

Supplementary Tables

Accurate Variant Detection across Non-amplified and Whole Genome Amplified DNA Using Targeted Next Generation Sequencing

Abdou ElSharawy¹, Jason Warner², Jeff Olson², Michael Forster¹, Markus B. Schilhabel¹, Darren Link², Stefan Rose-John³, Stefan Schreiber^{1,4}, Philip Rosenstiel¹, James Brayer² and Andre Franke^{1,*}

¹ Institute of Clinical Molecular Biology, Christian-Albrechts-University, Kiel, Germany

² RainDance Technologies, Inc. Lexington, Massachusetts, U.S.A.

³ Institute of Biochemistry, Christian-Albrechts-University, Kiel, Germany

⁴ First Medical Clinic, University Hospital, Schleswig-Holstein, Kiel, Germany

Email addresses

AE: a.sharawy@mucosa.de

JW: WARNERJ@raindancetech.com

JO: OLSONJ@raindancetech.com

MF: m.forster@ikmb.uni-kiel.de

MS: m.schilhabel@ikmb.uni-kiel.de

DL: dlink@raindancetechnologies.com

SR: rosejohn@biochem.uni-kiel.de

SS: s.schreiber@mucosa.de

PR: p.rosenstiel@mucosa.de

JP: brayerj@raindancetech.com

AF: a.franke@mucosa.de

Supplementary Table 1: Overview of Sample Processing: RainDance Sequence Enrichment, SOLiD Sequencing Library Construction and Sample Indexing, Emulsion PCR and Sequencing

Sample ID	HapMap Sample_ID	Family ID	Library ID	Bar Code	Emulsion ID	Sequencing ID
1	NA12003 (gDNA)	1420	759L	1	759em	759seq
2	NA12004 (gDNA)	1420	760L	2	760em	760seq
3	NA10838 (gDNA)	1420	761L	3	761em	761seq
4	NA11829 (gDNA)	1350	762L	4	762em	762seq
5	NA11830 (gDNA)	1350	763L	5	763em	763seq
6	NA10856 (gDNA)	1350	764L	6	764em	764seq
1-6_1	<i>6 DNAs pooled before library preparation</i>	1350 and 1420	768_1L	-	768_1Lem	768_1Lseq
1-6_2	<i>6 DNAs pooled before library preparation*</i>	1350 and 1420	768_2L	-	768_2Lem	768_2Lseq
7	<i>Indexed Sample: pooled before emPCR (pB)</i>	1350 and 1420	770L	1-4 & 6	770em	770seq
8	<i>Indexed samples: pooled after emPCR (pA)</i>	1350 and 1420	792L	1-6	792em	792seq
9	NA12003 (WGA gDNA)	1420	765L	9	765em	765seq
10	NA12004 (WGA gDNA)	1420	766L	10	766em	766seq
11	NA10838 (WGA gDNA)	1420	767L	11	767em	767seq

* Technical replicate of library 786_1.

- Boldfaced italic words indicate pooled samples/libraries.

- gDNA: genomic DNA; WGA: whole-genome amplification; emPCR: emulsion PCR.

Supplementary Table 2: RainDance Technologies 384 Member Primer Panel - gff

(A separate Excel sheet)

Supplementary Table 3: CLC Bio SNP Detection Parameters

Add conflict annotations	No
Alignment mode	local
Amplicon annotation	
Annotate consensus sequence	Yes
Annotate reference sequence	Yes
Colospace alignment	Yes
Colospace Alignment	Yes
Colospace error cost	3
Colospace Error Cost	3
Conflict resolution Vote	
Create Report	Yes
Create SequenceList	Yes
Create table	Yes
Deletion Cost	3
Genetic code translation	Standard
Guidance only	No
Insertion cost	3
Insertion Cost	3
Length	0.5
Mask reference sequence	
Match mode random	
Max alignments	1
Max distance	250
Maximum coverage	50000
Maximum expected variations (ploidy)	2
Maximum gap and mismatch count	2
Min distance	180

Minimum average quality	15
Minimum central quality	20
Minimum coverage	5
Minimum paired-end coverage	0
Minimum variant frequency (%)	10.0
Mismatch cost	2
Mismatch Cost	2
Read settings	
Reference sequences	chr21, chr22, chrY, chr20, chr19, chr18, chr17, chr16, chr15, chr14, chr13, chr12, chr11, chr10, chr9, chr8, chrX, chr7, chr6, chr5, chr4, chr3, chr2, chr1
Report type	1x, 5x, 10x, 20x, 40x, 80x, 100x
Score limit	8
Sequence masking	No
Similarity	0.8
Ungapped alignment	Yes
Use annotated amplicons	No
Variant count threshold	50
Window length	11

Supplementary Table 4: Coverage Metrics CLC bio Genomics Workbench (version 5.1)

HapMap	Sample Type	Library ID	Reads	Mapped		On-Target %		ADoC	C1	C10	C20	C30	C50	C100	Coverage
Sample ID															0.2X Mean
NA12003	gDNA	759L	36,453,208	19,290,598	52.92%	8,577,722	44.47%	2309.47	99.43%	98.47%	98.15%	97.92%	97.39%	96.24%	87.49%
	WGA	765L	36,986,943	20,508,825	55.45%	9,361,663	45.65%	2541.34	99.41%	98.56%	98.15%	97.83%	97.34%	96.54%	86.97%
	gDNA-pB	770L_BC1	10,031,809	5,627,776	56.10%	2,489,710	44.24%	671.32	98.82%	97.75%	96.79%	96.16%	94.66%	90.56%	87.63%
	gDNA-pA	792L_BC1	5,221,822	2,678,148	51.29%	1,142,288	42.65%	302.78	98.37%	96.43%	94.70%	92.87%	89.08%	77.89%	87.03%
NA12004	gDNA	760L	39,005,646	19,804,791	50.77%	9,852,432	49.75%	2606.79	99.25%	98.37%	98.03%	97.76%	97.37%	96.35%	87.66%
	WGA	766L	43,079,257	16,264,344	37.75%	6,035,720	37.11%	1515.41	99.22%	98.01%	97.45%	97.17%	96.55%	94.84%	86.28%
	gDNA-pB	770L_BC2	3,272,424	1,883,818	57.57%	987,460	52.42%	266.77	98.17%	96.45%	94.75%	92.74%	88.77%	76.85%	88.16%
	gDNA-pA	792L_BC2	6,023,113	3,088,315	51.27%	1,546,582	50.08%	408.18	98.40%	96.94%	95.64%	94.33%	91.75%	84.79%	87.41%
NA10838	gDNA	761L	35,573,703	14,695,823	41.31%	5,705,050	38.82%	1481.41	99.24%	98.20%	97.73%	97.27%	96.55%	94.89%	87.12%
	WGA	767L	43,140,968	21,584,861	50.03%	8,194,931	37.97%	2189.59	99.42%	98.54%	98.14%	97.80%	97.16%	96.04%	87.69%
	gDNA-pB	770L_BC3	7,114,637	3,869,662	54.39%	1,809,019	46.75%	489.74	98.70%	97.19%	95.95%	94.97%	92.83%	86.75%	87.12%
	gDNA-pA	792L_BC3	6,587,601	3,489,971	52.98%	1,600,827	45.87%	430.28	98.54%	97.00%	95.71%	94.49%	91.72%	85.08%	87.00%
NA11829	gDNA	762L	39,864,620	18,798,516	47.16%	7,830,282	41.65%	2099.65	99.47%	98.47%	98.09%	97.70%	97.15%	96.00%	87.03%
	gDNA-pB	770L_BC4*	638,985	354,599	55.49%	161,742	45.61%	43.75	95.99%	83.63%	66.47%	49.70%	28.12%	9.87%	86.54%

	gDNA-pA	792L_BC4*	677,767	335,666	49.53%	139,652	41.60%	36.98	95.38%	80.11%	59.39%	41.17%	22.45%	6.88%	85.65%
NA11830	gDNA	763L	38,654,830	20,144,033	52.11%	9,425,152	46.79%	2557.17	99.53%	98.61%	98.21%	97.92%	97.42%	96.45%	87.07%
	gDNA-pB	770L_BC5	Insufficient amount of material to run sample												
	gDNA-pA	792L_BC5	5,403,560	2,832,411	52.42%	1,296,850	45.79%	346.19	98.38%	96.87%	95.24%	93.64%	90.28%	81.46%	86.91%
NA10856	gDNA	764L	43,185,707	22,650,137	52.45%	9,489,382	41.90%	2574.94	99.53%	98.68%	98.24%	98.01%	97.58%	96.55%	86.90%
	gDNA-pB	770L_BC6	9,255,333	5,136,664	55.50%	2,156,020	41.97%	584.63	98.90%	97.62%	96.70%	95.74%	93.93%	89.00%	87.30%
	gDNA-pA	792L_BC6	6,435,690	3,407,875	52.95%	1,423,554	41.77%	381.03	98.64%	96.96%	95.54%	93.84%	90.87%	82.48%	86.57%
Pooled Samples	gDNA	768L_1	41,913,860	21,748,402	51.89%	9,833,007	45.21%	2627.27	99.55%	98.63%	98.22%	98.05%	97.67%	96.85%	87.01%
	gDNA	768L_2	41,958,628	22,079,022	52.62%	10,073,995	45.63%	2700.44	99.47%	98.66%	98.25%	98.08%	97.69%	96.90%	87.25%

Supplementary Table 5: An overview of all SNPs and genotypes detected

(A separate Excel sheet)

Supplementary Table 6: Sequence Data Generated using 454 FLX and Illumina of the Same Target Regions (172kb/384 exons)

Targeted Sequencing Results	454 FLX	Illumina
Total Reads	219,876	5,954,822
Reads Mapping to Target Regions	184,641	2,337,390
Average Mapped Reads per Amplicon	482	5,343
Percent of Target Amplicons Covered	99.7%	99.7%
Target Uniformity (<10-fold)	96%	98%

Supplementary Table 7: Sample Multiplexing Calculation

Library ID	Bar-code	Reads	Mapped to Genome	Mapped on Target
792L	BC1	5,221,822	31.60%	64.20%
	BC2	6,023,113	32.50%	72.90%
	BC3	6,587,601	34.00%	67.60%
	BC4	677,767	28.50%	66.40%
	BC5	5,403,560	32.80%	68.10%
	BC6	9,255,333	36.30%	61.30%
	Total Reads (BC1-BC6)	33,169,196		
	Average (BC1-BC6)		32.62%	66.75%
Total Reads per Octet				
Total Reads per Octet	A	33,169,196		
Average % of Reads Mapped to Genome	B	32.62%		
Mapped Reads	C	10,818,686	= A*B	
% of read on Target	D	66.75%		
Reads on Target	E	7,221,473	= C*D	
Read length (bases)	F	50		
Bases per Octet	G	361,073,648	= E*F	
Bases per Sample*	H	17,280,500		
Samples per Octet	I	20	= G/H	
Sample per Flow Cell	J	160	= I*8	
Samples per Run	K	320	= I*16	

*Bases per Sample = # of amplicon bases * 100