Table S1. Strains Used in This Study, Related to Experimental Procedures

Name	Genotype	Construction	Reference/Source
C. crescentus			
CB15N	Synchronizable variant of wild-type strain CB15		Evinger & Agabian, 1977
CS606	CB15N Δ*blaM*		West et al., 2002
EK61	CB15N P_{xyl}::P_{xyl}-*mcherry-CC1398*		Werner et al., 2009
EK363	CB15N P_{xyl}::P_{xyl}-*pstS-mcherry*	Transformation of CB15N with pEK301	This work
EK389	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*pstS-mcherry*	Transformation of SW51 with pEK301	This work
EK392	CB15N P_{xyl}::P_{xyl}-*yfp*	Transformation of CB15N with pXYFPN-5	This work
EK393	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*mcherry*	Transformation of SW51 with pXCHYN-5	This work
EK416	CB15N P_{xyl}::P_{xyl}-*mcherry*	Transformation of CB15N with pXCHYN-5	This work
EK417	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*yfp*	Transformation of SW51 with pXYFPN-5	This work
EK424	CB15N *pstS*::miniTn5LacZ P_{xyl}::P_{xyl}-*pstS-mcherry*	Transformation of EK425 with pEK301	This work
EK425	CB15N *pstS*::miniTn5LacZ	Transduction of kan^R from YB2811	This work
EK486	CB15N Δ*stpCD* P_{xyl}::P_{xyl}-*yfp*	Transformation of SS250 with pXYFPN-5	This work
EK487	CB15N Δ*stpCD* P_{xyl}::P_{xyl}-*mcherry*	Transformation of SS250 with pXCHYN-5	This work
GB255	CB15N *popZ::Ω*		Bowman et al., 2008
JK5	CB15N Δ*bacAB*		Kühn et al., 2010
MT304	CB15N Δ*pbpC*		Kühn et al., 2010
NR4042	CB15N Δ*stpX*		Viollier, P. H. (unpubl.)
PV5064	CB15N *stpX::stpX-gfp*		Hughes et al., 2010
SS141	CB15N Δ*stpA* P_{xyl}::P_{xyl}-*stpB-mcherry*	Transformation of SW49 with pSW32	This work
SS142	CB15N Δ*stpB* P_{xyl}::P_{xyl}-*stpA-mcherry*	Transformation of SW50 with pSW35	This work
SS160	CB15N *stpB-mCherry*	Gene replacement in CB15N with pSS109	This work
SS165	CB15N Δ*bla* P_{xyl}::P_{xyl}-*stpB-bla*	Transformation of CS606 with pSS120	This work
SS167	CB15N Δ*pbpC* P_{xyl}::P_{xyl}-*stpB-mcherry*	Transformation of MT304 with pSW32	This work
SS172	CB15N Δ*bla* P_{xyl}::P_{xyl}-*stpA-bla*	Transformation of CS606 with pSS119	This work
SS191	CB15N *ftsZ*::P_{xyl}-*ftsZ* P_{van}::P_{van}-*stpB-mcherry*	Transformation of YB1585 with pSS142	This work
SS213	CB15N Δ*stpX* P_{xyl}::P_{xyl}-*stpB-mcherry*	Transduction of kan^R from SW30 to NR4042	This work
SS214	CB15N *stpB::stpB-mcherry* pP_{xyl}-*stpAB*	Transformation of SS160 with pSW64	This work
SS216	CB15N Δ*stpAB* pP_{xyl}-TAT-*tdimer2*	Transformation of SW51 with pEJ216	This work
SS220	CB15N Δ*stpA stpB::stpB-His*	Transformation of SW49 with pSS187	This work
SS224	CB15N *divJ*::Ω P_{xyl}::P_{xyl}-*stpB-mcherry*	Transduction of $spec^R$ from YB3202 to SW30	This work
SS228	CB15N P_{xyl}::P_{xyl}-*stpC-mcherry*	Transformation of CB15N with pSS204	This work
SS233	CB15N *stpB::stpB-His*	Transformation of CB15N with pSS187	This work
SS234	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*stpD-gfp*	Transformation of SW51 with pSS202	This work
SS236	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*stpC-mcherry*	Transformation of SW51 with pSS204	This work
SS239	CB15N Δ*stpC*	Gene replacement in CB15N using pSS209	This work
SS240	CB15N Δ*stpC* P_{xyl}::P_{xyl}-*stpD-gfp*	Transformation of SS239 with pSS202	This work
SS243	CB15N *stpD*::*stpD-gfp* P_{xyl}::P_{xyl}-*stpA-mcherry*	Transformation of SW33 with pSS205	This work
SS244	CB15N *stpD*::*stpD-His*	Transformation of CB15N with pSS206	This work
SS247	CB15N *stpC*::*stpC-His*	Transformation of CB15N with pSS210	This work
SS248	CB15N *stpD*::*stpD-gfp*	Transformation of CB15N with pSS205	This work
SS250	CB15N Δ*stpCD*	Gene replacement in SS239 using pSS208	This work
SS252	CB15N Δ*stpD*	Gene replacement in CB15N using pSS208	This work
SS258	CB15N *stpB*::*stpB-mcherry* pBXMCS-2	Transformation of SS160 with pBXMCS-2	This work
SS263	CB15N Δ*stpD* P_{xyl}::P_{xyl}-*stpC-mcherry*	Transformation of SS252 with pSS204	This work
SS264	CB15N Δ*stpB* P_{xyl}::P_{xyl}-*stpD-gfp*	Transformation of SW50 with pSS202	This work
SS265	CB15N Δ*stpB* P_{xyl}::P_{xyl}-*stpC-mcherry*	Transformation of SW50 with pSS204	This work
SS269	CB15N *stpD*::*stpD-gfp* pP_{xyl}-*TATdimer2*	Transformation of SS248 with pEJ216	This work
SS272	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*gspG-mcherry*	Transformation of SW51 with pJK86	This work
SS273	CB15N Δ*bla* P_{xyl}::P_{xyl}-*stpC-blaM*	Transformation of CS606 with pSS220	This work
SS274	CB15N Δ*bla* P_{xyl}::P_{xyl}-*stpD-blaM*	Transformation of CS606 with pSS221	This work
SS277	CB15N *stpD*::*stpD-gfp* P_{xyl}::P_{xyl}-*gspG-mcherry*	Transformation of SS248 with pJK86	This work
SS281	CB15N Δ*bacAB* P_{xyl}::P_{xyl}-*stpB-mcherry*	Transformation of JK5 with pSW32	This work
SS283	CB15N *stpD*::*stpD-gfp* P_{xyl}::P_{xyl}-*elpS-mcherry*	Transformation of SS248 with pSW67	This work
SS284	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*elpS-mcherry*	Transformation of SW51 with pSW67	This work
SS292	CB15N *popZ::Ω* P_{xyl}::P_{xyl}-*stpB-mcherry*	Transformation of GB255 with pSW32	This work
SS294	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*malA-mcherry*	Transformation of SW51 with pSS227	This work

Table S1. Strains Used in This Study (continued)

Name	Relevant Genotype/Description	Construction	Reference/Source
SS297	CB15N *stpD*::*stpD-gfp* P_{xyl}::P_{xyl}-*malA-mcherry*	Transformation of SS248 with pSS227	This work
SS299	CB15N *stpD*::*stpD-gfp* P_{xyl}::P_{xyl}-*pstS-mcherry*	Transformation of SS248 with pJK101	This work
SS302	CB15N Δ*stpAB* P_{xyl}::P_{xyl}-*pstS-mcherry*	Transformation of SW51 with pJK101	This work
SS304	CB15N Δ*stpCD* pP_{xyl}-TAT-*tdimer2*	Transformation of SS250 with pEJ216	This work
SS388	CB15N *stpD*::*stpD-gfp stpB*::*stpB-mcherry*	Transformation of SS160 with pSS205	This work
SS389	CB15N *stpD*::*stpD-gfp stpC*::*stpC-mcherry*	Transformation of SS248 with pSS310	This work
SS412	CB15N *stpB*::*stpB-mcherry*	Transformation of CB15N with pSS309	This work
SS413	CB15N *stpC*::*stpC-mcherry*	Transformation of CB15N with pSS310	This work
SS414	CB15N Δ*stpA stpB*::*stpB-mcherry*	Transformation of SW49 with pSS309	This work
SS415	CB15N Δ*stpAB stpC*::*stpC-mcherry*	Transformation of SW51 with pSS310	This work
SW30	CB15N P_{xyl}::P_{xyl}-*stpB-mcherry*	Transformation of CB15N with pSW32	This work
SW33	CB15N P_{xyl}::P_{xyl}-*stpA-mcherry*	Transformation of CB15N with pSW35	This work
SW49	CB15N Δ*stpA*	Gene replacement in CB15N using pSW51	This work
SW50	CB15N Δ*stpB*	Gene replacement in CB15N using pSW52	This work
SW51	CB15N Δ*stpAB*	Gene replacement in CB15N using pSW53	This work
YB1585	CB15N *ftsZ*::P_{xyl}-*ftsZ*		Wang et al., 2001
YB2811	NY111d1 *pstS*::miniTn5LacZ		Ireland et al., 2002
YB3202	CB15N *divJ*::*Ω*		Pierce et al., 2006
YB5058	CB15N *stpX*::*StpX-gfp* P_{xyl}::P_{xyl}-*stpB-mCherry*	Transduction of $spec^R$ from PV5064	This work
YB5059	CB15N Δ*stpAB stpX*::*stpX-gfp*	Transduction of $spec^R$ from PV5064	This work
YB5231	CB15N Δ*stpX*		Hughes et al., 2010
A. excentricus			
CB48	Wild-type strain		Poindexter, 1964
SS309	CB48 Astex_0987::Astex_0987-*mcherry*	Transformation of CB48 with pSS229	This work
E. coli			
TOP10	General cloning strain		Invitrogen

Table S2. General Plasmids Used in This Study, Related to Experimental Procedures

Name	Description	Reference/Source
pBXMCS-2	Replicating plasmid for the inducible overexpression of genes in CB15N, Kan^R	Thanbichler et al., 2007
pCHYC-1	Plasmid for integrating genes encoding C-terminal fusions to the red fluorescent protein mCherry at the native gene locus, $Spec/Str^R$	Thanbichler et al., 2007
pEJ216	Replicating plasmid carrying TAT-dimer2 under control of P_{xyl}, Cam^R	Judd et al., 2005
pJAMY31	Kan^R ColE1 replicon, β-lactamase translational fusion vector	Alley, M. R. K. (unpubl.)
pNPTS138	*sacB*-containing suicide vector used for double homologous recombination, Kan^R	Alley, M. R. K. (unpubl.)
pTCYC-2	Plasmid for integrating C-terminal fusions to the tetracysteine tag at the native gene locus, $Spec/Str^R$	Thanbichler et al., 2007
pVCHYC-1	Plasmid for integrating genes encoding C-terminal fusions to the red fluorescent protein mCherry at the chromosomal *vanA* locus, $Spec/Str^R$	Thanbichler et al., 2007
pXCHYC-2	Plasmid for integrating genes encoding C-terminal fusions to the red fluorescent protein mCherry at the chromosomal *xylX* locus, Kan^R	Thanbichler et al., 2007
pXCHYC-5	Plasmid for integrating genes encoding C-terminal fusions to the red fluorescent protein mCherry at the chromosomal *xylX* locus, Tet^R	Thanbichler et al., 2007
pXCHYN-5	Plasmid for integrating genes encoding N-terminal fusions to the red fluorescent protein mCherry at the chromosomal *xylX* locus, Tet^R	Thanbichler et al., 2007
pXGFPC-2	Plasmid for integrating genes encoding C-terminal fusions to the green fluorescent protein GFP at the chromosomal *xylX* locus, Kan^R	Thanbichler et al., 2007
pXYFPC-5	Plasmid for integrating genes encoding C-terminal fusions to the yellow fluorescent protein eYFP at the chromosomal *xylX* locus, Tet^R	Thanbichler et al., 2007
pXYFPN-5	Plasmid for integrating genes encoding N-terminal fusions to the yellow fluorescent protein eYFP at the chromosomal *xylX* locus, Tet^R	Thanbichler et al., 2007

Table S3. Plasmids Generated in This Study, Related to Experimental Procedures

Name	Description	Construction
pEK301	pXCHYC-2 carrying *pstS*	a) amplification of the *pstS* gene using primers EK225/EK226, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR product into pXCHYC-2 cut with NdeI and EcoRI
pJK86	pXCHYC-2 carrying *gspG* (CCNA_00175)	a) amplification of *gspS* from genomic DNA using primers CC0176-NdeI-for/CC0176-EcoRI-rev, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR product into pXCHYC-2 cut with NdeI and EcoRI
pJK101	pXCHYC-2 carrying *pstS*	a) amplification of *pstS* from genomic DNA using primers pstS-for/pstS-rev-EcoRI, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR product into pXCHYC-2 cut with NdeI and EcoRI
pSS98	pSW32 carrying *stpB* downstream region	a) amplification of the *stpB* downstream sequence from genomic DNA using primers CC2476-1/-2, followed by restriction with BsrGI and NheI b) ligation into pSW32 cut with BsrGI and NheI
pSS109	pNTPS138-based plasmid for replacing native *stpB* with *stpB-mcherry*	a) isolation of *stpB*-mcherry from pSS98 by restriction with NdeI and NheI, followed by blunting of the fragment with T4 DNA polymerase b) ligation into pNPTS138 cut with EcoRV
pSS119	pXBlaMC-2 carrying *stpB*	a) isolation of *stpB* from pSW32 by restriction with NdeI and EcoRI b) ligation into pXBlaMC-2 cut with NdeI and EcoRI
pSS120	pXBlaMC-2 carrying *stpA*	a) isolation of *stpA* from pSW35 by restriction with NdeI and EcoRI b) ligation into pXBlaMC-2 cut with NdeI and EcoRI
pSS123	pTCYC-2 carrying *stpB*	a) isolation of *stpB* from pSW32 by restriction with NdeI and EcoRI b) ligation into pTCYC-2 cut with NdeI and EcoRI
pSS142	pVCHYC-1 carrying *stpB*	a) isolation of *stpB* from pSW32 by restriction with NdeI and EcoRI b) ligation into pVCHYC-1 cut with NdeI and EcoRI
pSS187	pTCYC-2-based plasmid carrying *stpB-His*	a) in-vitro assembly of a phosphorylated His_{10}-linker from AM_299f/AM_300r b) ligation into pSS123 cut with EcoRI
pSS202	pXGFPC-2 carrying *stpD*	a) amplification of *stpD* from genomic DNA using primers CCNA_2271-for/-rev, followed by restriction with NdeI and SacI b) ligation of the digested PCR product into pXGFPC-2 cut with NdeI and SacI
pSS204	pXCHYC-2 carrying *stpC*	a) amplification of *stpC* from genomic DNA using primers CCNA_02560-for/-rev, followed by restriction with NdeI and SacI b) ligation of the digested PCR product into pXCHYC-2 cut with NdeI and SacI
pSS205	pGFPC-1 carrying *stpD*	a) isolation of *stpD* from pSS202 by restriction with NdeI and EcoRI b) ligation into pGFPC-1 cut with NdeI and EcoRI
pSS206	pTCYC-2-based plasmid carrying *stpD-His*	a) isolation of *stpD* from pSS202 by restriction with NdeI and EcoRI b) ligation into pSS187 cut with NdeI and EcoRI
pSS208	pNTPS138-based plasmid for constructing an in-frame deletion in *stpD*	a) amplification of the *stpD* 5' region from genomic DNA using primers CCNA_02271-3/-4 b) amplification of the *stpD* 3' region from genomic DNA using primers CCNA_02271-5/-6 c) overlap extension PCR using both PCR products and primers CCNA_02271-3/-6, followed by restriction with HindIII and EcoRI c) ligation of the final PCR fragment into HindIII/EcoRI-cut pNPTS138
pSS209	pNTPS138-based plasmid for constructing an in-frame deletion in *stpC*	a) amplification of the *stpC* 5' region from genomic DNA using primers CCNA_02560-3/-4 b) amplification of the *stpC* 3' region from genomic DNA using primers CCNA_02560-5/-6 c) overlap extension PCR using both PCR products and primers CCNA_02560-3/-6, followed by restriction with HindIII and EcoRI c) ligation of the final PCR fragment into HindIII/EcoRI-cut pNPTS138
pSS210	pTCYC-2-based plasmid carrying *stpC-His*	a) isolation of *stpC* from pSS204 by restriction with NdeI and EcoRI b) ligation into pSS187 cut with NdeI and EcoRI
pSS220	pXBlaMC-2 carrying *stpC*	a) isolation of *stpC* from pSS204 by restriction with NdeI and EcoRI b) ligation into pXBlaMC-2 cut with NdeI and EcoRI
pSS221	pXBlaMC-2 carrying *stpD*	a) isolation of *stpD* from pSS202 by restriction with NdeI and EcoRI b) ligation into pXBlaMC-2 cut with NdeI and EcoRI
pSS227	pXCHYC-2 carrying *malA* (CCNA_02370)	a) amplification of *malA* from genomic DNA using primers CC2287-1/-2, followed by restriction with NdeI and SacI b) ligation of the digested PCR product into pXCHYC-2 cut with NdeI and SacI
pSS229	pCHYC-2 carrying *Astex_0987*	a) amplification of *Astex_0987* from genomic DNA using primers Astex_0987-2f/-3r, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR product into pCHYC-2 cut with NdeI and EcoRI
pSS309	pCHYC-2 carrying *stpB*	a) isolation of *stpB* from pSW32 by restriction with NdeI and EcoRI b) ligation into pXCHYC-2 cut with NdeI and EcoRI
pSS310	pCHYC-2 carrying *stpC*	a) isolation of *stpC* from pSS204 by restriction with NdeI and EcoRI b) ligation into pXCHYC-2 cut with NdeI and EcoRI

Table S3. Plasmids Generated in This Study (continued)

Name	Description	Construction
pSW32	pXCHYC-2 carrying *stpB*	a) amplification of the *stpB* gene using primers CC2476-for/CC2476-rev, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR products into pXCHYC-2 cut with NdeI and EcoRI
pSW35	pXCHYC-2 carrying *stpA*	a) amplification of the *stpA* gene using primers CC2477-uni/CC2477-rev, followed by restriction with NdeI and SacI b) ligation of the digested PCR products into pXCHYC-2 cut with NdeI and SacRI
pSW51	pNTPS138-based plasmid for constructing an in-frame deletion in *stpA*	a) amplification of the 5' region of *stpA* from genomic DNA using primers dCC2477-A-for/ dCC2477-B-rev, followed by restriction with BamHI and EcoRI b) amplification of the 3' region of *stpA* from genomic DNA using primers dCC2477-C-for/dCC2477-D-rev, followed by restriction with EcoRI and NheI c) ligation of both fragments into BamHI/NheI-cut pNPTS138
pSW52	pNTPS138-based plasmid for constructing an in-frame deletion in *stpB*	a) amplification of the 5' region of *stpB* from genomic DNA using primers dCC2476-A-for/dCC2476-B-rev, followed by restriction with BamHI and EcoRI b) amplification of the 3' region of *stpB* from genomic DNA using primers dCC2476-C-for/dCC2476-D-rev, followed by restriction with EcoRI and NheI c) ligation of both fragments into BamHI/NheI-cut pNPTS138
pSW53	pNTPS138-based plasmid for constructing an in-frame deletion in *stpAB*	a) amplification of the 5' region of *stpAB* from genomic DNA using primers d2476d2477-A-for/d2476d2477-B-rev, followed by restriction with BamHI and EcoRI b) amplification of the 3' region of *stpB* from genomic DNA using primers d2476d2477-C-for/ d2476d2477-D-rev, followed by restriction with EcoRI and NheI c) ligation of both fragments into BamHI/NheI-cut pNPTS138
pSW64 (P_{xyl}-*stpAB*)	pBXMCS-2 carrying *stpAB*	a) amplification of *stpAB* from genomic DNA using primers CC2477-uni/CC2476-rev2St, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR product into pBXMCS-2 cut with NdeI and EcoRI
pSW67	pXCHYC-2 carrying *elpS* (CCNA_00169)	a) amplification of *elpS* from genomic DNA using primers CC0170-C-for/CC0170-C-rev, followed by restriction with NdeI and EcoRI b) ligation of the digested PCR product into pXCHYC-2 cut with NdeI and EcoRI
pXBlaMC-2	Plasmid for integrating genes encoding C-terminal TEM-1 β-lactamase fusions at the chromosomal *xylX* locus, Kan^R	a) amplification of the TEM-1 β-lactamase gene from pJAMY31 using primers blaM-C_for/blaM-C_rev, followed by restriction with AgeI and NheI b) ligaton of the digested PCR product into pXGFPC-2 cut with AgeI and NheI

Table S4. Oligonucleotides Used in This Study, Related to Experimental Procedures

Note the different ORF numbers: CC2477 = CCNA_02562, CC2476 = CCNA_02561, CC0170 = CCNA_00169, CC0176 = CCNA_ 00175, CC2287=CCNA_02370.

Oligonucleotide	Sequence
AM_299f	AATTCCCATCACCACCATCATCACCATCACCACCACTAGT
AM_300r	GGGTAGTGGTGGTAGTAGTGGTAGTGGTGGTGATCATTAA
Astex_0987-2f	ATTAATTCATATGTCGGCCCAGACAGAAACCCGCTATG
Astex_0987-3r	ATGAATTCGATTTCGAGGCGACCAGCAGACGCGG
blaM-C_for	TTTTACCGGTCGGCCACCATGCACCCAGAAACGCTGGTGAAAGTAAAAG
blaM-C_rev	TTTTGCTAGCTTACCAATGCTTAATCAGTGAGGCACCTATC
CCNA_02271-for	TTAATTCATATGCGTCATCAAATGGCGCGTCGCG
CCNA_02271-rev	TAGAGCTCCGTGATGGGCGGCGGCGGCGTGCTTG
CCNA_02271-3	ATGAATTCGAACCAGACGACCTGAAGCGGCGCAG
CCNA_02271-4	CTTGTCCTTCACGCGACGCGCCATTTGATGAC
CCNA_02271-5	CGTCGCGTGAAGGACAAGCACGCCGCCGCCGC
CCNA_02271-6	TAAAGCTTCGGCGGTTTCCAGGTGATCGAGCA
CCNA_02560-for	TTAATTCATATGAGCAAGTCTGTTCGTAGCCGGCTGG
CCNA_02560-rev	TAGAGCTCCGCATCCGACGAGGCCCGCGCCGACG
CCNA_02560-3	ATGAATTCGAGTCAAGGCGACCGGCACGATCATG
CCNA_02560-4	GAAATTACGGGAAACGGCCAGCCGGCTACGAAC
CCNA_02560-5	GCCGTTTCCCGTAATTTCGTCGGCGCGGGCCTC
CCNA_02560-6	TAAAGCTTCTACGAGCAGGCGACGAAGCACCG
CC0170-C-for	AATTCATATGAAGCTGTATAGAAACCTAATCCTCATGAGCTGCG
CC0170-C-rev	AATTGAATTCTCGTTGGTCGGCAGCGACGTG
CC0176-NdeI-for	ATATCATATGTCGACCGCAAACGCTGAAACGAAAC
CC0176-EcoRI-rev	ATATGAATTCTCGCTCCAGTTGCCAATGTCGG
CC2287-3	GCCAATGATCGCGTGATCGGCG
CC2287-4	CATCCCGCAGGAAGCCATCATCG
CC2476-for	TTAACATATGCGCCGCCTCAGCCGCCTGCTGG
CC2476-rev	TAGAATTCGATCGAGGAGCTCCCCCTTGTTGAGGC
CC2477-uni	AAAACATATGCGCGAGGCCGGGGACGCAATTGC
CC2477-rev	TAGAGCTCCGTAATTCCCTTCGTTATACGGACGCCCGC
DCC2476-A-for	AATTGGATCCTCGGCCGTCCGAACACC
DCC2476-B-rev	AATTGAATTCGGCCCAAAGCGCCAGC
DCC2476-C-for	AATTGAATTCCGGTCCGCCGCCC
DCC2476-D-rev	AATTGCTAGCTTGAAGCAGCGGTTGTCGCC
CC2476-rev2St	AATTGAATTCTCATCGAGGAGCTCCCCCTTGT
CC2476-1	TATGTACAAGTAAGCAAGTCTGTTCGTAGCCGGCTGGC
CC2476-2	TATAGCTAGCCGACCAGCACCGTCCTCAGCATCC
DCC2477-B-rev	TTAAGAATTCGCGAAGGCGCGCAATT
DCC2477-C-for	TTAAGAATTCGATCGGGGCGGGCGT
DCC2477-D-for	TTAAGCTAGCACGCCGGGCTGGATCTTG
D2476d4277-A-for	TTAAGGATCCGGAGCTGGCCAATACGGC
D2476d4277-B-rev	TTAAGAATTCGCGAAGGCGCGCAATTG
D2476d4277-C-for	TTAAGAATTCGTTCCCGCGCCTCAACAAG
D2476d4277-D-for	TTAAGCTAGCGGGGGTGAAGATGCCGAG
EK225	TACTCATATGAACAAGCTCATCGGCGC
EK226	TACTGAATTCGCGTTCTTCGGCGCCGGCATCGG
pstS-for	ATATACATATGAACAAGCTCATCGGCGCGGTCGCC
pstS-rev-EcoRI	ATATGAATTCGCGTTCTTCGGCGCCGGCATC

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