

Supplemental Table 1: Strains used in this study.

| Strain | Genotype | Reference |
|---------------|---|------------------|
| SC5314 | Prototroph | (4) |
| RM1000 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> | (8) |
| CJC5 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::URA3-HIS1-GFP</i> | This study |
| CJC25 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::URA3-HIS1-GFP ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC26 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::ACT1p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC27 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::ARG1p-GFP-URA3-HIS1</i> | This study |
| CJC28 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::ARG1p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC29 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::ARG3p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC30 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::LYS1p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC31 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::LEU2p-GFP-URA3-HIS1</i> | This study |

| | | |
|-----------|---|------------|
| CJC32 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>RPS10/rps10::ARO4p-GFP-URA3-HIS1</i> | This study |
| HZY28 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>arg4::hisG/arg4::hisG arg81::HIS1/arg81::ARG4</i> | (7) |
| DSY3426-2 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>arg4::hisG/arg4::hisG arg83::URA3/arg83::ARG4</i> | (7) |
| DSY3233 | <i>ura3::λimm434/ura3::λimm434 gcn4::hisG/gcn4::hisG</i> | (7) |
| CJC33 | <i>ura3::imm434/ura3::imm434 his1::hisG/his1::hisG</i> <i>arg4::hisG/arg4::hisG arg81::HIS1/arg81::ARG4</i> <i>RPS10/rps10::ARG3p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC34 | <i>ura3::λimm434/ura3::λimm434 his1::hisG/his1::hisG</i> <i>arg4::hisG/arg4::hisG arg83::URA3/arg83::ARG4</i> <i>RPS10/rps10::ARG3p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| CJC35 | <i>ura3::λimm434/ura3::λimm434 gcn4::hisG/gcn4::hisG</i> <i>RPS10/rps10::ARG3p-GFP-URA3-HIS1</i> <i>ADH1/adh1::yCherry-SAT1</i> | This study |
| OXYellow | <i>ura3::λimm434/ura3:: λimm434</i> <i>CTA1p-GFP-URA3::hisG ADH1p-yCherry-SAT1</i> | (2) |
| ARGiallo | <i>ura3::λimm434/ura3:: λimm434</i> <i>ARG3p-GFP-URA3::hisG ADH1p-yCherry-SAT1</i> | This study |
| JRC12 | <i>arg1Δ::FRT/arg1Δ::FRT</i> | This study |
| JRC29 | <i>arg1Δ::FRT/arg1Δ::FRT-ARG1</i> | This study |
| JRC38 | <i>arg3Δ::FRT/arg3Δ::FRT</i> | This study |

| | | |
|-------|---|------------|
| JRC41 | <i>arg3Δ::FRT/arg3Δ::FRT-ARG3</i> | This study |
| JC50 | <i>ura3/ura3, his1/his1, hog1::LoxP-ura3- LoxP/hog1::LoxP-HIS1-LoxP Clp20 (URA3, HIS1)</i> | (6) |
| JC52 | <i>ura3/ura3, his1/his1, hog1::LoxP-ura3- LoxP/hog1::LoxP-HIS1-LoxP Clp20-HOG1 (URA3, HIS1)</i> | (6) |
| RGC1 | <i>RPS10/rps10::Clp10-ACT1p-GFP-SAT1</i> | This study |
| JRC47 | <i>arg1Δ::FRT/arg1Δ::FRT RPS10/rps10::Clp10-ACT1p-GFP-SAT1</i> | This study |
| JRC42 | <i>arg3Δ::FRT/arg3Δ::FRT RPS10/rps10::Clp10-ACT1p-GFP-SAT1</i> | This study |

Supplemental Table 2. Plasmids used in this study.

| Plasmid | Features | Reference |
|----------------|------------------------|------------------|
| CIp10 | CaURA3-RPS10 | (5) |
| CIp20 | CIp10-HIS1 | (3) |
| pGFP | CIp10-GFP | (1) |
| pACT1-GFP | CIp10-ACT1p-GFP | (1) |
| pCJ1 | pGFP-HIS1 | This study |
| pCJ2 | pACT1-GFP-HIS1 | This study |
| pCJ4 | pCJ1-ARG3p-GFP | This study |
| pCJ5 | pCJ1-ARG1p-GFP | This study |
| pADH1-mCherry | CIp-ADH1p-mCherry-SAT1 | (2) |
| pJRC28 | ARG1:FRT-SAT1-FRT | This study |
| pJRC40 | ARG3:FRT-SAT1-FRT | This study |
| pHZ130 | CIp-ACT1p-GFP-SAT1 | This study |

Supplemental References

1. **Barelle, C. J., C. L. Manson, D. M. MacCallum, F. C. Odds, N. A. Gow, and A. J. Brown.** 2004. GFP as a quantitative reporter of gene regulation in *Candida albicans*. *Yeast* **21**:333-340.
2. **Brothers, K. M., Z. R. Newman, and R. T. Wheeler.** 2011. Live imaging of disseminated candidiasis in zebrafish reveals role of phagocyte oxidase in limiting filamentous growth. *Eukaryot Cell* **10**:932-944.
3. **Dennison, P. M., M. Ramsdale, C. L. Manson, and A. J. Brown.** 2005. Gene disruption in *Candida albicans* using a synthetic, codon-optimised Cre-loxP system. *Fungal Genet Biol* **42**:737-748.
4. **Gillum, A. M., E. Y. Tsay, and D. R. Kirsch.** 1984. Isolation of the *Candida albicans* gene for orotidine-5'-phosphate decarboxylase by complementation of *S. cerevisiae* *ura3* and *E. coli* *pyrF* mutations. *Mol Gen Genet* **198**:179-182.
5. **Murad, A. M., P. R. Lee, I. D. Broadbent, C. J. Barelle, and A. J. Brown.** 2000. Clp10, an efficient and convenient integrating vector for *Candida albicans*. *Yeast* **16**:325-327.
6. **Smith, D. A., S. Nicholls, B. A. Morgan, A. J. Brown, and J. Quinn.** 2004. A conserved stress-activated protein kinase regulates a core stress response in the human pathogen *Candida albicans*. *Mol Biol Cell* **15**:4179-4190.
7. **Vandeputte, P., S. Pradervand, F. Ischer, A. T. Coste, S. Ferrari, K. Harshman, and D. Sanglard.** 2012. Identification and functional characterization of Rca1, a transcription factor involved in both antifungal susceptibility and host response in *Candida albicans*. *Eukaryot Cell* **11**:916-931.
8. **Wilson, R. B., D. Davis, and A. P. Mitchell.** 1999. Rapid hypothesis testing with *Candida albicans* through gene disruption with short homology regions. *J Bacteriol* **181**:1868-1874.

Figure S1. Misannotation of the *ARG1* gene. A) A transcriptional reporter generated based on the originally annotated start codon (at position 151,359 on Chromosome R) was not expressed on arginine-deficient YNB medium (left image). In contrast, a reporter using a second, in-frame, ATG 231 nt to the 3' (position 151,128) is strongly fluorescent in the absence of arginine (right image). Both images are GFP-DIC overlays. B) 5' RACE identifies one major and two minor transcriptional start sites at -48, -44, and -35 relative to the 3' ATG. C) A ClustalW alignment of the amino termini of Arg1p homologs from related species, using the originally annotated protein sequence of *C. albicans*, showing a 77 amino acid addition not homologous to the other species.

Figure S2. *ARG1* and *ARG3* respond specifically to arginine deprivation. *ARG1p-GFP* (CJC27) *ARG3p-GFP* (CJC29) cells were grown overnight in YPD and subculture in fresh YPD, YNB, or YNB supplemented with the indicated concentrations of L-arginine (R), L-leucine (L) and L-lysine (K). Cells were grown for 4 hours at 30°C. The images shown are GFP-DIC overlaid pictures.

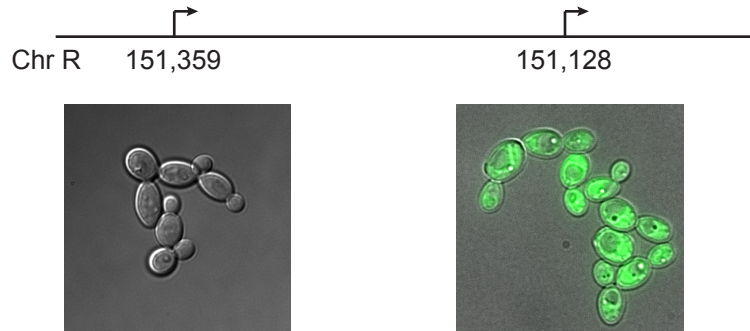
Figure S3. *ARG3* is induced in phagocytosed cells in vivo. Zebrafish were inoculated with the *ARG3-GFP/ADHI-yCherry* strain (ARGiallo) or the GFP-less control strain by microinjection into the hindbrain. At the indicated times, fish were anesthetized and immobilized in agarose prior to visualization by fluorescence microscopy. A) Representative images of phagocytosed ARGiallo cells. B) Ratiometric quantitation of GFP fluorescence intensity in intracellular versus extracellular cells over time.

Figure S4. *arg1*Δ and *arg3*Δ mutants are arginine auxotrophs. Wild-type (SC5314), *arg1*Δ (JRC12), *arg3*Δ (JRC38), and complemented (JRC29, JRC41) strains were diluted to an OD₆₀₀ of 0.1, and five-fold serial dilutions were spotted onto synthetic complete (SC) plates, synthetic dropout plates lacking arginine (SD-Arg) supplemented with or without additional arginine and grown at 30°C for 2 days.

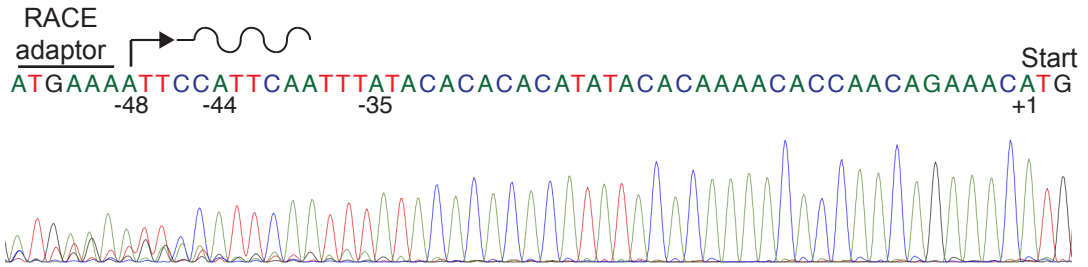
Figure S5. *arg* mutants are not sensitive to oxidative stress. Wild-type (SC5314), *hog1*Δ/Δ (JC50), *arg1*Δ (JRC12), *arg3*Δ (JRC38), and complemented (JC52, JRC29, and JRC41, respectively) strains were diluted to an OD₆₀₀ of 0.1 and five-fold serial dilutions were spotted onto YPD plates with the indicated concentrations of H₂O₂ or menadione and grown at 30°C for 3 days.

Supplemental Movie 1. The *ARG3p*-GFP reporter strain was co-cultured with macrophages and images were collected every three minutes for 3.5 hours, beginning 15 minutes after initiation of the co-culture. The movie shows the merged GFP-DIC image.

A.



B.



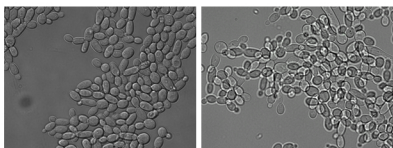
C.

| | | |
|--------------|--|-----|
| orf19.7469 | MTNSVIGQLKNFLTALNLLSIHSSRKEIFFFLNYKRDSKNLLNENFIYLFFFSFFSFLLF | 60 |
| CD36_25720 | ----- | |
| CTRG_01005 | ----- | |
| CLUG_04362 | ----- | |
| YOL058w | ----- | |
| CAGL0C05115g | ----- | |
| orf19.7469 | LFHSIYTHTYTQNTNRNMSKGKVCLAYSGGLDTSVILAWLLEQGYEVIAFLANIGQEEDF | 120 |
| CD36_25720 | -----MSKGVCLAYSGGLDTSVILAWLLEQGYEVIAFLANIGQEEDF | 43 |
| CTRG_01005 | -----MSKGVCLAYSGGLDTSVILAWLLEQGYEVIAFLANIGQEEDF | 43 |
| CLUG_04362 | -----MSKGVCLAYSGGLDTSVILAWLLEQGYEVVAFMANIGQEEDF | 43 |
| YOL058w | -----MSKGVCLAYSGGLDTSVILAWLLDQGYEVVAFMANVIGQEEDF | 43 |
| CAGL0C05115g | -----MSKGVCLAYSGGLDTSIILAWLLEQGYEVVAFMADVGQEEDF | 43 |
| | *****:*****:*****:**:~:***** | |

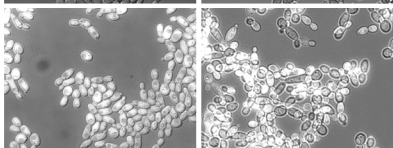
ARG3p

ARG1p

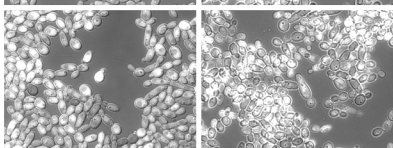
YPD



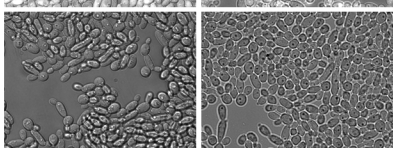
YNB



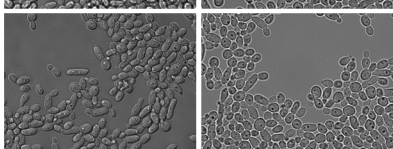
20 μ g/ml R



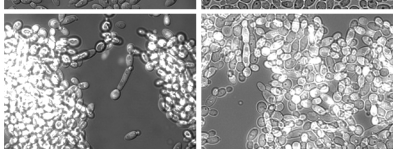
100 μ g/ml R



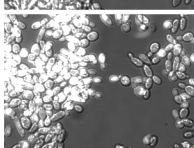
200 μ g/ml R



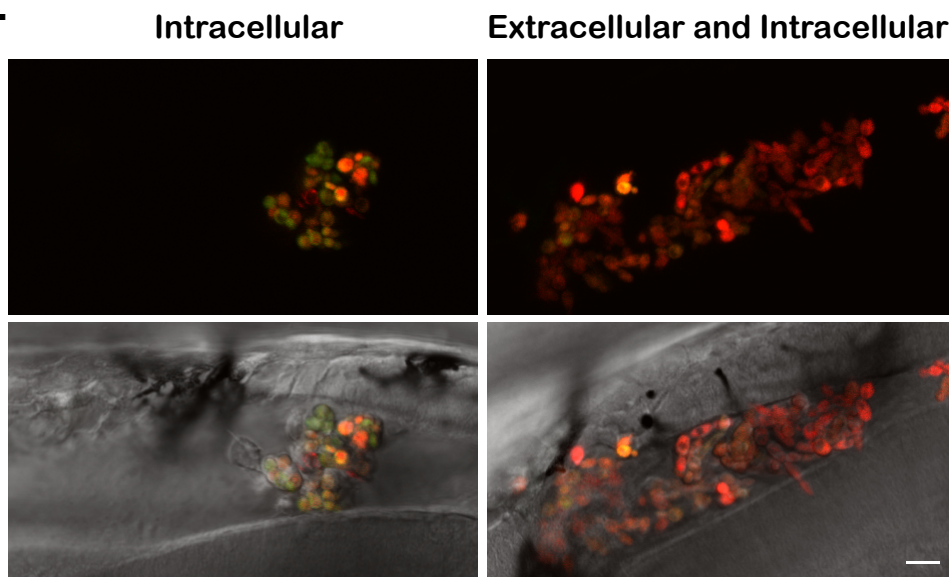
200 μ g/ml K



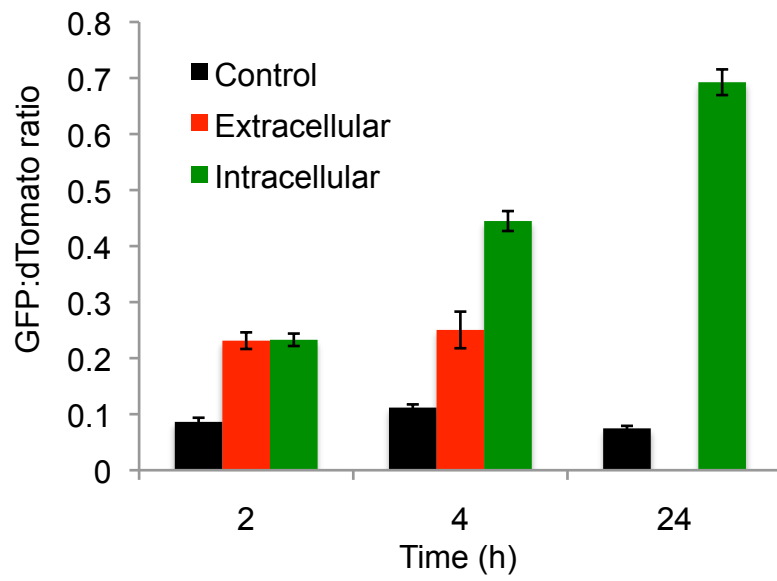
200 μ g/ml L



A.



B.



SC-ARG

