O-glycosylation modulates integrin and FGF signaling by influencing the secretion of basement membrane components

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Supplementary Figures S1 to S6 Supplementary Tables S1 and S2 Supplementary Methods



Supplementary Figure S1. The expression of *Galnts* **during SMG development.** (a) Relative expression levels of *Galnt* family members at E12, E13, E14, E15, E17, P1, P5 and adult (Adt) stages, as determined by qPCR are shown. The relative expression levels of each *Galnt* family member are expressed as fold changes in expression compared to E12 SMGs. CT values for each gene at E12 are shown in the top right corner of each graph. (b) Relative expression levels of each *Galnt* family member in E12 SMGs, as determined by qPCR. *Galnt1* is the most abundant isoform expressed at E12.





Supplementary Figure S2. Loss of *Galnt1* has no effect on kidney size at E12. (a) E12 kidneys from *Galnt1*^{+/+} (n=10), *Galnt1*^{+/-} (n=12) and *Galnt1*^{-/-} (n=10) embryos were isolated and measured (white dashed line). (b) The average perimeter of E12 kidneys from each genotype was graphed. No significant difference in kidney perimeter was seen.



Supplementary Figure S3. *Galnt1^{-/-}* SMGs remain smaller than *Galnt1^{+/+}* SMGs throughout development and into adulthood. (a, b) E16 SMGs from *Galnt1^{+/+}* (n=28), *Galnt^{+/-}* (n=25) and *Galnt1^{-/-}* (n=10) were collected and weighed. (c) The ratio of the weights of adult SMGs from male *Galnt1^{+/+}* (n=14), male *Galnt1^{-/-}* (n=6), female *Galnt1^{+/+}* (n=12) and female *Galnt1^{-/-}* (n=4) mice were calculated relative to total body weight. Student's *t*-test was used to calculate *P*-values. *, P<0.05; ***, P<0.001.



Supplementary Figure S4. Loss of *Galnt1* does not affect α 6 integrin receptors present along the basal region of epithelial cells of E12 SMGs. (a) *Galnt1^{+/+}*, *Galnt1^{+/-}* and *Galnt1^{-/-}* E12 SMGs were stained with α 6 integrin (red) to detect integrin receptors and E-cadherin (Ecad, green) as an internal control. The white dashed boxes represent areas of higher magnification that are shown in the last row of images. (b) The ratio of α 6 integrin staining over E-cadherin staining was calculated across 3-4 regions for each SMG. Average ratios were calculated for each genotype and graphed. 4 SMGs of each genotype from 3 independent crosses were examined. Scale bar=10 µm.



Supplementary Figure S5. The effects of disruption of N-linked glycosylation are distinct from those seen upon loss of *Galnt1*. (a) Loss of N-glycosylation after 5 ng ml⁻¹ tunicamycin (TM) treatment results in loss of SMG growth, which cannot be rescued by the addition of BM component Laminin. (b) Loss of N-glycosylation results in dispersed laminin $\alpha 1$ staining (Lam $\alpha 1$, red) and loss of E-cadherin staining (Ecad, green). (c) Loss of N-glycosylation causes disruption of $\alpha 6$ integrin staining (red) in epithelial cells and induction of apoptosis, as examined by caspase-3 staining (Casp3, green). Disruption of N-glycosylation causes ER stress-dependent splicing of Xbp1 (d), increases in ATF4 and the 50kDa cleaved form of ATF6 (p50ATF6) (e), and increases in the expression of ER stress genes (f). Expression was normalized to 29S. M=size markers. H₂O=control reaction without cDNA added. Scale bar=100 µm for a; and 10 µm for b and c. Student's *t*-test was used to calculate *P*-values. *, P<0.05; **, P<0.01.



Supplementary Figure S6. Laminin-111 addition to E12 SMGs. (a) $Galnt1^{+/+}$ (n=11), $Galnt1^{+/-}$ (n=20) and $Galnt1^{-/-}$ (n=9) E12 SMGs were untreated (Control) or treated with 80 µg ml⁻¹ laminin-111 (+ Laminin) for 48 hrs and glands were stained for laminin $\alpha 1$ (red) to demonstrate laminin accumulation along the BM. Intracellular accumulation is still observed in $Galnt1^{-/-}$ and $Galnt1^{+/-}$ SMG epithelial cells. (b) Expression of specific ER stress genes in $Galnt1^{-/-}$, $Galnt1^{+/-}$ and $Galnt1^{+/+}$ SMGs that were untreated (-) or treated with laminin-111 (+), as determined by qPCR. Expression was normalized to 29S. Scale bar = 10µm. Student's *t*-test was used to calculate *P*-values. *, P<0.05.

Gene	Primers
Galnt1	Sense: 5'- TCATCAAGAGCAGCGGCAAAGC -3'
	Anti-sense: 5'- ACAAGGCACATTCAGCAGAAACGG -3'
Galnt2	Sense: 5'- CGCCCTCTGCCTCCCTCTTTC -3'
	Anti-sense: 5'- TGATTGCTGCTTGCCCACTTGTTC -3'
Galnt3	Sense: 5'- TGCTACTCAGGGTGTCGTCCAG -3'
	Anti-sense: 5'- GCGTCACATGGCACTAAGTTTGG -3'
Galnt4	Sense: 5'- CCGCAATCGTATGTCCTGTCATCG -3'
	Anti-sense: 5'- AACGCCAGTCAAACCCACCAATC -3'
Galnt5	Sense: 5'- GCCGAGCAGAGATGGAAAGAAGG -3'
	Anti-sense: 5'- CTGGTGGTTGGGAGGTCATTGTG -3'
Galnt6	Sense: 5'- GTGTTGACCAGAAGTTCCG -3'
	Anti-sense: 5'- GATTTCATTCAGCAAGATGGC -3'
Galnt7	Sense: 5'- GGCTCGTGGTCCTCTGGTCTTC -3'
	Anti-sense: 5'- TCTCTGTCTTCCCTCATCCTGCTC -3'
Galnt10	Sense: 5'- CCGAGGCGAGGCTGCTTGG -3'
	Anti-sense: 5'- GGGTGACTGGGCTGGTGTGG -3'
Galnt11	Sense: 5'- CAGCAGTGGACCTTTGGGAAGAAC -3'
	Anti-sense: 5'- TGTTGAGAGGAGGAGCCATCGC -3'
Galnt12	Sense: 5'- CCGAGAGACCGTCCCAGAGAAC -3'
	Anti-sense: 5'- ACATTTCCTGCTGTGCTTGTGAAC -3'
Galnt13	Sense: 5'- CACCCGTCTTCAGTCTCCGTATTG -3'
	Anti-sense: 5'- GACATCAACAAGCACCCACATCAG -3'
Galnt14	Sense: 5'- GATGAGCGGCGGTATCTGAATGC -3'
	Anti-sense: 5'- GGTGATGATGATGCTGGTGTGAGG -3'
Galnt15	Sense: 5'- GGACTGGAGGACCGAAGAGGATG -3'
	Anti-sense: 5'- AGAGGATGACGCTGGCTGTAGG -3'
Galnt16	Sense: 5'- CGCCAATGCCATCGCCATCC -3'
	Anti-sense: 5'- GCTCGGTTGTCCTGCCATAAGTAG -3'
Galnt18	Sense: 5'- CCTGCCCTGCTCTCGGATTGC -3'
	Anti-sense: 5'- GCCTTGCGTGCGGTGATGTC -3'
Galnt19	Sense: 5'- TGGGTGTATGTTTGCGTGCTTGAG -3'
	Anti-sense: 5'- GCGTCCTTGTCCCTATCCACTGAG -3'

Supplementary Table S1. Mouse *Galnt* gene primer sequence for qPCR

Gene	Primers
Xbp1	Sense: 5'- GACGAGGTTCCAGAGGTG -3'
	Anti-sense: 5'- GAGGCAACAGTGTCAGAGT -3'
Canx	Sense: 5'- ACTGGTGGTGGTCATTGCCTTT -3'
	Anti-sense: 5'- GCACACGCTTAGCAGTTCTACCT -3'
Chop	Sense: 5'- CCTCGCTCTCCAGATTCCA -3'
	Anti-sense: 5'- GCCGCTCGTTCTCTCAG -3'
Edem1	Sense: 5'- GCTGGTCAGAGGGTAGGG -3'
	Anti-sense: 5'- GCAGGGAAGAGGCACTAGA -3'

Supplementary Table S2. Mouse ER stress gene primer sequence for qPCR

Supplementary Methods

Kidney size measurement. Kidneys were isolated from E12 *Galnt1^{+/+}*, *Galnt1^{+/-}*, *Galnt1^{-/-}* embryos and perimeters were measured using NIH ImageJ software.

E16 and adult SMG size measurements. E16 SMGs were dissected from $Galnt1^{+/+}$, $Galnt1^{+/-}$, $Galnt1^{-/-}$ embryos and weighted. Adult SMGs were isolated from 2-3 month old male and female $Galnt1^{+/+}$ and $Galnt1^{-/-}$ mice. Both SMG weight and total body weight were determined for each individual. Ratios of SMG weight/body weight were calculated and averaged to obtain the values graphed in Supplementary Fig. S3c.

Ex vivo SMG organ culture with Tunicamycin (TM). E12 SMGs were dissected from ICR time-pregnant mice and cultured as described in the main text. 5 ng ml⁻¹ TM (T7765, Sigma-Aldrich; 5 μ g ml⁻¹ stock solution in DMSO) was added to the culture medium and explants were cultured for 2 days⁵³. SMGs were photographed and harvested for qPCR and immunostaining as described in the main text. Cleaved caspase-3 (Asp175) antibody (#9661, Cell Signaling Technology; 1:100) was used to detect apoptosis by immunostaining.