

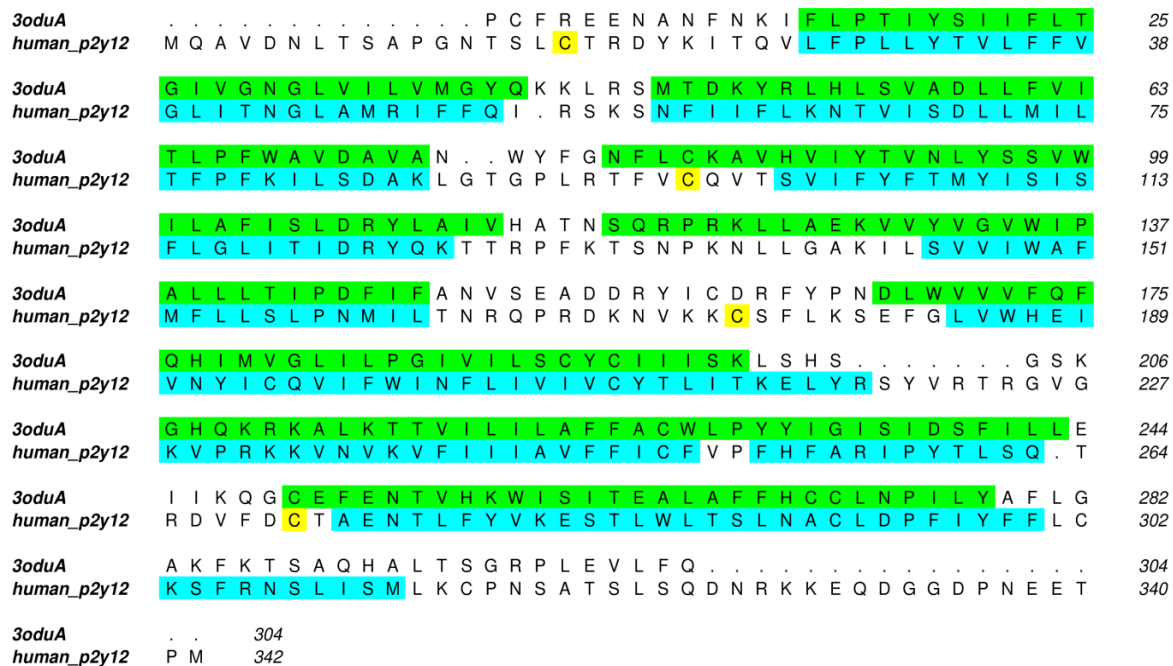
Supplementary Information – Molecular Pharmacology

Identification of determinants required for agonistic and inverse agonistic ligand properties at the ADP receptor P2Y₁₂

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Figure S1



Suppl. Figure S1. Sequence alignment of P2Y₁₂ with CXCR4.

The sequence of human P2Y₁₂ was aligned with the sequence of human CXCR4 (PDB ID: 3ODU) (Gupta et al., 2001) using CLUSTALW (Larkin et al., 2007). Transmembrane helical regions of the CXCR4 receptor are highlighted in green. Regions predicted to be transmembrane helical regions of P2Y₁₂ according to secondary structure prediction server PSIPRED (McGuffin et al., 2000) are highlighted in cyan. Cysteine residues known to form disulphide bonds are highlighted in yellow.

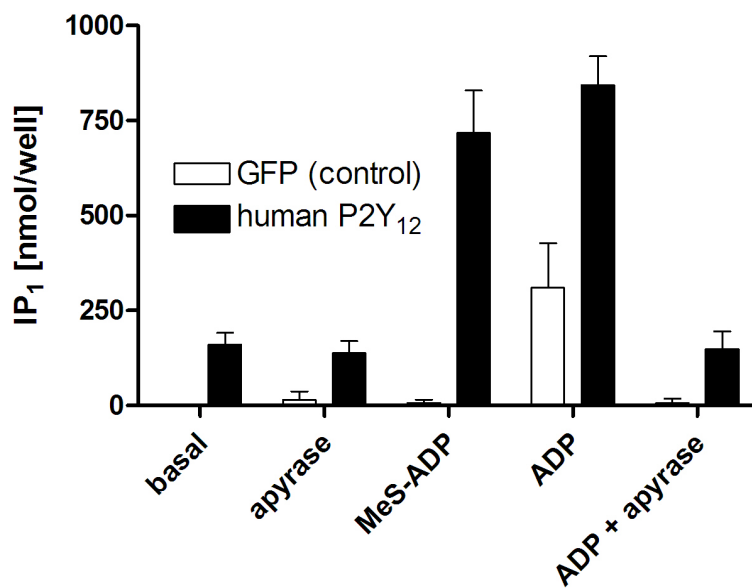
Figure S2



Suppl. Figure S2. Per residue change in free energy with and without ATP derivatives bound.

Rosetta was used to determine the difference in change in free energy ($\Delta\Delta G$) between P2Y₁₂ with and without ATP derivatives bound. Residues that demonstrated the largest difference in free energy change in respect to bound ADP, ATP, MeS-ADP, MeS-ATP, mant-ATP, mant-ADP, mant-dADP and mant-dATP are listed. For each ligand, residues with $\Delta\Delta G$ above the average energy change (-0.24) are in red, while those with energy change above average but still negative are in yellow and residues with a positive energy change are in green.

Figure S3



Suppl. Figure S3. Analysis of the functional impact of endogenous nucleotide released from CHO cells

To analyze whether endogenous nucleotide released from CHO cells contribute to basal activity of P2Y₁₂ we performed control experiments with CHO-K1 cells stably transfected with G α_{q14} . Intracellular inositol phosphate (IP₁) levels were determined with an immunological assay (cisbio Bioassays, IP-One ELISA, part-no. 72IP1PEA).

G α_{q14} -CHO-K1 cells transiently transfected with P2Y₁₂ presented an increased basal IP₁ level compared to cells transfected with the control plasmid (GFP). Incubation with 12.5 U/ml apyrase did not reduce this elevated IP₁ level. This clearly indicates that P2Y₁₂ does induce signal transduction by intrinsic active receptor conformation and not by nucleotides released from the cells into the medium. Proper P2Y₁₂ transfection was control by application of 100 μ M ADP and 100 μ M MeS-ADP. Proper apyrase function was demonstrated by loss of ADP action on P2Y₁₂.

Table

Suppl. Table S1. Purine compound library screening at the human WT P2Y₁₂

The activity of compounds (given in %) at the human P2Y₁₂ expressed in yeast is shown relative to the basal activity (OD_{600 nm} =0.089; set 0%) and the stimulation with 10 μM MeS-ADP (OD_{600 nm} =0.667; set 100%). Stimulation was measured after 24 h with 10 μM of the respective compound.

AMP -2.19	ADP 84.05	ATP 19.93	AP4 -0.29	cAMP -0.99
3'-dATP -2.02	7-Deaza-dAMP -1.33	NPE-caged-ATP -0.64	7-Deaza-dATP -1.33	DMB-caged-ATP 5.24
dATPαS -1.68	ADPβS 92	ATPγS 71.09	β-Methylene-APS -1.68	ApCp -0.81
AP3A -0.99	AP4A -1.5	AP5A -1.68	AP6A -1.68	AP4U -0.99
AP4(8I)G -2.37	AP5(8I)G -1.68	2'I-AMP -1.33	2'I-ADP -1.68	2'I-ATP -1.16
8Br-ADP 0.57	8Br-ATP -1.68	8Br-dATP -0.99	γ-[6-Aminoethyl]- ATP -1.5	2I-ATPγS 90.1
7-Deaza-7Br-dATP - 2.02	γ-Aminophenyl-ATP 2.99	γ-[(6-Aminoethyl)- imido]-ATP -1.16	γ-[(8-Aminooctyl)- imido]-ATP -1.16	N6-(4-Amino)butyl- ATP -1.33
EDA-ATP -3.75	γ-[6-Aminoethyl]- N6-Benzyl-ATP -2.02	2-Hydroxy-ATP -1.5	TNP-ADP 72.81	TNP-ATP 4.03
1-Methyl-AMP -0.99	1-Methyl-ADP 1.61	1-Methyl-ATP 0.05	dATP 0.57	ddATP 0.57
2'.5'-pAp -0.47	ara-ATP 0.4	AMPαS -0.12	ATPαS 2.3	dADPαS 0.4
ApCpp -1.33	AppCp 0.4	dApCpp -0.47	AppNp 0.57	AppNH2 76.79
AP5U -1.16	AP4T -0.29	AP5T -0.47	AP4G 0.22	AP5G 0.05
2'Br-ADP -1.16	2'Br-ATP -0.64	2'-Ome-ATP -0.81	mante-ATPγS -0.4	8Br-cAMP 0.05
2'F-AMP -0.81	2'F-ATP -0.47	2'Cl-ATP -0.81	BzBzATP -0.12	7-Deaza-7I-dATP -0.12
N6-(6-Amino)hexyl- ATP -0.81	N6-(6-Amino)hexyl- dATP -0.12	8-[(4-Amino)butyl]- amino-ATP -0.12	8-[(6-Amino)hexyl]- amino-ATP 0.57	EDA-ADP 1.95
mant-ADP 6.79	mant-ATP -0.47	mant-dATP -0.64	mant-N6-Methyl- ATP -0.29	ε-ATP 0.4
Adenine 0.6	Adenosine 0.75	IMP 0.15	UDP 2.54	Xanthine 0.75
GDP 1.94	GTP 0.75	GTPγS 0.45		