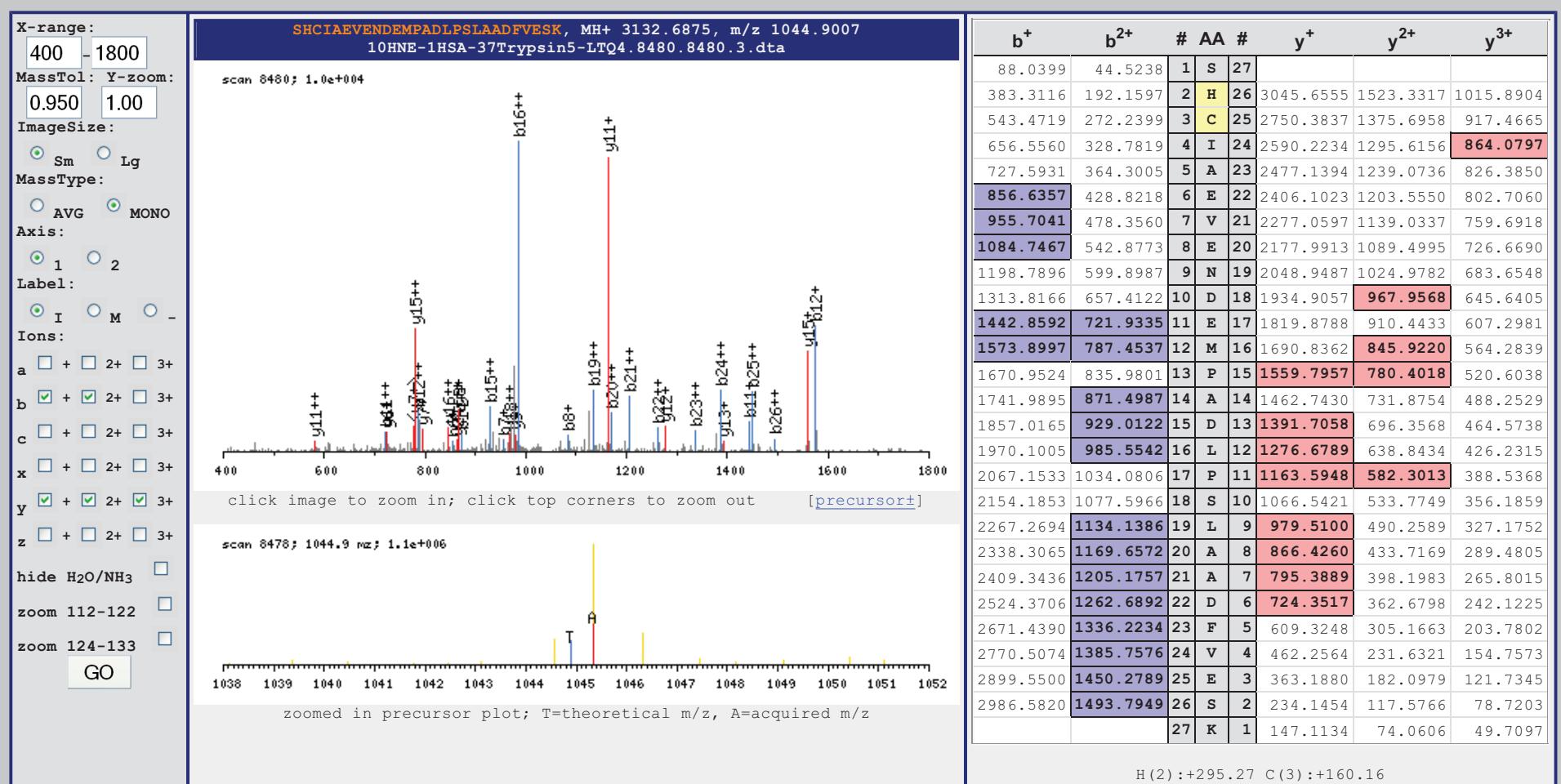
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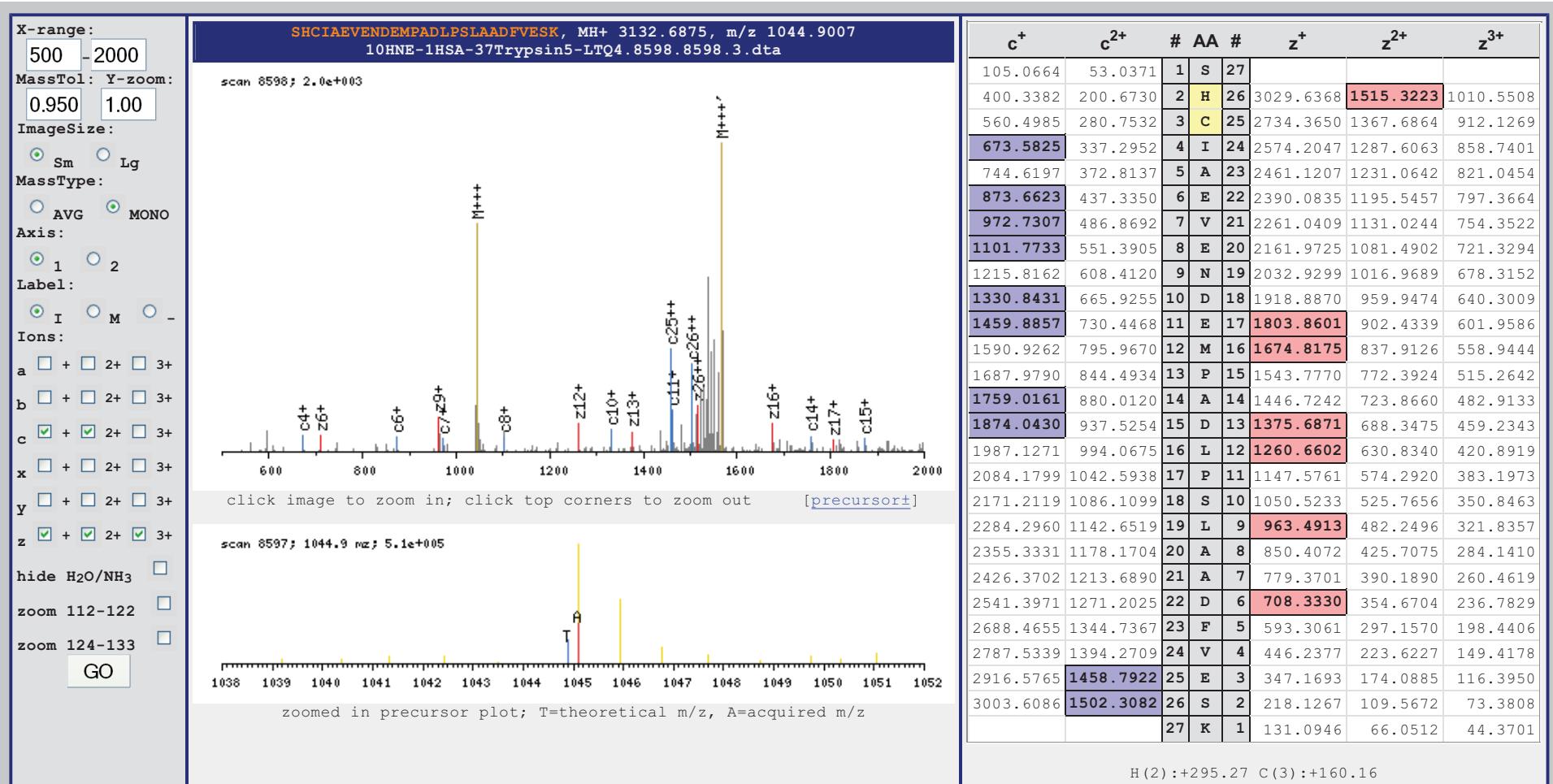
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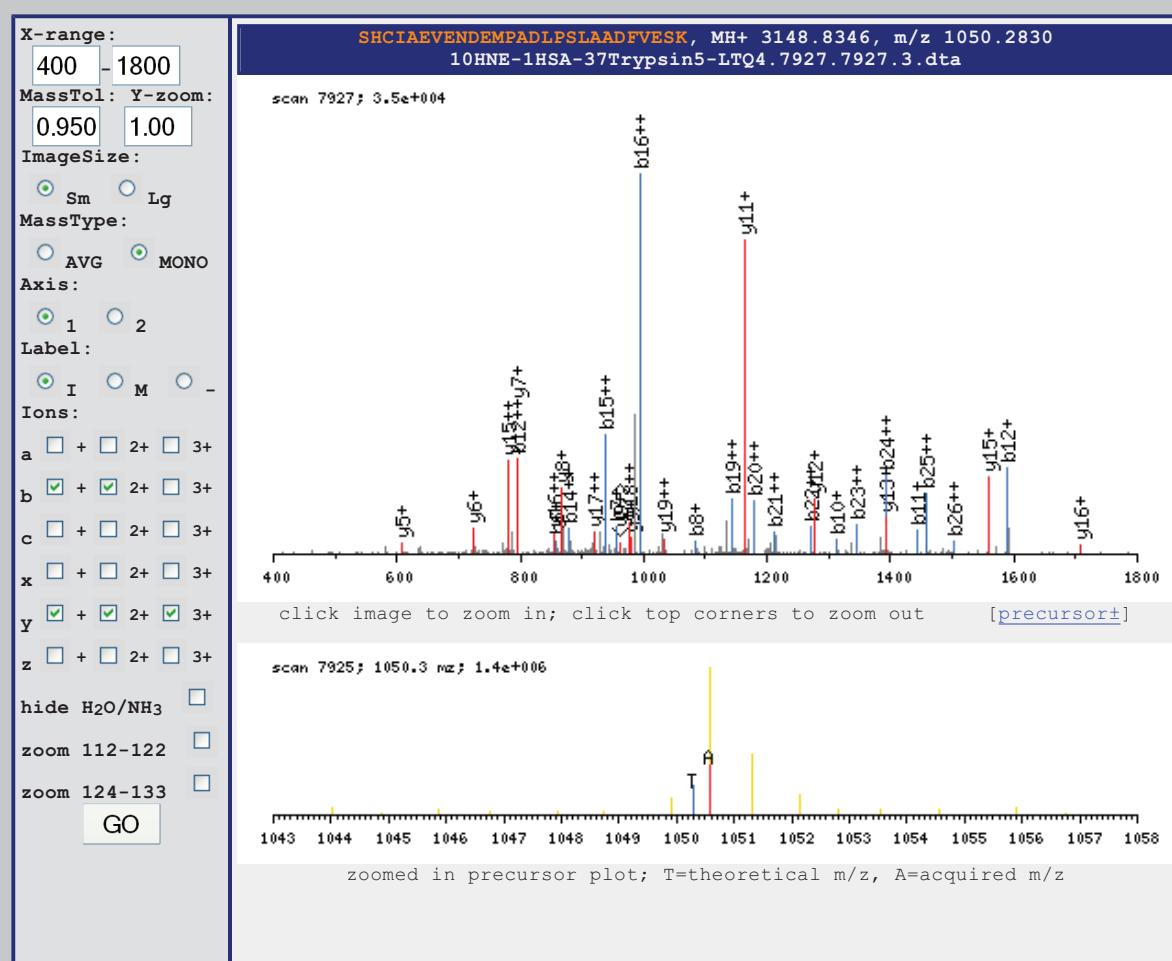
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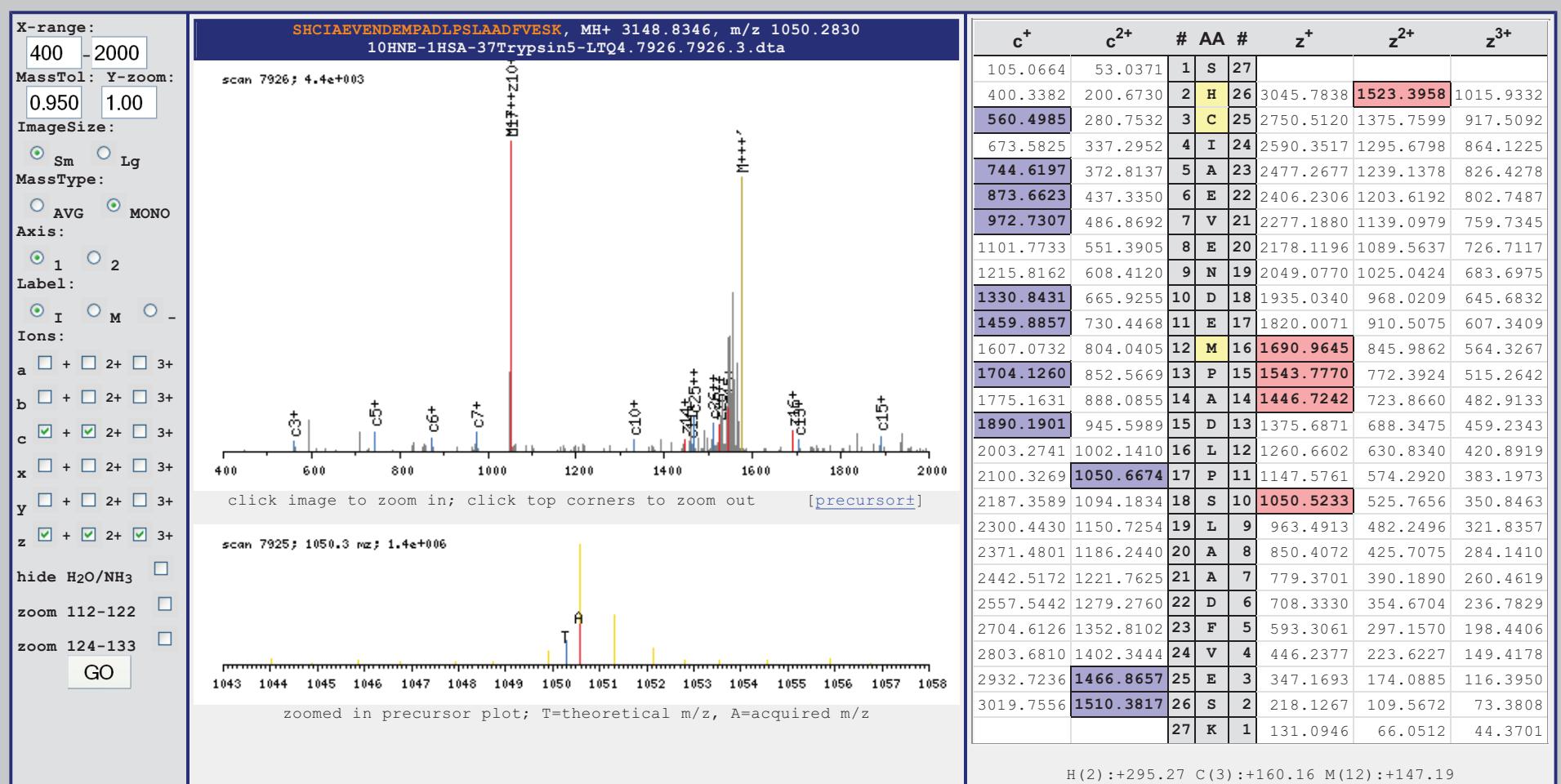


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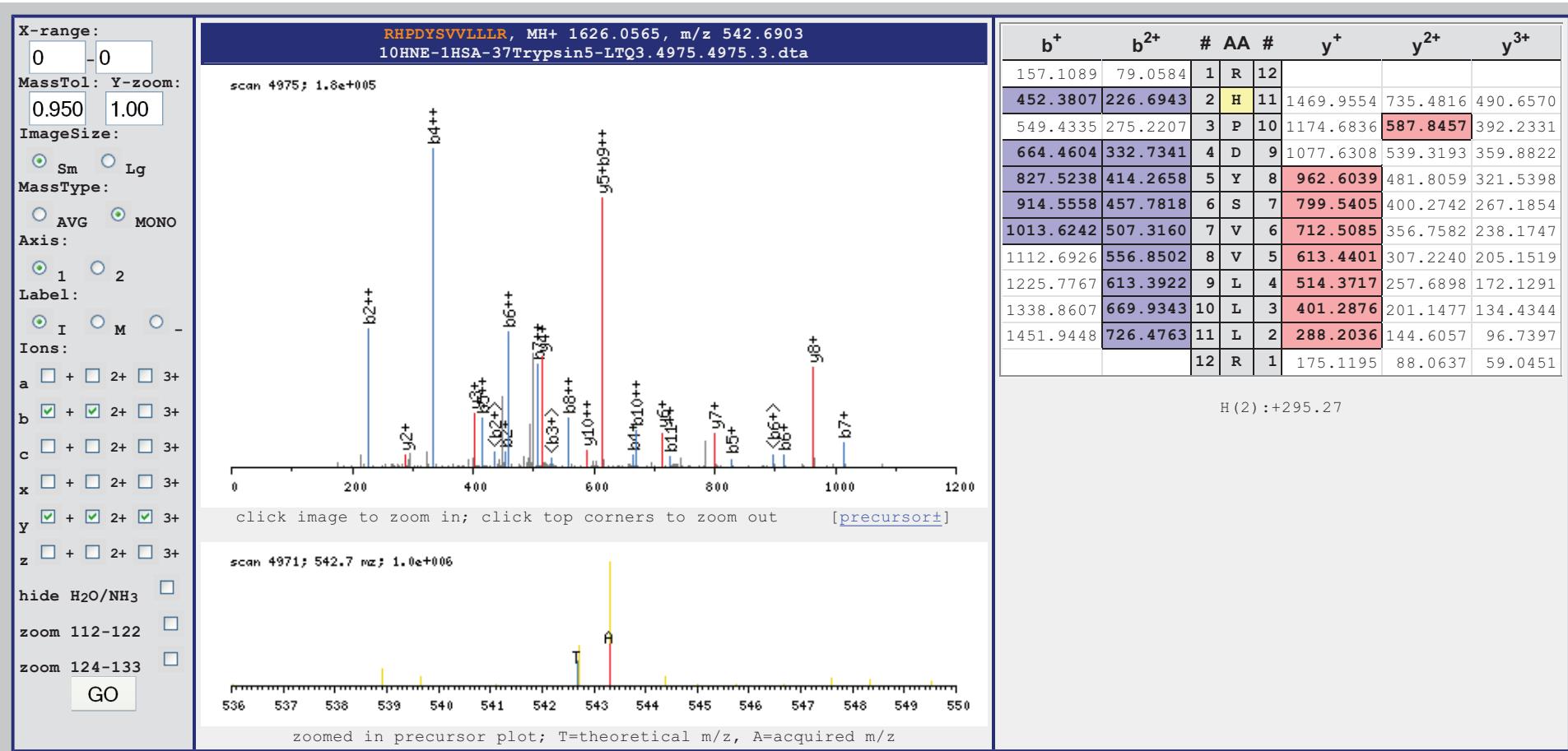


b <sup>+</sup>	b <sup>2+</sup>	#	AA	#	y <sup>+</sup>	y <sup>2+</sup>	y <sup>3+</sup>
88.0399	44.5238	1	S	27			
383.3116	192.1597	2	H	26	3061.8026	1531.4052	1021.2727
543.4719	272.2399	3	C	25	2766.5308	1383.7693	922.8488
656.5560	328.7819	4	I	24	2606.3705	1303.6891	869.4620
727.5931	364.3005	5	A	23	2493.2864	1247.1471	831.7674
856.6357	428.8218	6	E	22	2422.2493	1211.6286	808.0883
955.7041	478.3560	7	V	21	2293.2067	1147.1073	765.0741
1084.7467	542.8773	8	E	20	2194.1383	1097.5731	732.0513
1198.7896	599.8987	9	N	19	2065.0957	1033.0518	689.0371
1313.8166	657.4122	10	D	18	1951.0528	976.0303	651.0228
1442.8592	721.9335	11	E	17	1836.0258	918.5168	612.6805
1590.0467	795.5273	12	M	16	1706.9832	853.9955	569.6663
1687.0995	844.0536	13	P	15	1559.7957	780.4018	520.6038
1758.1366	879.5722	14	A	14	1462.7430	731.8754	488.2529
1873.1635	937.0857	15	D	13	1391.7058	696.3568	464.5738
1986.2476	993.6277	16	L	12	1276.6789	638.8434	426.2315
2083.3003	1042.1541	17	P	11	1163.5948	582.3013	388.5368
2170.3324	1085.6701	18	S	10	1066.5421	533.7749	356.1859
2283.4164	1142.2121	19	L	9	979.5100	490.2589	327.1752
2354.4535	1177.7307	20	A	8	866.4260	433.7169	289.4805
2425.4907	1213.2492	21	A	7	795.3889	398.1983	265.8015
2540.5176	1270.7627	22	D	6	724.3517	362.6798	242.1225
2687.5860	1344.2969	23	F	5	609.3248	305.1663	203.7802
2786.6544	1393.8311	24	V	4	462.2564	231.6321	154.7573
2915.6970	1458.3524	25	E	3	363.1880	182.0979	121.7345
3002.7291	1501.8684	26	S	2	234.1454	117.5766	78.7203
		27	K	1	147.1134	74.0606	49.7097

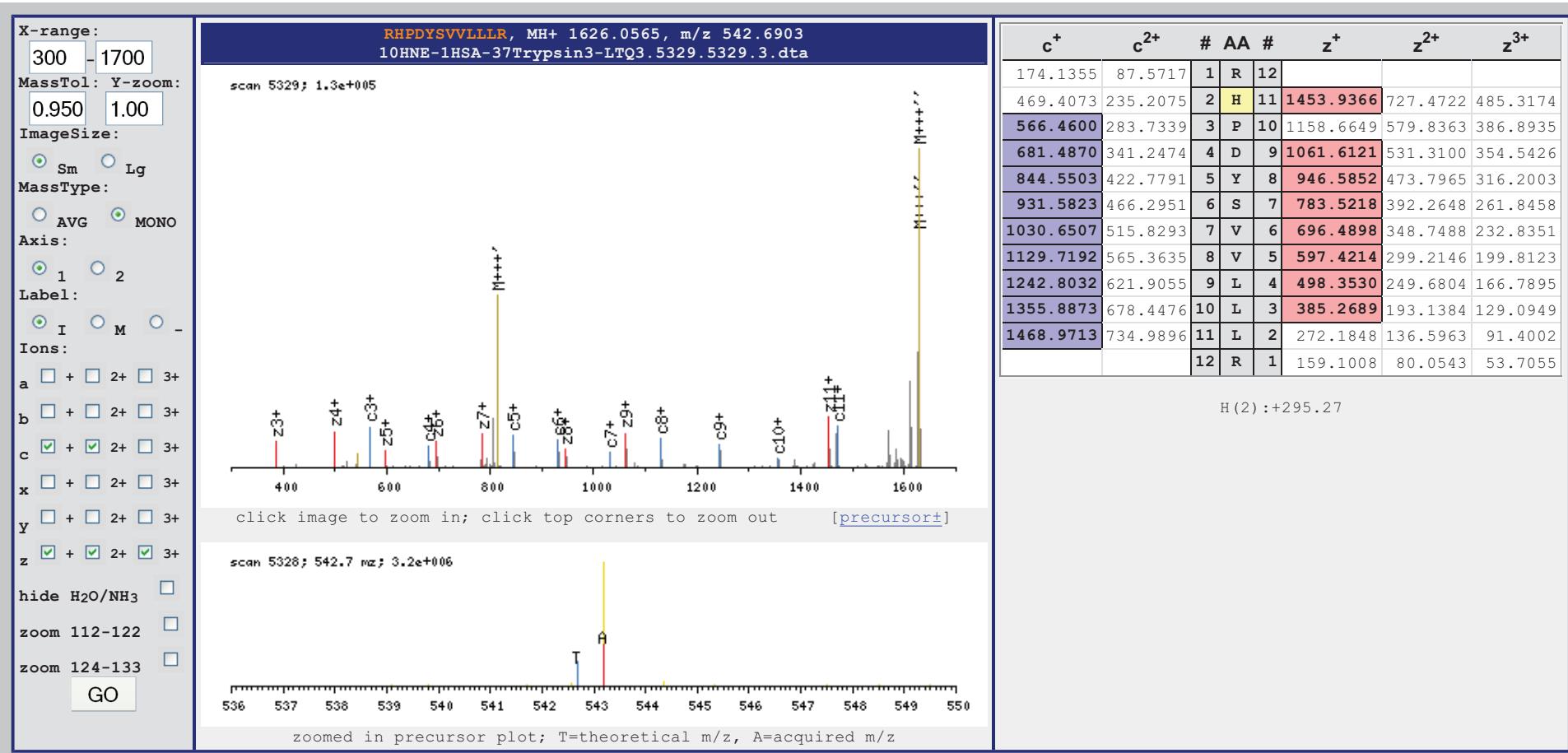
H(2):+295.27 C(3):+160.16 M(12):+147.19



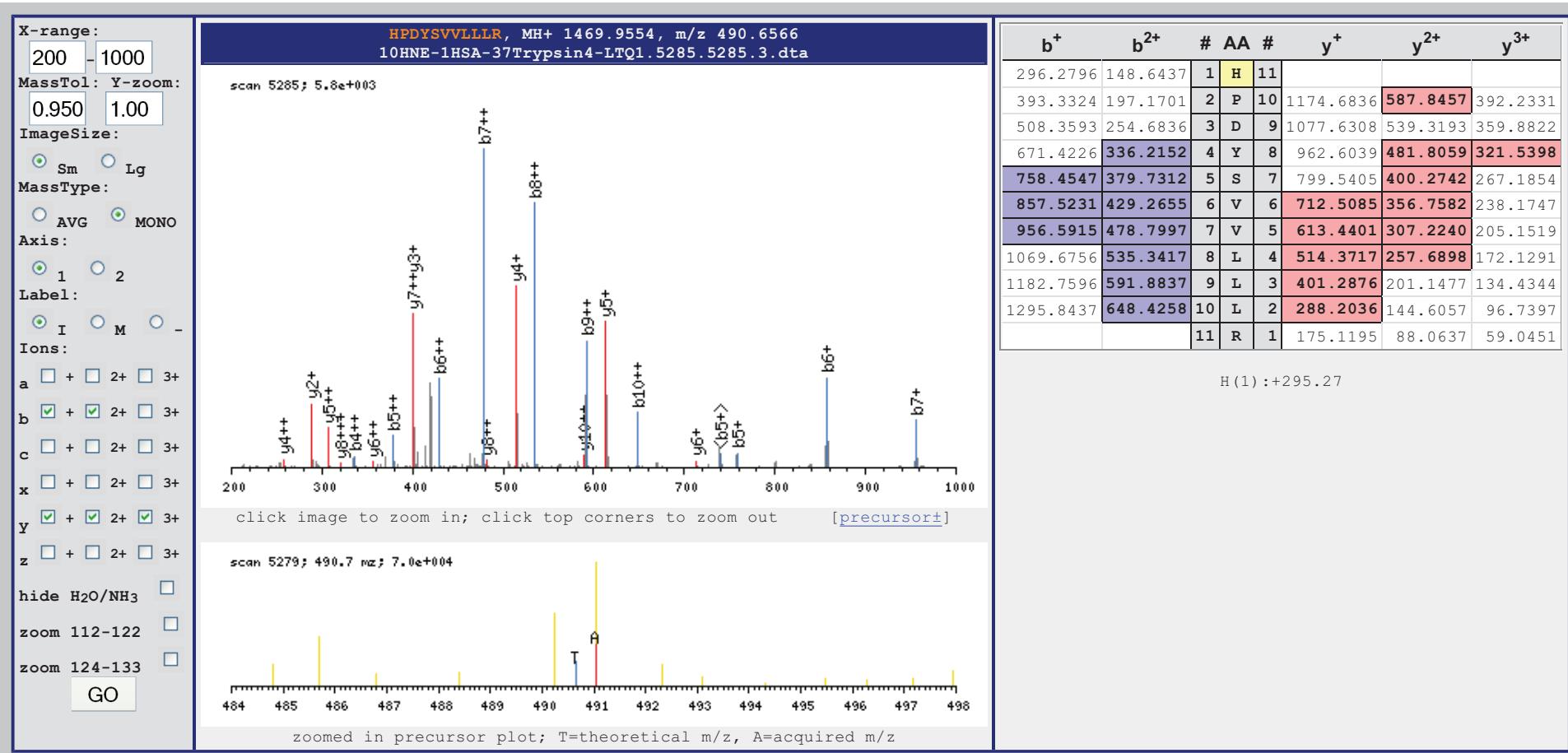
COMET Spectrum View by J.Eng (c) ISB 2001  
(TPP v4.4 VUVUZELA rev 1, Build 201010121551 (MinGW))

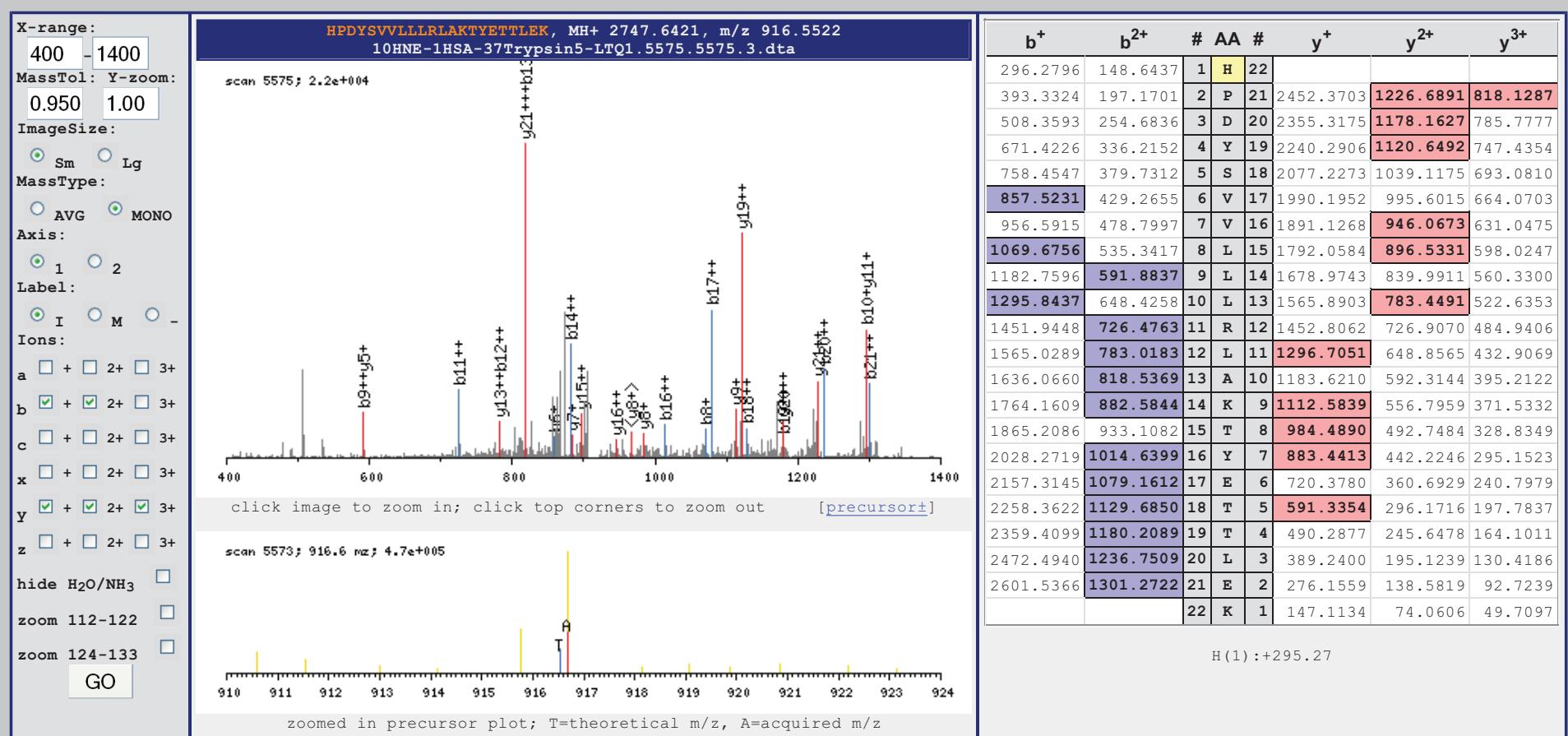


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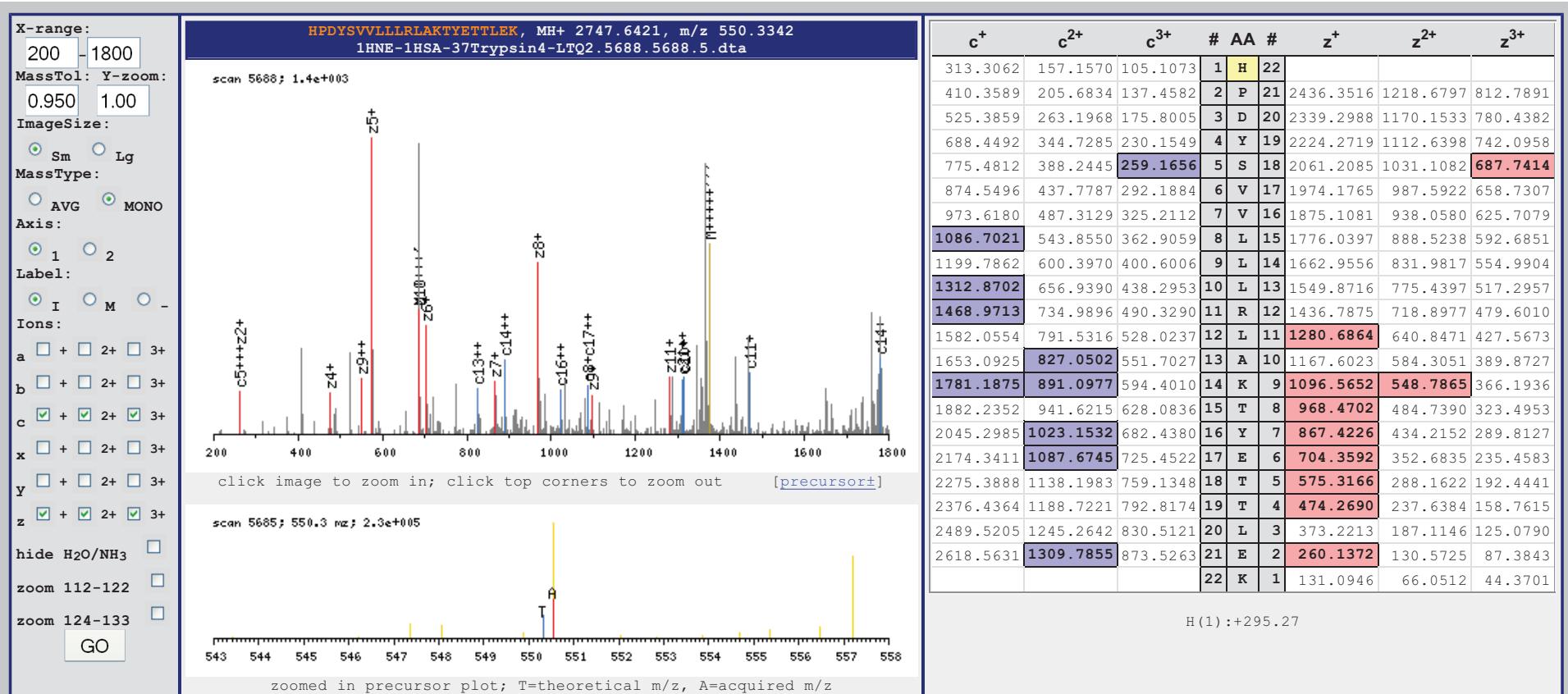


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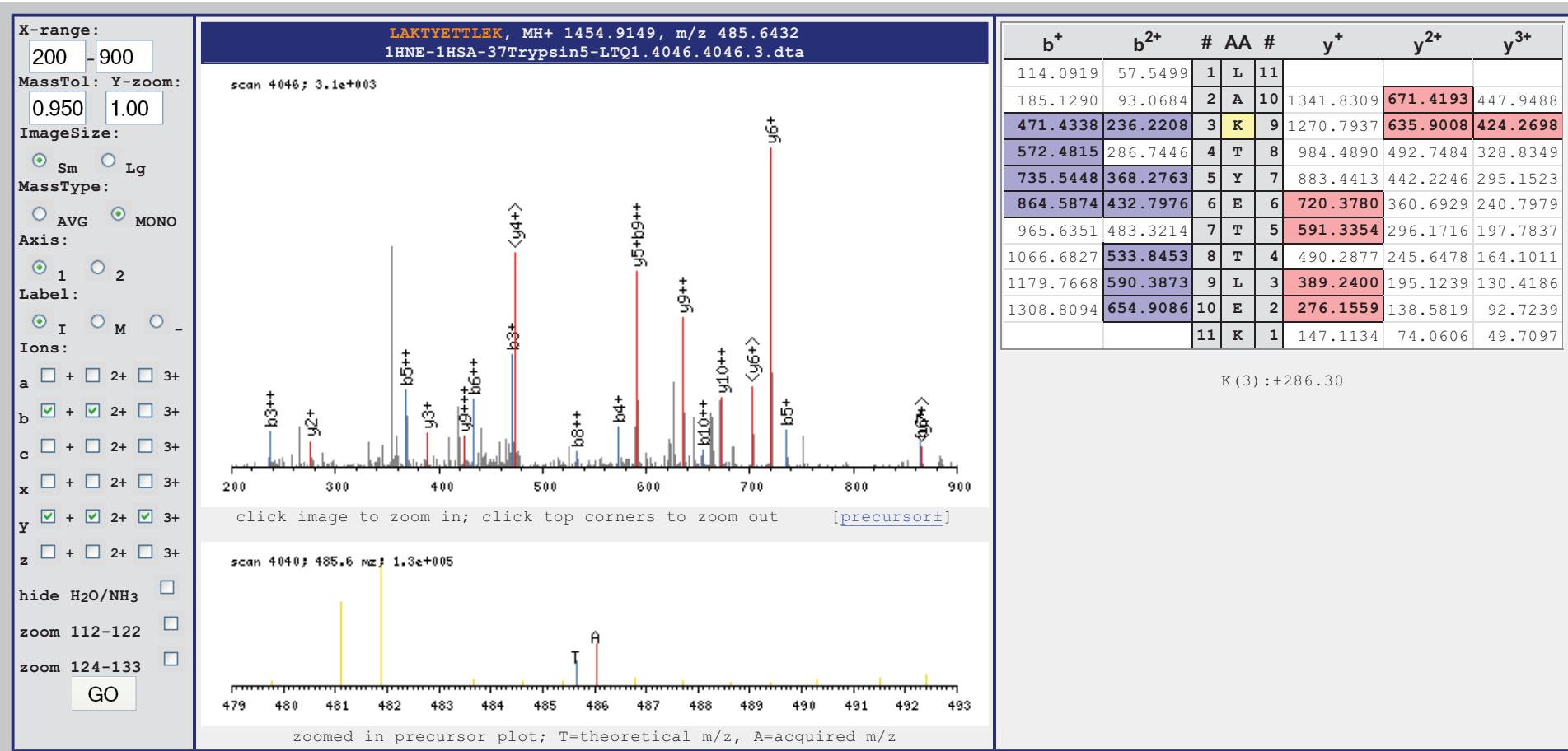




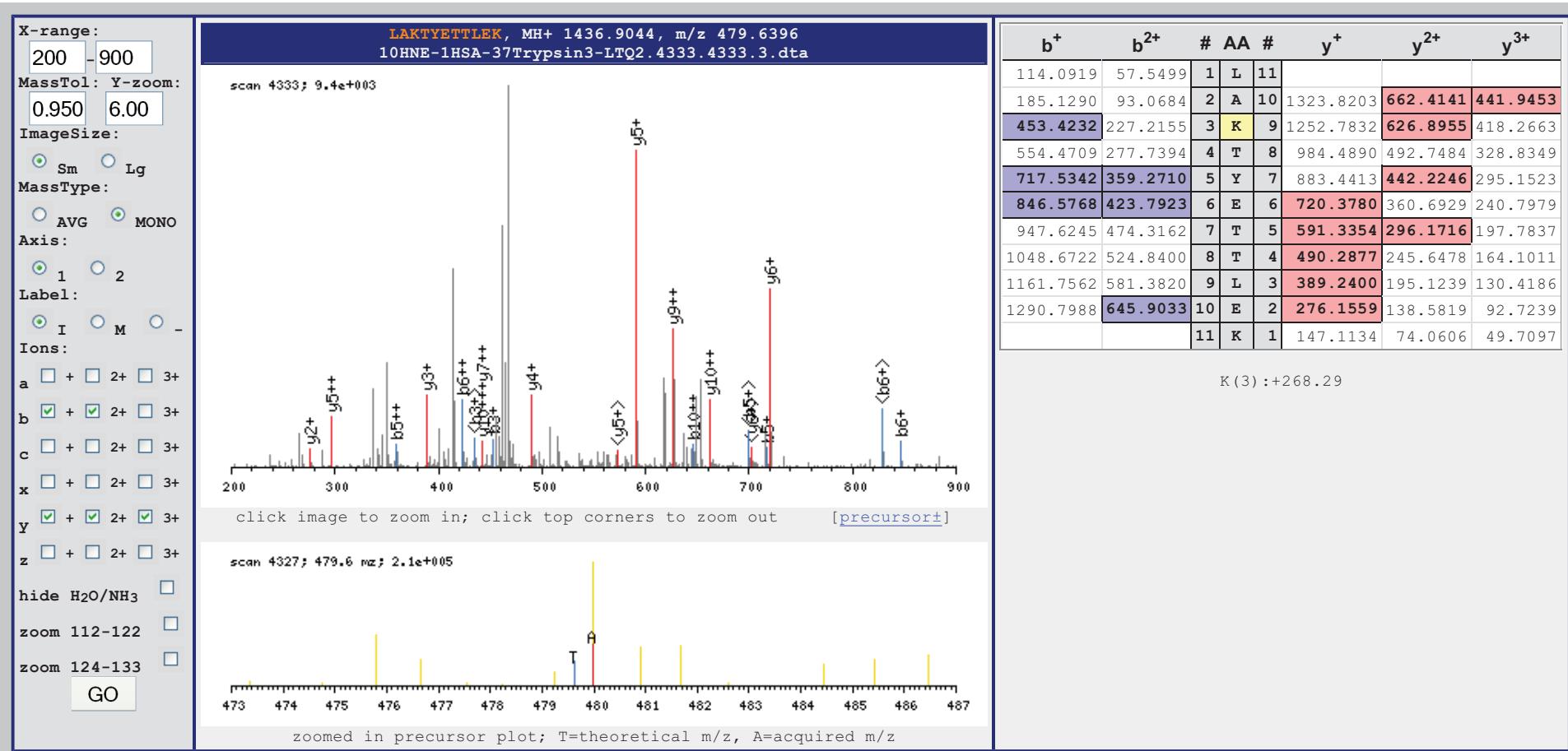
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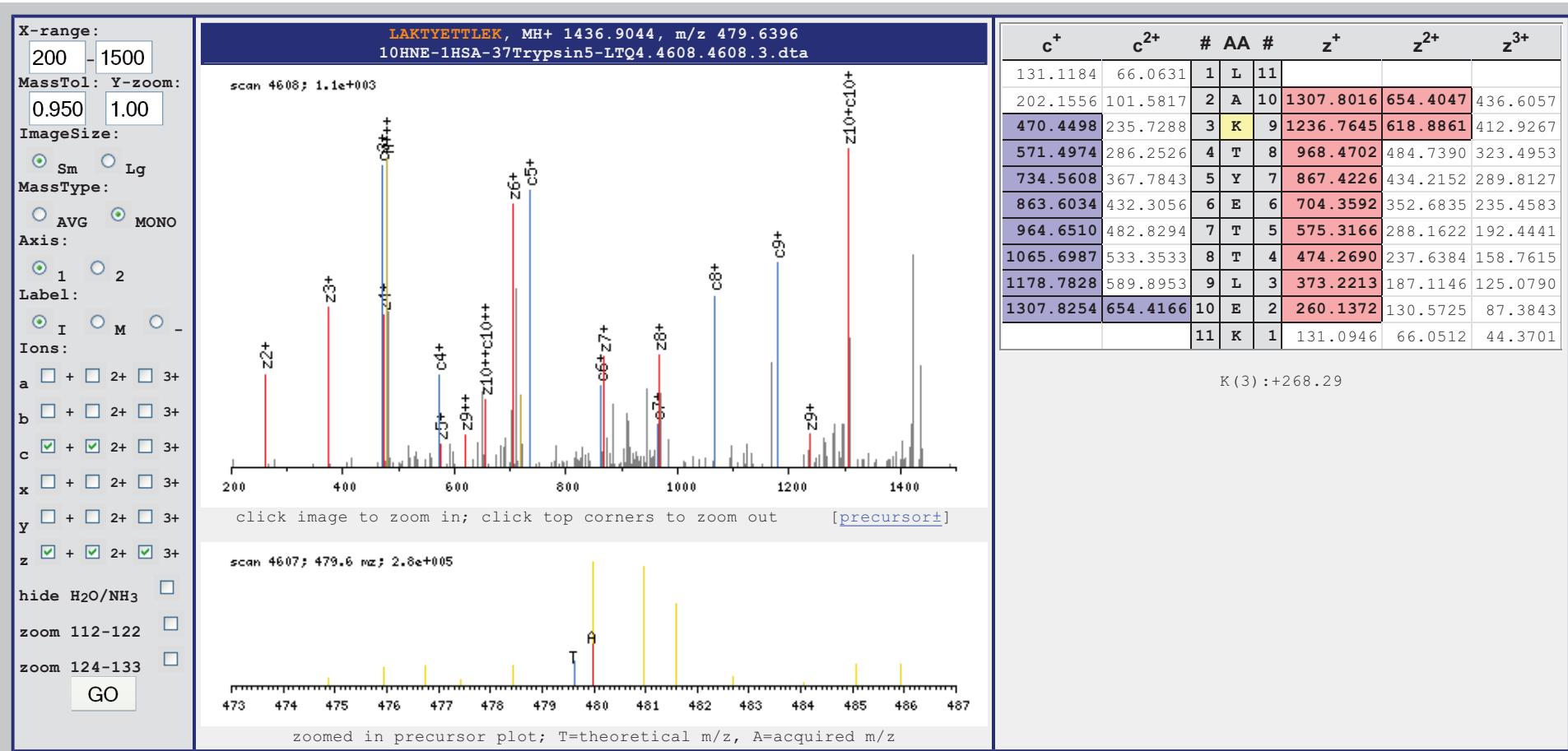
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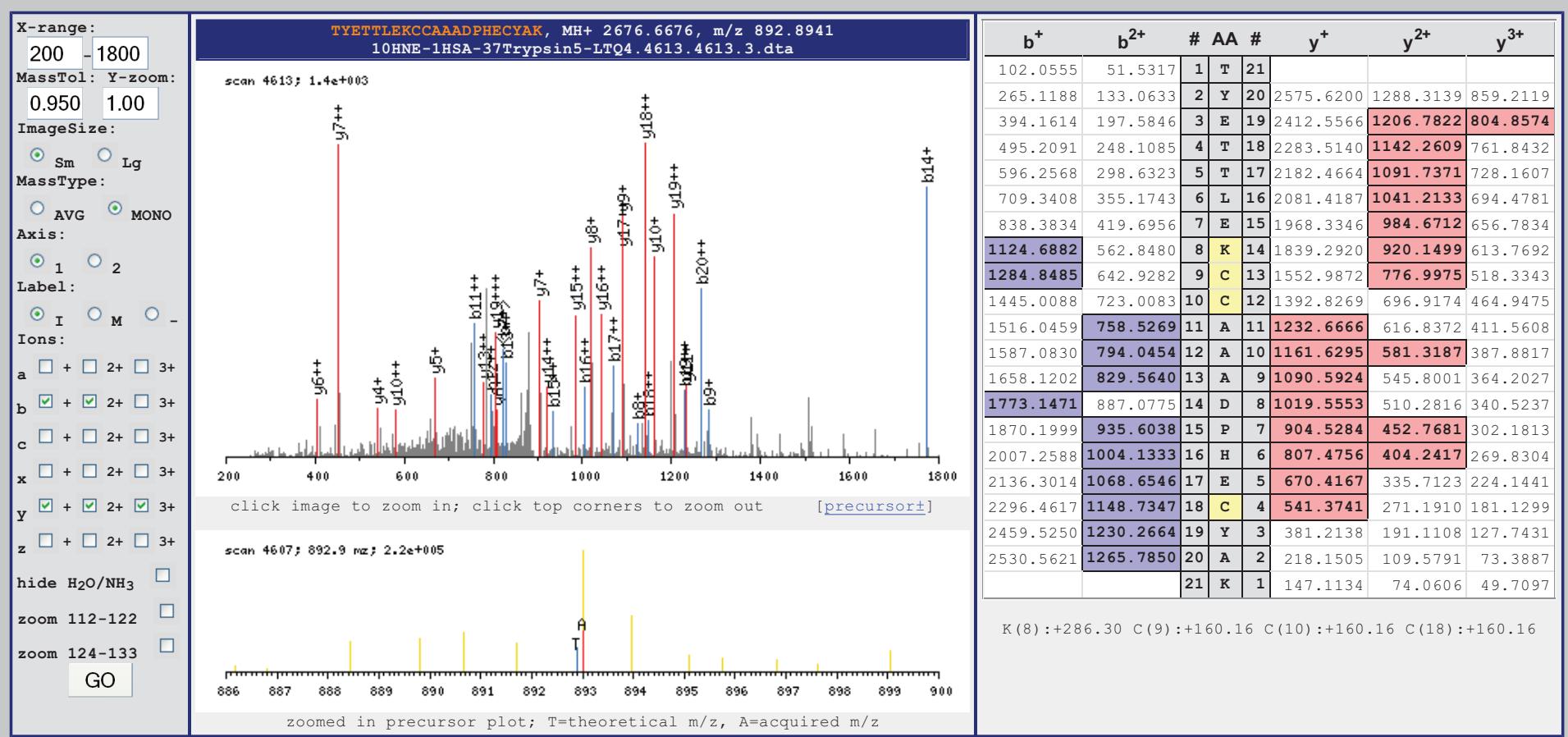
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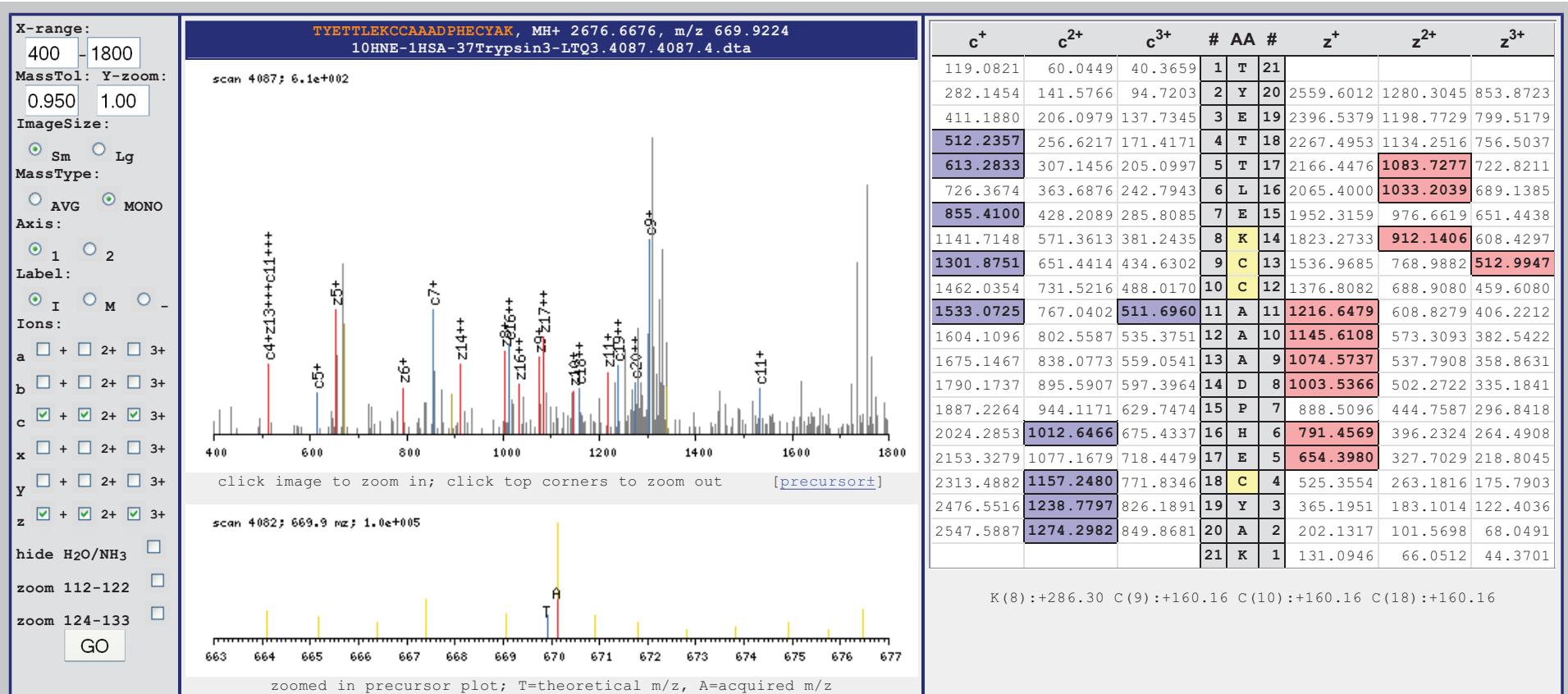
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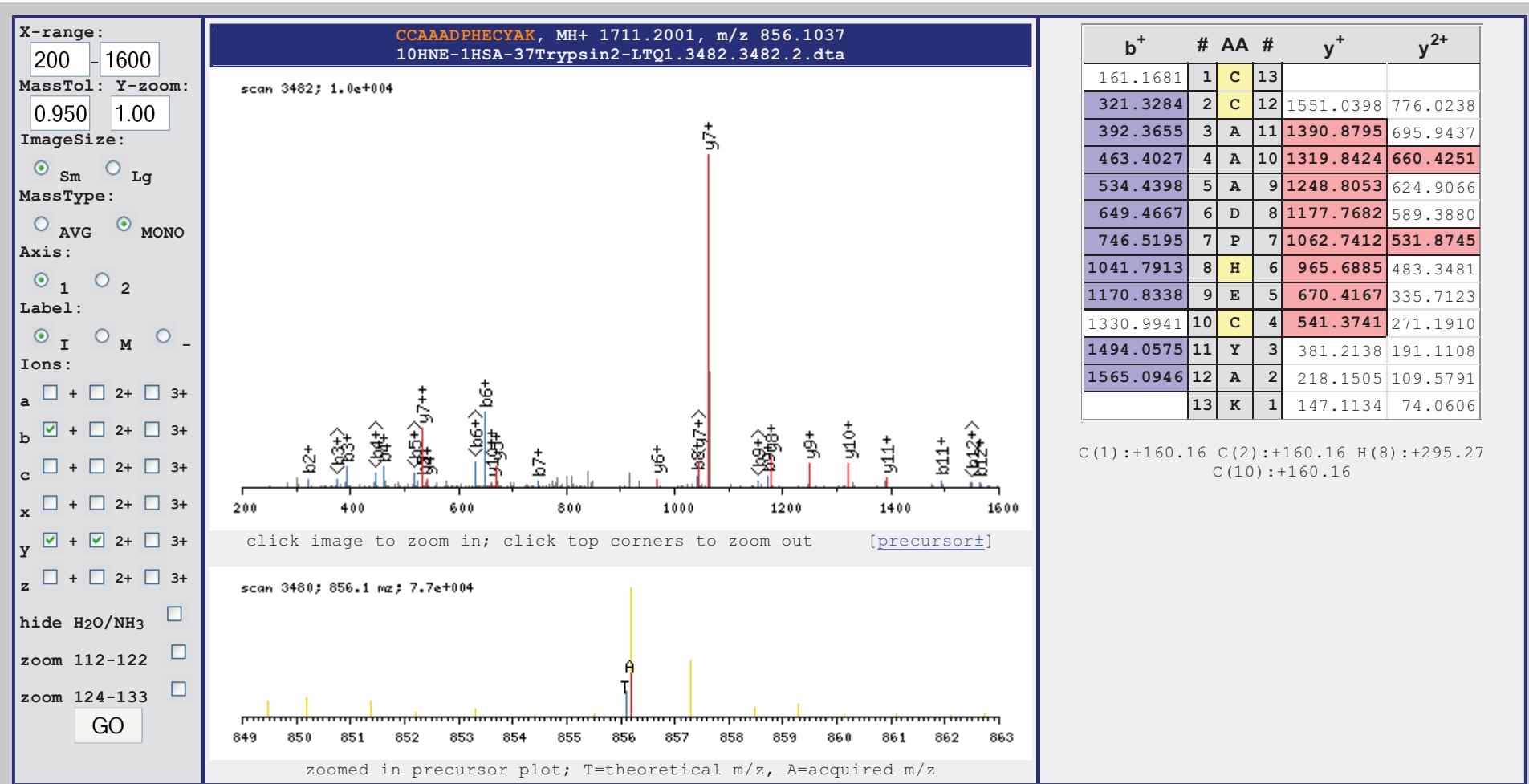
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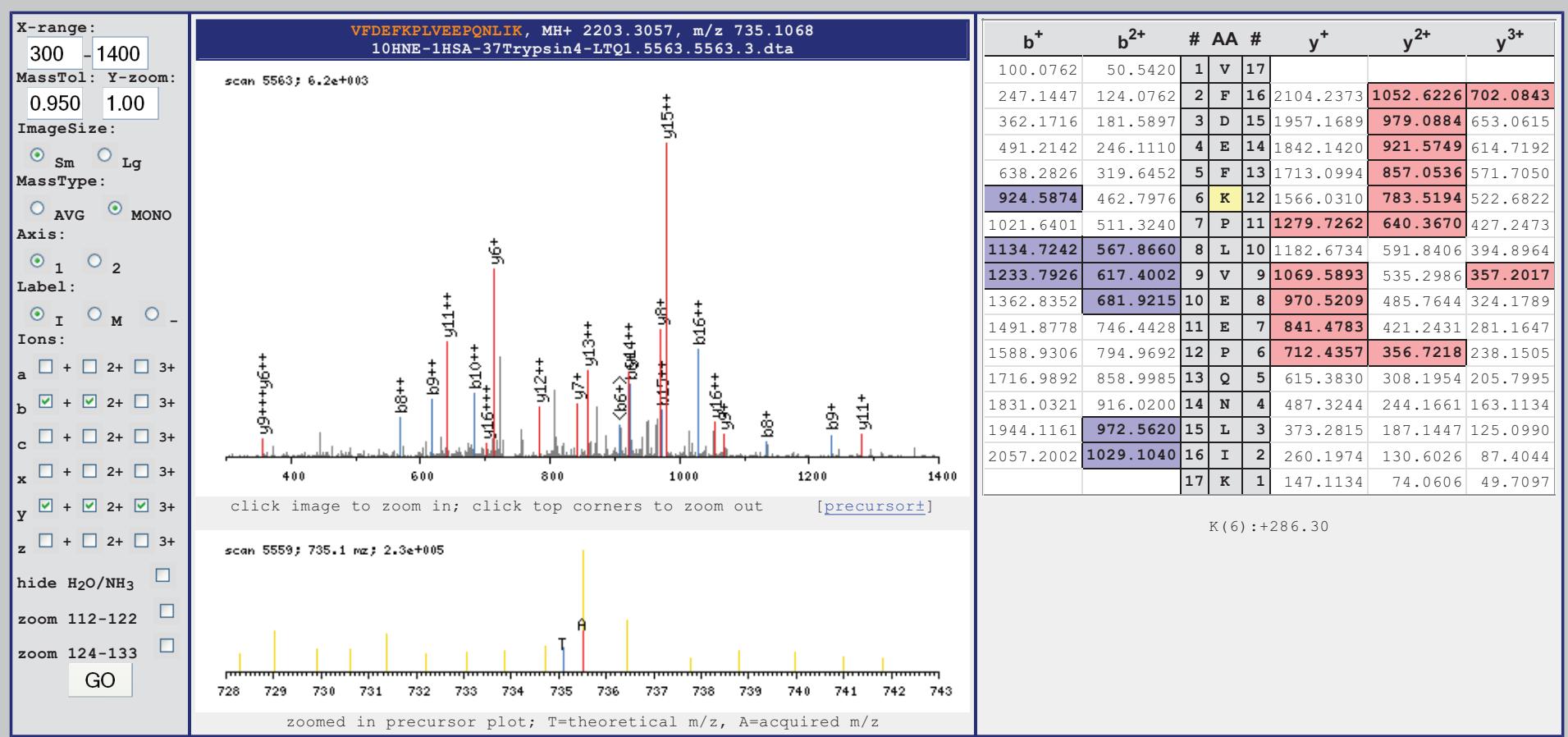
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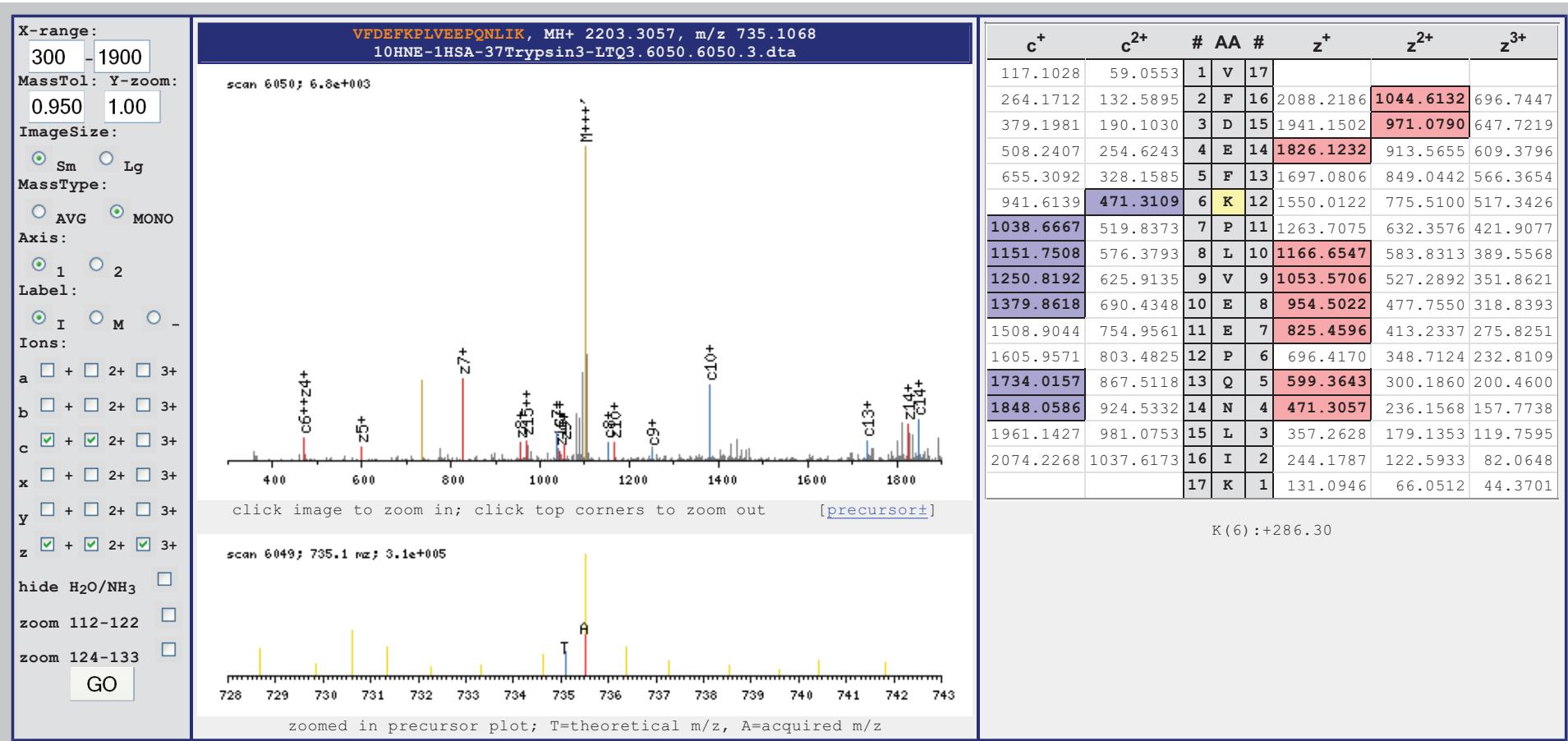
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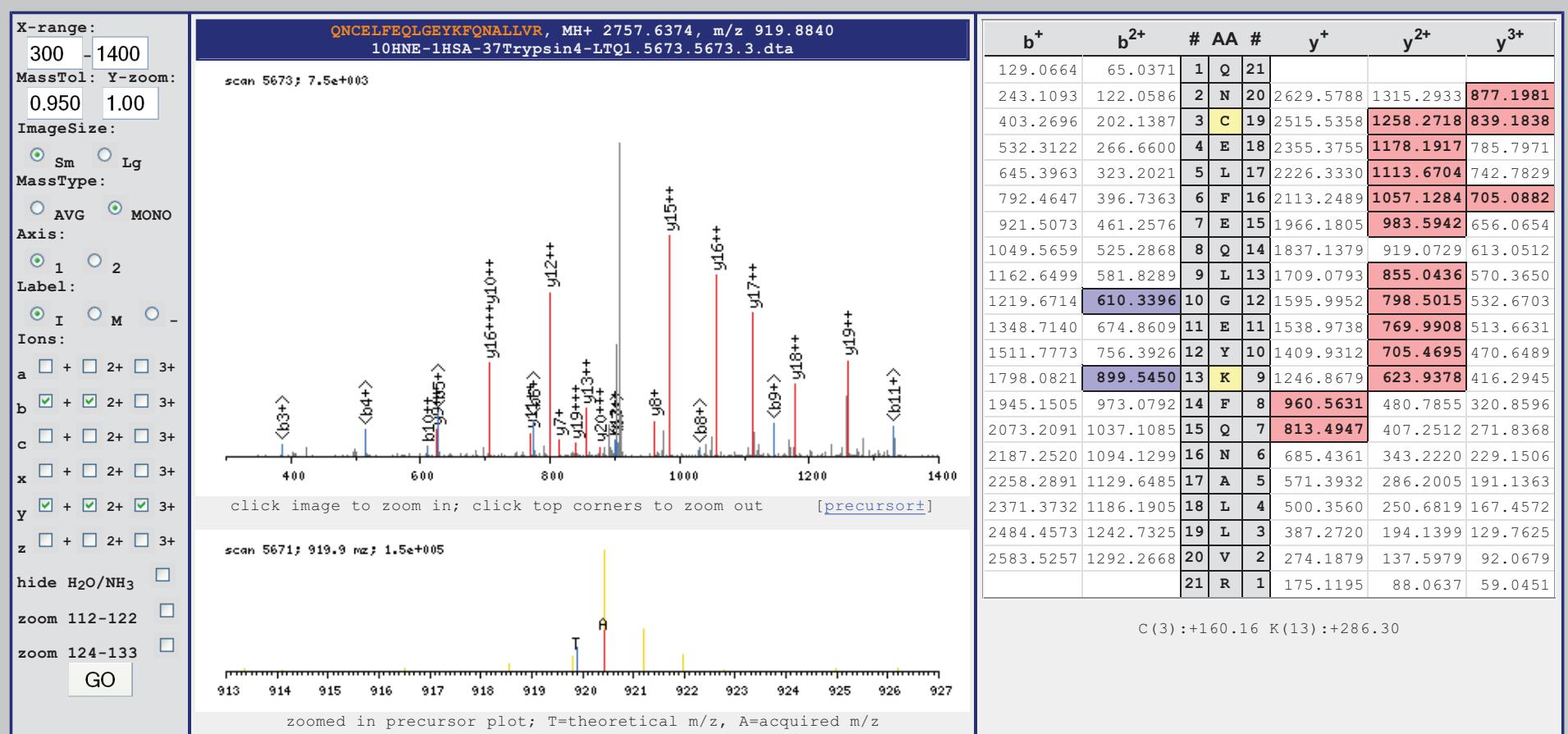
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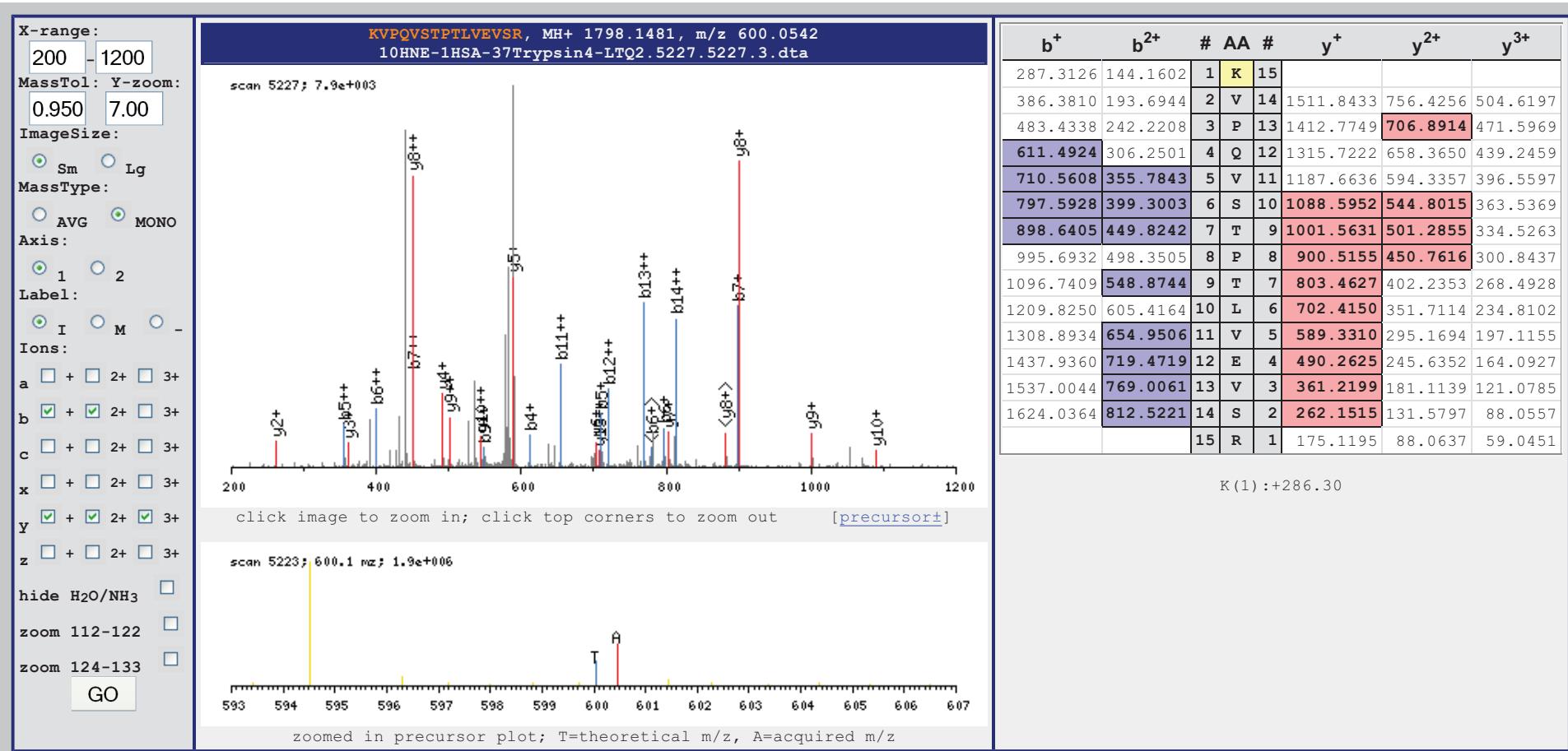
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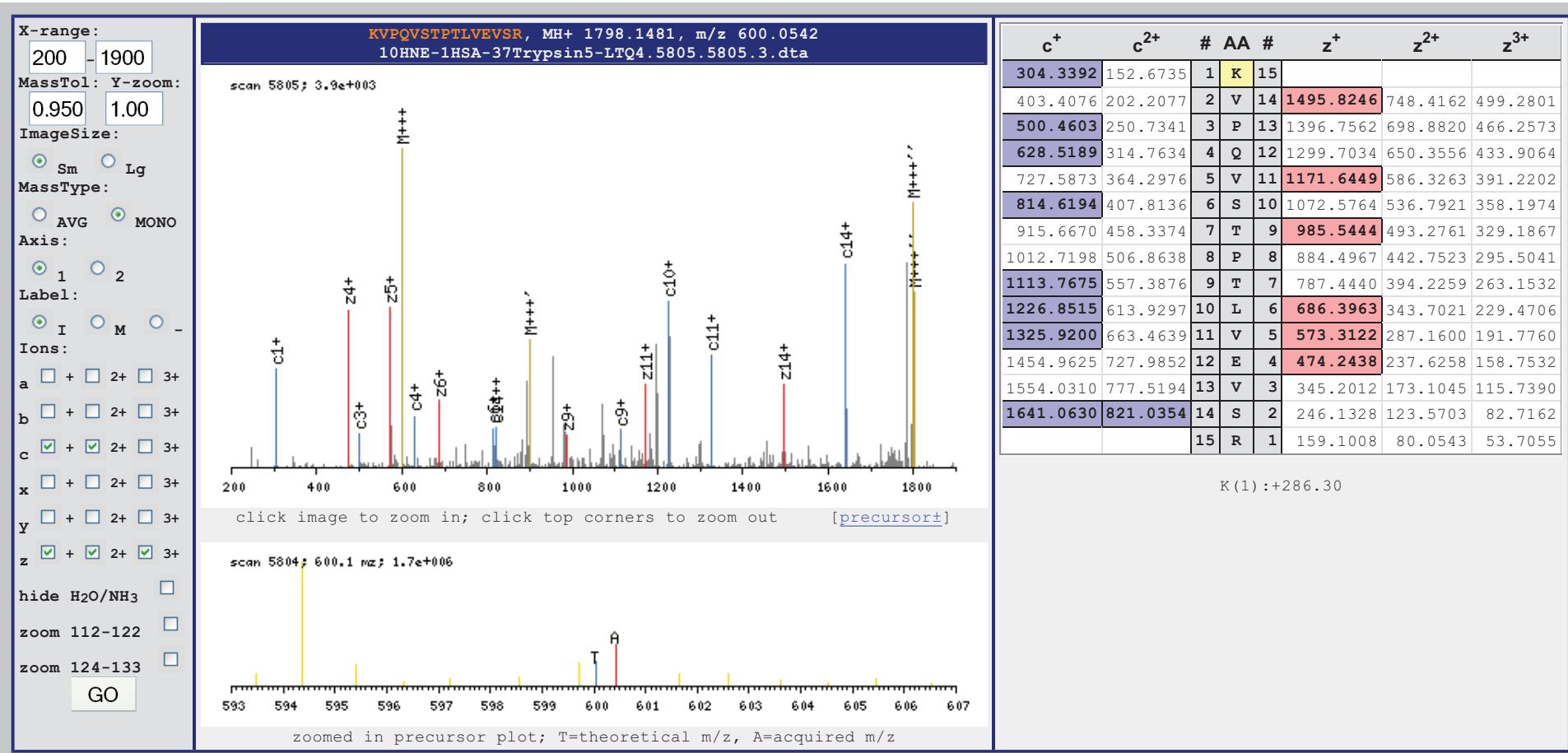
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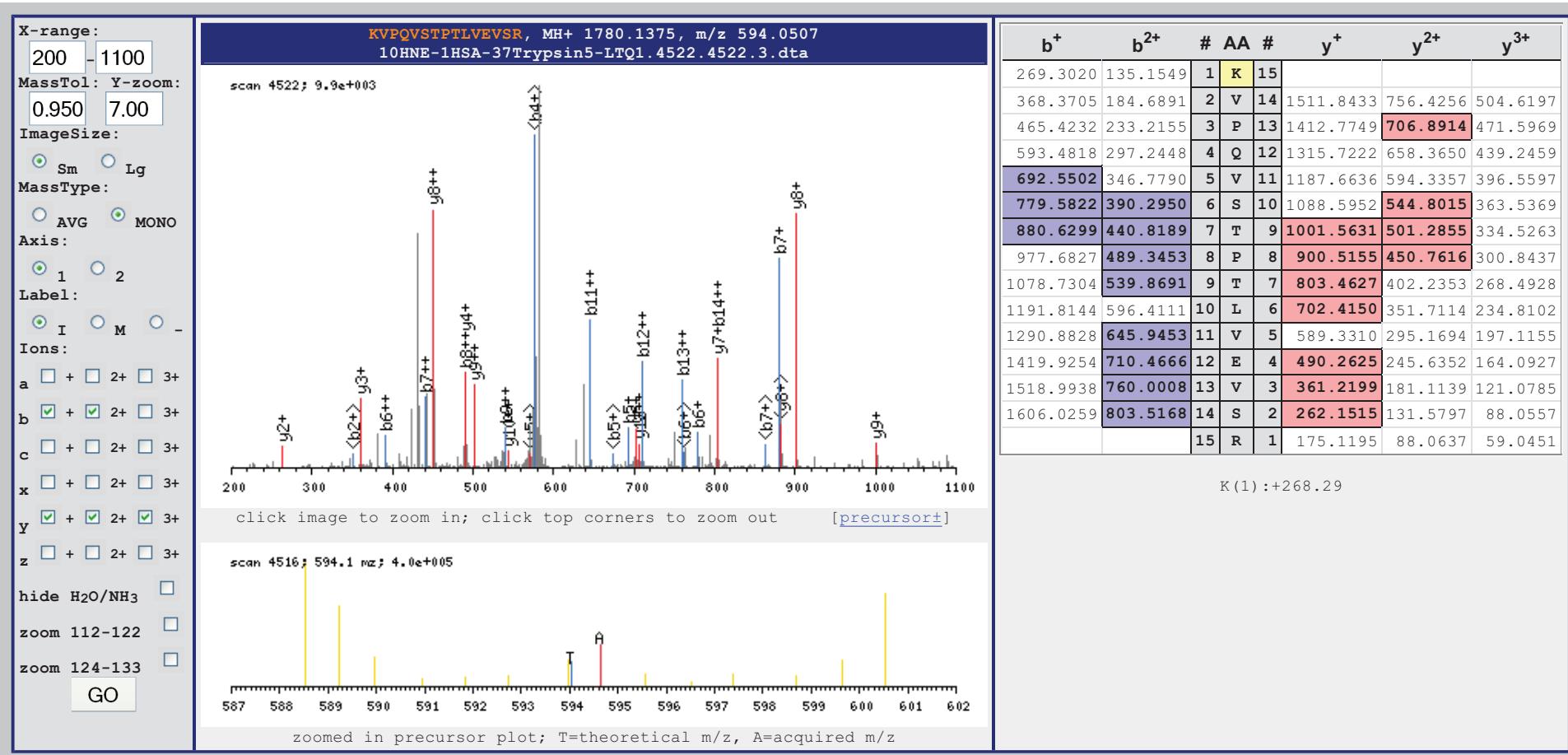
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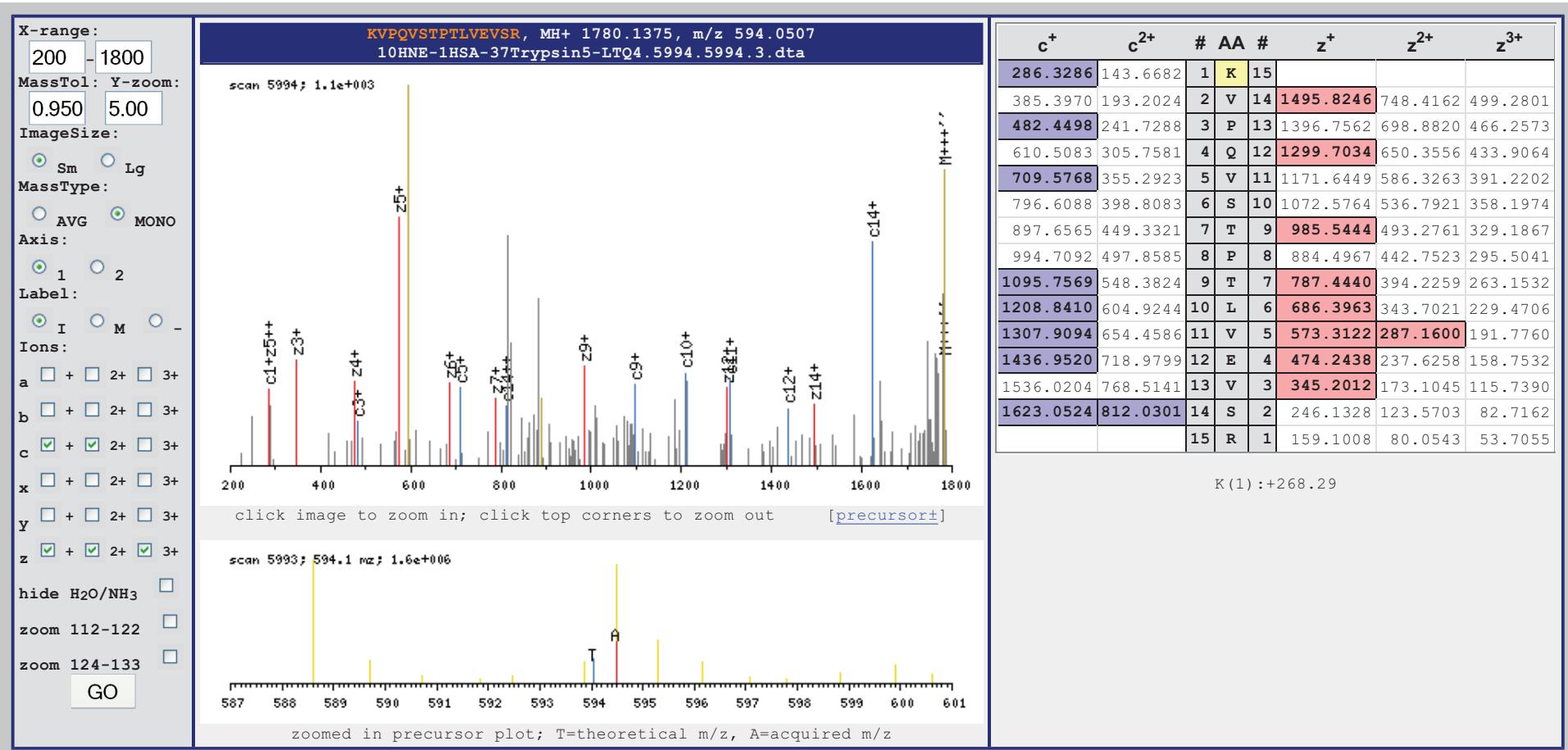
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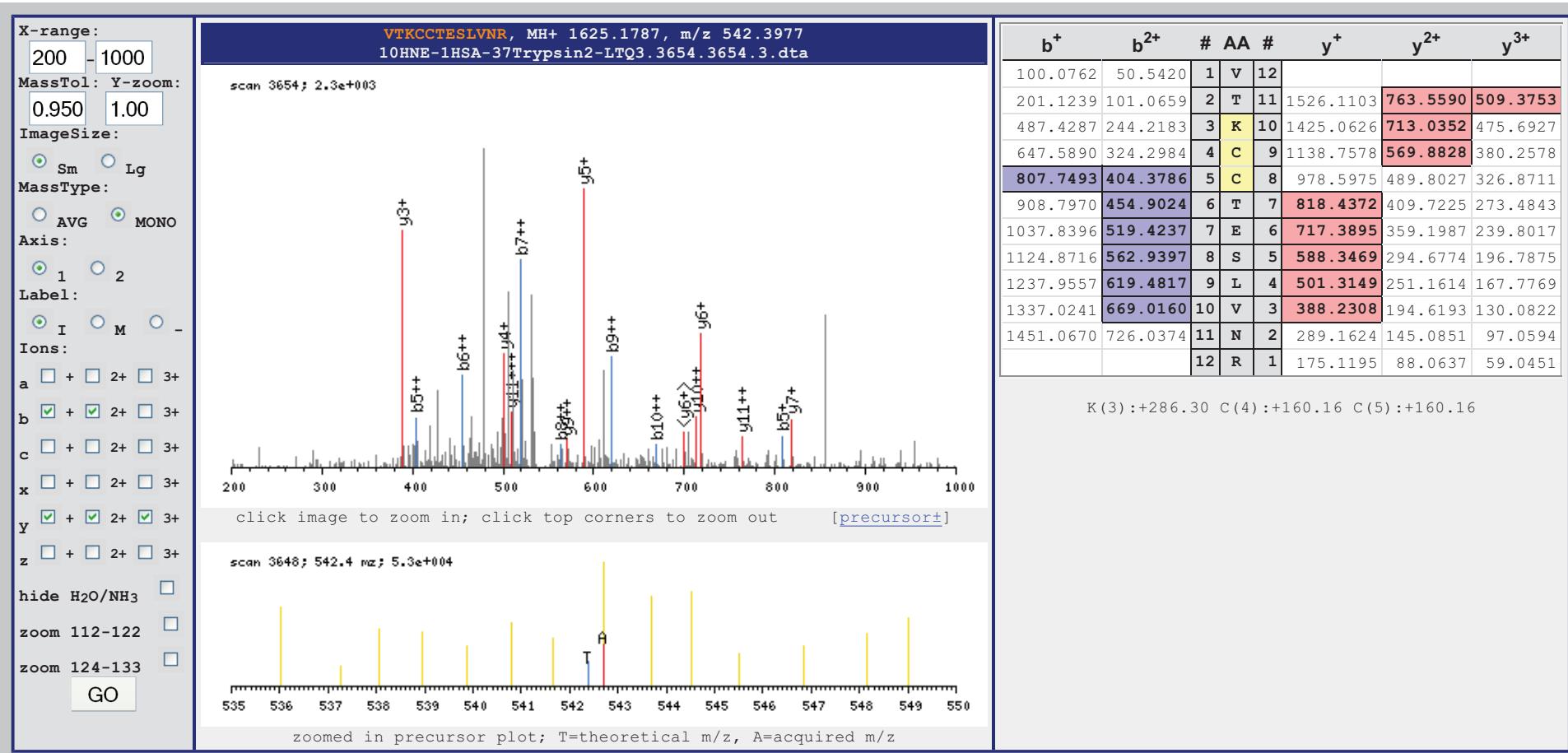
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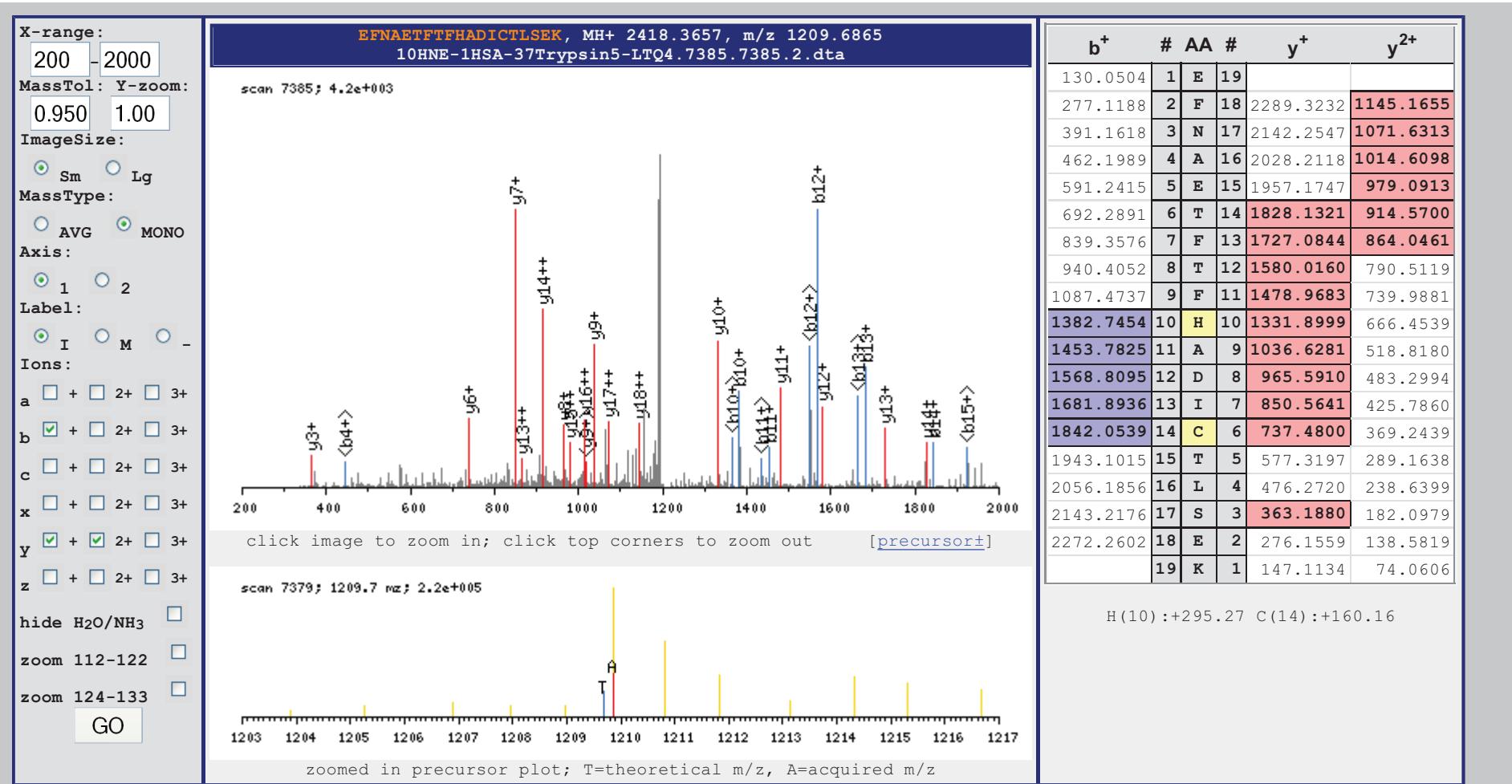
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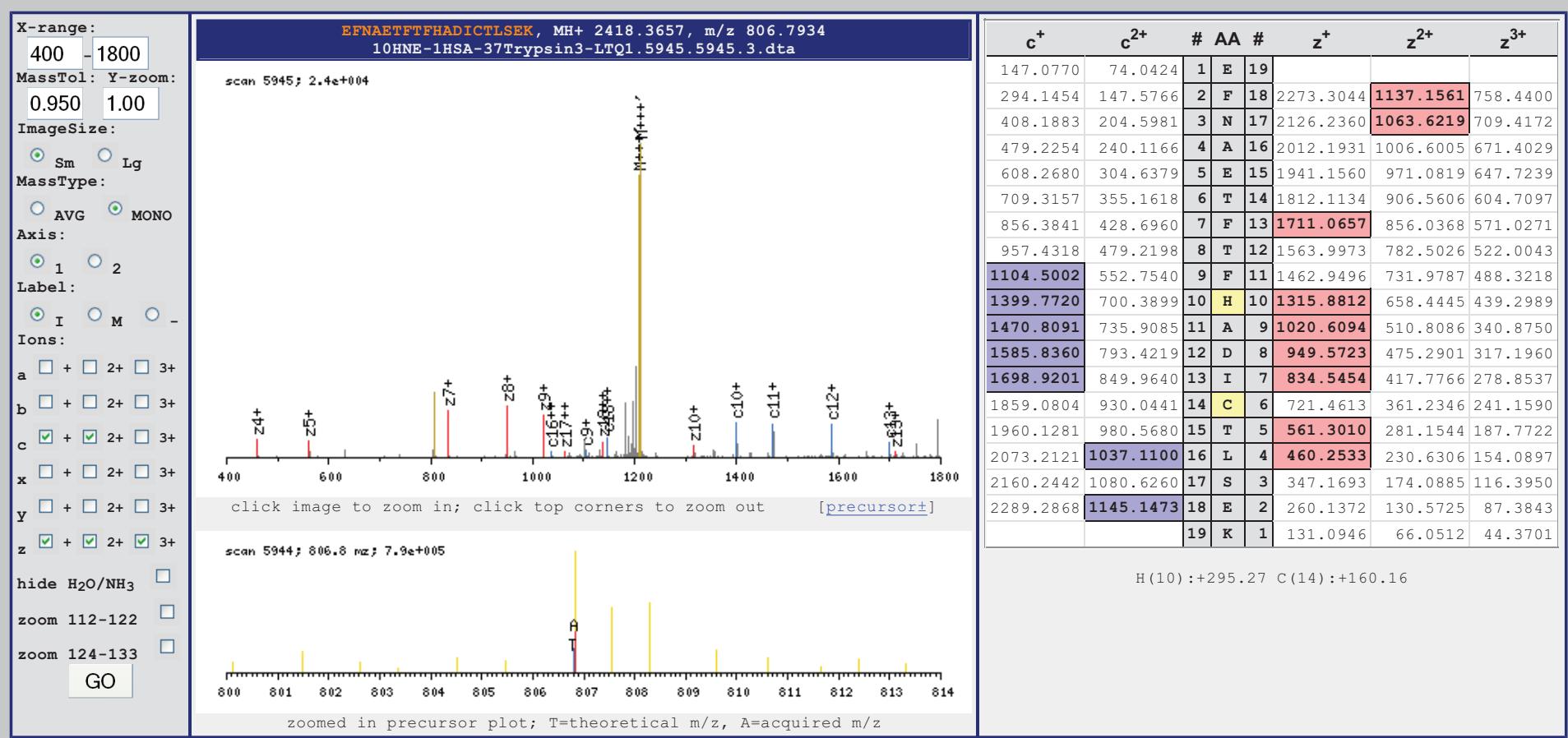
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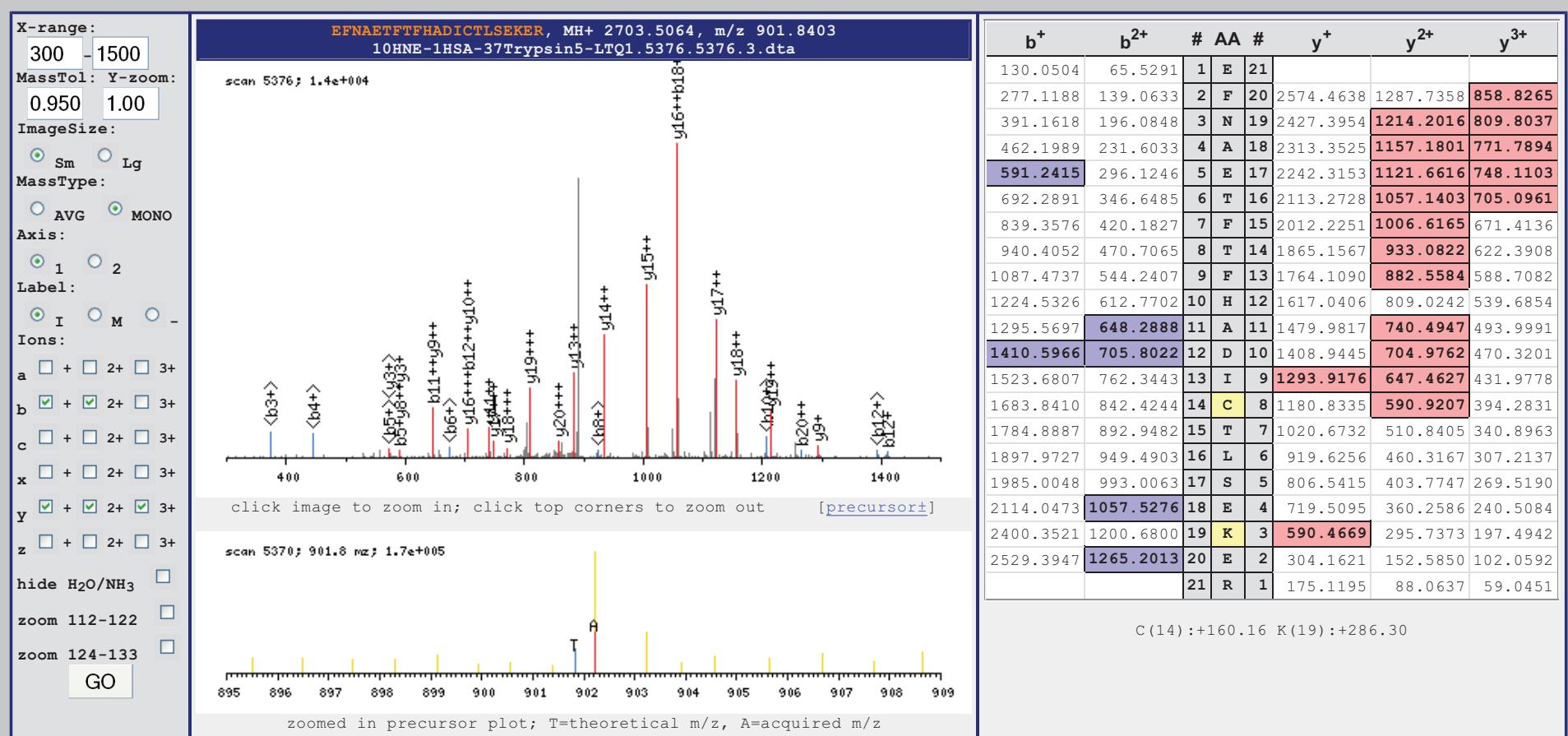
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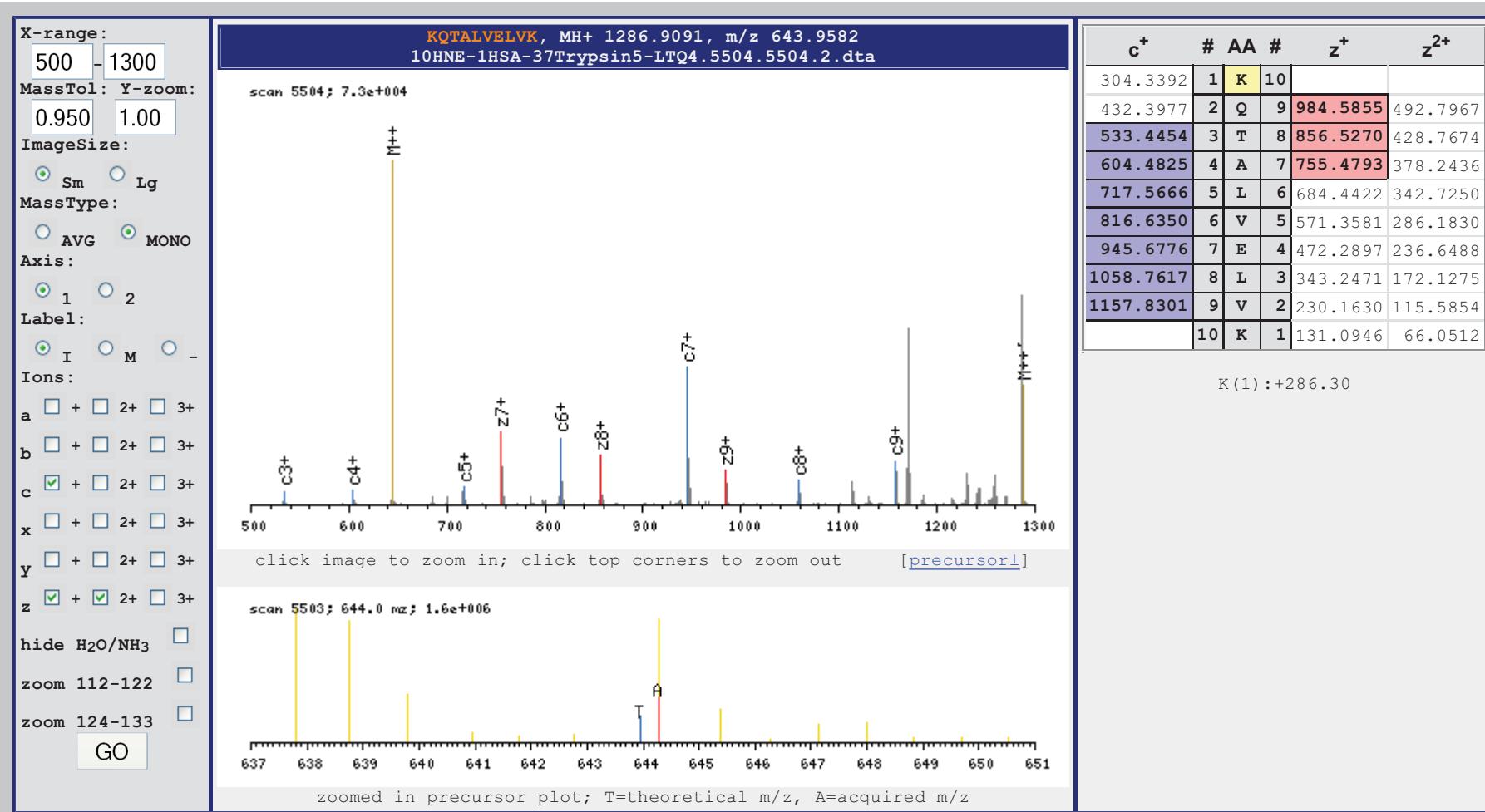
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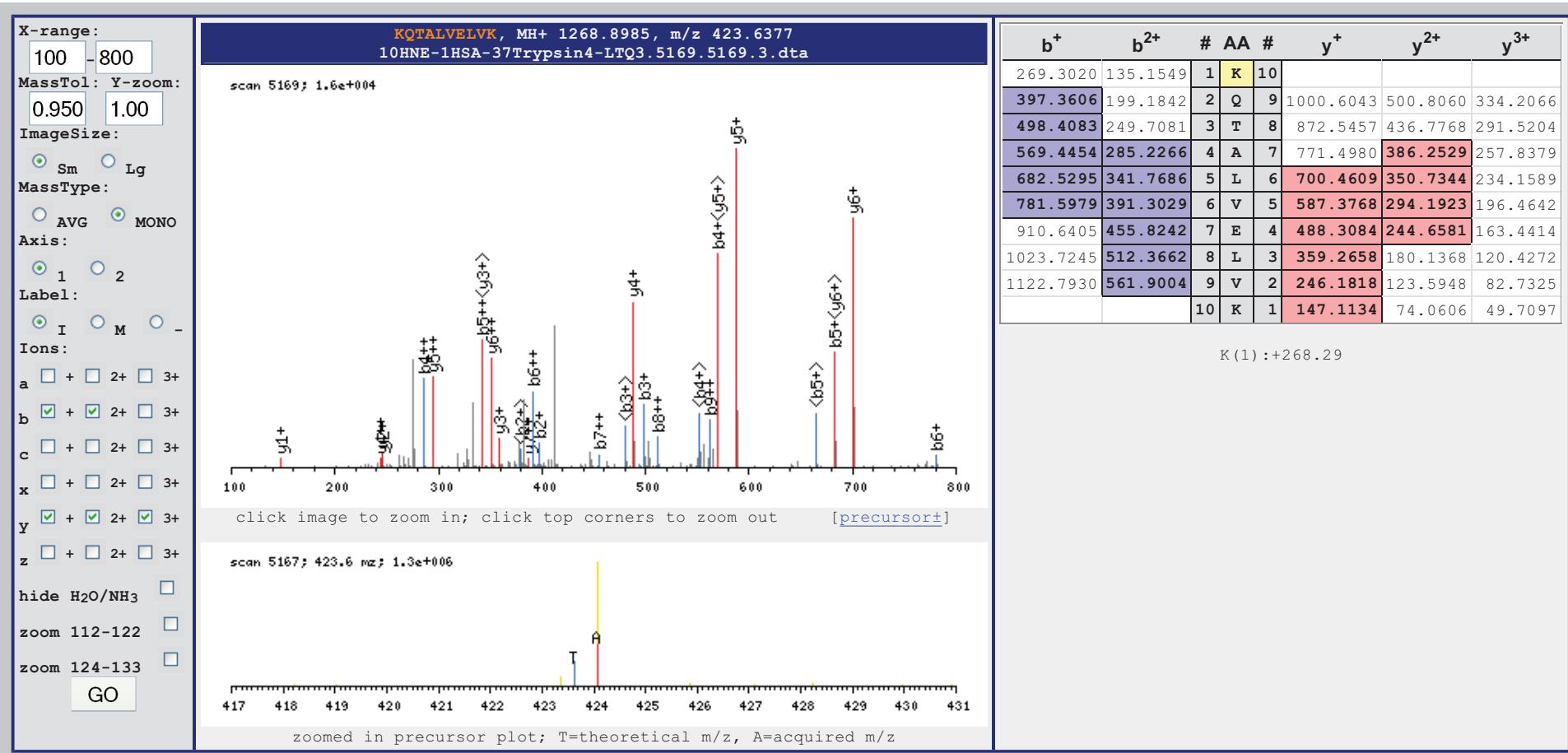
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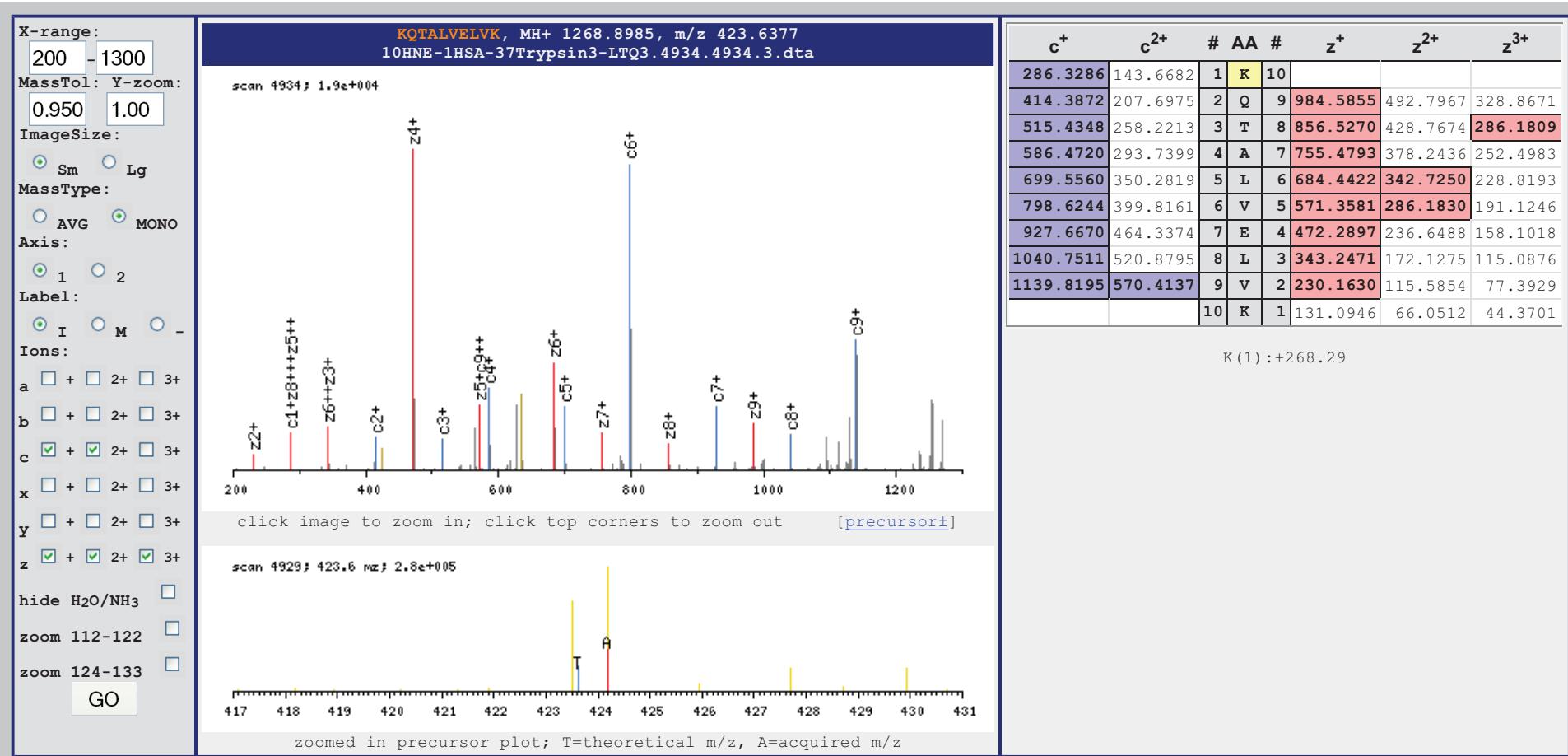
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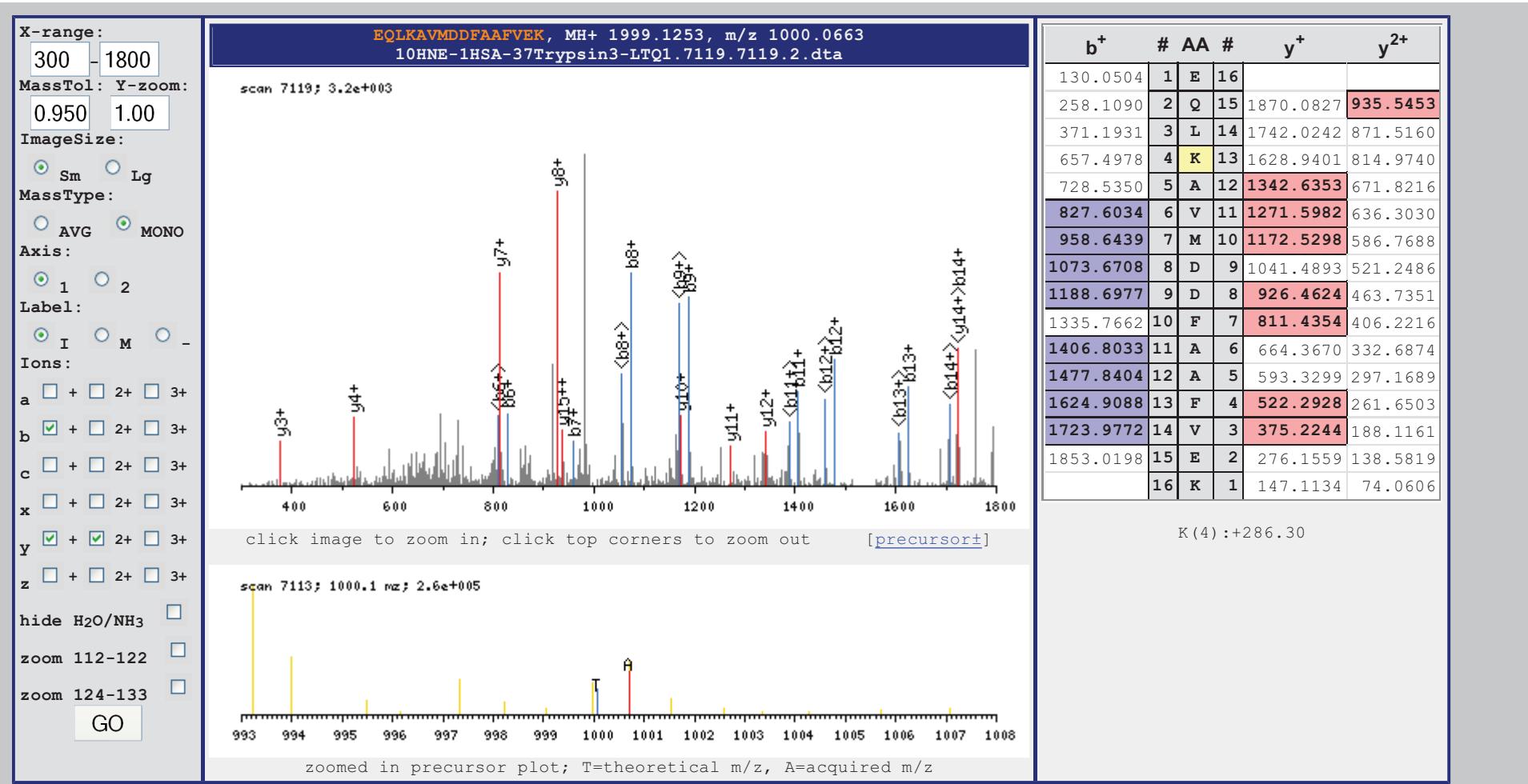
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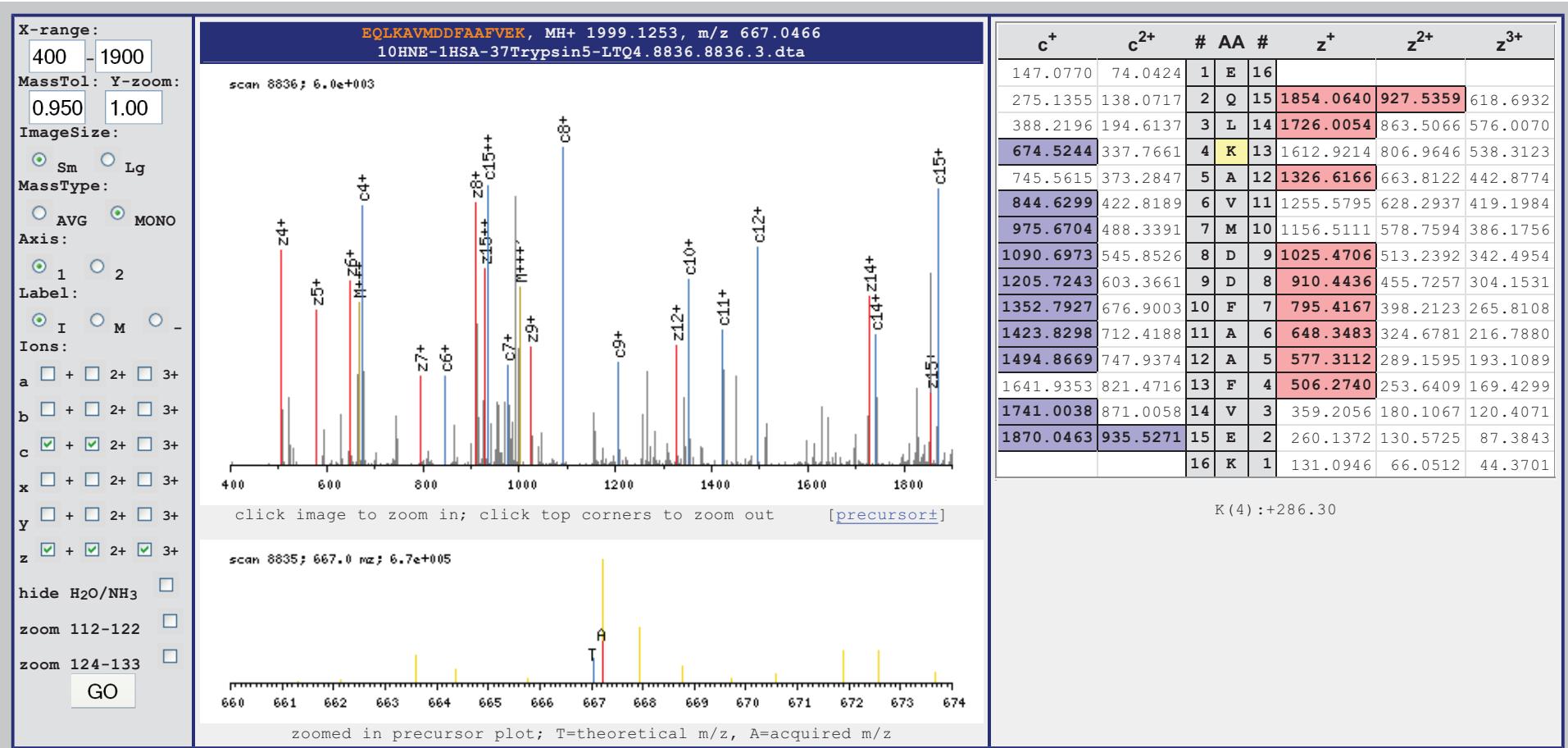
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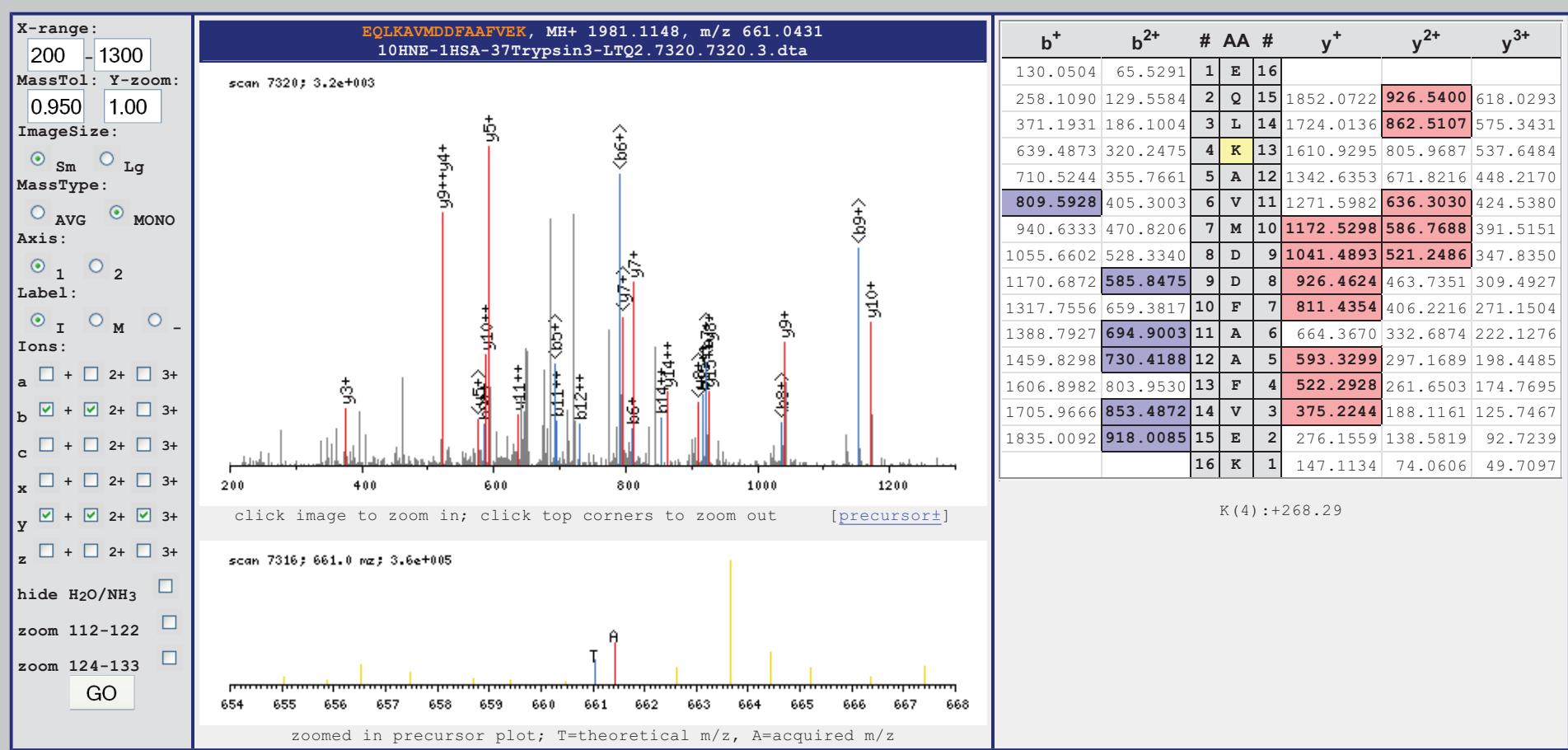
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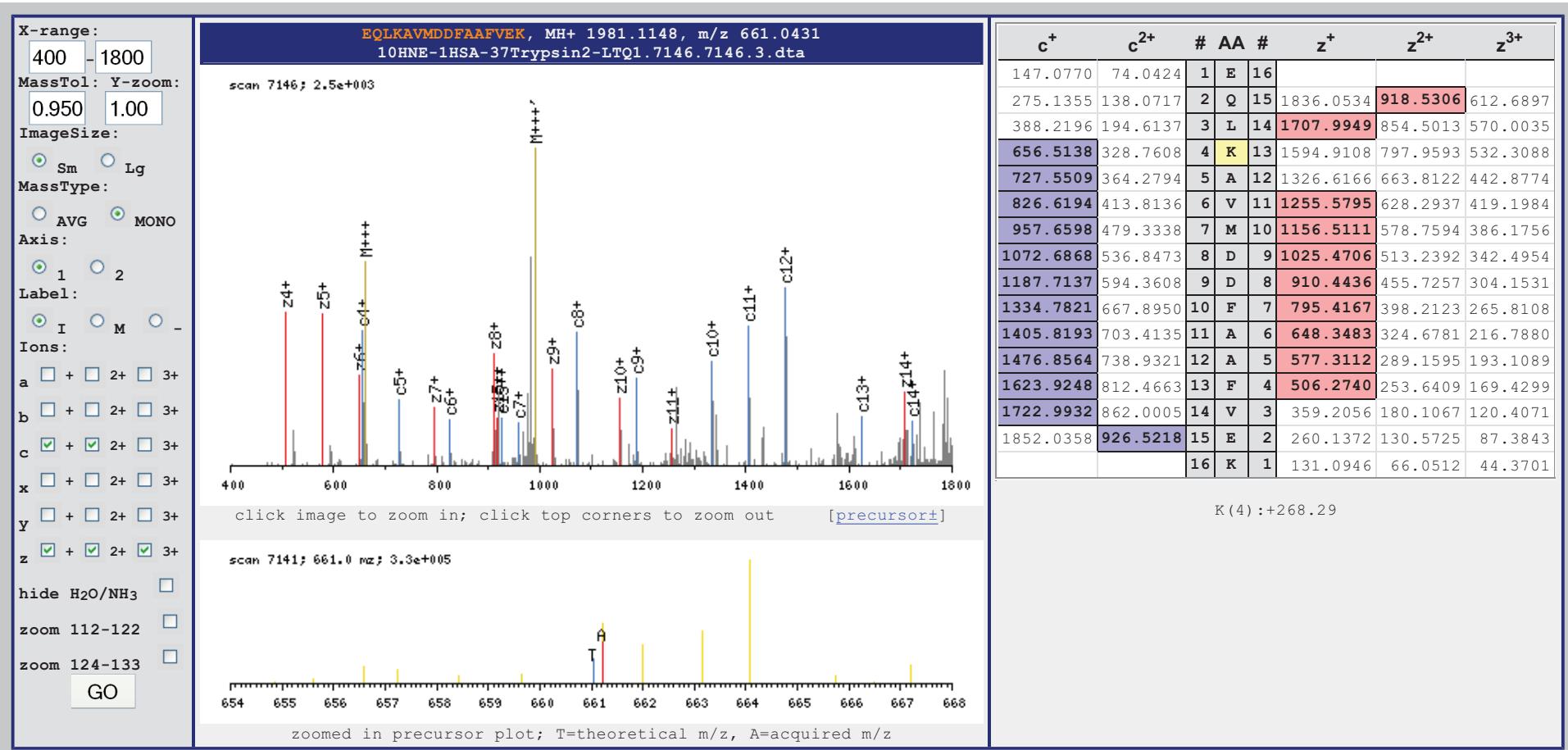
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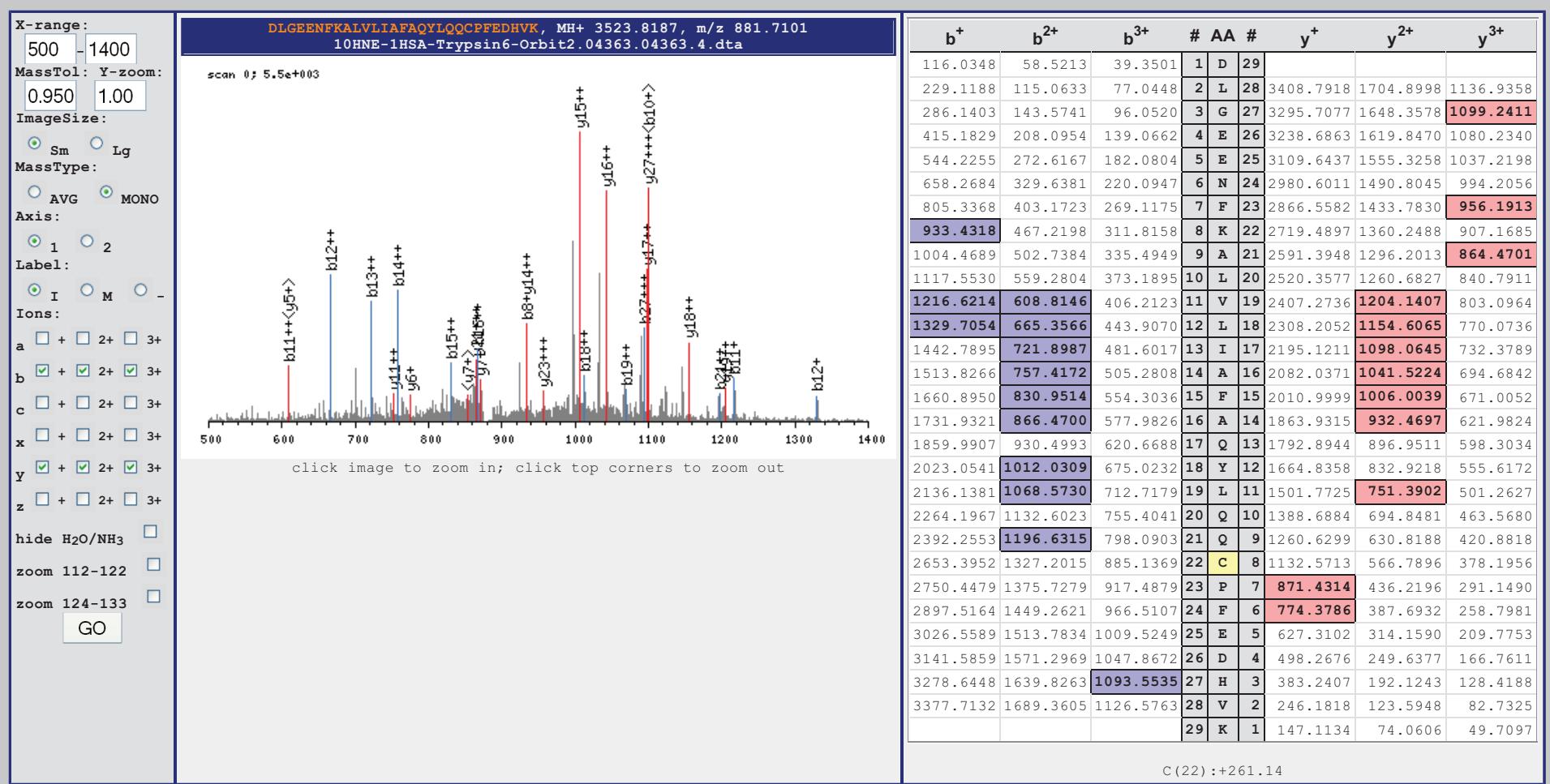
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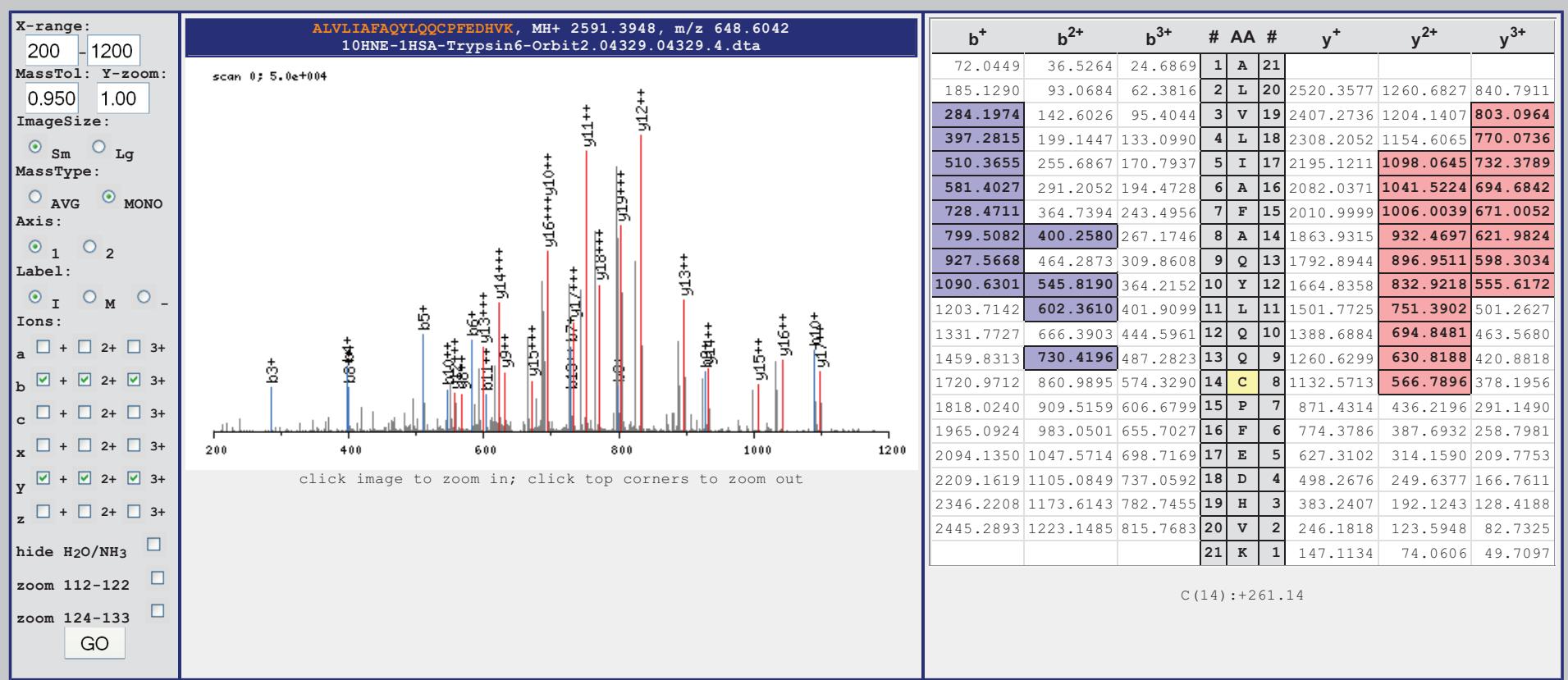
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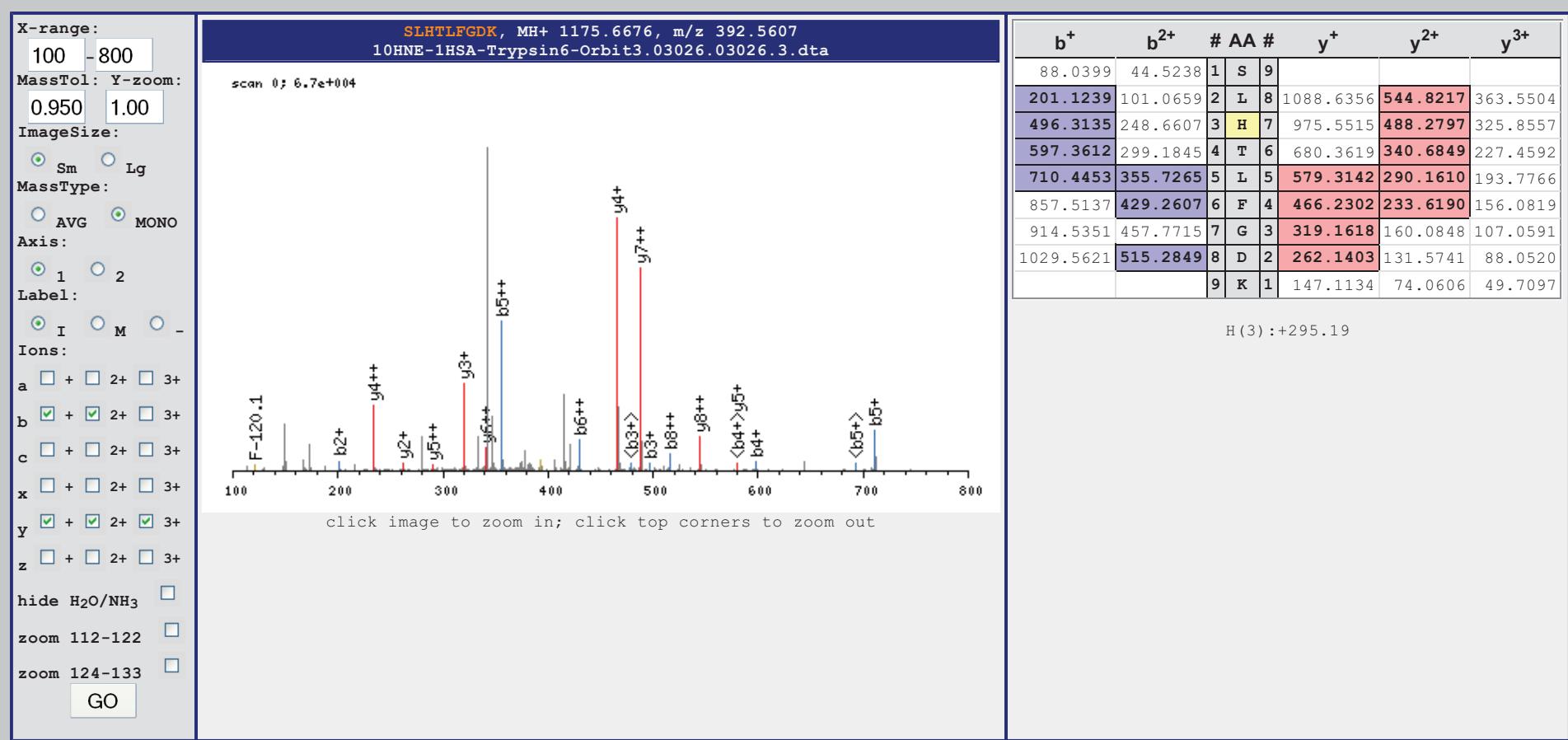
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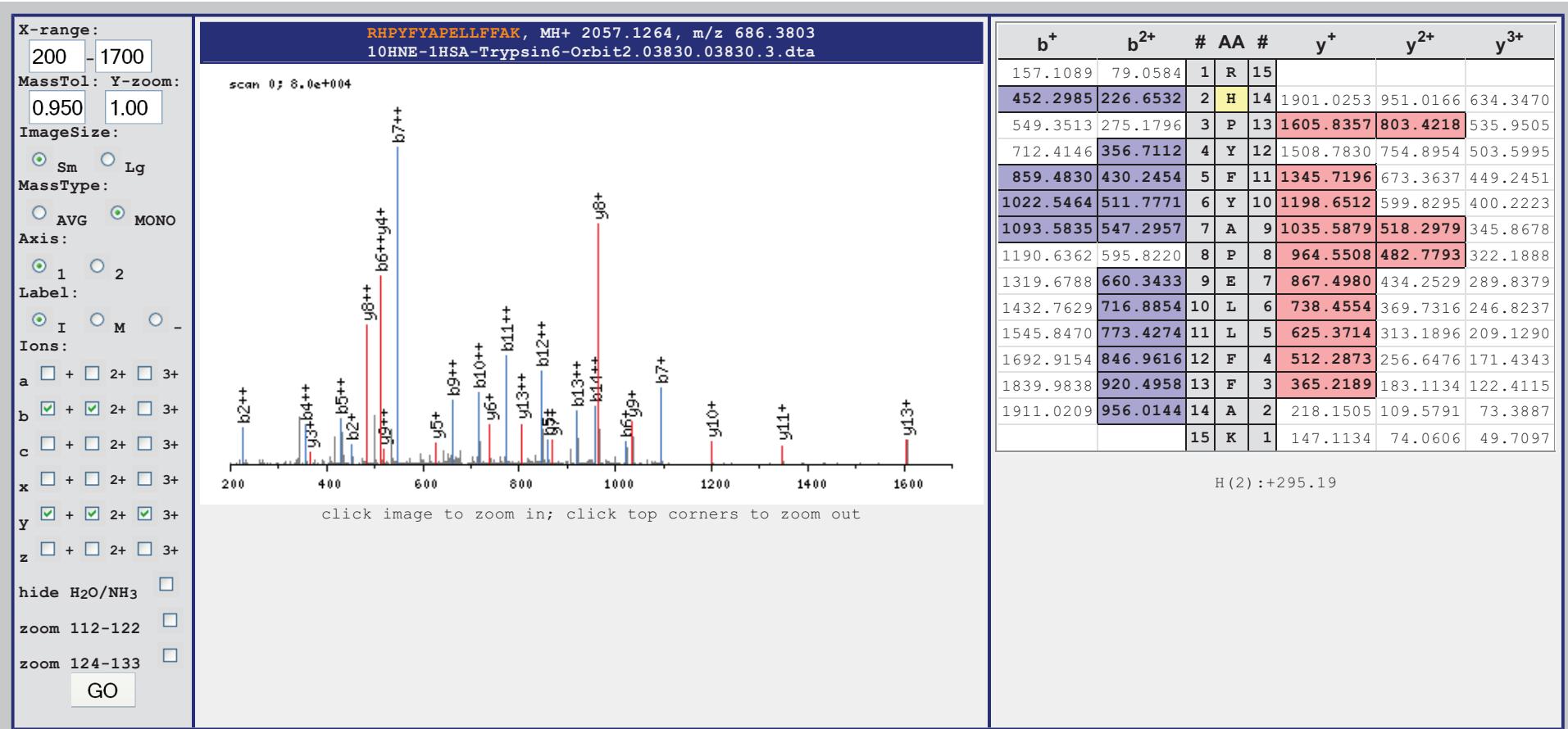
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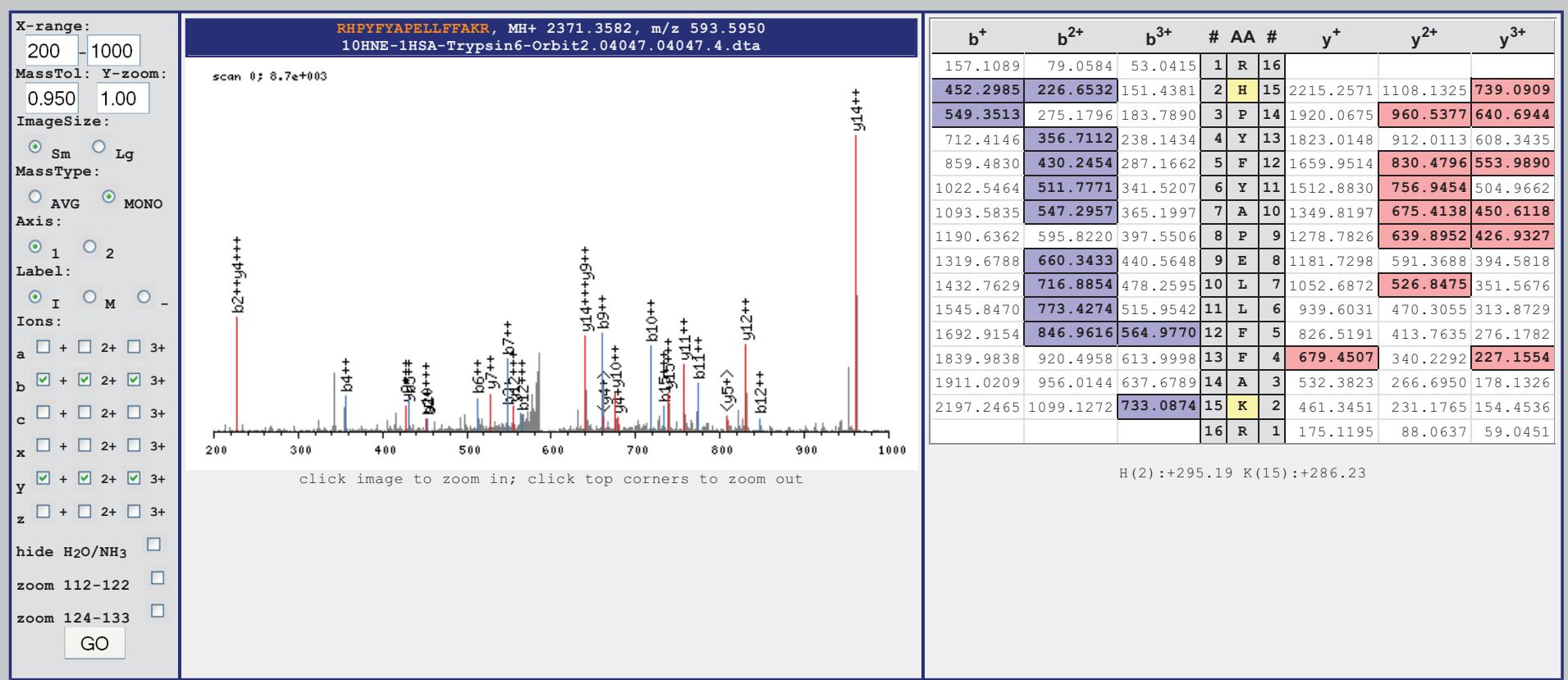
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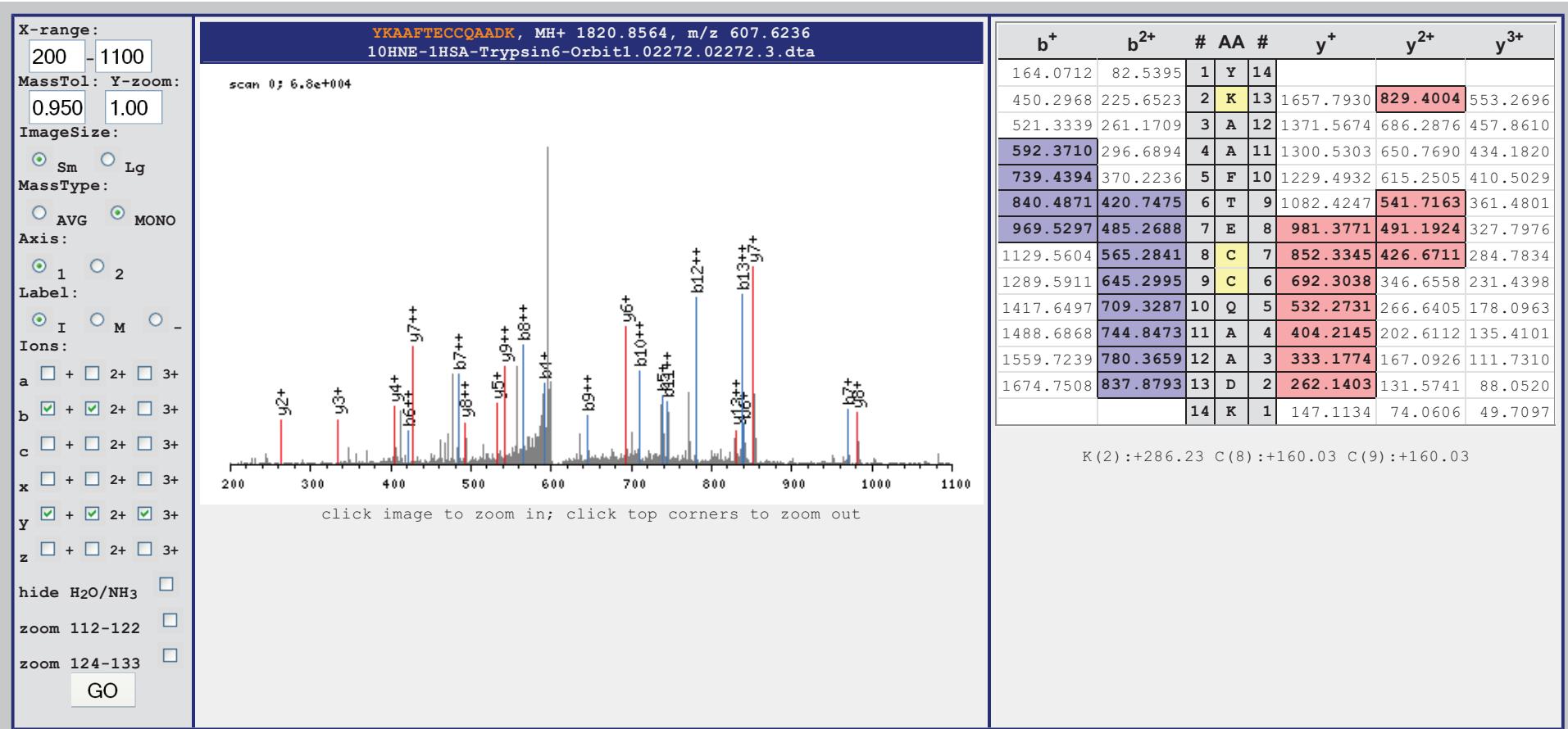
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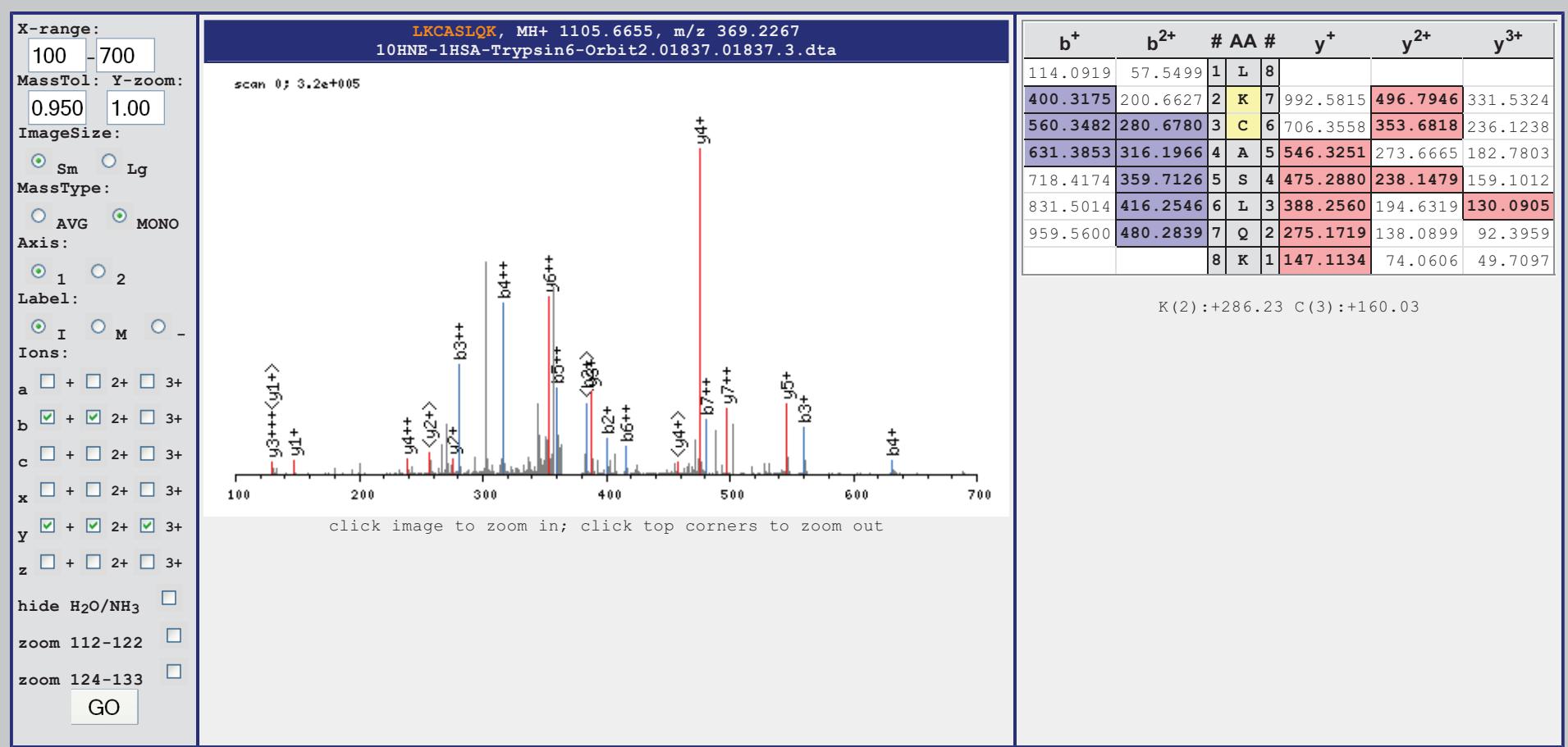
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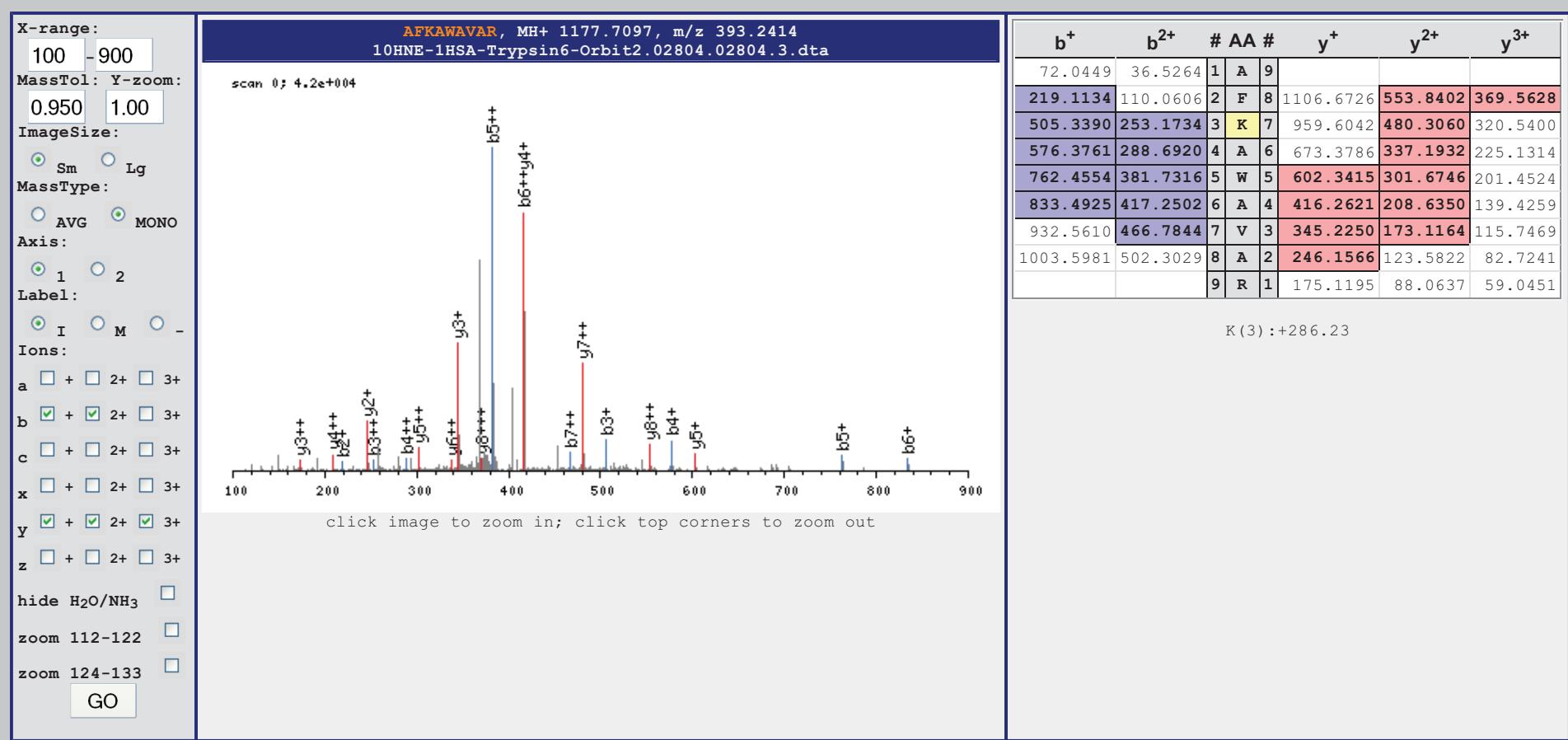
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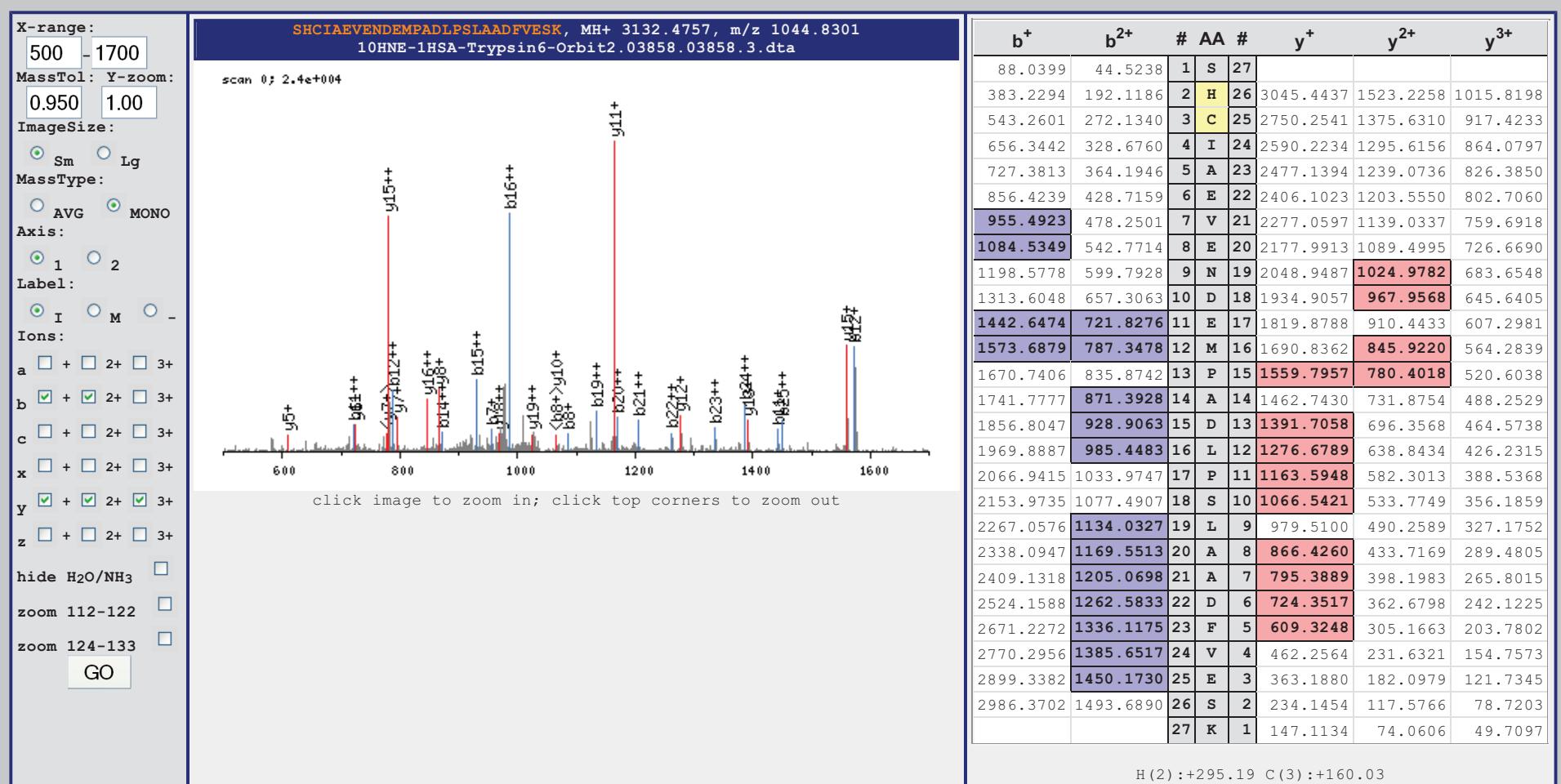
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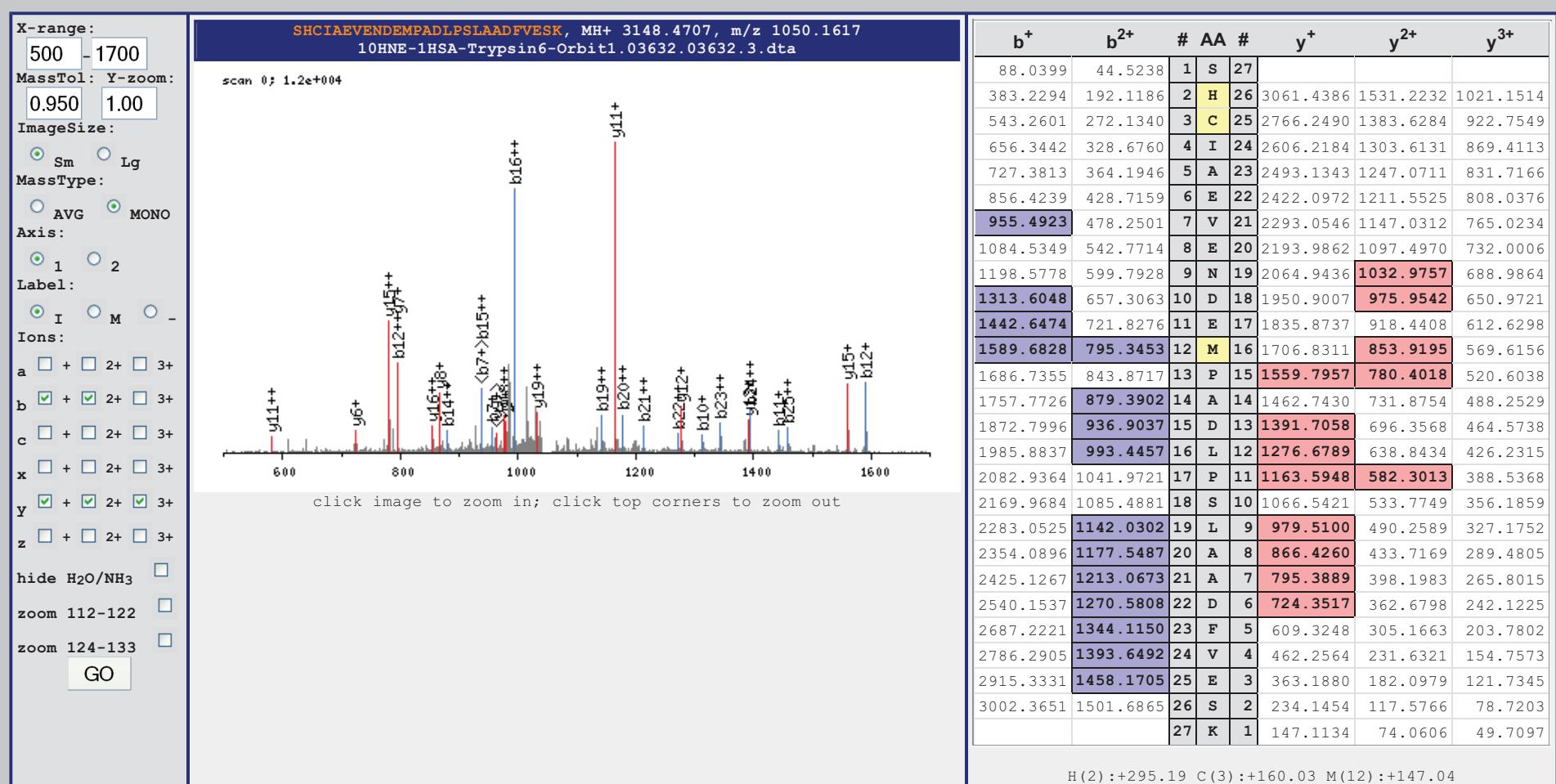
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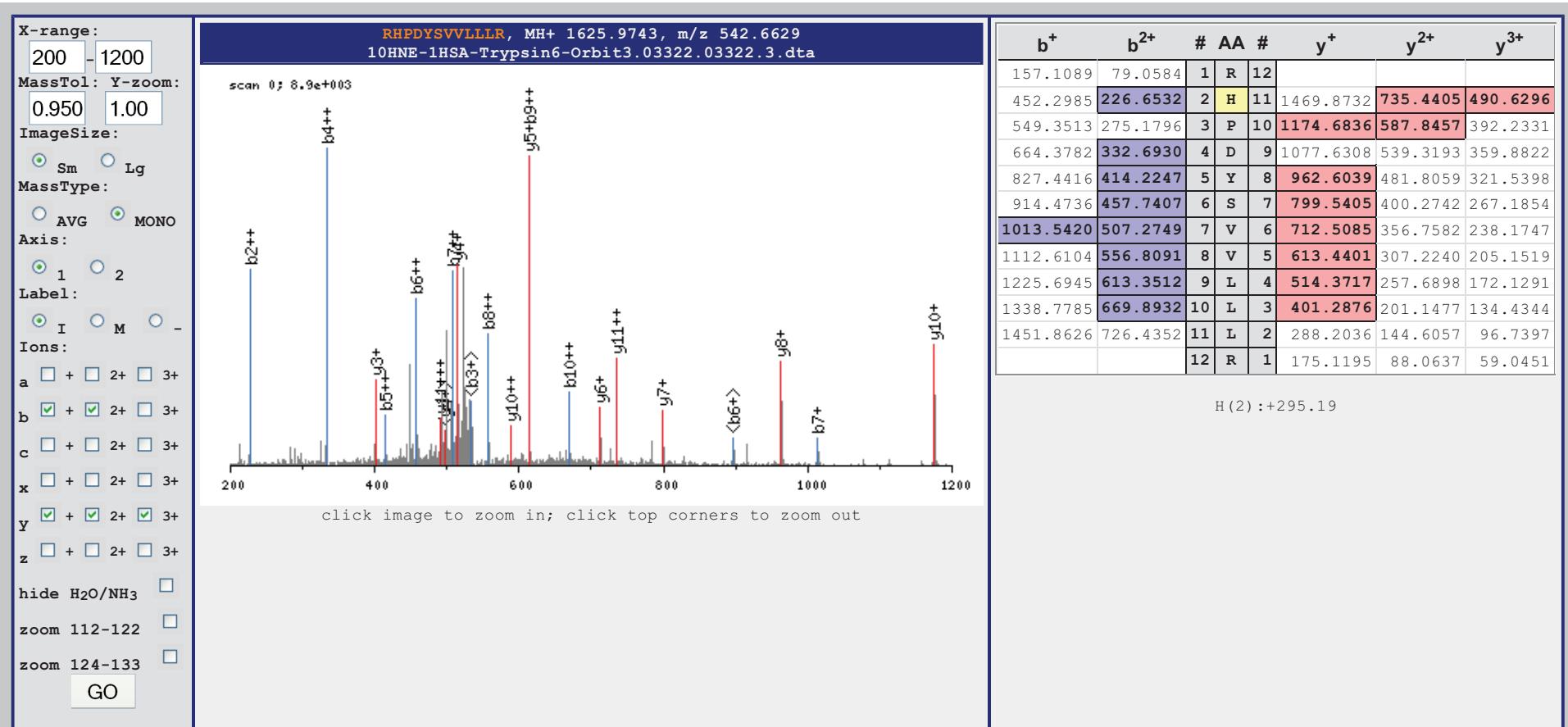
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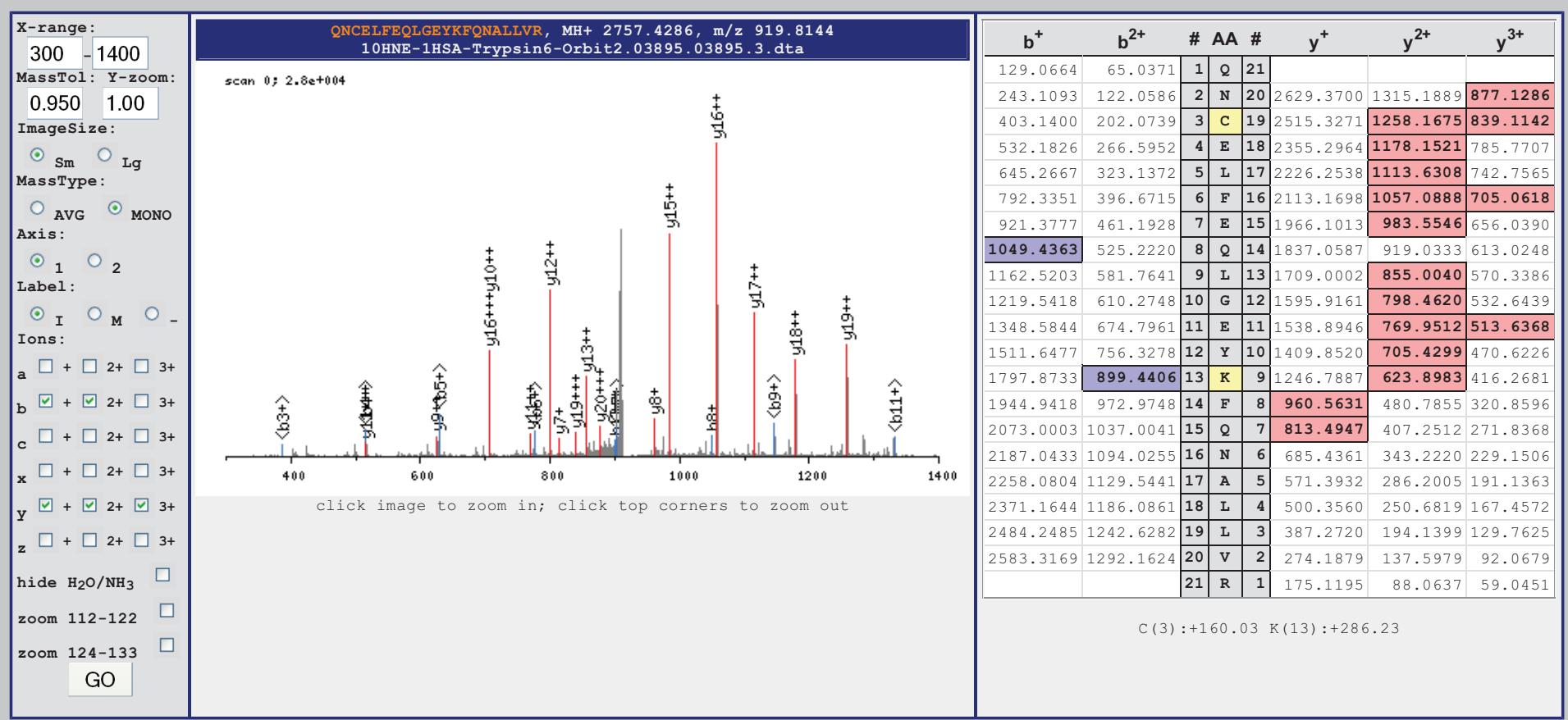
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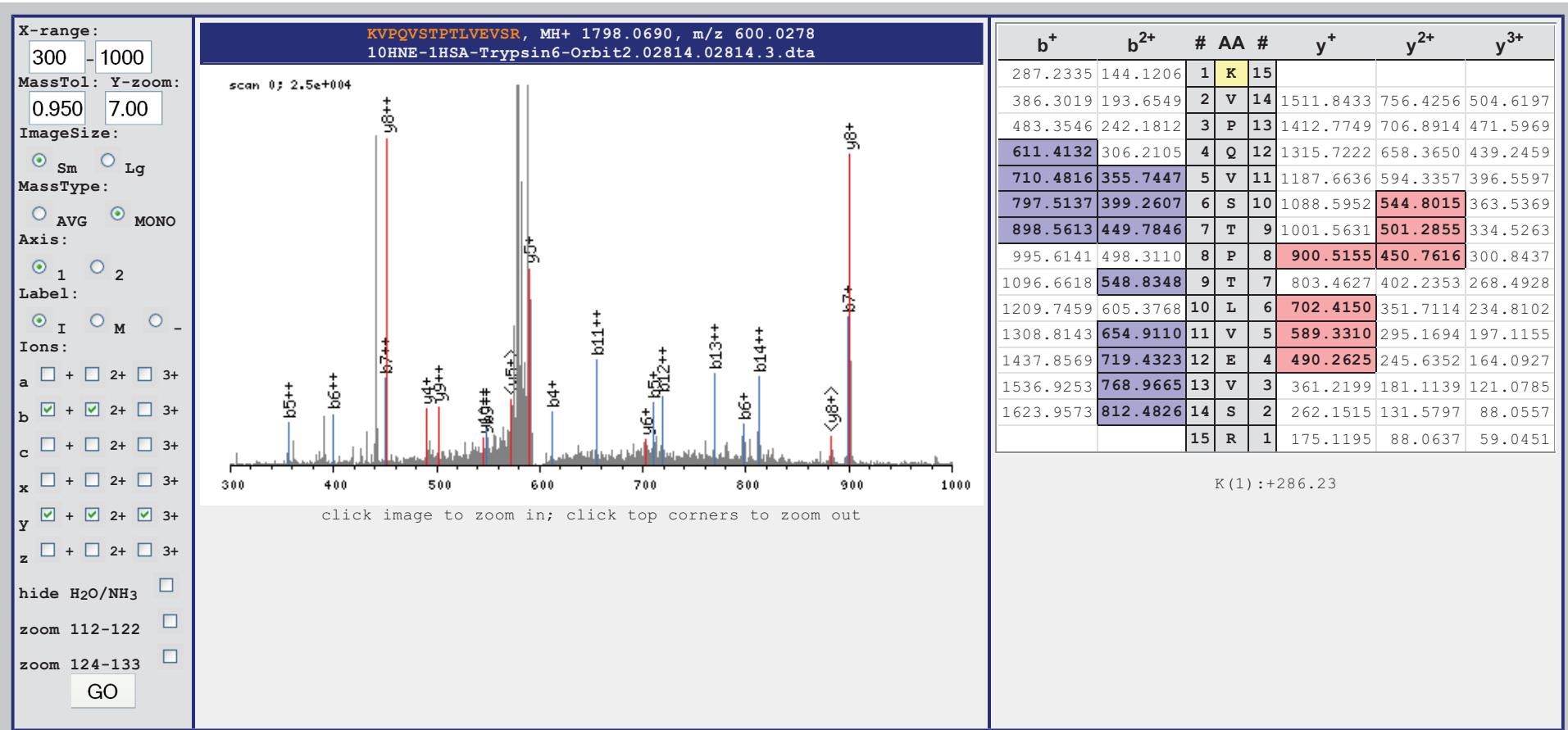
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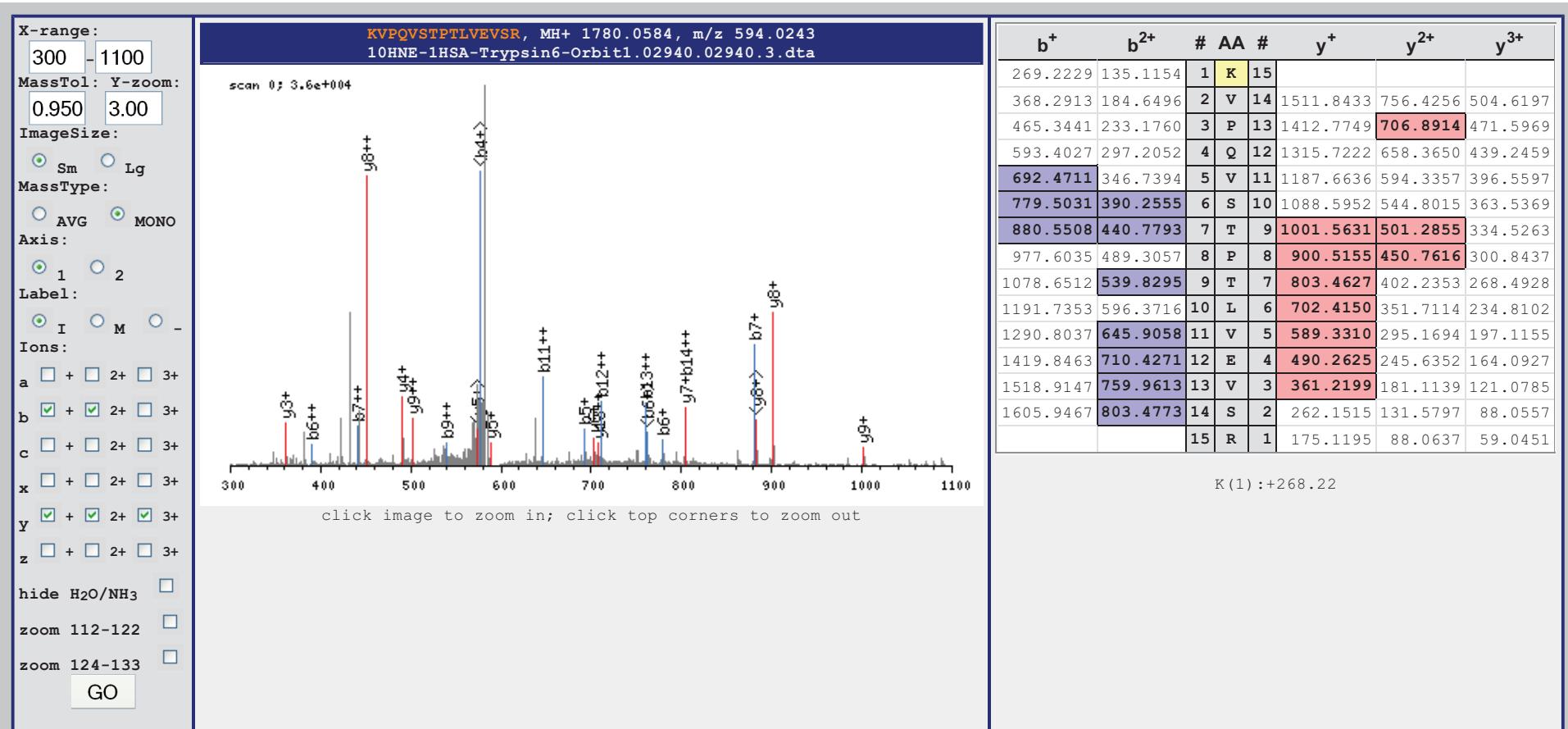
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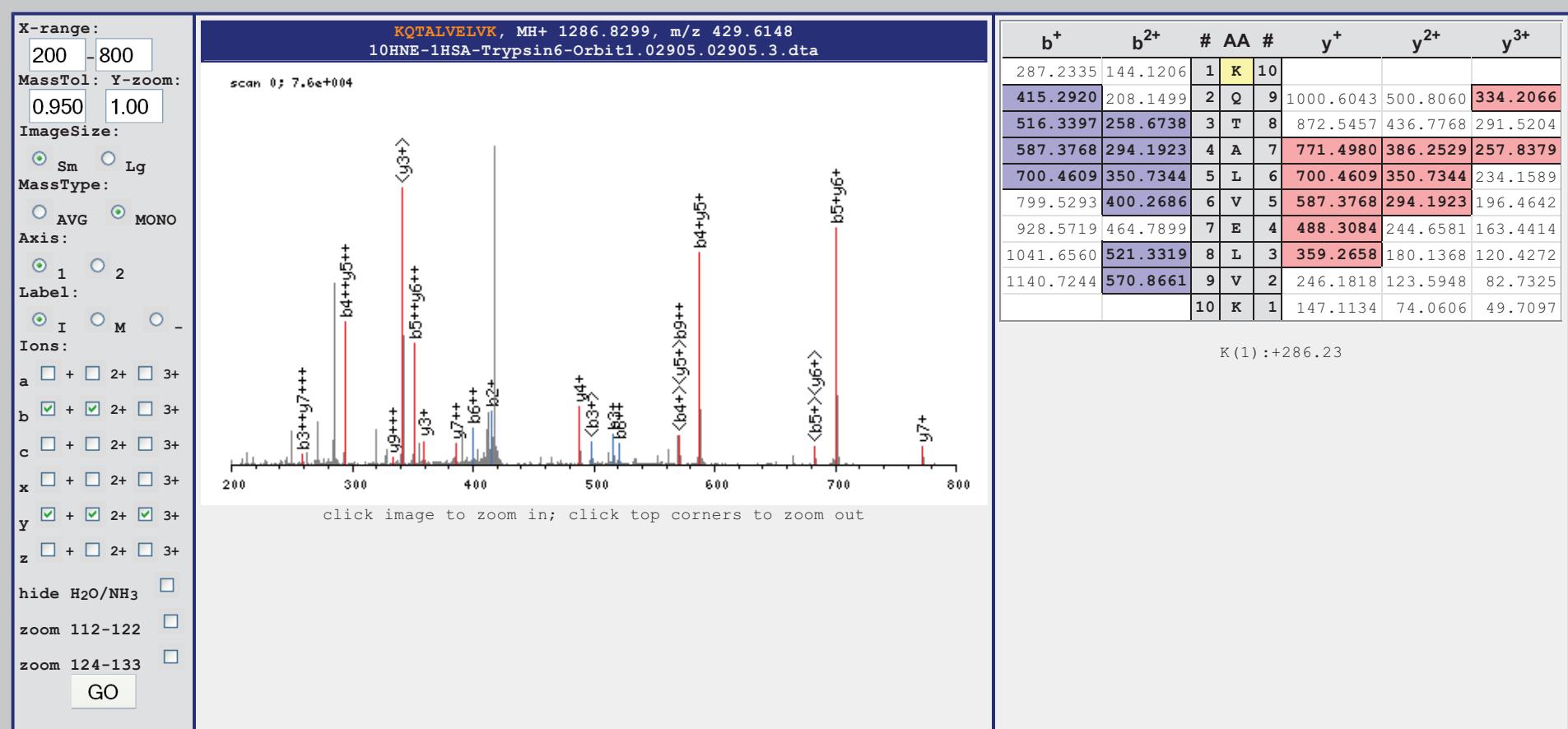
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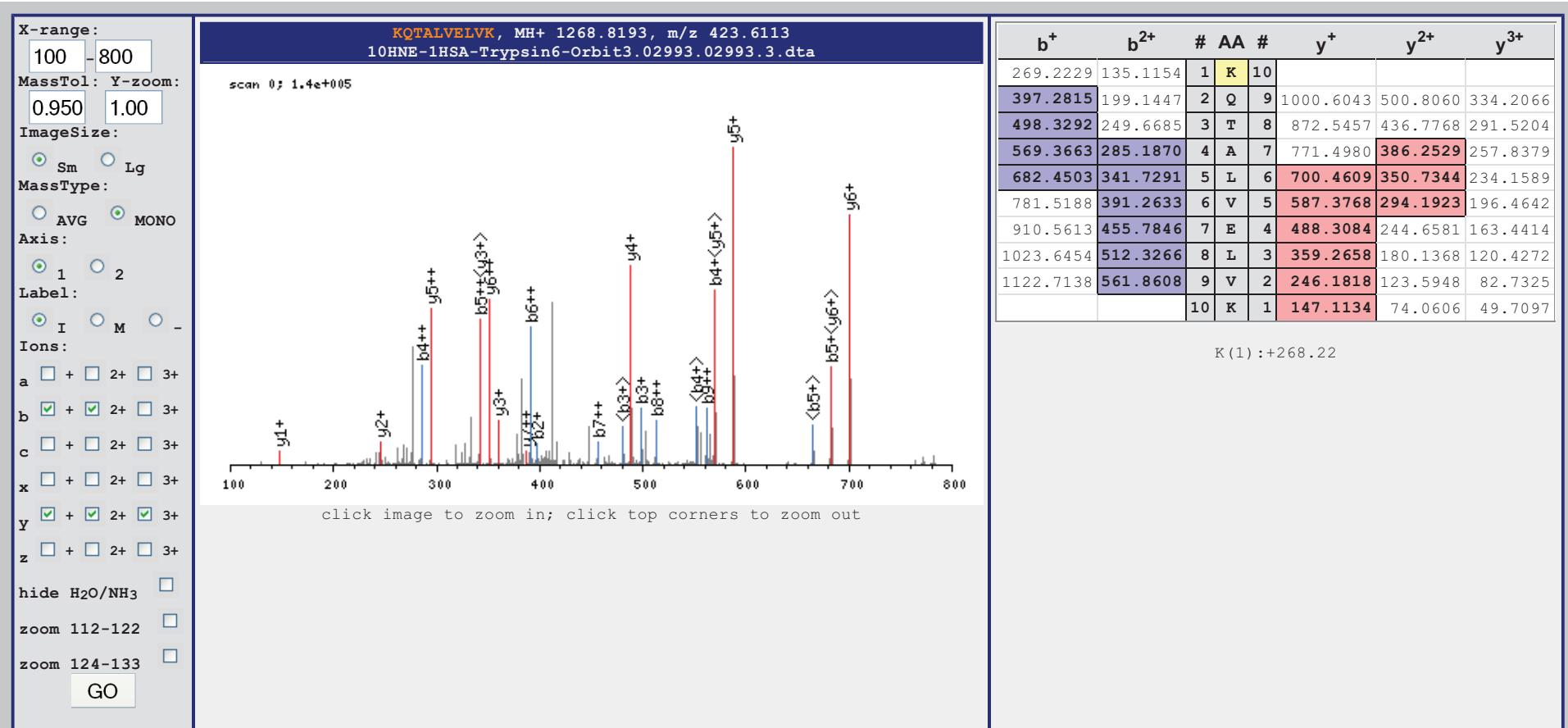
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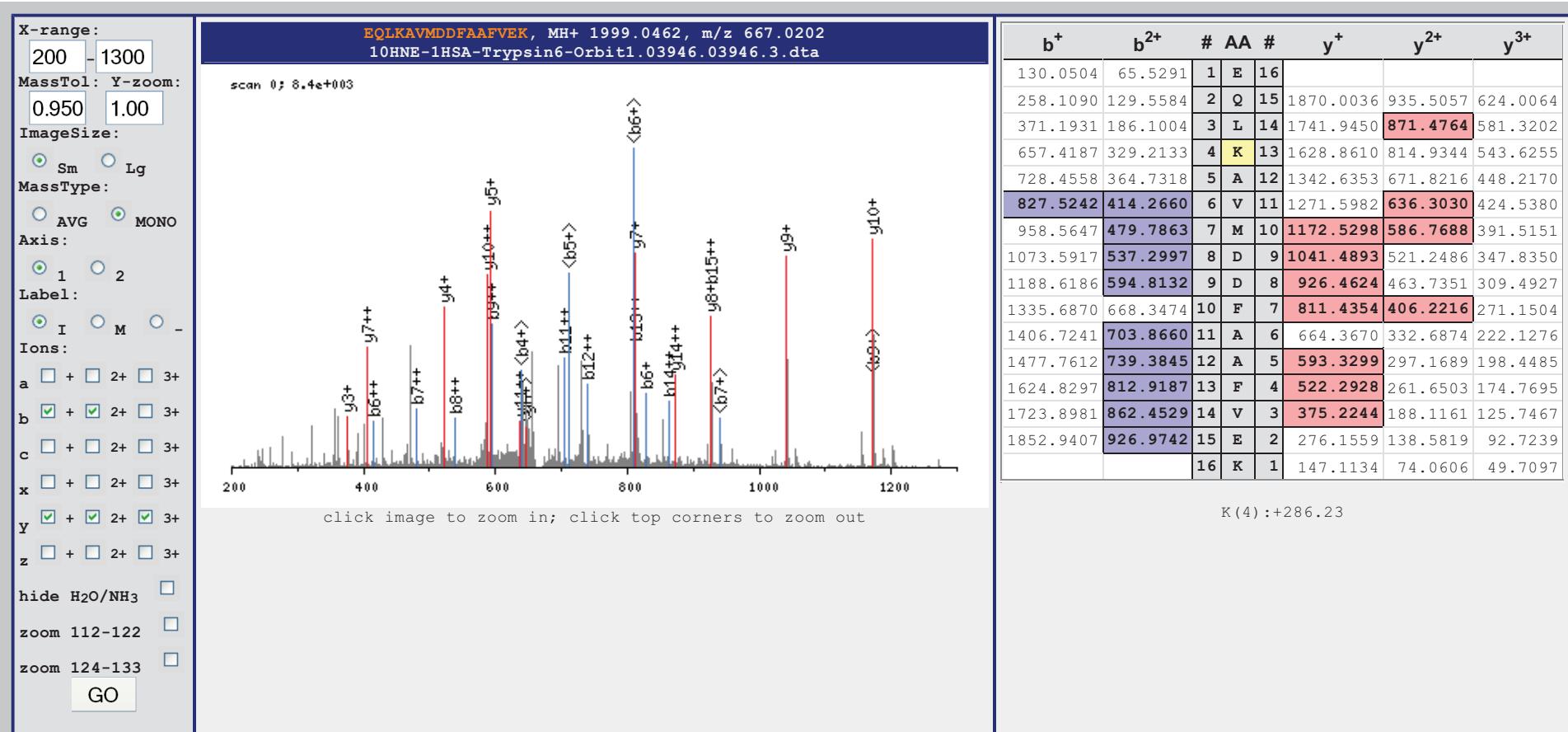
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